

Ancestral state reconstruction (incl. diagnostics)

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Analysis of BayesTraits output

Output from each independent run should be saved into its own subdirectory within the BayesTraits directory.

Set your working directory to the parent directory containing all of your MCMC analyses

```
knitr::opts_knit$set(root.dir = '..' )
```

1) Read in data as mcmc objects

Read in your traces and make sure they are named correctly

```
path <- ("BayesTraits/")
directories <- c("run1/", "run2/")
filename <- "IschMultiState.Log.txt"

for (i in directories){
  x = c(path, i, filename)
  assign(paste("run", which(directories==i), sep=""),
        as.data.frame(read.delim(paste(x, collapse = ""), header=T, skip =
        129,
        na.strings = "--")))
}
```

Set the variables that will be used to create your MCMC object

```
thin <- run1$Iteration[2] - run1$Iteration[1]
start <- min(run1$Iteration)
end <- max(run1$Iteration)
```

Remove variables we are not interested in

```
output <- run1[ , !(names(run1) %in% c("X", "Model.string"))]
```

Divide rates by 100 to get realistic values (because ScaledTrees to 0.001 to prevent rates getting too small)

```
for(i in 6:ncol(output[,1:17])) {
  output[,i] <- output[,i]/100
}
```

Call the “coda” package for creating multiple mcmc objects

```
require(coda)
```

Create mcmc object

```
outputmc <- mcmc(data = output, start = start, end = end, thin = thin)
```

Calculate the percentage of iterations in which there was only a single parameter

```
(sum(outputmc[,4] == 1, na.rm = TRUE)/length(outputmc[,4]))*100
```

```
## [1] 92.1
```

Summarize each parameter into mean and HPD interval of percent support for each state per node

```
nodes_output <- outputmc[,22:181]
summary_table <- matrix(0,3,ncol(nodes_output))
rownames(summary_table) <- c("Mean", "LowerHPD", "UpperHPD")
colnames(summary_table) <- colnames(nodes_output)

for(i in 1:ncol(nodes_output)){
  summary_table[1,i] <- mean(nodes_output[,i], na.rm = TRUE)
  hpd <- HPDinterval(nodes_output[,i])
  summary_table[2,i] <- hpd[1]
  summary_table[3,i] <- hpd[2]
}
```

Reshape data into mean posterior value by node (row) and state (column)

```
mean_state_per_node <- data.frame("node" = 42:81, matrix(summary_table
  [1,], ncol = 4, byrow = TRUE))
colnames(mean_state_per_node) <- c("node", "FMA", "FPD", "FMH", "FPT")
print(mean_state_per_node)
```

