

Figure 6 A-D & Sup

RAC

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below code was used to process countfiles on cluster for snoRNAs

```
knitr::opts_chunk$set(tidy.opts=list(width.cutoff=60), tidy=TRUE)
```

```
#!/usr/bin/env Rscript
#
#suppressMessages(library(dplyr))
#suppressMessages(library(tidy))
#suppressMessages(library(fuzzyjoin))
#suppressMessages(library(stringr))
#args = commandArgs(trailingOnly=TRUE)
#
#ANNOTATION_BED_FILEPATH<-args[1]
#EXPRESSION_VECTOR_FILEPATH<-args[2]
#IN_FILE_PATH<-args[3]
#HOST_GENE_INDEX_FILEPATH<-args[4]
#OUTFILE_PATH<-args[5]
#
##Function to process count files -----
#wrangle_bed_counts <- function(dataframe) {
#  dataframe %>%
#    #the gene ID is complicated and has different number of columns for some snoRNAs, best to label by
#    mutate(snoRNAlocation = case_when(
#      grepl("::intronic::", geneID) ~ "intronic",
#      grepl("::nonintronic::", geneID) ~ "non_intronic"
#    )) %>%
#    mutate(region = case_when(
#      grepl("intron3ss", geneID) ~ "intron3ss",
#      grepl("intron5ss", geneID) ~ "intron5ss",
#      grepl("mature3end", geneID) ~ "mature3end",
#      grepl("mature5end", geneID) ~ "mature5end",
#    )) %>%
#    mutate(snoRNAtype = case_when(
#      grepl("SNORD|snoU|U3|U8|snoMe28S-Am2634|snoMBII|snoZ|snosnR66", geneID) ~ "cdBox",
#      grepl("SNORA|ACA|RNU105C|RNU105B", geneID) ~ "HACA",
#      grepl("SCARNA", geneID) ~ "scaRNA",
#      TRUE ~ "other"
#    )) %>%
#    mutate(geneID = as.character(geneID)) %>%
#    #fuzzy_left join allows string matching rather than exact matching. Noticed more output afterwards
#    fuzzy_left_join(host_gene_index, by=c("geneID" = "geneID"), match_fun = str_detect) %>%
#    separate(hostGeneID, into=c("hostGeneID", "transcriptBiotype", "intronNumber", "distFromTSS"), sep = "
```

```

#   unique() %>%
#   #left join the expression vector using specific predefined columns.
#   left_join(expression_vector, by = c("hostGeneID" = "geneID")) %>%
#   mutate(norm_count = count/ctrl_RNAseq_expr) %>%
#   select(-count, -ctrl_RNAseq_expr) %>%
#   group_by(Sample, region, snoRNAType, snoRNALocation, DistToLandmark) %>%
#   summarise(sum_RNAseq_norm_count_norm_annotation_number = sum(norm_count)) %>%
#   left_join(number_of_intron_annotations) %>%
#   mutate(sum_RNAseq_norm_count_norm_annotation_number = sum_RNAseq_norm_count_norm_annotation_number,
# }
#
#
#
#
##Load expression vector -----
#print("load expression vector")
#
#load(EXPRESSION_VECTOR_FILEPATH)
#
#expression_vector<-left_join(
#   (as.data.frame(ctrl_RNAseq_expr) %>%
#     tibble::rownames_to_column(var = "geneID")),
#   (as.data.frame(ctrl_TTseq_expr) %>%
#     tibble::rownames_to_column(var = "geneID"))
#) %>%
#   select(-ctrl_TTseq_expr) %>%
#   mutate(ctrl_RNAseq_expr = case_when(
#     ctrl_RNAseq_expr ==0 ~ min(ctrl_RNAseq_expr[ctrl_RNAseq_expr > 0]),
#     TRUE ~ ctrl_RNAseq_expr
#   ))
#
#
##load annobed -----
#print("load annotation bed")
#
#annoBed<-read.table(ANNOTATION_BED_FILEPATH, sep = "\t", header = F)
#print(ncol(annoBed))
#colnames(annoBed)<-c("chr", "start", "end", "geneID", "score", "strand")
#
#number_of_intron_annotations<-
#   annoBed %>%
#   mutate(snoRNALocation = case_when(
#     grepl(":::intronic", geneID) ~ "intronic",
#     grepl(":::nonintronic", geneID) ~ "non_intronic"
#   )) %>%
#   mutate(snoRNAType = case_when(
#     grepl("SNORD/snoU/U3/U8/snoMe28S-Am2634/snoMBII/snoZ/snosnR66", geneID) ~ "cdBox",
#     grepl("SNORA/ACA/RNU105C/RNU105B", geneID) ~ "HACA",
#     grepl("SCARNA", geneID) ~ "scaRNA",
#     TRUE ~ "other"
#   )) %>%
#   group_by(snoRNALocation, snoRNAType) %>%
#   summarise(intron_count =n())

