# Package 'optima'

July 26, 2023

Title Tapestri Single-Cell Data Analysis

Version 0.1.0

Description The Tapestri platform offers DNA and protein analysis at the single cell level. Integrating both types of data is beneficial for studying multiple cell populations in heterogeneous microenvironments, such as cancer cells. We present optima, an R package for the processing and analysis of data generated from the Tapestri platform. This package provides streamlined functionality for raw data filtering, integration, normalization, transformation, and visualization. Insights gained from the optima package help users to identify unique cell populations and uncover surface protein expression patterns. The results generated by optima help researchers elucidate dynamic changes at the single cell level in heterogeneous microenvironments.

License 'Artistic-2.0'
Encoding UTF-8
Roxygen list(markdown = TRUE)
RoxygenNote 7.2.3
Imports compositions,
 dbscan,
 httr,
 jsonlite,
 methods,
 pheatmap,
 rhdf5,
 umap

# R topics documented:

notate Variant		2
lculatePloidy		2
awHeatmap		3
oowPlot		3
terVariant	4	4
adSignature		5
tClones		6
tCNVmtx		
tDNAmtx		
tInfo		8
tPloidyMtx		8
tProteinMtx	(	Ç

2 calculatePloidy

normalizeCNV normalizeProtein .																	
optima-class														 			
plotPloidy														 			
$plot Protein Feature \ . \\$																	
$plot Variant Feature \ . \\$														 			
readHdf5														 			
reduceDim														 			

Index 15

annotate Variant

Variant annotation

## Description

This function takes variant names as input and returns annotation for annotation table for all variant IDs in a data frame.

## Usage

```
annotateVariant(variant.names)
```

## **Arguments**

variant.names Input variant IDs, can be a vector.

## Value

A data frame with annotation for all input variant IDs.

## **Examples**

annotateVariant(variants\_id)

calculatePloidy

CNV ploidy calculation function

## **Description**

The function uses the normalized CNV matrix to calculate the ploidy for each CNV locus.

## Usage

```
calculatePloidy(optima.obj, diploid.cell)
```

## Arguments

optima.obj optima object.

diploid.cell The cell type that should be considered as diploid cell. This cell type should be

one of cell types in the cell.labels vector.

drawHeatmap 3

#### Value

optima object with normalized CNV

## **Examples**

```
calculatePloidy(optima.obj)
```

drawHeatmap

Heatmap function

#### **Description**

This function creates a heatmap using matrix data. Matrix data can be DNA or protein.

## Usage

```
drawHeatmap(optima.obj, omic.type)
```

## **Arguments**

optima.obj

optima object.

omic.type

Type of data for heat map. Potential values "dna" and "protein". If "dna" is specified, then the VAF data will be used for plot. If "protein" is specified, then

protein expression values will be used.

## Value

Heat map visualization.

#### **Examples**

drawHeatMap()

elbowPlot

Generate elbow plot

## **Description**

This function generate an elbow plot. The plot is useful for determining how many PCs will be used for dimension reduction The input matrix can be the DNA matrix in an optima object.

## Usage

```
elbowPlot(input.mtx)
```

## **Arguments**

input.mtx

Input matrix

4 filterVariant

#### Value

an elbow plot

## **Examples**

```
elbowPlot(example.matrix)
```

filterVariant

DNA Variant filter function

## **Description**

This function uses multiple matrices imported from the h5 file to conduct quality filtering. This includes the sequencing depth matrix, genotype matrix, variant allele frequency matrix, genotype quality matrix The function returns an optima object that has been filtered with variant/cells. In addition, the returned optima object's variant.filter label is changed to "filtered". This function is usually applied before protein and CNV analysis.

