

Design, Synthesis, and Synergistic Activity of Eight-Membered Oxabridged Neonicotinoid Analogues

Xiao Zhang, Yiping Wang, Zhiping Xu, Xusheng Shao, Zewen Liu, Xiaoyong Xu, Peter Maienfisch, and Zhong Li*



Cite This: *J. Agric. Food Chem.* 2021, 69, 3005–3014



Read Online

ACCESS |



Metrics & More



Article Recommendations



Supporting Information

ABSTRACT: Insecticide synergists are sought-after due to their potential in improving the pesticide control efficacy with a reduced dose of an active ingredient. We previously reported that a *cis*-configuration neonicotinoid (IPPA08) exhibited specific synergistic activity toward neonicotinoid insecticides. In this study, we synthesized a series of structural analogues of IPPA08 by converting the pyridyl moiety of IPPA08 into phenyl groups, via facile double-Mannich condensation reactions between nitromethylene compounds and glutaraldehyde. All of the oxabridged neonicotinoid compounds were found to increase the toxicity of imidacloprid against *Aphis craccivora*. Notably, compound **25** at 0.75 mg/L lowered the LC₅₀ value of imidacloprid against *A. craccivora* by 6.54-fold, while a 3.50-fold reduction of the LC₅₀ value was observed for IPPA08. The results of bee toxicity test showed that compound **25** display selectivity in its effects on imidacloprid toxicity against the honey bee (*Apis mellifera* L.). In summary, replacing the pyridyl ring with a phenyl ring was a viable approach to obtain a novel synergist with oxabridged moiety for neonicotinoid insecticides.

KEYWORDS: neonicotinoid, insecticide synergist, imidacloprid, selectivity

INTRODUCTION

Since the introduction of imidacloprid in the 1990s, neonicotinoid insecticides have played crucial roles in pest control due to their safety profile, insecticidal spectrum, and systemic properties.^{1–3} Currently, neonicotinoids represent the largest insecticide group, accounting for over 25% of the global insecticide market.⁴ However, the abusive use of neonicotinoids in the past three decades has brought about tough challenges, such as resistance and toxicity to honey bees.^{5,6} Also, neonicotinoids are considered to have side effects on insect pollinators at sublethal concentrations through the potential uptake from crops and wild plants.^{7,8} Recently, these concerns about sublethal effects of neonicotinoids on bees have led to a reevaluation of the overall biological safety of neonicotinoids in Europe and elsewhere.⁹ Insecticide synergists are a potential solution for these issues. They can greatly enhance the activity of the existing pesticides and therefore lower the necessary dose of the active ingredient.

Cycloxaprid is a new class of neonicotinoid with a unique oxabridged substructure (Figure 1). It exhibited excellent activity, high selectivity, and good safety profiles, and was

recently approved in China.^{10–14} Interestingly, an eight-membered oxabridged homologue of cycloxaprid, i.e., IPPA08 (Figure 1), was almost inactive to insects and yet exhibited synergistic effects on traditional neonicotinoids, including imidacloprid, acetamiprid, thiacloprid, and clothianidin. We also find that IPPA08 presented little effects on the toxicities of dinotefuran and cycloxaprid. Also, IPPA08 did not display selectivity in its effects on imidacloprid toxicity against the honey bee (*A. mellifera* L.).^{15,16} Therefore, we sought to modify the oxabridged structure, by changing the peripheral pyridyl ring into a phenyl ring, to further improve its synergistic activity as well as with selectivity and safety toward honey bees.

Herein, a focused library of eight-membered oxabridged compounds were synthesized from nitromethylene analogues and glutaraldehyde, and their structure–activity relationships (SARs) were thoroughly tested.

MATERIALS AND METHODS

Chemicals. Cycloxaprid, paichongding, and IPPA08 were synthesized and purified according to a literature procedure.¹⁷ Other insecticides were purchased from Sigma-Aldrich (St. Louis, MO).

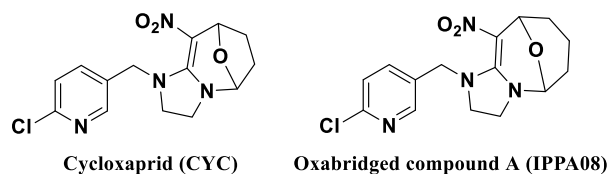


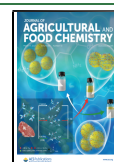
Figure 1. Oxabridged neonicotinoid compounds cycloxaprid (CYC) and IPPA08.

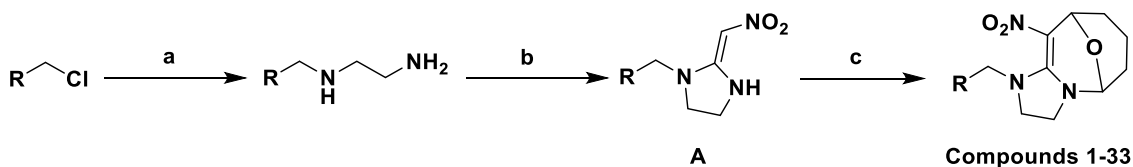
Received: July 27, 2020

Revised: December 23, 2020

Accepted: February 5, 2021

Published: March 2, 2021



Scheme 1. Synthetic Routes of Target Compounds^a

^aReagents and conditions: (a) ethane-1,2-diamine, CH₃CN, 0–5 °C; (b) 1,1-bis(methylthio)-2-nitroethene, CH₃CH₂OH, refluxing; (c) glutaraldehyde, concentrated HCl, rt.

Table 1. Compounds 1–33

| | | | | | |
|--|--|--|--|--|--|
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |
| | | | | | |

Experiment Design. First, we tested the standard curve of imidacloprid and calculated the LC₃₀ (0.997 mg/L) and LC₅₀ (1.72 mg/L) values. To screen out candidates with significant synergistic effect, the mortality of a mixture of imidacloprid (1 mg/L) with the new compounds at low concentrations (0.5, 1, 2, 4 mg/L) against *Aphis craccivora* was analyzed. To obtain the initial synergistic concentration **a** and the maximum synergistic concentration **b** (the minimum concentration when the mortality reaches the maximum), a further refined concentration gradient was performed. Finally, we assessed the LC₅₀ of imidacloprid when the candidate compound is combined with imidacloprid at the initial synergistic concentration **a** and maximum synergistic concentration **b**, and compared with the blank to calculate the synergism ratio.

Insects and Bioassay. Cowpea Aphid (*A. craccivora*) is a susceptible strain from our laboratory, and the bioassay was carried out using the leaf dip bioassay.¹⁸ Aqueous solution of each compound was prepared, to which 0.1% Triton X-100 (0.1 mg/L) as a surfactant and dimethyl sulfoxide (DMSO) as a solvent were added. The mixture was then diluted with 0.1% Triton X-100 (0.1 mg/L) to obtain a series of concentrations of 4, 2, 1, and 0.5 mg/L and so on. Several adults of aphid were selected, avoiding light, and given a starvation treatment about an hour in the dark. Then, they were let to eat broad bean seedling until their mouthparts pierce the bean sprout (2–3 h). Then, the leaves of the horsebean plant, infested with 20–30 aphids, were dipped for 5 s in each concentration three times. This

procedure was repeated three times in each group. After treatment, the burgeoning shoots were placed in a conditioned room (25 ± 1 °C, 50% RH). Water containing Triton X-100 (0.1 mg/L) was used as control. The mortality was recorded in 48 h. The data obtained were analyzed using Polo software (LeOra Software, Inc., Cary, NC) to determine the LC₅₀ values.

Bee Toxicity Assay. Honey bees (*Apis mellifera* L.) were provided by Chinese Academy of Agricultural Sciences (Beijing, China), and a bioassay was performed following the Organization for Economic Cooperation and Development (OECD) method for contact toxicity and oral toxicity tests.¹⁹

Contact Assay. Honey bees (*A. mellifera* L.) were put into a dryer and anesthetized by 5 mL of diethyl ether for 3 min before the test. Then, solutions of different concentrations were added dropwise on the pronotum of the bees by a 1.00 μL microdropper. The bees were enclosed in the cage in time before the bees fully recovered and were fed with 33% honey water. The cage was put on the laboratory table and covered by a black cloth, and acetone was used as control.

Uptake Assay. Degrease cotton was dipped in diluted solutions of the chemicals, to which was added Tween 80 and diluted by 33% honey water until saturation. Then, the degrease cotton was spread on a gauze net in the cage and a 50 mL beaker was placed on the degrease cotton so that the honey bees (*A. mellifera* L.) could suck the liquid. The cage was placed on a laboratory table and covered by a black cloth. The amount of acetone with Tween 80 was the same as

the maximum concentration of diluted solution. It was used as control.

The mortality rates and poisoning symptoms were recorded 24 and 48 h after treatment. The test data were processed by SPSS12.0 to obtain the LD₅₀ (bee contact toxicity) for 24 and 48 h, and LD₅₀ (bee oral toxicity) for 24 h and 48 h and 95% confidence intervals.