```

```

#
##load index file -----
#print("load index file")
#
#host_gene_index<-read.table(HOST_GENE_INDEX_FILEPATH, sep = "\t", header = F)
#colnames(host_gene_index)<-c("geneID", "hostGeneID")
#host_gene_index$geneID<-as.character(host_gene_index$geneID)
#
#
##load count file -----
#print("load count file")
#
#int_exon_junction_coverage<-read.table(IN_FILE_PATH, sep = "\t", header = F)
#colnames(int_exon_junction_coverage)<-c("Sample", "chr", "start", "end", "geneID", "DistToLandmark", "s
#
#int_exon_junction_coverage<-
# wrangle_bed_counts(int_exon_junction_coverage)
#
##rbind the files together and write to output filename
#
#OUTFILE<-int_exon_junction_coverage
#
#write.table(OUTFILE, OUTFILE_PATH, quote = F, sep = "\t", row.names = F)

# suppressMessages(library(dplyr))
# suppressMessages(library(tidyr))
# suppressMessages(library(stringr))
# suppressMessages(library(ggplot2)) dir.create('figs/')
# dir.create('figs/snoRNA_coverage_plots') load in the file
# annotation_file<-read.table('snoRNAs.GRCh38andrefGene.mature.bed')
# colnames(annotation_file)<-c('chr', 'start', 'end',
# 'geneID', 'score', 'strand') count number of snoRNAs to
# provide a number to normalise to. annotation_number<-
# annotation_file %>% mutate(snoRNAlocation = case_when(
# grepl(':::intronic', geneID) ~ 'intronic',
# grepl(':::nonintronic', geneID) ~ 'non_intronic' )) %>%
# mutate(snoRNAtype = case_when(
# grepl('SNORD|snoU|U3|U8|snoMe28S-Am2634|snoMBII|snoZ|snosnR66',
# geneID) ~ 'cdBox', grepl('SNORA|ACA|RNU105C|RNU105B',
# geneID) ~ 'HACA', grepl('SCARNA', geneID) ~ 'scaRNA',
# TRUE ~ 'other' )) %>% group_by(snoRNAlocation) %>%
# summarise(n=n())

suppressMessages(library(dplyr))
suppressMessages(library(tidyr))
suppressMessages(library(stringr))
suppressMessages(library(ggplot2))
library(extrafont)

## Registering fonts with R
# read in count file

snoRNA_and_host_introns <- read.table("../data/xiCLIP_all_hg38_snoRNAs_and_host_introns_no_gene_norm