##	node	FMA	FPD	FMH	FPT
## 1	42	6.565790e-02	7.504201e-02	8.101822e-01	0.0491178671
## 2	43	9.274950e-02	4.728775e-02	8.072559e-01	0.0527068645
## 3	44	5.120039e-02	2.728475e-02	8.809508e-01	0.0405640207
## 4	45	2.428807e-02	9.627171e-03	9.463384e-01	0.0197463551
## 5	46	3.531183e-02	1.580538e-02	9.208026e-01	0.0280801793
## 6	47	1.777336e-01	9.270024e-02	6.508177e-01	0.0787484276
## 7	48	2.463743e-01	1.411997e-01	5.161916e-01	0.0962344616
## 8	49	6.830667e-02	1.899536e-01	6.900512e-01	0.0516884563
## 9	50	4.539266e-01	8.295473e-02	3.791511e-01	0.0839675974
## 10	51	9.017519e-02	2.134361e-01	6.248634e-01	0.0715252852
## 11	52	7.370373e-02	7.672139e-02	7.984371e-01	0.0511377819
## 12	53	5.701542e-02	4.663767e-02	8.502526e-01	0.0460943197
## 13	54	7.428305e-02	8.988538e-02	7.773171e-01	0.0585144220
## 14	55	4.909961e-02	2.407956e-02	8.880385e-01	0.0387823455
## 15	56	1.440219e-01	1.880322e-01	5.867415e-01	0.0812043822
## 16	57	2.189587e-01	6.865205e-01	3.383505e-02	0.0606857260
## 17	58	1.877773e-02	9.503892e-01	1.187817e-02	0.0189549118
## 18	59	2.286832e-02	9.437721e-01	1.160026e-02	0.0217593602
## 19	60	4.599682e-03	9.919882e-01	7.357941e-04	0.0026762946
## 20	61	5.574331e-03	9.926921e-01	3.473139e-04	0.0013862433
## 21	62	2.074524e-02	9.777176e-01	1.973757e-04	0.0013397804
## 22	63	1.448938e-04	9.996814e-01	3.456607e-05	0.0001391503
## 23	64	4.213800e-01	5.763215e-01	2.195850e-04	0.0020788851
## 24	65	1.178999e-02	9.719373e-01	5.399090e-03	0.0108736573

```
## 25 66 4.907636e-02 4.437551e-02 8.674836e-01 0.0390645205
## 26 67 8.528026e-02 1.910135e-01 6.581645e-01 0.0655417725
## 27 68 2.977692e-02 1.298142e-02 9.334053e-01 0.0238363410
## 28 69 9.058939e-02 7.211650e-01 9.644028e-02 0.0918053710
## 29 70 5.035718e-02 8.524114e-01 4.144335e-02 0.0557881091
## 30 71 3.314916e-02 9.046198e-01 1.843635e-02 0.0437947336
## 31 72 2.809980e-02 9.144222e-01 1.122436e-02 0.0462536739
## 32 73 2.755034e-02 9.070550e-01 7.038726e-03 0.0583558772
## 33 74 6.801236e-02 6.955341e-01 2.294490e-02 0.2135086116
## 34 75 4.420434e-02 4.400526e-01 1.863992e-03 0.5138790764
## 35 76 1.906032e-04 2.075660e-05 9.546388e-06 0.9997790836
## 36 77 6.699462e-05 6.618309e-06 3.974343e-06 0.9999223906
## 37 78 2.914782e-02 9.224737e-01 1.985723e-02 0.0285212553
## 38 79 2.071764e-02 9.467702e-01 1.134417e-02 0.0211680267
## 39 80 2.172570e-02 9.440430e-01 1.254825e-02 0.0216830240
## 40 81 3.666653e-03 9.915628e-01 1.192997e-03 0.0035775245
```

2) Build tree displaying ancestral state reconstruction (Fig. 5)

Call “treeio” and “ggtree” packages for building phylogenetic tree

```
require(treeio)
require(ggtree)
```

Read in StarBEAST2 summary tree

```
beast_tree <- read.beast("StarBEAST2/summary.tree")
```

Create a new tip label variable in tree data

```
relabelling <- data.frame("label" = fortify(beast_tree)$label[1:41])
relabelling$label2 <- sub("_", "□", relabelling$label)
relabelling$label2 <- plyr::revalue(relabelling$label2,
                                   c("Amorphostigma□armstrongi" = "Ischnura□
                                     armstrongi",
                                     "Amorphostigma□sp." = "Ischnura□sp.",
                                     "Rhodischnura□nursey" = "Ischnura□
                                     nursey"))
```

```
beast_tree <- full_join(beast_tree, relabelling, by = "label")
```

Read in female morph state data

```
extant_state <- read.table("BayesTraits/FMorph.txt")
```

Call “dplyr” for manipulating dataframe

```
require(dplyr)
require(dplyr)
```

Create dataframe for pie charts at tips (female morph state of extant species as proposed by the literature) and add it to tree data

```
extant_state <- data.frame("node"= 1:41,
                           arrange(extant_state, match(extant_state$V1,
                                                           fortify(beast_tree)$
                                                           label[1:41])))
```

Reorder states into (FMA, FMH, FPD, FPT)

```
extant_state$V2 <- revalue(as.character(extant_state$V2), c("0"="FMA", "1"
  ="FPD", "2"="FMH", "3"="FPT"))
extant_state$V2 <- ordered(extant_state$V2, levels = c("FMA", "FMH", "FPD",
  "FPT"))
```

