

Assignment 3: Data Exploration

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Change “Student Name, Section #” on line 3 (above) with your name and section number.
2. Work through the steps, **creating code and output** that fulfill each instruction.
3. Be sure to **answer the questions** in this assignment document.
4. When you have completed the assignment, **Knit** the text and code into a single PDF file.
5. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai. Add your last name into the file name (e.g., “FirstLast_A03_DataExploration.Rmd”) prior to submission.

The completed exercise is due on <>.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. **Be sure to add the stringsAsFactors = TRUE parameter to the function when reading in the CSV files.**

```
setwd

## function (dir)
## .Internal(setwd(dir))
## <bytecode: 0x15821abf8>
## <environment: namespace:base>
("~/Documents/Duke Spring 2022/Environmental Data Analytics/Environmental_Data_Analytics_2022")

## [1] "~/Documents/Duke Spring 2022/Environmental Data Analytics/Environmental_Data_Analytics_2022"
getwd()

## [1] "/Users/rachelgordon/Documents/Duke Spring 2022/Environmental Data Analytics/Environmental_Data_
library(tidyverse)
Neonics <- read.csv(
  "./Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv", stringsAsFactors = TRUE)
Litter <- read.csv(
  "./Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv", stringsAsFactors=TRUE)
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency’s ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used

widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: It's important to study the ecotoxicology of neonicotinoids on insects because neonicotinoids can have detrimental risks to bees and other pollinators, negatively affecting their immune system and making them more susceptible to parasites and disease. Neonicotinoids are especially toxic to bees, as they become present in pollen and nectar when they are absorbed by the plant, posing risks to the bees and other pollinators when they feed on the plant. As neonicotinoids are important for managing pests that are resistant to other insecticides, it's important to understand how using them for agricultural purposes may have negative impacts on bee and other pollinator populations.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: It is important to study litter and woody debris that falls to forest grounds as they play a major role in forest biodiversity, nutrient cycling, and habitats for terrestrial organisms. Specifically, the coarseness of the litter and woody debris has an impact on the forest ecosystem, as more coarse debris can provide nesting, foraging, and shelter services to the various organisms living in the forest.

4. How is litter and woody debris sampled as part of the NEON network? Read the `NEON_Litterfall_UserGuide.pdf` document to learn more. List three pieces of salient information about the sampling methods here:

Answer: * The litter and woody debris are collected from elevated and ground traps * Ground traps are sampled once per year, whereas the sampling frequency of elevated traps depends on the vegetation present at the site, with sampling methods being 1x every 2week or 1x every 1-2 sites for deciduous forest sites vs. evergreen sites, respectively * Litter and fine woody debris sampling is done at terrestrial NEON sites that have woody vegetation >2m tall and sampling only occurs in tower plots

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
#Using the dim() function to determine how many rows and columns there are.
dim(Neonics)
```

```
## [1] 4623 30
```