```

```

header = T)

head(snoRNA_and_host_introns)

##           Sample      region snoRNAType snoRNALocation DistToLandmark
## 1 ALYREF_1_DMSO_3endOfRead2 intron3ss      cdBox      intronic      -100
## 2 ALYREF_1_DMSO_3endOfRead2 intron3ss      cdBox      intronic      -99
## 3 ALYREF_1_DMSO_3endOfRead2 intron3ss      cdBox      intronic      -98
## 4 ALYREF_1_DMSO_3endOfRead2 intron3ss      cdBox      intronic      -97
## 5 ALYREF_1_DMSO_3endOfRead2 intron3ss      cdBox      intronic      -96
## 6 ALYREF_1_DMSO_3endOfRead2 intron3ss      cdBox      intronic      -95
##      sum_count intron_count sum_count_norm_annotation_number
## 1 1175.6097          393          2.9913734
## 2 1315.9832          393          3.3485577
## 3  407.9558          393          1.0380555
## 4  140.3714          393          0.3571792
## 5  109.6652          393          0.2790464
## 6  741.3364          393          1.8863521

# group by snoRNA type, location and each bin and the sum
# the count. This is then joined to the appropriate
# annotation number and further normalised

# wrangle in the rest of the factors
snoRNA_and_host_introns_wrangled_sum_for_graph <- snoRNA_and_host_introns %>%
  separate(Sample, c("Protein", "Rep", "Timepoint", "Read")) %>%
  mutate(region = factor(region, c("intron5ss", "mature5end",
    "mature3end", "intron3ss")), Timepoint = factor(Timepoint,
    c("negative", "PBSDRB", "t00", "t05", "t10", "t15", "t20",
    "t40", "t60", "DMSO"))) %>%
  mutate(Timepoint_f = case_when(Timepoint == "PBSDRB" ~ "t00",
    TRUE ~ as.character(Timepoint))) %>%
  mutate(Timepoint_f = factor(Timepoint_f, levels = c("negative",
    "t00", "t05", "t10", "t15", "t20", "t40", "t60", "DMSO")))

head(snoRNA_and_host_introns_wrangled_sum_for_graph)

##      Protein Rep Timepoint      Read      region snoRNAType snoRNALocation
## 1  ALYREF    1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 2  ALYREF    1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 3  ALYREF    1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 4  ALYREF    1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 5  ALYREF    1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 6  ALYREF    1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
##      DistToLandmark sum_count intron_count sum_count_norm_annotation_number
## 1             -100 1175.6097          393          2.9913734
## 2             -99 1315.9832          393          3.3485577
## 3             -98  407.9558          393          1.0380555
## 4             -97  140.3714          393          0.3571792
## 5             -96  109.6652          393          0.2790464
## 6             -95  741.3364          393          1.8863521
##      Timepoint_f
## 1      DMSO

```

```
## 2      DMSO
## 3      DMSO
## 4      DMSO
## 5      DMSO
## 6      DMSO
```

```
figure_data <- snoRNA_and_host_introns_wrangled_sum_for_graph %>%
  filter(snoRNAType %in% c("cdBox", "HACA") & snoRNALocation ==
         "intronic" & Timepoint != "negative" & Protein == "RBM7" &
         region %in% c("mature3end", "intron3ss"))

head(figure_data)
```