Join state data to consensus tree data

```
beast_tree <- full_join(beast_tree, extant_state, by = "node")
```

Create dataframe for pie charts at nodes (mean posterior estimate for each state at each node and percentage of posterior that significantly predicted the dominant state)

```
pie_labels <- data.frame(mean_state_per_node,
  "x" = fortify(beast_tree)$x[42:81],
  "y" = fortify(beast_tree)$y[42:81])
```

Reorder pie_label columns into (node, FMA, FMH, FPD, FPT, x, y)

```
pie_labels <- pie_labels[, c(1, 2, 4, 3, 5, 6, 7)]
```

Call “ggplot2”, “ggtree” and “ggimage” for drawing phylogeny

```
require(ggplot2)
require(ggtree)
require(ggimage)
```

Create phylogeny with female morph states as pies and confidence percentage as text labels at nodes

```
p <- ggtree(beast_tree) +
  geom_tiplab(aes(label = label2), offset = 1) +
  coord_cartesian(clip="off") +
  geom_tippoint(aes(x = x+0.7, color = V2), size = 3) +
  theme_tree(plot.margin=margin(6, 150, 6, 20), legend.position = c(0.1,
    0.9)) +
  scale_color_manual(values = c("FMA"="#56B4E9", "FMH"="#009E73", "FPD"="#D55E00", "FPT"="#CC79A7"),
    labels = c("FM(A)",
      "FM(H)",
      "FP(D)",
      "FP(T)"),
    name = "Female State")
```

```
pies <- nodepie(pie_labels, cols=2:5, color=c("FMA"="#56B4E9", "FMH"="#009E73", "FPD"="#D55E00", "FPT"="#CC79A7"), alpha=0.8)
```

```
tree <- inset(p, pies, width=0.09, height=0.09)
```

```
ggsave("Fig4.pdf", tree, path = "Figures", width = 7, height = 9)
```

3) Prepare data for diagnostics

Remove any parameters that are not of further interest (i.e. anything that is not an iteration, likelihood or rate parameter)

```
attributes(run1)$names
```