6. Using the `summary` function on the "Effect" column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
#Using the summary() function to determine the most common effects studied.
summary(Neonics$Effect)
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##           12           102           360              11
##      Cell(s)      Development      Enzyme(s) Feeding behavior
##           9           136           62              255
##      Genetics      Growth      Histology      Hormone(s)
##          82           38           5              1
## Immunological      Intoxication      Morphology      Mortality
##          16           12           22             1493
```

##	Physiology	Population	Reproduction
##	7	1803	197

Answer: The most common effects studied are Population, Mortality, and Behavior. As this data is helping study how neonicotinoids may be impacting insects, population and mortality are important to study as they are both effects linked to death/the health of a population. As neonicotinoids are known to be toxic to pollinators, studying the mortality and population of them can provide insight into the health of the populations. Additionally, neonicotinoids impact the nervous system functions, so studying behavior is also important to see if neonicotinoid exposure can be linked to behavior changes in pollinator populations that may be driven by nervous system disruptions.

- Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

#Using the summary() function to identify the six most commonly studied species in the dataset
`summary(Neonics$Species.Common.Name)`

##	Honey Bee	Parasitic Wasp
##	667	285
##	Buff Tailed Bumblebee	Carniolan Honey Bee
##	183	152
##	Bumble Bee	Italian Honeybee
##	140	113
##	Japanese Beetle	Asian Lady Beetle
##	94	76
##	Euonymus Scale	Wireworm
##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle

##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17	17
##	Hemlock Woolly Adelgid Lady Beetle	Hemlock Woolly Adelgid
##	16	16
##	Mite	Onion Thrip
##	16	16
##	Western Flower Thrips	Corn Earworm
##	15	14
##	Green Peach Aphid	House Fly
##	14	14
##	Ox Beetle	Red Scale Parasite
##	14	14
##	Spined Soldier Bug	Armoured Scale Family
##	14	13
##	Diamondback Moth	Eulophid Wasp
##	13	13
##	Monarch Butterfly	Predatory Bug
##	13	13
##	Yellow Fever Mosquito	Braconid Parasitoid
##	13	12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12
##	Pea Aphid	Pond Wolf Spider

##		12		12
##	Spotless Ladybird Beetle		Glasshouse Potato Wasp	
##		11		10
##	Lacewing		Southern House Mosquito	
##		10		10
##	Two Spotted Lady Beetle		Ant Family	
##		10		9
##	Apple Maggot		(Other)	
##		9		670

Answer: The six most commonly studied species in the dataset are Honey Bees, Parasitic Wasps, Buff Tailed Bumblebees, Carniolan Honey Bees, Bumble Bees, and Italian Honeybees. All of these species are part of the bee or wasp family, which are all important for studying neonicotinoids because as mentioned earlier, neonicotinoids pose a major risk and can be toxic to pollinators, such as bees/wasps.

8. Concentrations are always a numeric value. What is the class of Conc.1..Author. in the dataset, and why is it not numeric?

```
#Finding class of Conc.1..Author
class(Neonics$Conc.1..Author)
```

```
## [1] "factor"
```

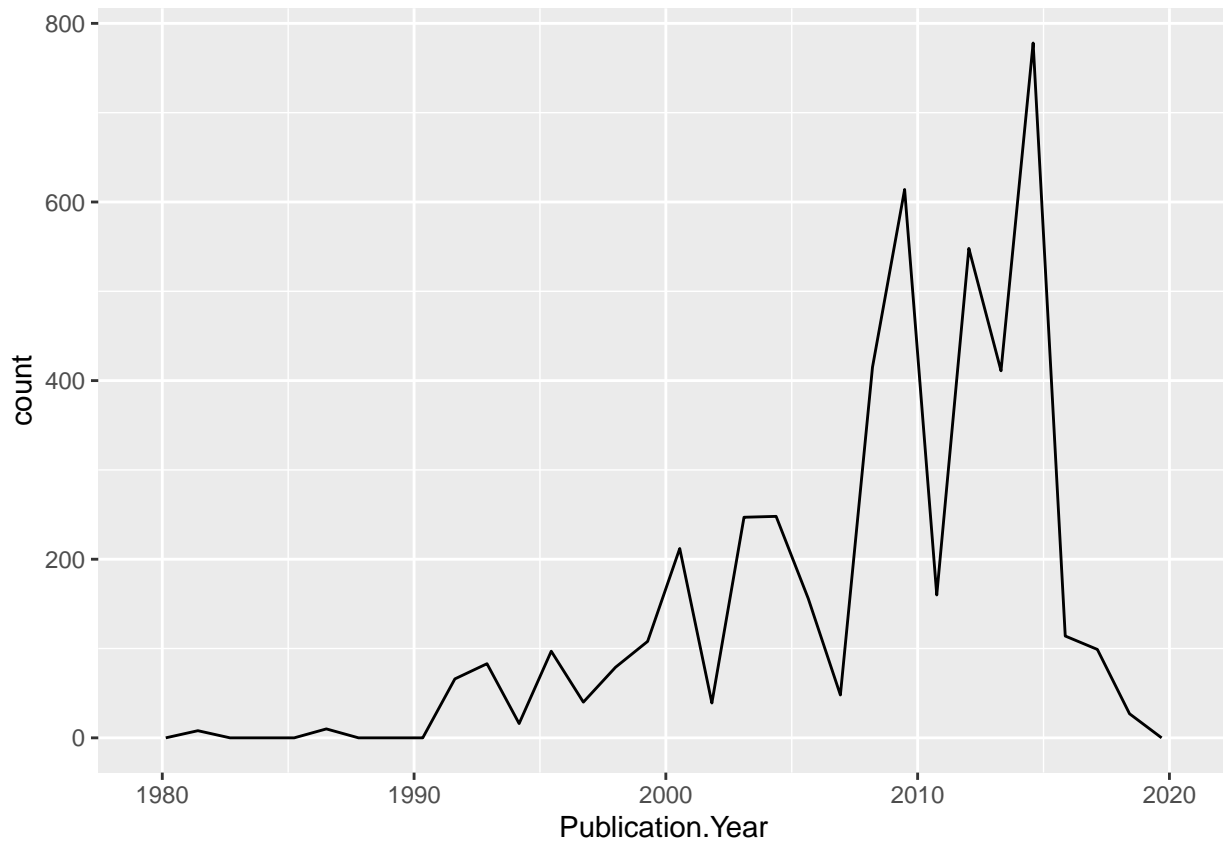
Answer: The class of Conc.1..Author is 'factor'. The class is not numeric because some of the entries in the data have '.' or '/' in them. The data will only be categorized as numeric by R if they're all in numeric format and don't contain any '.', '/', or other symbols/characters that aren't numbers. Since some of them do, R recognizes them as factors instead.

Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
#Using ggplot() and geom_freqpoly() to generate a plot of the # of studies by pub year
ggplot(Neonics)+
geom_freqpoly(aes(x=Publication.Year,))
```