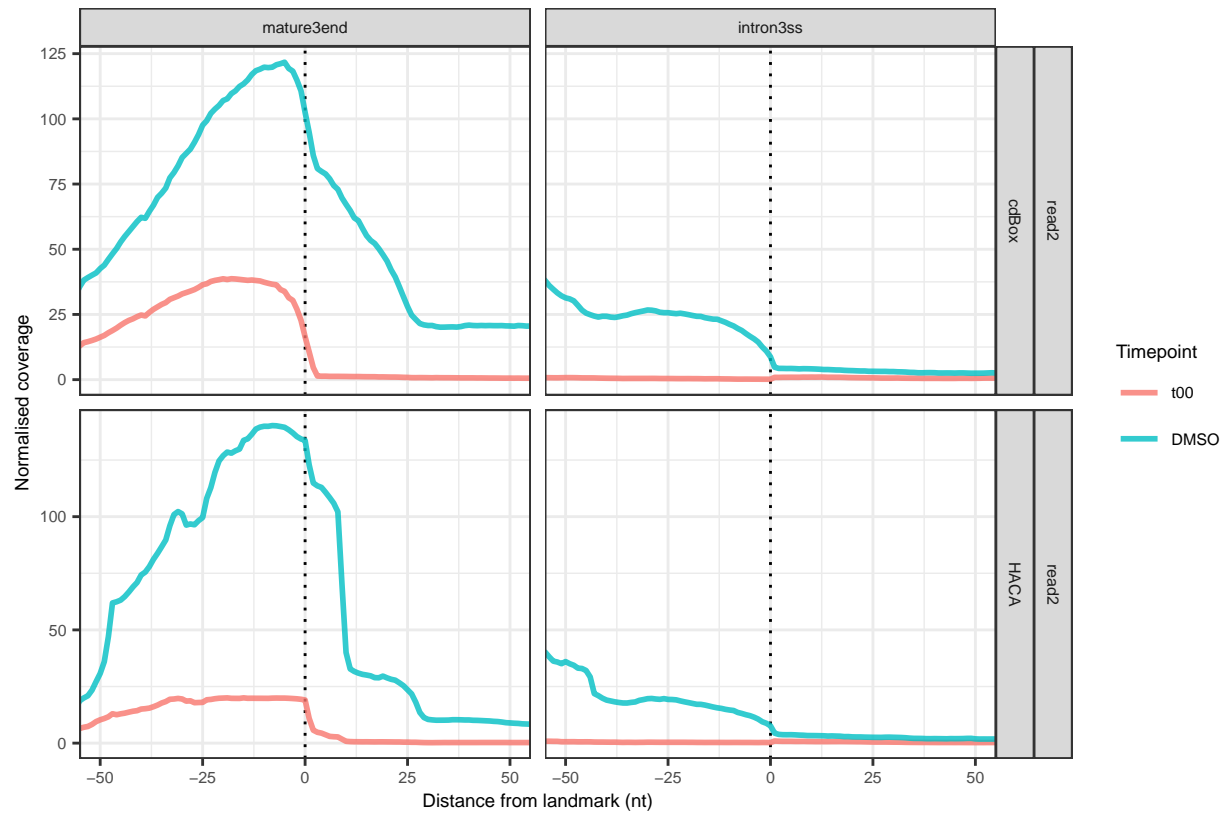
```
## Protein Rep Timepoint      Read      region snoRNAType snoRNALocation
## 1   RBM7   1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 2   RBM7   1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 3   RBM7   1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 4   RBM7   1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 5   RBM7   1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## 6   RBM7   1      DMSO 3endOfRead2 intron3ss      cdBox      intronic
## DistToLandmark sum_count intron_count sum_count_norm_annotation_number
## 1              -100 817.5904           393                2.0803827
## 2              -99 524.9302           393                1.3357003
## 3              -98 376.2774           393                0.9574489
## 4              -97 283.3694           393                0.7210417
## 5              -96 329.8234           393                0.8392453
## 6              -95 548.1572           393                1.3948020
## Timepoint_f
## 1      DMSO
## 2      DMSO
## 3      DMSO
## 4      DMSO
## 5      DMSO
## 6      DMSO
```

#Figure 6 A-D. Calculate coverage of read2 and 3'CLIP at nucleotide resolution over 101nt windows centred on the 3' end of snoRNAs or its corresponding downstream intron-exon junction. SnoRNAs are stratified by class (H/ACA or cdBox). Normalisation: read coverage over snoRNA window to host gene expression, aggregate reads, and divide by the total number of snoRNA annotations.

fig 6 a

```
# Figure 6 A whole read plots
figure_data %>%
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "read2" &
         Timepoint %in% c("t00", "DMSO")) %>%
  ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
                        size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
                                                    col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
                                                size = 1) + facet_grid(Read + snoRNAType ~ region, scales = "free_y") +
  theme_bw() + coord_cartesian(xlim = c(-50, 50)) + xlab("Distance from landmark (nt)") +
  ylab("Normalised coverage") + labs(subtitle = "Fig 6. a - Controls; whole read") +
  theme(text = element_text(size = 7, family = "Arial"))
```