##	[1]	"Iteration"	"Lh"	"Tree.No"
##	[4]	"No.Off.Parmeters"	"No.Off.Zero"	"Model.string"
##	[7]	"q01"	"q02"	"q03"
##	[10]	"q10"	"q12"	"q13"
##	[13]	"q20"	"q21"	"q23"
##	[16]	"q30"	"q31"	"q32"
##	[19]	"Root.P.0."	"Root.P.1."	"Root.P.2."
##	[22]	"Root.P.3."	"RecNode42.P.0."	"RecNode42.P.1."
##	[25]	"RecNode42.P.2."	"RecNode42.P.3."	"RecNode43.P.0."
##	[28]	"RecNode43.P.1."	"RecNode43.P.2."	"RecNode43.P.3."
##	[31]	"RecNode44.P.0."	"RecNode44.P.1."	"RecNode44.P.2."
##	[34]	"RecNode44.P.3."	"RecNode45.P.0."	"RecNode45.P.1."
##	[37]	"RecNode45.P.2."	"RecNode45.P.3."	"RecNode46.P.0."
##	[40]	"RecNode46.P.1."	"RecNode46.P.2."	"RecNode46.P.3."
##	[43]	"RecNode47.P.0."	"RecNode47.P.1."	"RecNode47.P.2."
##	[46]	"RecNode47.P.3."	"RecNode48.P.0."	"RecNode48.P.1."
##	[49]	"RecNode48.P.2."	"RecNode48.P.3."	"RecNode49.P.0."
##	[52]	"RecNode49.P.1."	"RecNode49.P.2."	"RecNode49.P.3."
##	[55]	"RecNode50.P.0."	"RecNode50.P.1."	"RecNode50.P.2."
##	[58]	"RecNode50.P.3."	"RecNode51.P.0."	"RecNode51.P.1."
##	[61]	"RecNode51.P.2."	"RecNode51.P.3."	"RecNode52.P.0."
##	[64]	"RecNode52.P.1."	"RecNode52.P.2."	"RecNode52.P.3."
##	[67]	"RecNode53.P.0."	"RecNode53.P.1."	"RecNode53.P.2."
##	[70]	"RecNode53.P.3."	"RecNode54.P.0."	"RecNode54.P.1."
##	[73]	"RecNode54.P.2."	"RecNode54.P.3."	"RecNode55.P.0."
##	[76]	"RecNode55.P.1."	"RecNode55.P.2."	"RecNode55.P.3."
##	[79]	"RecNode56.P.0."	"RecNode56.P.1."	"RecNode56.P.2."
##	[82]	"RecNode56.P.3."	"RecNode57.P.0."	"RecNode57.P.1."
##	[85]	"RecNode57.P.2."	"RecNode57.P.3."	"RecNode58.P.0."
##	[88]	"RecNode58.P.1."	"RecNode58.P.2."	"RecNode58.P.3."
##	[91]	"RecNode59.P.0."	"RecNode59.P.1."	"RecNode59.P.2."
##	[94]	"RecNode59.P.3."	"RecNode60.P.0."	"RecNode60.P.1."
##	[97]	"RecNode60.P.2."	"RecNode60.P.3."	"RecNode61.P.0."
##	[100]	"RecNode61.P.1."	"RecNode61.P.2."	"RecNode61.P.3."
##	[103]	"RecNode62.P.0."	"RecNode62.P.1."	"RecNode62.P.2."
##	[106]	"RecNode62.P.3."	"RecNode63.P.0."	"RecNode63.P.1."
##	[109]	"RecNode63.P.2."	"RecNode63.P.3."	"RecNode64.P.0."
##	[112]	"RecNode64.P.1."	"RecNode64.P.2."	"RecNode64.P.3."
##	[115]	"RecNode65.P.0."	"RecNode65.P.1."	"RecNode65.P.2."
##	[118]	"RecNode65.P.3."	"RecNode66.P.0."	"RecNode66.P.1."
##	[121]	"RecNode66.P.2."	"RecNode66.P.3."	"RecNode67.P.0."
##	[124]	"RecNode67.P.1."	"RecNode67.P.2."	"RecNode67.P.3."
##	[127]	"RecNode68.P.0."	"RecNode68.P.1."	"RecNode68.P.2."
##	[130]	"RecNode68.P.3."	"RecNode69.P.0."	"RecNode69.P.1."
##	[133]	"RecNode69.P.2."	"RecNode69.P.3."	"RecNode70.P.0."
##	[136]	"RecNode70.P.1."	"RecNode70.P.2."	"RecNode70.P.3."
##	[139]	"RecNode71.P.0."	"RecNode71.P.1."	"RecNode71.P.2."
##	[142]	"RecNode71.P.3."	"RecNode72.P.0."	"RecNode72.P.1."
##	[145]	"RecNode72.P.2."	"RecNode72.P.3."	"RecNode73.P.0."
##	[148]	"RecNode73.P.1."	"RecNode73.P.2."	"RecNode73.P.3."
##	[151]	"RecNode74.P.0."	"RecNode74.P.1."	"RecNode74.P.2."
##	[154]	"RecNode74.P.3."	"RecNode75.P.0."	"RecNode75.P.1."
##	[157]	"RecNode75.P.2."	"RecNode75.P.3."	"RecNode76.P.0."
##	[160]	"RecNode76.P.1."	"RecNode76.P.2."	"RecNode76.P.3."

```
## [163] "RecNode77.P.0." "RecNode77.P.1." "RecNode77.P.2."
## [166] "RecNode77.P.3." "RecNode78.P.0." "RecNode78.P.1."
## [169] "RecNode78.P.2." "RecNode78.P.3." "RecNode79.P.0."
## [172] "RecNode79.P.1." "RecNode79.P.2." "RecNode79.P.3."
## [175] "RecNode80.P.0." "RecNode80.P.1." "RecNode80.P.2."
## [178] "RecNode80.P.3." "RecNode81.P.0." "RecNode81.P.1."
## [181] "RecNode81.P.2." "RecNode81.P.3." "RJRates...Mean"
## [184] "X"
```

```
for (i in 1:length(directories)){
  temp <- eval(parse(text=paste("run", i, "[,c(1,2,7:22)]", sep = "")))
  assign(paste("run", i, sep = ""), temp)
}
```