Instruments. All melting points were obtained with a Buchi Melting Point B540 and are uncorrected. Nuclear magnetic resonance spectra were recorded on a Bruker AM-400 spectrometer with CDCl₃ or DMSO-*d*₆ as the solvent (¹H at 400 MHz and ¹³C at 100 MHz) and TMS as the internal standard. Chemical shifts are reported in parts per million (δ). Coupling constants (*J*) are reported in hertz (Hz). High-resolution electron mass spectra (electrospray ionization time-of-flight (ESI-TOF)) were performed on a Micromass liquid chromatography-TOF (LC-TOF) spectrometer. High-resolution mass spectra (HRMS) EI were recorded under electron impact (70 eV) condition using a Micromass GCT CA 055 instrument. Analytical thin-layer chromatography (TLC) was carried out on precoated plates (silica gel 60 F₂₅₄), and spots were visualized with ultraviolet (UV) light. All other solvents and reagents were obtained from commercial sources without further purification.

RESULTS

Synthesis. The synthetic route of the target compounds is summarized in Scheme 1. The structures of the title compounds were well characterized by ¹H NMR, ¹³C NMR, and HRMS (ESI).

General Procedure for the Preparation of Compounds. A mixture of starting material A (nitromethylene analogues) (2 mmol) and concentrated hydrochloric acid (0.1 mL) in MeCN (6 mL) was stirred at room temperature. After 5 min of stirring, a 25% glutaraldehyde aqueous solution (1 mL) was added slowly. Then, the reaction mixture was stirred at 0 °C to room temperature and monitored by TLC. After completion, the pH value of the reaction mixture was adjusted to 7–8 by saturated aqueous NaHCO₃ and the solvent was removed under reduced pressure. The resulting mixture was extracted with dichloromethane (3 × 20 mL), and the organic layer was washed with brine, dried over anhydrous sodium sulfate, filtered, and concentrated. The residue was purified by silica gel column chromatography (DCM/MeOH as eluent) to afford the pure products (1–33, Table 1). The data of 1–33 are shown as follows.

1-Methyl-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (1). Yield, 80%; mp, 109.8–110.5 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 5.11–4.99 (m, 2H), 3.86–3.56 (m, 4H), 3.14 (s, 3H), 1.84 (d, *J* = 13.4 Hz, 1H), 1.76–1.65 (m, 3H), 1.58–1.49 (m, 1H), 1.47–1.36 (m, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 156.45, 104.44, 81.46, 68.64, 51.73, 45.97, 38.62, 28.56, 26.49, 14.45 ppm. HRMS (ESI) calcd for C₁₀H₁₆N₃O₃ [M + H]⁺, 226.1192, found, 226.1193.

1-Benzyl-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (2). Yield, 81%; mp, 154.5–155.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.43–7.25 (m, 5H), 5.16–5.04 (m, 2H), 4.98 (d, *J* = 15.2 Hz, 1H), 4.78 (d, *J* = 15.2 Hz, 1H), 3.75–3.56 (m, 4H), 1.85–1.66 (m, 4H), 1.54–1.50 (m, 1H), 1.44–1.30 (m, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 156.02, 137.01, 128.45, 127.75, 127.39, 105.35, 81.25, 68.52, 53.60, 49.15, 45.80, 28.23, 26.70, 14.33 ppm. HRMS (ESI) calcd for C₁₆H₂₀N₃O₃ [M + H]⁺, 302.1505, found, 302.1506.

1-(2-Fluorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (3). Yield, 85%; mp, 144.1–144.8 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.38–7.35 (m,

2H), 7.23–7.19 (m, 2H), 5.12 (s, 1H), 5.03 (s, 1H), 4.97 (d, *J* = 15.8 Hz, 1H), 4.89 (d, *J* = 15.8 Hz, 1H), 3.82–3.54 (m, 4H), 1.80–1.61 (m, 4H), 1.52 (d, *J* = 13.8 Hz, 1H), 1.38–1.35 (m, 1H) ppm. ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -118.36 (s, 1F) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 161.31, 155.99 (d, *J*_{C-F} = 255.8 Hz, 1C), 129.88, 129.44, 124.59 (d, *J*_{C-F} = 19.2 Hz, 1C), 123.87, 115.21 (d, *J*_{C-F} = 3.9 Hz, 1C), 105.33, 81.24, 68.43, 49.44, 47.95, 45.83, 28.16, 26.78, 14.21 ppm. HRMS (ESI) calcd for C₁₆H₁₉N₃O₃F [M + H]⁺, 320.1410, found, 320.1411.

1-(3-Fluorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (4). Yield, 86%; mp, 129.3–130.7 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.53–7.03 (m, 4H), 5.07–5.01 (m, 3H), 4.75 (d, *J* = 15.6 Hz, 1H), 3.68 (d, *J* = 18.2 Hz, 4H), 1.90–1.29 (m, 6H) ppm. ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -113.32 (s, 1F) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 163.42, 156.08 (d, *J*_{C-F} = 255.1 Hz, 1C), 140.09, 130.32 (d, *J*_{C-F} = 9.8 Hz, 1C), 123.69 (d, *J*_{C-F} = 4.6 Hz, 1C), 114.44, 114.12, 105.30, 81.23, 68.45, 53.40, 49.37, 45.79, 28.25, 26.70, 14.32 ppm. HRMS (ESI) calcd for C₁₆H₁₉N₃O₃F [M + H]⁺, 320.1410, found, 320.1411.

1-(4-Fluorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (5). Yield, 76%; mp, 143.3–144.1 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.42–7.36 (m, 2H), 7.23–7.16 (m, 2H), 5.11 (s, 1H), 5.06 (d, *J* = 2.4 Hz, 1H), 4.96 (d, *J* = 15.0 Hz, 1H), 4.72 (d, *J* = 15.0 Hz, 1H), 3.73–3.57 (m, 4H), 1.85–1.65 (m, 4H), 1.53 (d, *J* = 13.8 Hz, 1H), 1.43–1.29 (m, 1H) ppm. ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -115.23 (s, 1F) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 161.52, 155.97 (d, *J*_{C-F} = 249.2 Hz, 1C), 133.16, 129.89 (d, *J*_{C-F} = 10.5 Hz, 1C), 115.18 (d, *J*_{C-F} = 5.0 Hz, 1C), 105.34, 81.23, 68.48, 52.96, 49.14, 45.77, 28.23, 26.69, 14.33 ppm. HRMS (ESI) calcd for C₁₆H₁₉N₃O₃F [M + H]⁺, 320.1410, found, 320.1415.

1-(2-Chlorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (6). Yield, 86%; mp, 130.4–131.7 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.50–7.44 (m, 2H), 7.38–7.33 (m, 2H), 5.15 (s, 1H), 5.06 (d, *J* = 2.2 Hz, 1H), 4.94 (d, *J* = 16.4 Hz, 1H), 4.85 (d, *J* = 16.4 Hz, 1H), 3.82–3.61 (m, 4H), 1.88–1.66 (m, 4H), 1.64–1.40 (m, 2H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 156.46, 134.61, 131.68, 129.17, 129.00, 128.79, 127.26, 105.03, 81.30, 68.46, 52.55, 49.48, 45.91, 28.35, 26.76, 14.37 ppm. HRMS (ESI) calcd for C₁₆H₁₉N₃O₃³⁵Cl [M + H]⁺, 336.1115, found, 336.1113; calcd for C₁₆H₁₉N₃O₃³⁷Cl [M + H]⁺, 338.1085, found, 338.1089.

1-(3-Chlorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (7). Yield, 91%; mp, 132.7–133.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.39–7.34 (m, 4H), 5.13 (s, 1H), 5.07 (s, 1H), 5.00 (d, *J* = 15.4 Hz, 1H), 4.71 (d, *J* = 15.4 Hz, 1H), 3.76–3.58 (m, 4H), 1.86–1.66 (m, 4H), 1.54 (d, *J* = 13.8 Hz, 1H), 1.48–1.32 (m, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 156.11, 139.69, 133.08, 130.24, 127.57, 127.30, 126.38, 105.30, 81.21, 68.44, 53.42, 49.37, 45.76, 28.22, 26.72, 14.31 ppm. HRMS (ESI) calcd for C₁₆H₁₈N₃O₃³⁵ClNa [M + Na]⁺, 358.0934, found, 358.0933; calcd for C₁₆H₁₈N₃O₃³⁷ClNa [M + Na]⁺, 360.0905, found, 360.0903.