## Usage

```
filterVariant(
  optima.obj,
  min.dp = 10,
  min.gq = 30,
  vaf.ref = 5,
  vaf.hom = 95,
  vaf.het = 35,
  min.cell.pt = 50,
  min.mut.cell.pt = 1
)
```

## **Arguments**

optima.obj	An optima object with raw data unfiltered.
min.dp	Minimum depth, defaults to 10. Read depth ranges from 0 to positive infinity. When to change: If your sample ended up with more cells being sequenced than planned, then you may have less reads per cell. In such cases, you could try to lower this parameter to 7, 8 or 9. It is not common to increase this parameter above 10.
min.gq	Minimum genotype quality. The default value is 30. The possible range of GQ score is 0-99. This GQ score is derived from GATK. A higher score means a more confident genotype call. A lower score means genotype call with low confidence. When to change: You may consider decreasing the threshold when the reads sequencing quality is low overall.
vaf.ref	If reference call vaf (GT=0) is larger than vaf.ref, then value in genotype call matrix is converted to GT=3.
vaf.hom	If homozygous call vaf (GT=2) is smaller than vaf.hom, then value in genotype call matrix is converted to GT=3.

findSignature 5

vaf.het

If heterozygous call vaf (GT=1) is smaller than vaf.ref, then value in genotype call matrix is converted to GT=3. If heterozygous call vaf (GT=1) is smaller than vaf.ref, then value in genotype call matrix is converted to GT=3. The default value is 35. For heterozygous cell, there should be 50/50 in two alleles, respectively. However, due to sampling error, there are chances when we sequenced more reads in one allele than the other. This parameter allows for tolerating such situations. We don't normally change this parameter.

min.cell.pt

Minimum threshold for cell percentage that has valid variant call (GT = 0, 1 or 2) after applying the filter. Minimum threshold for cell percentage that has valid variant call (GT = 0, 1 or 2) after applying the filter. The default value is 50%. This means for one variant; we need at least 50% of cells have a valid variant call. When to change: If the variant of interest is in a high GC content region, then PCR amplification is hard. In such cases, you may choose to decrease the percent to 30 or 40 so that your interested variant could come through the filter.

min.mut.cell.pt

Minimum threshold for cell percentage that has mutated genotype (GT = 1 or 2) after applying the filter. The default is 1, corresponds to 1%. This filter is used to remove false positives. When to change: If you know the variant is rare in the data, then you could try lower threshold to try to keep the variant in your dataset.

#### Value

An optima object, The DNA data in the object is filtered, the variant.filter label is "filtered". Meanwhile, the protein matrix and CNV matrix is also updated so that only cells withstand DNA variant filter are kept.

#### **Examples**

filterVariant(optima.obj)

findSignature

Identify signature protein function

## **Description**

This function compares protein levels for a input cell type against all other cells using t test. This function returns a data frame ranked by FDR adjusted p-value. If two cell types specified, then this function compares protein expression between the two cell types.

#### Usage

```
findSignature(optima.obj, cell.type, cell.type.2 = "all")
```

#### **Arguments**

optima.obj optima object.

cell.type Input cell type to compare protein level to all other cell types.

cell.type.2 Second cell type to compare.

6 getClones

## Value

Data frame of all proteins p-values comparing protein levels of input cell type to all other cell types. In addition, it provides the mean difference between Input cell type and other cells. If mean difference is positive, this mean more expression in the cell type.

## **Examples**

```
findSignature(optima.obj, cell.type)
```

getClones

Clustering variant function

## **Description**

This function identifies cell clones based on DNA variant data.

## Usage

```
getClones(optima.obj, eps = 1, minPts = 100, num.PC = 5, plot = FALSE)
```

## **Arguments**

optima.obj	optima object.
eps	size/radius of the epsilon neighborhood. This argument will passed to dbscan function.
minPts	number of minimum points required in the eps neighborhood for core points, including the point itself. This argument will passed to dbscan function.
plot	if True, a UMAP plot will be generated based on the dimension reduction result from variant matrix. Default is FALSE.

## Value

A list, the first element in the list is the optima object with labels assigned based on dbscan. the second element in the list is the dimension reduction result based on VAF matrix.

```
getClones(my.obj)
```

getCNVmtx 7

getCNVmtx

The getter function for CNV matrix

## Description

This function returns the CNV matrix within the optima object.