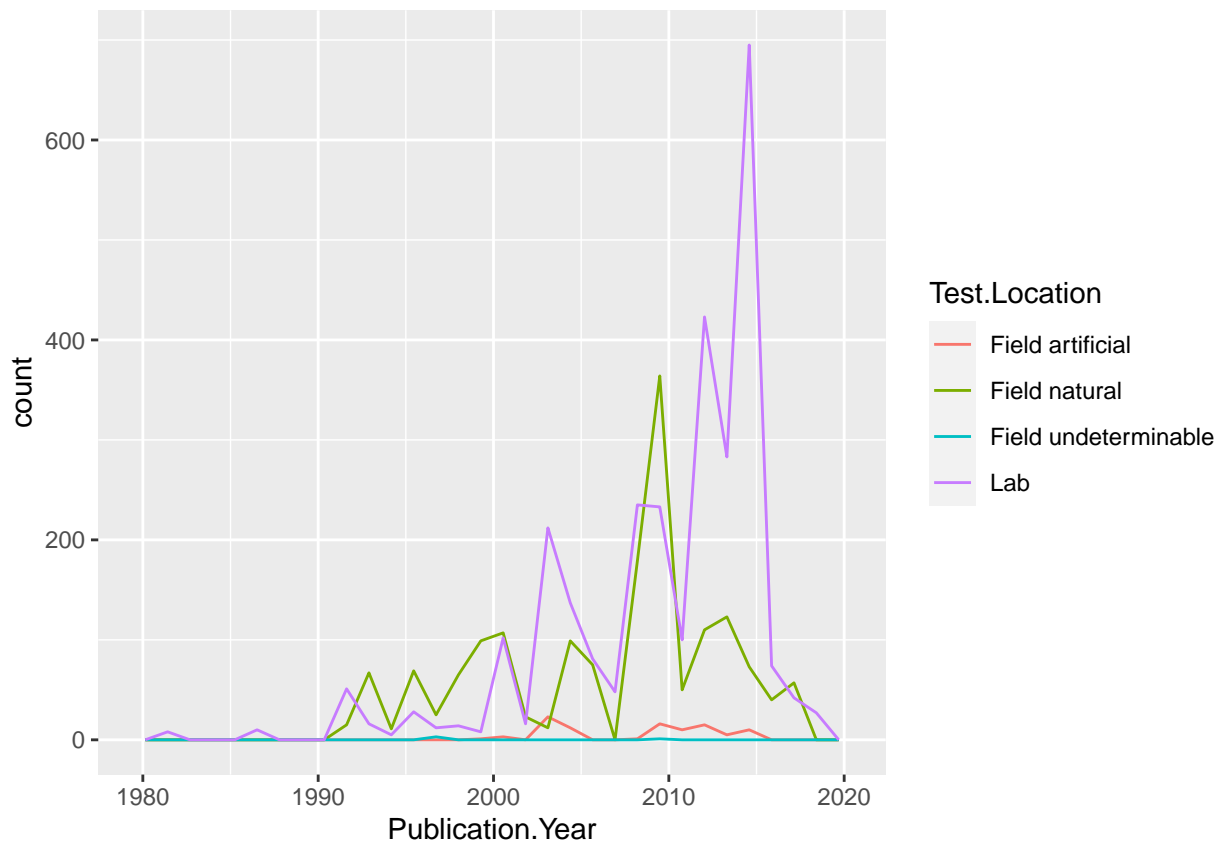
```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
#Using ggplot() and geom_freqpoly() to add a color feature for different test locations
ggplot(Neonics)+
geom_freqpoly(aes(x=Publication.Year, color = Test.Location))
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

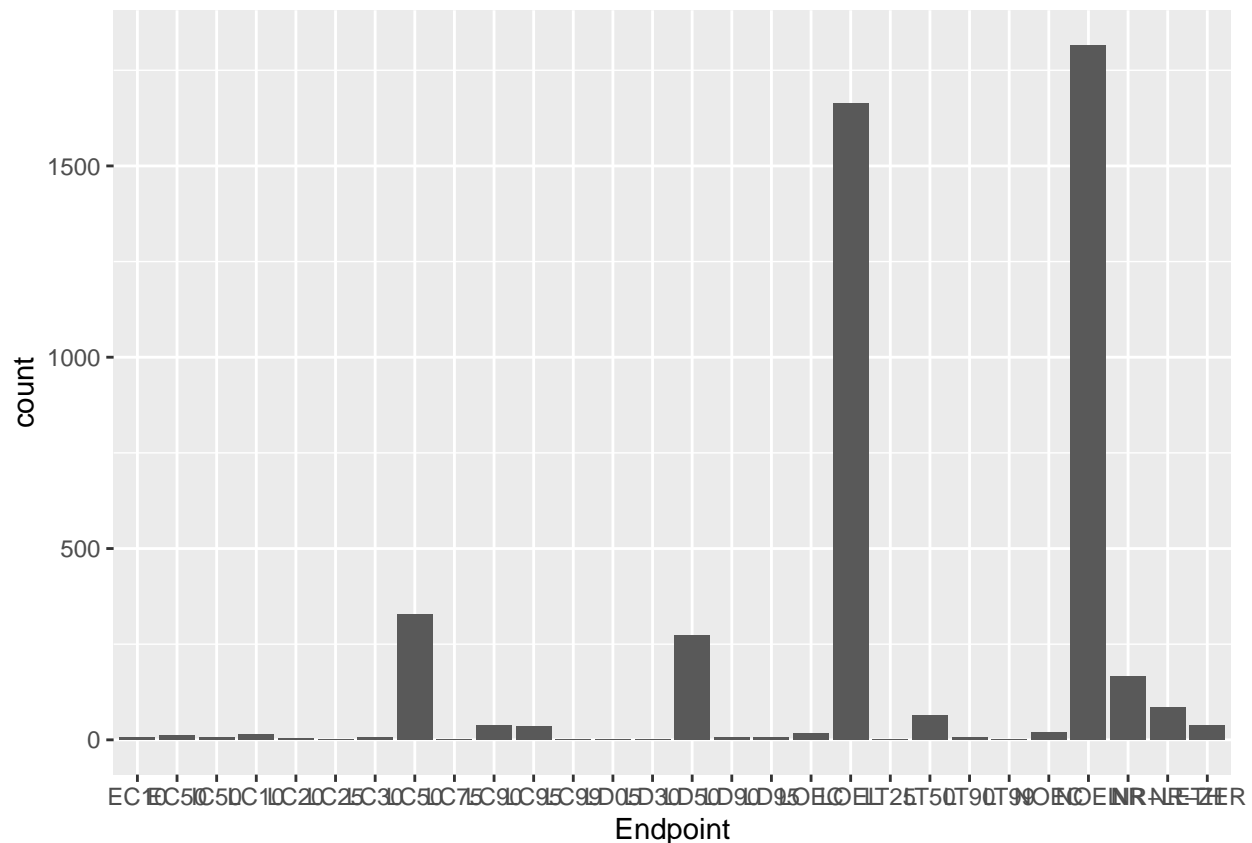


Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are lab and field natural, with very few tests located at field undeterminable and field artificial. The test location trends have differed over time, with lab being most common from 1980-1990, then field natural being more common between 1990-2000, and eventually lab returning as the more common test location from 2000-2020.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