Make your mcmc objects and throw away the burnin

```
for (i in 1:length(directories)){
  temp <- eval(parse(text=paste("run", i, sep = "")))
  assign (paste ("mcmc",i ,sep = ""),
          mcmc(data = temp , start = start, end = end, thin = thin))
}
```

Put them into a list

```
mh.list <-mcmc.list()
for (i in 1:length(directories)){
  temp <- eval(parse(text=paste("mcmc", i, sep = "")))
  mh.list[[i]] <- temp
}
```

2) Diagnostics for a single run

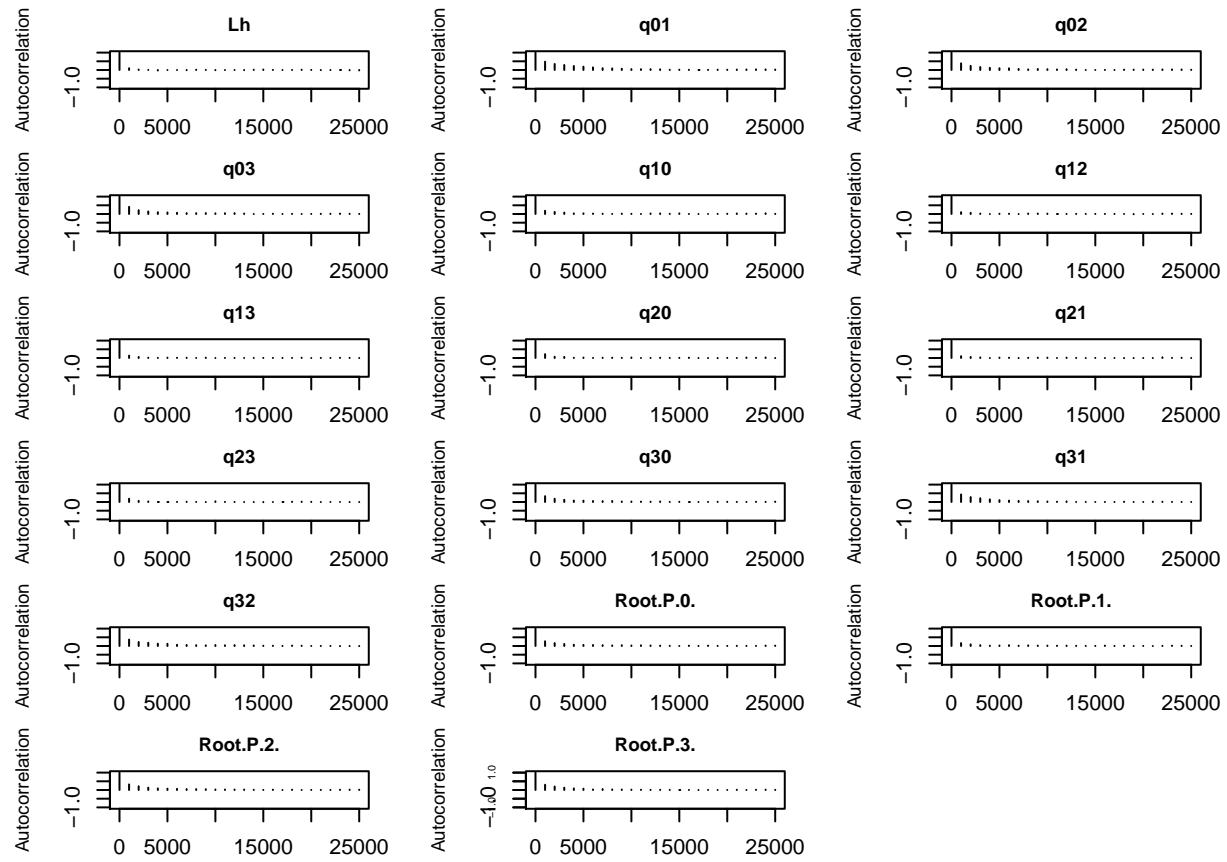
a) **Autocorrelation** Calculate autocorrelation between draws

```
diag(autocorr(mcmc1)[2, , ])
```

```
## Iteration      Lh      q01      q02      q03      q10      q12
      q13
## 0.9996250 0.1020494 0.4791905 0.3840774 0.4103537 0.1877256 0.1036359
      0.1402739
##      q20      q21      q23      q30      q31      q32 Root.P.0.
      Root.P.1.
## 0.2205123 0.1036719 0.1889591 0.3361016 0.4347158 0.3690565 0.2731704
      0.1685609
## Root.P.2. Root.P.3.
## 0.3224923 0.3019637
```