1-(4-Chlorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (8). Yield, 83%; mp, 139.0–141.4 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.42–7.36 (m, 2H), 7.23–7.16 (m, 2H), 5.11 (s, 1H), 5.06 (d, *J* = 2.4 Hz, 1H), 4.96 (d, *J* = 15.0 Hz, 1H), 4.72 (d, *J* = 15.0 Hz, 1H),

3.73–3.57 (m, 4H), 1.85–1.65 (m, 4H), 1.53 (d, $J = 13.8$ Hz, 1H), 1.43–1.29 (m, 1H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.03, 136.11, 131.94, 129.65, 128.36, 105.31, 81.24, 68.47, 53.17, 49.25, 45.79, 28.27, 26.67, 14.35 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{19}\text{N}_3\text{O}_3^{35}\text{Cl}$ $[\text{M} + \text{H}]^+$, 336.1115, found, 336.1114; calcd for $\text{C}_{16}\text{H}_{19}\text{N}_3\text{O}_3^{37}\text{Cl}$ $[\text{M} + \text{H}]^+$, 338.1085, found, 338.1087.

1-(2-Bromobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (9). Yield, 84%; mp, 142.3–143.1 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.68–7.60 (m, 1H), 7.49–7.39 (m, 2H), 7.30–7.21 (m, 1H), 5.15 (s, 1H), 5.07 (d, $J = 2.4$ Hz, 1H), 4.89 (d, $J = 16.6$ Hz, 1H), 4.79 (d, $J = 16.6$ Hz, 1H), 3.80–3.60 (m, 4H), 1.87–1.67 (m, 4H), 1.61–1.47 (m, 2H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.53, 136.16, 132.39, 129.01, 128.97, 127.79, 121.86, 104.99, 81.31, 68.46, 55.16, 49.50, 45.93, 28.40, 26.74, 14.41 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{19}\text{N}_3\text{O}_3^{79}\text{Br}$ $[\text{M} + \text{H}]^+$, 380.0610, found, 380.0609; calcd for $\text{C}_{16}\text{H}_{19}\text{N}_3\text{O}_3^{81}\text{Br}$ $[\text{M} + \text{H}]^+$, 382.0589, found, 382.0588.

1-(3-Bromobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (10). Yield, 72%; mp, 140.9–142.1 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.56 (s, 1H), 7.52–7.49 (m, 1H), 7.38–7.30 (m, 2H), 5.13 (s, 1H), 5.06 (d, $J = 2.2$ Hz, 1H), 5.00 (d, $J = 15.4$ Hz, 1H), 4.69 (d, $J = 15.4$ Hz, 1H), 3.74–3.59 (m, 4H), 1.87–1.66 (m, 4H), 1.54 (d, $J = 13.8$ Hz, 1H), 1.47–1.30 (m, 1H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.11, 139.94, 130.54, 130.46, 130.20, 126.78, 121.72, 105.30, 81.20, 68.44, 53.37, 49.35, 45.76, 28.21, 26.74, 14.31 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{19}\text{N}_3\text{O}_3^{79}\text{Br}$ $[\text{M} + \text{H}]^+$, 380.0610, found, 380.0609; calcd for $\text{C}_{16}\text{H}_{19}\text{N}_3\text{O}_3^{81}\text{Br}$ $[\text{M} + \text{H}]^+$, 382.0589, found, 382.0588.

1-(4-Bromobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (11). Yield, 77%; mp, 149.1–150.0 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.63–7.47 (m, 2H), 7.31 (d, $J = 8.4$ Hz, 2H), 5.11 (s, 1H), 5.06 (d, $J = 2.6$ Hz, 1H), 4.94 (d, $J = 15.2$ Hz, 1H), 4.71 (d, $J = 15.2$ Hz, 1H), 3.78–3.58 (m, 4H), 1.87–1.64 (m, 4H), 1.54 (d, $J = 13.8$ Hz, 1H), 1.47–1.28 (m, 1H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.04, 136.55, 131.27, 129.99, 120.46, 105.30, 81.25, 68.47, 53.25, 49.26, 45.79, 28.28, 26.67, 14.36 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_3^{79}\text{BrNa}$ $[\text{M} + \text{Na}]^+$, 402.0429, found, 402.0430; calcd for $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_3^{81}\text{BrNa}$ $[\text{M} + \text{Na}]^+$, 404.0409, found, 404.0408.

1-(2-Methylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (12). Yield, 80%; mp, 141.3–142.1 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.32–7.15 (m, 4H), 5.14 (s, 1H), 5.08 (d, $J = 2.6$ Hz, 1H), 4.90 (d, $J = 16.0$ Hz, 1H), 4.82 (d, $J = 16.0$ Hz, 1H), 3.77–3.52 (m, 4H), 2.25 (s, 3H), 1.91–1.67 (m, 4H), 1.53–1.48 (m, 2H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.38, 135.39, 134.99, 130.08, 127.04, 126.97, 125.92, 105.03, 81.40, 68.62, 51.95, 48.98, 45.85, 28.43, 26.67, 18.55, 14.48 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_3\text{O}_3\text{Na}$ $[\text{M} + \text{Na}]^+$, 338.1481, found, 338.1482.

1-(3-Methylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (13). Yield, 75%; mp, 111.2–111.9 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.24 (t, $J = 7.4$ Hz, 1H), 7.19–7.06 (m, 3H), 5.09 (d, $J = 15.2$ Hz, 2H), 4.97 (d, $J = 15.0$ Hz, 1H), 4.70 (d, $J = 15.0$ Hz, 1H), 3.74–3.53 (m, 4H), 2.30 (s, 3H), 1.87–1.63 (m, 4H), 1.53 (d, $J = 13.8$ Hz, 1H), 1.46–1.29 (m, 1H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.04, 137.55, 136.91, 128.43, 128.35, 128.04, 124.86, 105.34, 81.23, 68.52, 53.55, 49.04, 45.76, 28.17, 26.78, 20.99,

14.28 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_3\text{O}_3\text{Na}$ $[\text{M} + \text{Na}]^+$, 338.1481, found, 338.1482.

1-(4-Methylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (14). Yield, 89%; mp, 122.9–124.1 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.21–7.17 (m, 4H), 5.37–5.29 (m, 1H), 4.96 (d, $J = 14.8$ Hz, 1H), 4.83 (d, $J = 14.8$ Hz, 1H), 3.75–3.64 (m, 1H), 3.63–3.45 (m, 4H), 2.34 (s, 3H), 2.16 (d, $J = 13.6$ Hz, 1H), 1.94–1.83 (m, 2H), 1.68–1.65 (m, 3H) ppm. ^{13}C NMR (101 MHz, CDCl_3) δ 156.53, 137.77, 133.27, 129.53, 128.34, 106.72, 83.02, 69.74, 54.48, 48.37, 46.72, 29.56, 26.48, 21.15, 15.01 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_3\text{O}_3\text{Na}$ $[\text{M} + \text{Na}]^+$, 338.1481, found, 338.1483.

1-(2-Methoxybenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (15). Yield, 77%; mp, 127.5–128.7 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.32–7.24 (m, 1H), 7.19 (d, $J = 7.4$ Hz, 1H), 7.00 (d, $J = 8.0$ Hz, 1H), 6.91 (t, $J = 7.4$ Hz, 1H), 5.08 (s, 1H), 5.02 (s, 1H), 4.81 (s, 2H), 3.78 (d, $J = 8.4$ Hz, 3H), 3.76–3.61 (m, 2H), 3.59–3.52 (m, 2H), 1.79–1.60 (m, 4H), 1.49 (d, $J = 13.8$ Hz, 1H), 1.40–1.22 (m, 1H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.93, 156.15, 128.90, 128.83, 124.44, 120.21, 110.61, 105.33, 81.35, 68.53, 55.18, 49.51, 49.18, 45.96, 28.07, 26.90, 14.12 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_3\text{O}_4\text{Na}$ $[\text{M} + \text{Na}]^+$, 354.1430, found, 354.1431.

1-(3-Methoxybenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (16). Yield, 81%; mp, 135.3–135.9 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.31–7.25 (m, 1H), 6.92–6.84 (m, 3H), 5.12 (s, 1H), 5.07 (d, $J = 2.2$ Hz, 1H), 4.93 (d, $J = 15.0$ Hz, 1H), 4.77 (d, $J = 15.0$ Hz, 1H), 3.74 (s, 3H), 3.73–3.58 (m, 4H), 1.84–1.65 (m, 4H), 1.53 (d, $J = 14.0$ Hz, 1H), 1.42–1.32 (m, 1H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 159.34, 156.02, 138.59, 129.55, 119.89, 113.31, 112.86, 105.42, 81.26, 68.52, 54.94, 53.45, 49.12, 45.83, 28.21, 26.73, 14.32 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_3\text{O}_4\text{Na}$ $[\text{M} + \text{Na}]^+$, 354.1430, found, 354.1431.