```
#Creating bar graph of Endpoint counts
ggplot(Neonics)+
  geom_bar(aes(x=Endpoint))
```



Answer: The two most common end points are NOEL and LOEL. The NOEL (terrestrial database usage) stands for no-observable-effect-level, meaning that the highest dose produces effects not significantly different from responses of controls according to the author's report statistical test. The LOEL (terrestrial database usage) stands for lowest-observable-effect-level, meaning that the lowest dose produces effects that were significantly different from responses of controls.

Explore your data (Litter)

12. Determine the class of `collectDate`. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
#Determining the class of collectDate
class(Litter$collectDate)
```

```
## [1] "factor"
```

```
#Changing from Factor class to Date class
Litter$collectDate <- as.Date(Litter$collectDate)
#Confirming class of collectDate is now date
class(Litter$collectDate)
```

```
## [1] "Date"
```

```
#Using 'unique' to determine which dates litter was sampled in August 2018.
unique(Litter$collectDate)
```

```
## [1] "2018-08-02" "2018-08-30"
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?


```
#Using 'unique' function to determine how many different types of plots there are by plotID
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

```
#Using 'summary' function to determine the difference between 'unique' and 'summary'
summary(Litter$plotID)
```

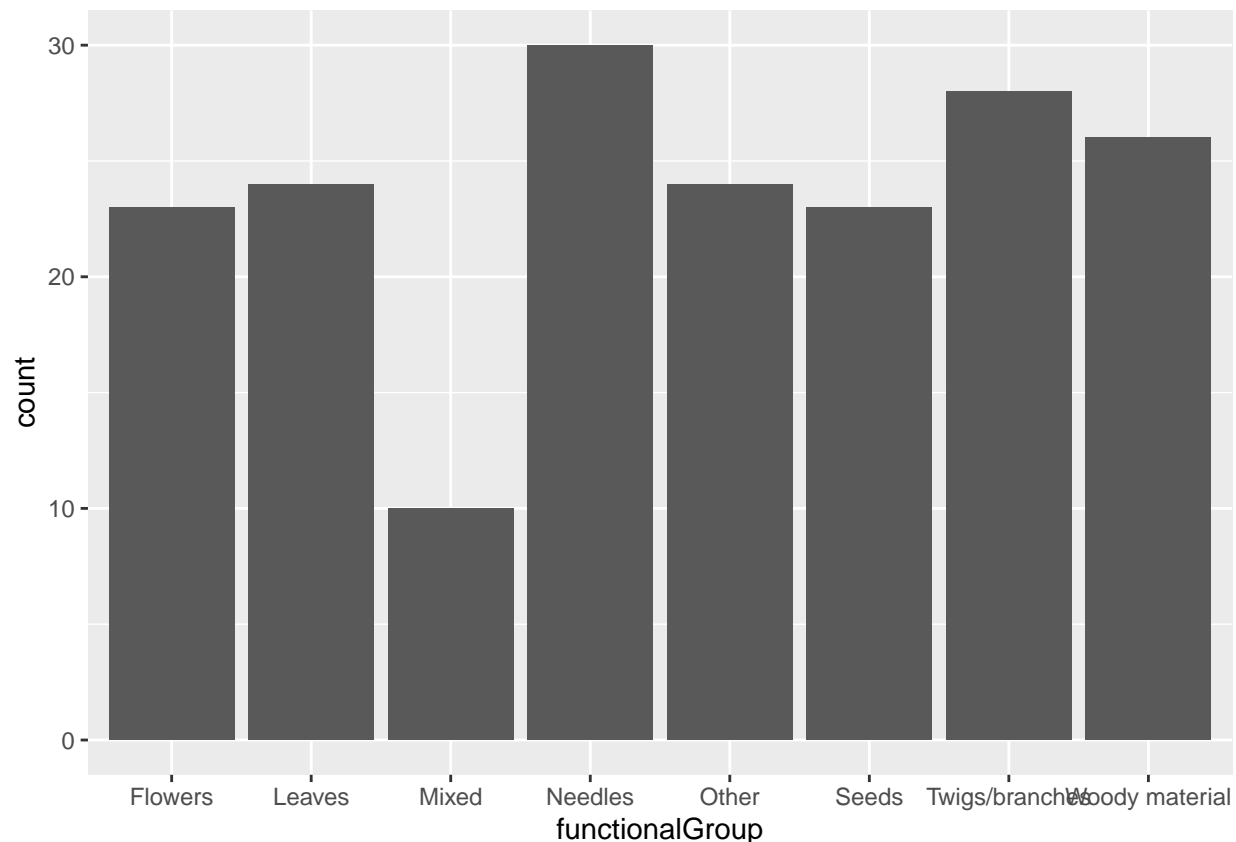
```
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061
##      20      19      18      15      14      8      16      17
## NIWO_062 NIWO_063 NIWO_064 NIWO_067
##      14      14      16      17
```