## Usage

```
getCNVmtx(optima.obj)
```

## **Arguments**

```
optima.obj
```

optima object.

#### Value

A matrix that contains CNV data in the optima object. The row names are cell id, the column names are CNV IDs.

## **Examples**

```
getCNVmtx(my.obj)
```

getDNAmtx

The getter function for VAF matrix

## Description

This function returns the VAF matrix within the optima object.

## Usage

```
getDNAmtx(optima.obj)
```

## **Arguments**

```
optima.obj
```

optima object.

## Value

A matrix that contains VAF data in the optima object. The row names are cell id, the column names are variant ID.

```
getDNAmtx(my.obj)
```

8 getPloidyMtx

getInfo

Single variant ID annotation function

## **Description**

Returns annotation from one variant ID. This function is not visable to users.

## Usage

```
getInfo(variant)
```

## Arguments

variant

variant name in a string

#### Value

Annotation information for one specific variant ID.

## **Examples**

```
getInfo(variant_id)
```

getPloidyMtx

The getter function for Ploidy matrix

## Description

This function returns the Ploidy matrix within the optima object.

## Usage

```
getPloidyMtx(optima.obj)
```

## **Arguments**

```
optima.obj
```

optima object.

## Value

A matrix that contains Ploidy data in the optima object. The row names are cell id, the column names are CNV IDs.

```
getPloidyMtx(my.obj)
```

getProteinMtx 9

getProteinMtx

The getter function for Protein matrix

## Description

This function returns the Protein matrix within the optima object.

## Usage

```
getProteinMtx(optima.obj)
```

## **Arguments**

```
optima.obj optima object.
```

#### Value

A matrix that contains Protein data in the optima object. The row names are cell IDs, the column names are protein IDs.

## **Examples**

```
getProteinMtx(my.obj)
```

normalizeCNV

CNV normalization function

## Description

The function normalizes the CNV matrix to correct for column-wise and row-wise variation and updates the optima object amp.normalize.method from "unnormalized" to "normalized".

## Usage

```
normalizeCNV(optima.obj)
```

## **Arguments**

```
optima.obj optima object.
```

#### Value

optima object with normalized CNV and amp.normalize.method updated to "normalized".

```
normalizeCNV(optima.obj)
```

10 optima-class

normalizeProtein

Protein matrix normalization

#### **Description**

The function normalizes protein matrix within an optima object using CLR method.

#### **Usage**

```
normalizeProtein(optima.object)
```

#### **Arguments**

```
optima.obj optima object.
```

#### Value

An optima object with protein matrix being normalized and protein.normalize.method label updated to "normalized".

## **Examples**

```
normalizeProtein(optima.object)
```

optima-class

optima object

#### **Description**

An optima object contains DNA, protein and CNV for Tapestri platform single cell sequencing data.

## Value

Object containing DNA, protein and CNV single cell sequencing data.

#### **Slots**

meta.data User-defined metadata can be kept with the object.

cell.ids A vector of cell IDs/barcodes from Tapestri. This vector should contain unique IDs.

cell.labels A vector that is used to store the cell type information for each cell.

variants A vector of variant IDs.

variant.filter A string that keeps track of if optima object is being QC filtered on its variant matrix.

vaf.mtx Variant matrix.

gt.mtx Genotype matrix, The integers within matrix are reference call GT=0, heterozygous call GT=1, homozygous call GT=2, no calls GT=3.

dp.mtx Sequencing depth matrix.

gq.mtx Genotype quality.

plotPloidy 11

```
amps A vector of CNV locus.
```

amp.normalized.method A string that keeps track of if optima object is being normalized on its CNV matrix.

```
amp.mtx CNV matrix.
```

ploidy.mtx Ploidy matrix.

proteins A vector of surface protein ID.

protein.normalize.method A string that keeps track of if optima object is being normalized on its protein matrix.

protein.mtx Protein matrix

## **Examples**

setClass()

plotPloidy

Ploidy scatter plot function

## Description

For a specified cell type, this function creates a scatter plot indicating ploidy for different CNV loci.