1-(4-Methoxybenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (17). Yield, 86%; mp, 130.7–132.5 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.29–7.24 (m, 2H), 6.94–6.90 (m, 2H), 5.15–5.03 (m, 2H), 4.90 (d, $J = 14.6$ Hz, 1H), 4.71 (d, $J = 14.6$ Hz, 1H), 3.78–3.71 (m, 3H), 3.71–3.54 (m, 4H), 1.85–1.44 (m, 7H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 158.68, 155.87, 129.31, 128.73, 113.84, 105.38, 81.26, 68.55, 55.02, 52.89, 48.83, 45.76, 28.21, 26.71, 14.32 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_3\text{O}_4\text{Na}$ $[\text{M} + \text{Na}]^+$, 354.1430, found, 354.1431.

2-((10-Nitro-2,3,6,7,8,9-hexahydro-5,9-epoxyimidazo[1,2-*a*]azocin-1(5*H*)-yl)methyl)benzonitrile (18). Yield, 83%; mp, 135.2–136.4 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.85 (dd, $J = 8.0, 1.0$ Hz, 1H), 7.78–7.73 (m, 1H), 7.60 (d, $J = 8.0$ Hz, 1H), 7.49 (t, $J = 7.6$ Hz, 1H), 5.21–5.03 (m, 3H), 4.96 (d, $J = 16.6$ Hz, 1H), 3.84–3.67 (m, 4H), 1.85–1.69 (m, 4H), 1.55–1.50 (m, 2H) ppm. ^{13}C NMR (100 MHz, DMSO- d_6) δ 156.26, 141.26, 133.27, 132.98, 127.93, 127.84, 117.25, 109.84, 105.15, 81.27, 68.39, 53.16, 49.94, 45.85, 28.36, 26.74, 14.32 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{19}\text{N}_4\text{O}_3$ $[\text{M} + \text{H}]^+$, 327.1457, found, 327.1458.

3-((10-Nitro-2,3,6,7,8,9-hexahydro-5,9-epoxyimidazo[1,2-*a*]azocin-1(5*H*)-yl)methyl)benzonitrile (19). Yield, 89%; mp, 144.8–145.7 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 7.85–7.74 (m, 2H), 7.69 (d, $J = 7.8$ Hz, 1H), 7.58 (t, $J = 7.8$ Hz, 1H), 5.13 (s, 1H), 5.09–4.99 (m, 2H), 4.73 (d, $J = 15.6$ Hz, 1H), 3.79–3.61 (m, 4H), 1.84–1.66 (m, 4H), 1.55 (d, $J = 13.8$

Hz, 1H), 1.46–1.39 (m, 1H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.19, 138.97, 132.50, 131.17, 131.06, 129.55, 118.77, 111.29, 105.23, 81.22, 68.41, 53.55, 49.56, 45.79, 28.28, 26.70, 14.33 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{18}\text{N}_4\text{O}_3\text{Na}$ $[\text{M} + \text{Na}]^+$, 349.1277, found, 349.1276.

4-((10-Nitro-2,3,6,7,8,9-hexahydro-5,9-epoxyimidazo[1,2-a]azocin-1(5H)-yl)methyl)benzonitrile (**20**). Yield, 72%; mp, 140.9–142.2 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.88–7.81 (m, 2H), 7.54 (d, J = 8.4 Hz, 2H), 5.14 (s, 1H), 5.06–5.03 (m, 2H), 4.81 (d, J = 16.0 Hz, 1H), 3.79–3.60 (m, 4H), 1.84–1.66 (m, 4H), 1.57–1.52 (m, 1H), 1.48–1.37 (m, 1H) ppm. ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ 156.21, 143.24, 132.26, 128.39, 118.80, 109.94, 105.23, 81.25, 68.41, 53.97, 49.62, 45.84, 28.33, 26.65, 14.37 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{19}\text{N}_4\text{O}_3$ $[\text{M} + \text{H}]^+$, 327.1457, found, 327.1456.

10-Nitro-1-(4-nitrobenzyl)-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**21**). Yield, 70%; mp, 146.1–146.8 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 8.24 (d, J = 8.8 Hz, 2H), 7.63 (d, J = 8.8 Hz, 2H), 5.17–5.05 (m, 4H), 4.85 (d, J = 16.2 Hz, 1H), 3.79–3.62 (m, 4H), 1.79–1.72 (m, 3H), 1.55 (s, 1H), 1.51–1.41 (m, 2H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.30, 146.69, 145.48, 128.58, 123.43, 105.23, 81.25, 68.41, 53.94, 49.65, 45.87, 28.36, 26.65, 14.39 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{18}\text{N}_4\text{O}_3\text{Na}$ $[\text{M} + \text{Na}]^+$, 369.1175, found, 369.1176.

10-Nitro-1-(4-(trifluoromethyl)benzyl)-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**22**). Yield, 81%; mp, 128.8–129.5 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.78–7.53 (m, 4H), 5.18–4.96 (m, 3H), 4.82 (d, J = 15.8 Hz, 1H), 3.79–3.61 (m, 4H), 1.85–1.65 (m, 4H), 1.60–1.35 (m, 2H) ppm. ^{19}F NMR (376 MHz, $\text{DMSO}-d_6$) δ -60.78 (s, 3F) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 157.01, 143.25 (d, J = 249.5 Hz, 1C), 132.20, 128.34 (t, J = 7.4 Hz, 1C), 118.77, 109.96 (d, J = 3.8 Hz, 1C), 105.13, 81.22, 68.45, 53.94, 49.63, 45.85, 28.31, 26.66, 14.29 ppm. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{19}\text{N}_3\text{O}_3\text{F}_3$ $[\text{M} + \text{H}]^+$, 370.1379, found, 370.1376.

10-Nitro-1-((6-(trifluoromethyl)pyridin-3-yl)methyl)-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**23**). Yield, 73%; mp, 147.1–148.4 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 8.75 (d, J = 1.2 Hz, 1H), 8.06 (dd, J = 8.0, 1.2 Hz, 1H), 7.92 (d, J = 8.0 Hz, 1H), 5.15 (s, 1H), 5.08 (d, J = 16.2 Hz, 2H), 4.83 (d, J = 16.2 Hz, 1H), 3.82–3.65 (m, 4H), 1.85–1.68 (m, 4H), 1.55 (d, J = 13.8 Hz, 1H), 1.49–1.35 (m, 1H) ppm. ^{19}F NMR (376 MHz, $\text{DMSO}-d_6$) δ -66.36 (s, 3F) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 162.91, 156.08, 135.93, 131.36, 130.52, 129.09 (q, J = 22.8 Hz, 1C), 124.83, 105.01, 81.59, 68.79, 48.92, 45.94, 45.31, 28.23, 26.88, 14.35 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{18}\text{N}_4\text{O}_3\text{F}_3$ $[\text{M} + \text{H}]^+$, 371.1331, found, 371.1332.

1-([1,1'-Biphenyl]-4-ylmethyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**24**). Yield, 84%; mp, 129.6–130.8 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.79–7.56 (m, 4H), 7.55–7.33 (m, 5H), 5.19–4.96 (m, 3H), 4.83 (d, J = 15.2 Hz, 1H), 3.79–3.58 (m, 4H), 1.89–1.65 (m, 4H), 1.63–1.34 (m, 2H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.06, 139.76, 139.25, 136.27, 128.92, 128.38, 127.41, 126.72, 126.59, 105.35, 81.27, 68.53, 53.39, 49.22, 45.82, 28.28, 26.70, 14.37 ppm. HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{24}\text{N}_3\text{O}_3$ $[\text{M} + \text{H}]^+$, 378.1818, found, 378.1816.

1-(4-(tert-Butyl)benzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**25**). Yield, 85%; mp, 137.9–138.5 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.44–7.33 (m, 2H), 7.25–7.21 (m, 2H), 5.13–5.04 (m, 2H), 4.92

(d, J = 15.0 Hz, 1H), 4.77 (d, J = 15.0 Hz, 1H), 3.74–3.55 (m, 4H), 1.83–1.63 (m, 4H), 1.51–1.48 (m, 2H), 1.28 (s, 9H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.03, 149.76, 133.97, 127.44, 125.20, 99.49, 81.24, 68.52, 53.20, 49.14, 45.79, 34.20, 31.11, 28.22, 26.71, 14.31 ppm. HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{28}\text{N}_3\text{O}_3$ $[\text{M} + \text{H}]^+$, 358.2131, found, 358.2132.

1-(4-Isopropylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**26**). Yield, 92%; mp, 140.2–141.3 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.34–7.14 (m, 4H), 5.09–5.06 (m, 2H), 4.92 (d, J = 15.0 Hz, 1H), 4.75 (d, J = 15.0 Hz, 1H), 3.76–3.54 (m, 4H), 2.94–2.80 (m, 1H), 1.83–1.44 (m, 6H), 1.22–1.15 (m, 6H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.01, 147.53, 134.33, 127.75, 126.35, 105.33, 81.24, 68.51, 53.29, 49.11, 45.78, 33.08, 28.21, 26.71, 23.85, 14.31 ppm. HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{26}\text{N}_3\text{O}_3$ $[\text{M} + \text{H}]^+$, 344.1974, found, 344.1976.