Answer: There were 12 plots sampled at Niwot Ridge. The information provided from ‘unique’ gives you an overview of how many unique levels there are for one specific column. The ‘summary’ function differs, as it provides a count of how many entries/occurrences of each of these unique levels there are in the dataset.

14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

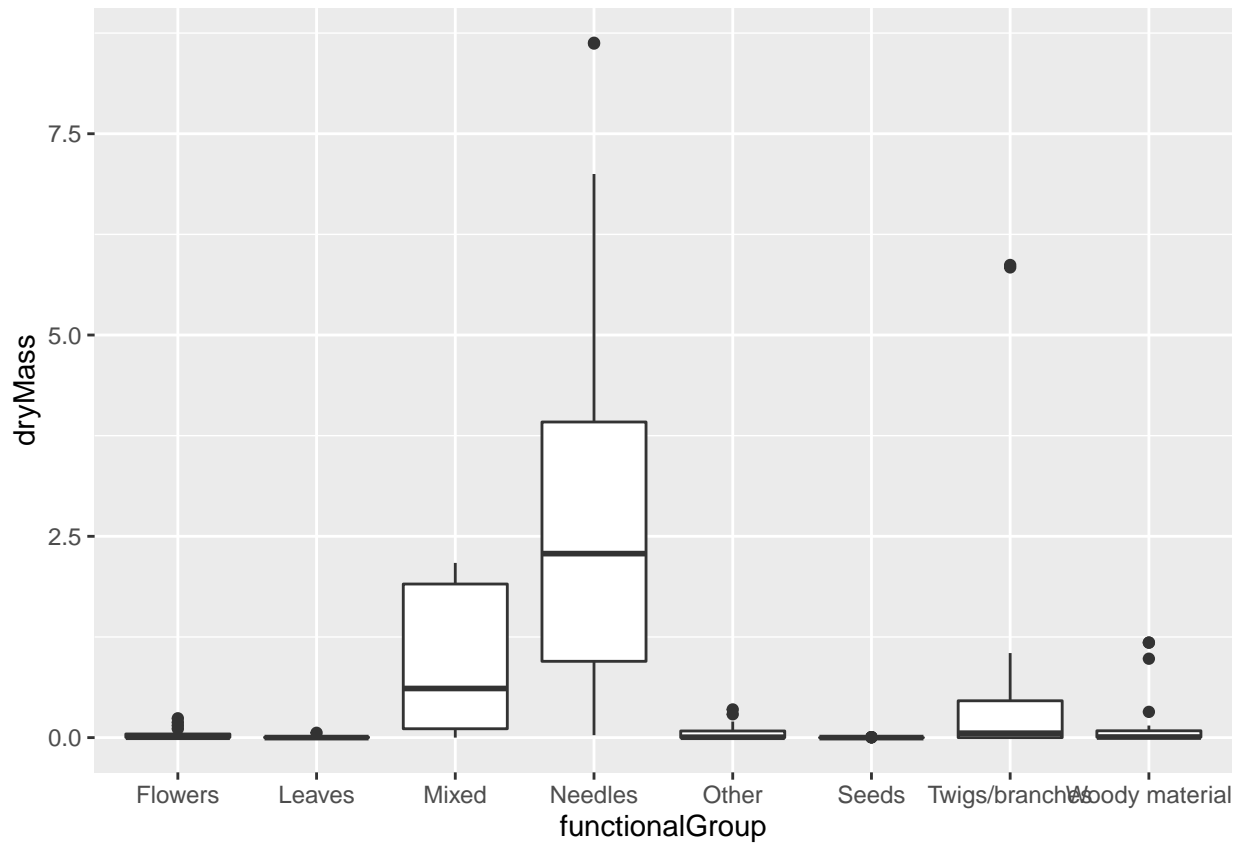
```
#Creating a bar graph of functionalGroup counts
```

```
ggplot(Litter)+
  geom_bar(aes(x=functionalGroup,))
```