Fig 6. a – Controls; whole read



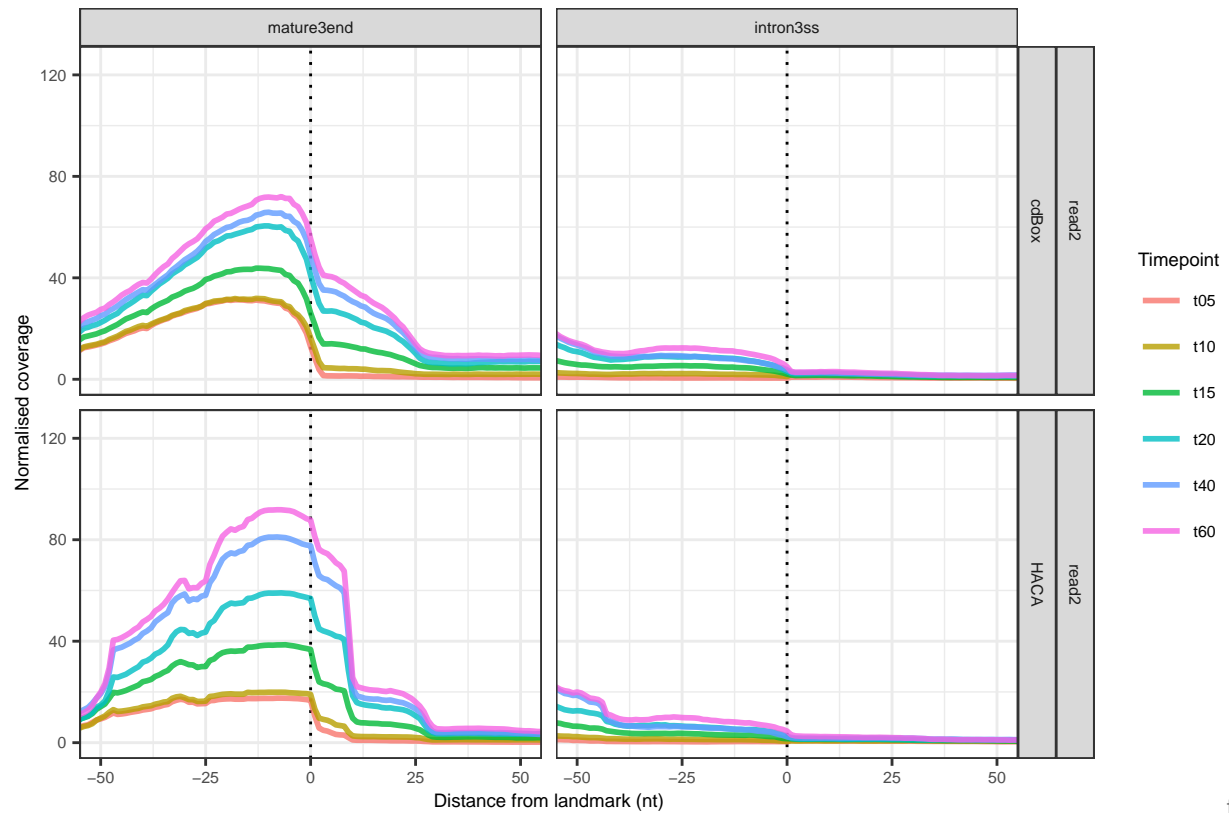
fig

6 b

```
# Figure 6 B
dummy_data <- figure_data %>%
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "read2" &
    Timepoint %in% c("t00", "DMSO"))

# whole read plots
figure_data %>%
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "read2" &
    !(Timepoint %in% c("t00", "DMSO"))) %>%
  ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
    size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
    col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
    size = 1) + facet_grid(Read + snoRNAType ~ region, scales = "free_y") +
  theme_bw() + coord_cartesian(xlim = c(-50, 50), ylim = c(0,
    125)) + xlab("Distance from landmark (nt)") + ylab("Normalised coverage") +
  labs(subtitle = "Fig 6. b - Time course; whole read") + theme(text = element_text(size = 7,
    family = "Arial"))
```