1-(2,6-Dichlorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**27**). Yield, 75%; mp, 134.4–134.9 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.71–7.56 (m, 2H), 7.53–7.42 (m, 1H), 5.17–5.01 (m, 3H), 4.95 (d, J = 14.6 Hz, 1H), 3.57–3.33 (m, 4H), 1.90–1.46 (m, 6H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.01, 135.95, 131.37, 130.51, 129.09, 105.04, 81.55, 68.80, 48.95, 45.95, 45.35, 28.22, 26.90, 14.30 ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_3^{35}\text{Cl}_2$ $[\text{M} + \text{H}]^+$, 370.0725, found, 370.0724; calcd for $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_3^{35}\text{Cl}^{37}\text{Cl}$ $[\text{M} + \text{H}]^+$, 372.0696, found, 372.0699; calcd for $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_3^{37}\text{Cl}_2$ $[\text{M} + \text{H}]^+$, 374.0666, found, 374.0671.

1-(2,6-Difluorobenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**28**). Yield, 72%; mp, 143.3–144.8 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.51–7.41 (m, 1H), 7.21–7.11 (m, 2H), 5.15–5.05 (m, 3H), 5.05–4.99 (m, 1H), 3.73–3.59 (m, 2H), 3.58–3.45 (m, 2H), 1.83–1.66 (m, 4H), 1.58–1.48 (m, 1H), 1.42–1.27 (m, 1H) ppm. ^{19}F NMR (376 MHz, $\text{DMSO}-d_6$) δ -114.24 (s, 1F), -114.27 (s, 1F) ppm. HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_3\text{F}_2$ $[\text{M} + \text{H}]^+$, 338.1316, found, 338.1315.

1-(2,6-Dimethylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**29**). Yield, 84%; mp, 130.7–131.7 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.18–7.12 (m, 3H), 5.13 (d, J = 2.8 Hz, 1H), 5.07 (s, 1H), 4.84 (d, J = 14.4 Hz, 1H), 4.68 (d, J = 14.4 Hz, 1H), 3.59–3.47 (m, 1H), 3.47–3.28 (m, 2H), 3.25–3.13 (m, 1H), 2.31 (s, 6H), 1.91 (d, J = 13.2 Hz, 1H), 1.81–1.67 (m, 3H), 1.63–1.40 (m, 3H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 155.96, 137.80, 131.66, 128.48, 128.15, 104.97, 81.82, 68.98, 47.67, 46.07, 45.37, 28.54, 26.53, 19.49, 14.54 ppm. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{23}\text{N}_3\text{O}_3\text{Na}$ $[\text{M} + \text{Na}]^+$, 352.1637, found, 352.1636.

1-(2,5-Dimethylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**30**). Yield, 80%; mp, 129.4–130.6 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.07 (d, J = 8.6 Hz, 2H), 7.00 (d, J = 7.6 Hz, 1H), 5.13 (s, 1H), 5.08 (d, J = 2.4 Hz, 1H), 4.90 (d, J = 15.8 Hz, 1H), 4.72 (d, J = 15.8 Hz, 1H), 3.73–3.51 (m, 4H), 2.26 (s, 3H), 2.19 (s, 3H), 1.89–1.66 (m, 4H), 1.55–1.49 (m, 2H) ppm. ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δ 156.40, 134.76, 134.67, 132.30, 130.05, 127.92, 127.62, 105.03, 81.37, 68.62, 51.91, 48.81, 45.81, 28.34, 26.79, 20.72, 18.13, 14.43 ppm. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{24}\text{N}_3\text{O}_3$ $[\text{M} + \text{H}]^+$, 330.1818, found, 330.1819.

1-(3,4-Dimethylbenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-a]azocine (**31**). Yield, 88%; mp, 137.1–137.8 °C; ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ 7.15–6.99 (m, 3H), 5.16–5.04 (m, 2H), 4.92 (d, J = 14.8 Hz, 1H),

Table 2. Mortality of a Mixture of Imidacloprid (1 mg/L) with the New Compounds 1–33 at Low Concentrations (0.5–4 mg/L) against *A. craccivora*^a

| compound | R | mortality (co-applied with imidacloprid) (%) | | | |
|----------|---|--|------------|------------|------------|
| | | 0.5 mg/L | 1 mg/L | 2 mg/L | 4 mg/L |
| 1 | H | nt | 35.7 ± 3.9 | 66.6 ± 2.4 | 71.8 ± 3.7 |
| 2 | C ₆ H ₅ | nt | 41.2 ± 2.3 | 72.1 ± 2.8 | 79.8 ± 4.2 |
| 3 | 2-F-C ₆ H ₅ | nt | 39.2 ± 2.0 | 69.5 ± 1.7 | 76.4 ± 1.7 |
| 4 | 3-F-C ₆ H ₅ | nt | 44.5 ± 3.1 | 76.5 ± 3.2 | 77.9 ± 1.7 |
| 5 | 4-F-C ₆ H ₅ | nt | 53.9 ± 1.2 | 81.6 ± 1.9 | 85.3 ± 1.6 |
| 6 | 2-Cl-C ₆ H ₅ | nt | 34.0 ± 5.5 | 69.8 ± 1.3 | 74.6 ± 1.8 |
| 7 | 3-Cl-C ₆ H ₅ | nt | 38.9 ± 2.4 | 74.3 ± 2.1 | 80.1 ± 1.3 |
| 8 | 4-Cl-C ₆ H ₅ | nt | 55.3 ± 1.8 | 72.5 ± 1.8 | 84.4 ± 2.1 |
| 9 | 2-Br-C ₆ H ₅ | nt | 36.7 ± 2.3 | 74.8 ± 2.0 | 80.9 ± 1.7 |
| 10 | 3-Br-C ₆ H ₅ | nt | 44.4 ± 1.9 | 76.9 ± 2.5 | 81.7 ± 1.8 |
| 11 | 4-Br-C ₆ H ₅ | nt | 45.3 ± 1.5 | 73.6 ± 2.2 | 75.4 ± 3.9 |
| 12 | 2-Me-C ₆ H ₅ | nt | 39.0 ± 1.9 | 75.6 ± 1.7 | 76.4 ± 2.4 |
| 13 | 3-Me-C ₆ H ₅ | nt | 54.1 ± 2.7 | 75.4 ± 1.2 | 82.4 ± 2.3 |
| 14 | 4-Me-C ₆ H ₅ | 59.8 ± 2.4 | 82.9 ± 2.5 | 88.5 ± 3.1 | 91.4 ± 1.9 |
| 15 | 2-OMe-C ₆ H ₅ | nt | 39.2 ± 1.5 | 80.7 ± 3.2 | 81.3 ± 1.5 |
| 16 | 3-OMe-C ₆ H ₅ | nt | 55.0 ± 2.0 | 87.6 ± 2.8 | 88.4 ± 2.3 |
| 17 | 4-OMe-C ₆ H ₅ | 63.0 ± 1.1 | 84.8 ± 2.5 | 90.4 ± 1.8 | 93.4 ± 1.5 |
| 18 | 2-CN-C ₆ H ₅ | nt | 39.0 ± 1.5 | 76.1 ± 1.6 | 81.2 ± 2.5 |
| 19 | 3-CN-C ₆ H ₅ | nt | 38.2 ± 1.2 | 82.9 ± 2.1 | 85.7 ± 3.0 |
| 20 | 4-CN-C ₆ H ₅ | nt | 40.4 ± 2.6 | 85.6 ± 2.9 | 87.3 ± 1.0 |
| 21 | 4-NO ₂ -C ₆ H ₅ | nt | 55.7 ± 1.2 | 79.7 ± 1.5 | 82.5 ± 2.2 |
| 22 | 4-CF ₃ -C ₆ H ₅ | nt | 58.5 ± 4.0 | 81.7 ± 2.7 | 83.0 ± 1.9 |
| 23 | 6-CF ₃ -pyridyl | nt | 49.6 ± 2.4 | 86.0 ± 1.8 | 88.7 ± 1.8 |
| 24 | 4-Ph-C ₆ H ₅ | 34.6 ± 1.2 | 71.9 ± 1.9 | 88.2 ± 1.3 | 92.8 ± 1.3 |
| 25 | 4- <i>t</i> -Bu-C ₆ H ₅ | 77.0 ± 2.0 | 94.2 ± 0.7 | 97.0 ± 2.2 | 98.1 ± 2.7 |
| 26 | 4- <i>i</i> -Pr-C ₆ H ₅ | 56.5 ± 2.0 | 82.0 ± 4.4 | 90.1 ± 1.9 | 92.5 ± 1.6 |
| 27 | 2,6-(Cl) ₂ -C ₆ H ₃ | nt | 51.6 ± 1.1 | 69.8 ± 2.4 | 77.7 ± 6.0 |
| 28 | 2,6-(F) ₂ -C ₆ H ₃ | nt | 54.7 ± 1.5 | 76.0 ± 1.8 | 80.9 ± 2.0 |
| 29 | 2,6-(Me) ₂ -C ₆ H ₃ | nt | 60.8 ± 2.8 | 81.7 ± 1.1 | 87.6 ± 3.9 |
| 30 | 2,5-(Me) ₂ -C ₆ H ₃ | nt | 64.6 ± 1.7 | 81.9 ± 3.8 | 86.2 ± 1.1 |
| 31 | 3,4-(Me) ₂ -C ₆ H ₃ | nt | 64.7 ± 1.1 | 82.0 ± 1.4 | 87.3 ± 1.5 |
| 32 | 3,4-(OMe) ₂ -C ₆ H ₃ | nt | 60.0 ± 1.9 | 81.4 ± 1.6 | 86.3 ± 1.8 |
| 33 | 2-chlorothiazol-5-yl | nt | 48.8 ± 2.4 | 74.5 ± 2.9 | 84.4 ± 1.6 |
| IPPA08 | | 70.6 ± 2.1 | 86.6 ± 3.1 | 90.1 ± 2.5 | 93.0 ± 0.9 |