Present autocorrelation in plot form

```
par(mfrow=c(6,3), mar=c(2,4,2,2)+0.1, cex.main = 0.9, cex.lab = 0.9)
autocorr.plot(mcmc1[, -1], lag.max = 25, auto.layout = F)
axis(2, cex.axis=0.5)
```



b) Effective sample size Calculate effective sample size

```
effectiveSize(mcmc1[, -1])
```

```
##          Lh          q01          q02          q03          q10          q12          q13
##          q20
## 6696.540 1461.169 2050.247 2145.036 2634.353 4986.226 5561.923
## 4351.389
##          q21          q23          q30          q31          q32 Root.P.0. Root.P.1.
## Root.P.2.
## 5505.977 4534.217 2454.345 2082.622 1884.090 2674.737 3951.874
## 2423.264
## Root.P.3.
## 2661.313
```

3) Test for convergence of multiple runs

a) Gelman-Rubin statistic Calculate Gelman-Rubin diagnostic of convergence

```
gelman.diag(mh.list, confidence = 0.95, transform=TRUE, autoburnin=FALSE,
            multivariate=FALSE)
```

```
## Potential scale reduction factors:
##
##          Point est. Upper C.I.
```

## Iteration	NaN	NaN
## Lh	1.00	1.00
## q01	1.01	1.01
## q02	1.01	1.02
## q03	1.00	1.01
## q10	1.01	1.01
## q12	1.02	1.02
## q13	1.00	1.00
## q20	1.03	1.03
## q21	1.01	1.01
## q23	1.00	1.01
## q30	1.00	1.00
## q31	1.00	1.00
## q32	1.01	1.01
## Root.P.0.	1.00	1.00
## Root.P.1.	1.00	1.00
## Root.P.2.	1.00	1.00
## Root.P.3.	1.00	1.00