## Usage

```
plotPloidy(optima.obj, cell.type)
```

## **Arguments**

optima.obj optima object.

cell.type String that indicates which cell type,

## Value

optima object with normalized CNV.

```
plotPloidy()
```

12 plotVariantFeature

plotProteinFeature Plot specific protein levels

## **Description**

This function create a plot based on protein level dimension reduction result. Within the plot, Each cell was colored based on the protein level

#### Usage

```
plotProteinFeature(optima.obj, protein.name, reduceDim.obj)
```

## **Arguments**

optima.obj optima object.

protein.name the specific protein user interested

reduceDim.obj dimension reduction result returned by reduceDim() function.

#### Value

A scatter plot based on dimension reduction result. Each cell is colored based on the protein level. The more expression.

## **Examples**

```
plotProteinFeature(my.obj, "CD11b", protein.reduceDim)
```

plotVariantFeature Plot specific variant VAF levels

## **Description**

This function create a plot based on VAF dimension reduction result. Within the plot, Each cell was colored based on the VAF

## Usage

```
plotVariantFeature(optima.obj, vaf.name, reduceDim.obj)
```

## **Arguments**

optima.obj optima object.

vaf.name the specific variant name user interested

 ${\tt reduceDim.obj} \quad {\tt dimension} \ {\tt reduction} \ {\tt result} \ {\tt returned} \ {\tt by} \ {\tt reduceDim}() \ {\tt function}.$ 

## Value

A scatter plot based on dimension reduction result. Each cell is colored based on the protein level. The more expression.

readHdf5

#### **Examples**

```
plotVariantFeature(my.obj, "chr4:106190862:T/C", vaf.reduceDim)
```

readHdf5

H5 file to optima object function

## **Description**

This function read in a h5 file and return one optima object. The h5 file can be found in the Tapestri pipeline software output. The .h5 file contains all necessary data needed for single cell DNA and protein analysis.

## Usage

```
readHdf5(directory, sample.name, omic.type = "DNA+protein")
```

## **Arguments**

directory Directory for the input h5 file.

sample.name A sample name that will be used naming visualizations.

omic.type This parameter indicates if the data set is DNA only or DNA+protein. It has two

possible values: "DNA+protein" or "DNA". The default value is "DNA+protein".

## Value

optima object

## **Examples**

readHdf5("path/to/my/file.h5")

reduceDim

Dimension reduction function

## **Description**

This function reduces dimensions for a data matrix, such data matrix can be protein or DNA matrix in an optima object.

## Usage

```
reduceDim(input.mtx, num.PC = 5)
```

## **Arguments**

input.mtx Input optima object.

num. PC The number of PCs being used for preserving variation. Default is 5

14 reduceDim

## Value

List containing PCA result and umap result derived from first 5 PCs

## Examples

reduceDim(example.matrix)

# **Index**

* DNA
filterVariant,4
* being
calculatePloidy, 2
* cell.type
findSignature, 5
* directory
readHdf5, 13
* filter
filterVariant,4
* optima.obj,
findSignature, 5
* optima.obj
calculatePloidy, 2
normalizeCNV, 9
normalizeProtein, 10
plotPloidy, 11
* ploidy.mtx
calculatePloidy, 2
* updated
calculatePloidy, 2
* variant
annotate $Variant, 2$
getInfo, 8
* with
calculatePloidy, 2
annotate $Variant, 2$
calculatePloidy, 2
drawHeatmap, 3
elbowPlot, 3
filterVariant,4
findSignature, 5
.03
getClones, 6
getCNVmtx, 7
getDNAmtx, 7
getInfo, 8
getPloidyMtx, 8
<pre>getProteinMtx,9</pre>

```
normalizeCNV, 9
normalizeProtein, 10
{\tt optima}\,({\tt optima-class}),\,10
optima-class, 10
plotPloidy, 11
{\tt plotProteinFeature}, \\ 12
plotVariantFeature, 12
readHdf5, 13
reduceDim, 13
```