^aEffects of compounds 1–33 and IPPA08 on imidacloprid (1 mg/L) toxicity against *A. craccivora*. Data are mean of at least six independent experiments ± standard error of the mean (SEM). Imidacloprid alone at a concentration of 1 mg/L gives 29% mortality. nt = not test.

4.66 (d, *J* = 14.8 Hz, 1H), 3.69–3.52 (m, 4H), 2.20 (s, 6H), 1.87–1.62 (m, 4H), 1.53 (d, *J* = 13.8 Hz, 1H), 1.45–1.28 (m, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 155.97, 136.17, 135.31, 134.24, 129.53, 129.09, 125.33, 105.35, 81.23, 68.54, 53.22, 48.84, 45.74, 28.17, 26.77, 19.39, 19.01, 14.29 ppm. HRMS (ESI) calcd for C₁₈H₂₃N₃O₃Na [M + Na]⁺, 352.1637, found, 352.1638.

1-(3,4-Dimethoxybenzyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (32). Yield, 70%; mp, 121.9–123.0 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 6.91–6.87 (m, 3H), 5.14–5.04 (m, 2H), 4.86 (d, *J* = 14.8 Hz, 1H), 4.71 (d, *J* = 14.8 Hz, 1H), 3.73 (d, *J* = 6.4 Hz, 6H), 3.70–3.53 (m, 4H), 1.84–1.64 (m, 4H), 1.51 (d, *J* = 13.8 Hz, 1H), 1.38–1.27 (m, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆) δ 155.95, 148.66, 148.24, 129.15, 120.33, 111.67, 105.54, 81.29, 68.55, 55.46, 55.30, 53.11, 48.85, 45.87, 28.17, 26.77, 14.31 ppm. HRMS calcd for C₁₈H₂₄N₃O₅ [M + H]⁺, 362.1716, found, 362.1717.

1-((2-Chlorothiazol-5-yl)methyl)-10-nitro-1,2,3,5,6,7,8,9-octahydro-5,9-epoxyimidazo[1,2-*a*]azocine (33). Yield, 84%; mp, 134.9–135.7 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.48 (s,

1H), 5.31 (s, 1H), 5.24 (d, *J* = 15.4 Hz, 1H), 4.97 (s, 1H), 4.78 (d, *J* = 15.4 Hz, 1H), 3.85–3.74 (m, 1H), 3.73–3.58 (m, 3H), 2.12 (d, *J* = 13.4 Hz, 1H), 1.99–1.78 (m, 2H), 1.77–1.51 (m, 4H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ 155.68, 154.16, 140.49, 135.56, 106.97, 82.71, 69.36, 49.40, 48.31, 46.17, 29.43, 26.47, 14.93 ppm. HRMS (ESI) calcd for C₁₃H₁₅N₄O₃S³⁵ClNa [M + Na]⁺, 365.0451, found, 365.0452; calcd for C₁₃H₁₅N₄O₃S³⁷ClNa [M + Na]⁺, 367.0422, found, 367.0422.

BIOLOGICAL RESULTS

Synergistic Effect on Imidacloprid. The mortality caused by the oxabridged compounds and their effects on imidacloprid toxicity against *A. craccivora* were assessed. None of the oxabridged homologues showed any mortality at a low concentration (10 mg/L). When the oxabridged compounds were applied at concentrations of 4 and 2 mg/L together with 1 mg/L imidacloprid, the toxicity against *A. craccivora* strongly increased (Table 2). Moreover, even at a low concentration of 0.5 mg/L, four compounds (14, 17, 25, and 26) possessing an electron-donating group (EDG) at position 4 of the phenyl

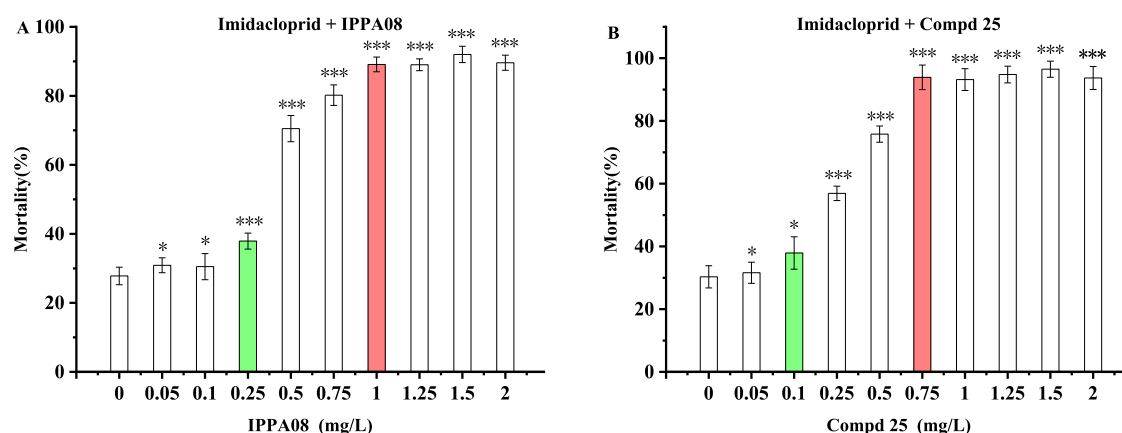
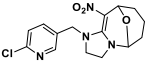
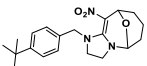


Figure 2. (A) Effects of IPPA08 on imidacloprid (1 mg/L) toxicity against *A. craccivora*. (B) Effects of compound 25 on imidacloprid (1 mg/L) toxicity against *A. craccivora*. Data are means of at least six independent experiments \pm SEM. Significant differences from blank control: * p < 0.05, ** p < 0.01, *** p < 0.001.

Table 3. Synergistic Effects of IPPA08 and Compound 25 on Imidacloprid Toxicity Against *A. craccivora*^a

| compound | concentration ^b | Imidacloprid LC ₅₀ | 95% CI ^c | slope | SR ^d |
|--|----------------------------|-------------------------------|---------------------|-------|-----------------|
|  | 0 | 1.72 | 1.19-2.47 | 2.20 | 1 |
| | 0.25 | 0.606*** | 0.446-0.828 | 1.91 | 2.84 |
| | 0.75 | 0.492*** | 0.360-0.665 | 1.57 | 3.50 |
| | 1 | 0.382*** | 0.328-0.443 | 1.77 | 4.50 |
|  | 0 | 1.72 | 1.19-2.47 | 2.20 | 1 |
| | 0.1 | 0.687*** | 0.536-0.885 | 2.02 | 2.50 |
| | 0.25 | 0.490*** | 0.369-0.646 | 1.66 | 3.51 |
| | 0.75 | 0.263*** | 0.179-0.363 | 1.71 | 6.54 |

^aData are the means of at least six independent experiments. ^bIPPA08 and compound 25 at concentrations in mg/L showed no insecticidal activities against *A. craccivora*. ^cCI = confidence interval (CI can be used to estimate population parameters; the narrower the scope of CI, the better the reliability of population parameters estimated by sample indicators). ^dSR = synergism ratio. Significant differences from blank control: * p < 0.05, ** p < 0.01, *** p < 0.001.

ring increased the mortality of *A. craccivora* by imidacloprid and exhibited obvious synergy. In particular, compound 25 showed higher synergistic activity than IPPA08. In addition, compared to compound 24, compound 25 was more effective, which indicated that electronic factor could be the major factor instead of steric factor.