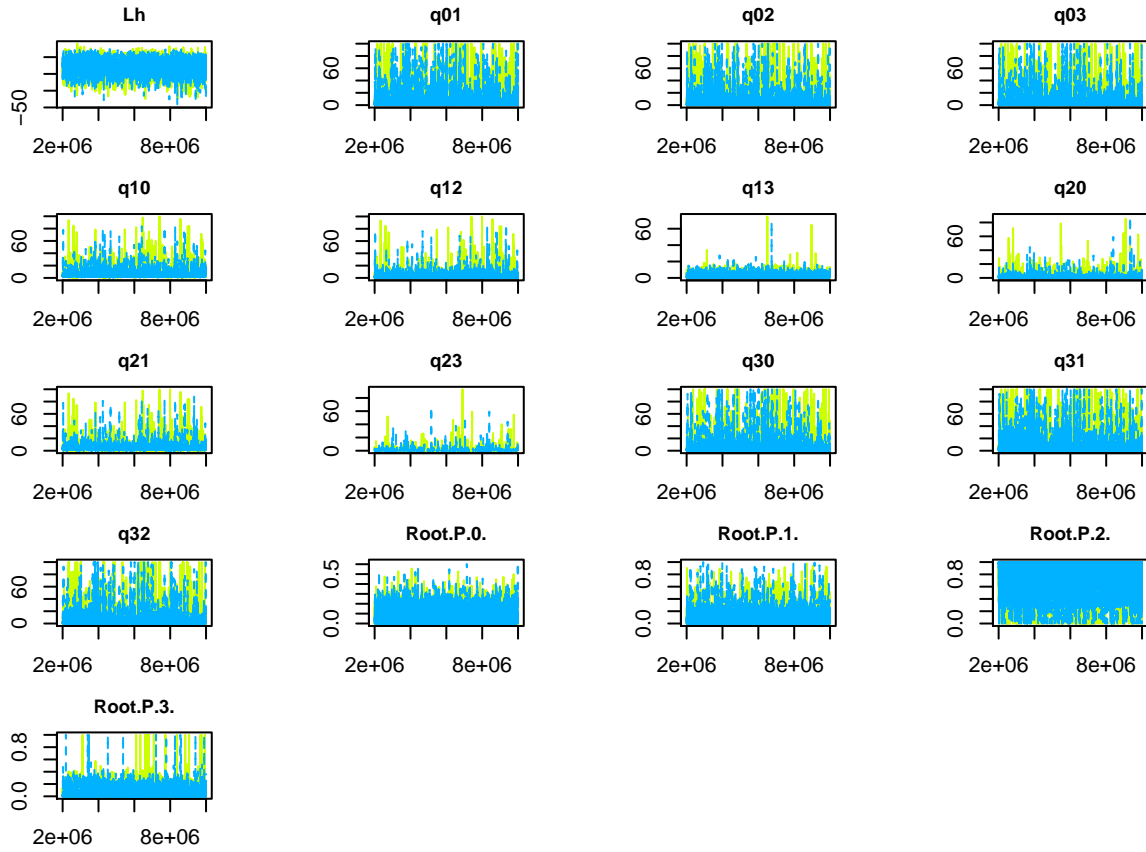
b) **Plotting traces** Get data into correct format for plotting traces from both runs simultaneously

```
for (i in 2:length(run1)){
  for (j in 1:length(directories)){
    temp <- eval(parse(text=paste("mcmc", j, "[,i]", sep = "")))
    assign(paste("trace_", "mcmc", j, colnames(run1[i]), sep = ""), temp)
  }
}

temp_list = mcmc.list()
for (i in 2:length(run1)){
  for (j in 1:length(directories)){
    temp_list[[j]] <- eval(parse(text=paste("trace_mcmc", j , colnames(
      run1[i]),
      sep = "")))
    assign(paste("trace_" , colnames(run1[i]), sep = ""), temp_list)
  }
}
```

Plot traces

```
par(mfrow=c(5,4), mar = c(2,4,2,2)+0.1, cex.main = 0.9)
for (i in 2:length(run1)){
  temp <- eval(parse(text = paste("trace_", colnames(run1[i]), sep = ""))
)
  traceplot(temp,col = rainbow(3, start=0.2, end=0.9))
  title(main = colnames(run1[i]), ylab = "□")
}
```

c) **Plotting density curves** Get data into correct format for plotting density curves of both runs and throw out burnin

```
reshaping <- function(x) {
  y <- rbind(t(x[,-1]))
  result <- setNames(as.data.frame.table(y),c("variable","run","value"))
}
```

```
for (i in 1:length(directories)){
  temp <- eval(parse(text=paste("run", i, sep = "")))
  reshape <- reshaping(temp)
  reshape$run <- as.factor(rep(i, nrow(reshape)))
  assign(paste("reshape", i, sep = ""), reshape)
}
```

```
plot_df <- rbind(reshape1, reshape2)
```

Call the “ggplot2” package for plotting and “ggforce” for paginate function

```
require(ggplot2)
require(ggforce)
```

Plot by each parameter

```
p <- ggplot(plot_df, aes(x= value, fill = run, colour = run)) +
  geom_density(alpha = 0.1) +
  theme_bw(base_size = 12)+
```

```

theme(legend.position = "none", strip.text.x = element_text(size = 6),
      axis.text.y = element_text(size=5), axis.text.x = element_text(
        size=5),
      panel.grid.major = element_blank(), panel.grid.minor = element_
        blank())+
labs(x="Parameter_value", y = "Density") +
facet_wrap(~variable, scales = "free", ncol = 4)

```

p