15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
#Creating a boxplot of dryMass by functionalGroup
ggplot(Litter)+
  geom_boxplot(aes(x=functionalGroup, y=dryMass))
```

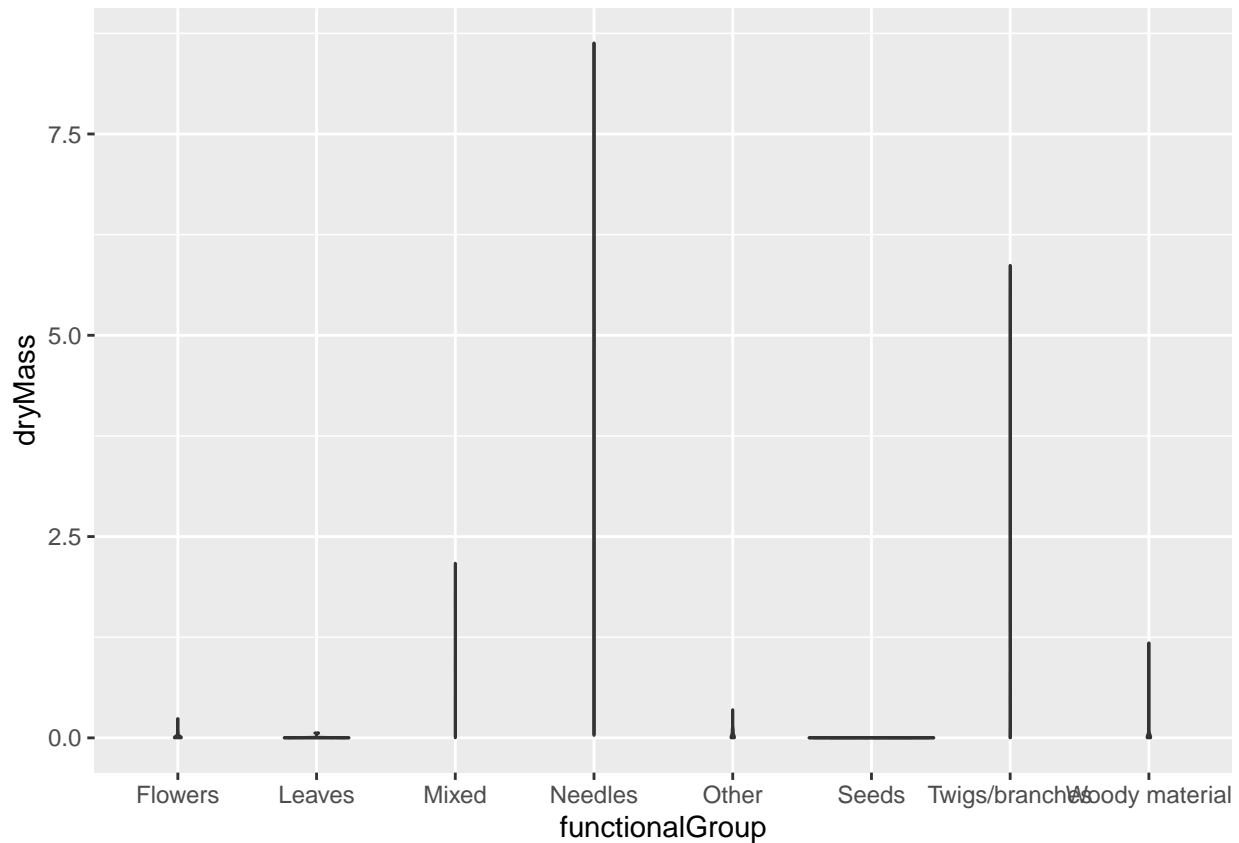


```
#Creating a violin plot of dryMass by functionalGroup
ggplot(Litter)+
  geom_violin(aes(x=functionalGroup, y=dryMass), draw_quantiles = c(0.25, 0.5, 0.75))
```

```
## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values
```

```
## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values
```

```
## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: The boxplot is a more effective visualization option because we only want the summary statistics of the data, and the boxplot is effective at visualizing the summary points of the dryMass data by functionalGroup. The violin plot is not as effective in this case, as it shows the entire distribution of the data, which is not as useful to us in this case.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Based on the plots, it is clear that 'Needles' has the highest biomass at these sites, followed by the 'Mixed' functional group.