Further study showed that the mortality of *A. craccivora* by imidacloprid (29% at 1 mg/L) could be increased to 90% with 1.0 mg/L IPPA08, at which IPPA08 alone showed no insecticidal activity (Figure 2A). Similarly, 30% mortality of *A. craccivora* by 1 mg/L imidacloprid was increased to 97% when combined with compound 25 at 0.75 mg/L, a concentration at which compound 25 itself caused no mortality (Figure 2B). Compared to IPPA08, compound 25 presented higher effects to synergize imidacloprid at the same co-applied concentrations (0.25, 0.75 mg/L). For a co-applied concentration at 0.25 mg/L, IPPA08 and compound 25 increased the toxicity of imidacloprid against *A. craccivora* by 2.84- and 3.51-fold, respectively. For a co-applied concentration at 0.75 mg/L, IPPA08 and compound 25 increased the toxicity of imidacloprid against *A. craccivora* by 3.50- and 6.54-fold, respectively. Even though IPPA08 and compound 25 showed optimal co-applied concentrations of 1 and 0.75 mg/L, respectively, compound 25 (6.54-fold) showed a higher synergism ratio than IPPA08 (4.50-fold) (Table 3).

Influence of IPPA08 and Compound 25 on the Toxicities of Other Neonicotinoids. The effects of IPPA08 and compound 25 on the toxicities of other neonicotinoids were also tested, including nitenpyram, dinotefuran, acetamiprid, thiamethoxam, paichongding, and cycloxyaprid. For *A. craccivora*, IPPA08 and compound 25 at low concentrations (causing no toxicity by itself) increased the mortalities caused by fixed concentrations (1 mg/L) of nitenpyram, acetamiprid, and thiamethoxam, and compound 25 showed higher effectiveness as a synergist (Figure 3A,C,D). However, IPPA08 and compound 25 showed little impact on the toxicities of dinotefuran and paichongding against *A. craccivora* (Figure 3B,E). Interestingly, compound 25 showed weak effects on cycloxyaprid toxicity against *A. craccivora* at 4 mg/L (Figure 3F). IPPA08 and compound 25 showed good stability under these conditions, excluding the possibility that the synergistic effects were caused by metabolites (Table 4).

Safety of Compound 25 to Honey Bee. Bee mortalities by the independent use of imidacloprid or by the combined use of imidacloprid and compound 25 (1:1) were tested (Table 5). The mixture of imidacloprid and compound 25 did not exhibit a higher toxicity toward the bee than the imidacloprid alone. However, for *A. craccivora*, the mortality by imidacloprid (30% at 1 mg/L) could be increased to 90% with 1 mg/L compound 25 (Figure 2B). These results

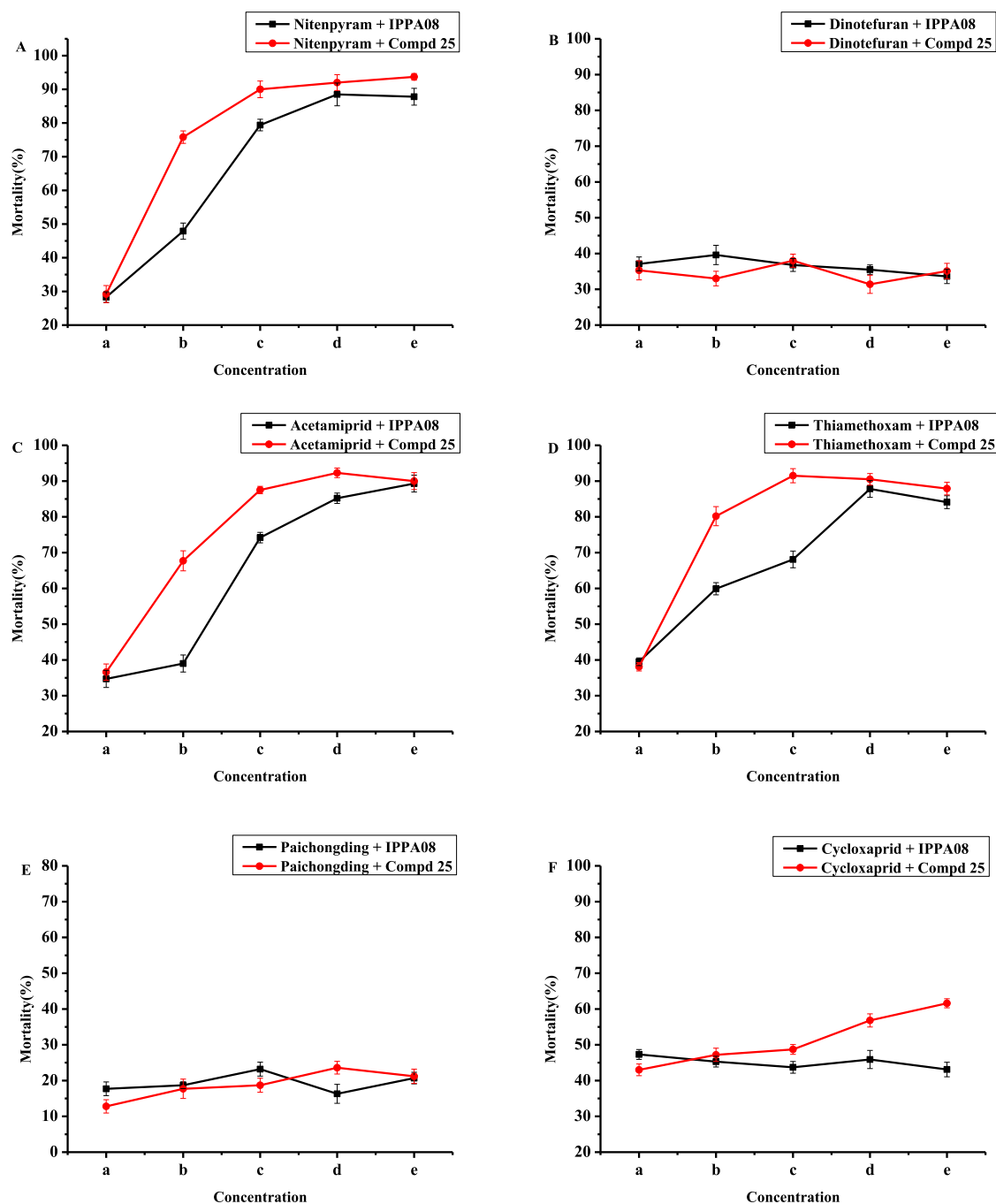


Figure 3. Effects of IPPA08 and compound 25 on the toxicity of neonicotinoid insecticides, including nitenpyram, dinotefuran, acetamiprid, thiamethoxam, paichongding, and cycloxaprid. The concentration for each neonicotinoid insecticide was 1 mg/L, and the concentrations of IPPA08 and compound 25 were (a–e) 0, 0.5, 1, 2, and 4 mg/L, respectively. Data are mean of at least six independent experiments \pm SEM.

Table 4. Stability of IPPA08 and Compound 25 in PBS at Different pHs^a

| compound | pH | detection time | degradation rate (%) |
|--------------------|-----|----------------|----------------------|
| IPPA08 | 4.0 | 3 days | 1.65 |
| | 7.0 | 7 days | 0.809 |
| | 9.0 | 5 days | 0.501 |
| 25 (<i>t</i> -Bu) | 4.0 | 3 days | 1.49 |
| | 7.0 | 7 days | 0.527 |
| | 9.0 | 5 days | 0.333 |

^aStability was detected by UPLC.

indicated that compound 25 displayed selectivity in its effects on imidacloprid toxicity against the honey bee (*A. mellifera* L.).

DISCUSSION

Synergists can improve the pesticidal performance of insecticides, such as increasing insecticidal activity, reducing application frequency, and decreasing the rate of resistance development.²⁰ In our previous study, we found that IPPA08, a novel neonicotinoid with a unique oxabridged substructure, has proven strong synergistic effects on neonicotinoid insecticides, which is currently the most crucial class of insecticides.^{15,16} To discover new synergists with higher

Table 5. Bee Toxicity (*A. mellifera* L.) of Imidacloprid and a Mixture of Imidacloprid with Compound 25^{a,b}

| compound | bee toxicity | times | toxic regression equation | LD ₅₀ (μg/worker) | 95% CI | toxicological grade |
|----------------|------------------|-------|---------------------------|------------------------------|-------------|---------------------|
| 25 + IMI (1:1) | contact toxicity | 24 h | F | F | F | |
| | | 48 h | $y = 1.018x + 2.341$ | 0.118 | 0.069–0.228 | II |
| | oral toxicity | 24 h | $y = 1.797x + 0.690$ | 1.93 | 1.54–2.57 | II |
| | | 48 h | $y = 1.057x + 2.970$ | 1.06 | 0.624–2.60 | II |
| IMI | contact toxicity | 24 h | F | F | F | |
| | | 48 h | $y = 0.999x + 1.015$ | 0.096 | 0.053–0.363 | II |
| | oral toxicity | 24 h | F | F | F | |
| | | 48 h | $y = 1.383x + 0.234$ | 0.677 | 0.555–0.858 | II |
| 25 | contact toxicity | 24 h | F | F | F | |
| | | 48 h | | >12 | | IV |
| | oral toxicity | 24 h | F | F | F | |
| | | 48 h | | >27 | | IV |

^aF, failed to measure, which means that some of the bee in the test group escaped. ^bThe grade of the bee toxicity, very toxicity: I; high toxicity: II; moderate toxicity: III; low toxicity: IV.

effectiveness, a series of eight-membered oxabridged neonicotinoid analogues were synthesized. Biological evaluation revealed that all of these compounds possess significant synergistic effects on imidacloprid toxicity against cowpea aphid (*A. craccivora*) under the concentration of 4 and 2 mg/L. The structure–activity relationships (SARs) showed that compound 25 having an electron-donating group (EDG) *t*-Bu at position 4 of the phenyl ring exhibited excellent synergistic effects for imidacloprid activity. Interestingly, compound 25 presented a higher degree of synergism than IPPA08, along with a notable selectivity in its effects on imidacloprid toxicity against the honey bee (*A. mellifera* L.). These results indicate that replacement of the pyridyl ring by a phenyl ring was a successful strategy to obtain a novel synergist with improved effectiveness. Further studies on field-testing of the title compounds are ongoing.

Our previous investigation found that at low concentrations, IPPA08 can act as a positive allosteric modulator of noncanonical interfaces and likely slow the decay of currents through stabilizing the open-channel state caused by the action of imidacloprid on canonical interfaces.¹⁶ Based on our present results, we propose two hypotheses. First, the origin of the differential synergistic activity of compound 25 and IPPA08 toward *A. craccivora* and *A. mellifera* L. likely resides in the target itself. Second, it is possible that the synergistic mechanism of compound 25 is different from that of IPPA08 considering their structural difference, i.e., phenyl ring and pyridyl ring. We are currently working to gain experimental evidence to further support such hypotheses. In this work, we only provided some implicit cues for such hypotheses. At the same time, our present study provides a unique strategy to alleviate the dilemma from neonicotinoids. When imidacloprid or thiamethoxam are co-applied with compound 25, at the same insecticidal activity level, the amount of imidacloprid or thiamethoxam was reduced. The reduction in the use of neonicotinoids indirectly improves the safety of honey bees as well as decreases the risk of environmental residue by neonicotinoids. Moreover, compound 25 is expected to be developed as a novel synergist, which is selective and safe for honey bee.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jafc.0c04786>.

¹H NMR and ¹³C NMR spectral data of compounds 1–33 (PDF)

■ AUTHOR INFORMATION

Corresponding Author

Zhong Li – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China; orcid.org/0000-0001-6665-5040; Phone: +86-21-64253540; Email: lizhong@ecust.edu.cn; Fax: +86-21-64252603

Authors

Xiao Zhang – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China

Yiping Wang – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China

Zhiping Xu – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China

Xusheng Shao – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China; orcid.org/0000-0002-2579-7533

Zewen Liu – Key Laboratory of Integrated Management of Crop Diseases and Pests (Ministry of Education), College of Plant Protection, Nanjing Agricultural University, Nanjing, Jiangsu 210095, China; orcid.org/0000-0003-1084-9449

Xiaoyong Xu – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China; orcid.org/0000-0001-9679-3153

Peter Maiefisch – Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China; Research Portfolio Manager Insecticides and Seedcare, Syngenta Crop Protection AG, Basel CH-4002, Switzerland

Complete contact information is available at:

<https://pubs.acs.org/doi/10.1021/acs.jafc.0c04786>

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work was financial supported by National Key Research and Development Program of China (2017YFD0200500) and Innovation Program of Shanghai Municipal Education Commission (2017-01-07-00-02-E00037).

■ REFERENCES

- (1) Elbert, A.; Haas, M.; Springer, B.; Thielert, W.; Nauen, R. Applied aspects of neonicotinoid uses in crop protection. *Pest Manage. Sci.* **2008**, *64*, 1099–1105.
- (2) Jeschke, P.; Nauen, R. Neonicotinoids-from zero to hero in insecticide chemistry. *Pest Manage. Sci.* **2008**, *64*, 1084–1098.
- (3) Nauen, R.; Jeschke, P.; Copping, L. In Focus: neonicotinoid insecticides. *Pest Manage. Sci.* **2008**, *64*, 1081.
- (4) Bass, C.; Denholm, I.; Williamson, M. S.; Nauen, R. The global status of insect resistance to neonicotinoid insecticides. *Pestic. Biochem. Physiol.* **2015**, *121*, 78–87.
- (5) Crossthwaite, A. J.; Rendine, S.; Stenta, M.; Slater, R. Target-site resistance to neonicotinoids. *J. Chem. Biol.* **2014**, *7*, 125–128.
- (6) Raine, N. E.; Gill, R. J. Tasteless pesticides affect bees in the field. *Nature* **2015**, *521*, 38–40.
- (7) Navarro-Roldán, M. A.; Gemenio, C. Sublethal effects of neonicotinoid insecticide on calling behavior and pheromone production of tortricid moths. *J. Chem. Ecol.* **2017**, *43*, 881–890.
- (8) Alkassab, A. T.; Kirchner, W. H. Sublethal exposure to neonicotinoids and related side effects on insect pollinators: honeybees, bumblebees, and solitary bees. *J. Plant Dis. Prot.* **2017**, *124*, 1–30.
- (9) Fairbrother, A.; Purdy, J.; Anderson, T.; Fell, R. Risk of neonicotinoid insecticides to honeybees. *Environ. Toxicol. Chem.* **2014**, *33*, 719–731.
- (10) Shao, X.; Lee, P. W.; Liu, Z.; Xu, X.; Li, Z.; Qian, X. cis-Configuration: a new tactic/rationale for neonicotinoid molecular design. *J. Agric. Food Chem.* **2011**, *59*, 2943–2949.
- (11) Shao, X.; Fu, H.; Xu, X.; Xu, X.; Liu, Z.; Li, Z.; Qian, X. Divalent and oxabridged neonicotinoids constructed by dialdehydes and nitromethylene analogues of imidacloprid: design, synthesis, crystal structure, and insecticidal activities. *J. Agric. Food Chem.* **2010**, *58*, 2696–2702.
- (12) Cui, L.; Sun, L.; Yang, D.; Yan, X.; Yuan, H. Effects of cycloxaprid, a novel cis-nitromethylene neonicotinoid insecticide, on the feeding behaviour of *Sitobion avenae*. *Pest Manage. Sci.* **2012**, *68*, 1484–1491.
- (13) Zhang, J.; Fu, Q.; Wang, H.; Li, J.; Wang, W.; Yang, Z.; Zhang, S.; Ye, Q.; Li, C.; Li, Z. Enantioselective uptake and translocation of a novel chiral neonicotinoid insecticide cycloxaprid in *Youdonger* (*Brassica campestris* subsp. *Chinensis*). *Chirality* **2013**, *25*, 686–691.
- (14) Liu, X.; Xu, X.; Li, C.; Zhang, H.; Fu, Q.; Shao, X.; Ye, Q.; Li, Z. Degradation of chiral neonicotinoid insecticide cycloxaprid in flooded and anoxic soil. *Chemosphere* **2015**, *119*, 334–341.
- (15) Bao, H.; Shao, X.; Zhang, Y.; Deng, Y.; Xu, X.; Liu, Z.; Li, Z. Specific Synergist for Neonicotinoid Insecticides: IPPA08, a cis-Neonicotinoid Compound with a Unique Oxabridged Substructure. *J. Agric. Food Chem.* **2016**, *64*, 5148–5155.
- (16) Bao, H.; Shao, X.; Zhang, Y.; Cheng, J.; Wang, Y.; Xu, X.; Fang, J.; Liu, Z.; Li, Z. IPPA08 allosterically enhances the action of imidacloprid on nicotinic acetylcholine receptors. *Insect Biochem. Mol. Biol.* **2016**, *79*, 36–41.
- (17) Shao, X.; Fu, H.; Xu, X.; Xu, X.; Liu, Z.; Li, Z.; Qian, X. Divalent and oxabridged neonicotinoids constructed by dialdehydes and nitromethylene analogues of imidacloprid: Design, synthesis, crystal structure, and insecticidal activities. *J. Agric. Food Chem.* **2010**, *58*, 2696–2702.
- (18) Mohan, M.; Gujar, G. T. Local variation in susceptibility of the diamondback moth, *Plutella xylostella* (Linnaeus) to insecticides and role of detoxification enzymes. *Crop Prot.* **2003**, *22*, 495–504.
- (19) Organisation for Economic Co-operation and Development (OECD). Test No. 213: Honeybees, acute oral toxicity test. *OECD Guidelines for the Testing of Chemicals*; OECD: Paris, France, 2004; Vol. 1, pp 1–8.
- (20) Tabashnik, B. E. Managing resistance with multiple pesticide tactics: Theory, evidence, and recommendations. *J. Econ. Entomol.* **1989**, *82*, 1263–1269.