Fig 6. b – Time course; whole read



fig

6 c

Figure 6 C whole read plots, 3'CLIP

figure_data %>%

```
filter(snoRNAType %in% c("HACA", "cdBox") & Read == "3endOfRead2" &
       (Timepoint %in% c("t00", "DMSO"))) %>%
```

```
ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
                      size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
                                                  col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
                      size = 1) + facet_grid(Read + snoRNAType ~ region, scales = "free_y") +
  theme_bw() + coord_cartesian(xlim = c(-50, 50)) + xlab("Distance from landmark (nt)") +
  ylab("Normalised coverage") + labs(subtitle = "Fig 6. c - Controls; 3'CLIP") +
  theme(text = element_text(size = 7, family = "Arial"))
```

Fig 6. c – Controls; 3'CLIP

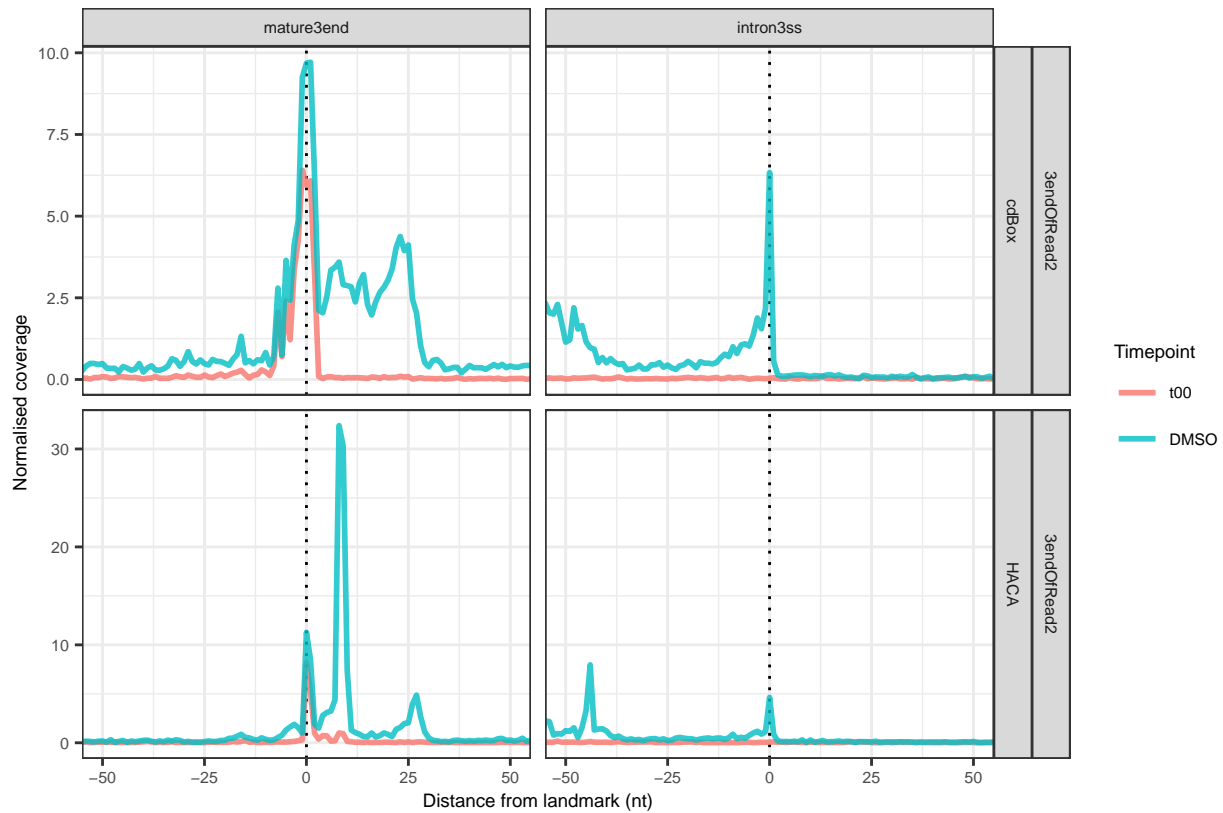


Figure 6 D invisible data to keep yaxis same

```
dummy_data <- figure_data %>%
```

```
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "3endOfRead2" &
         (Timepoint %in% c("t00", "DMSO")))
```

whole read plots

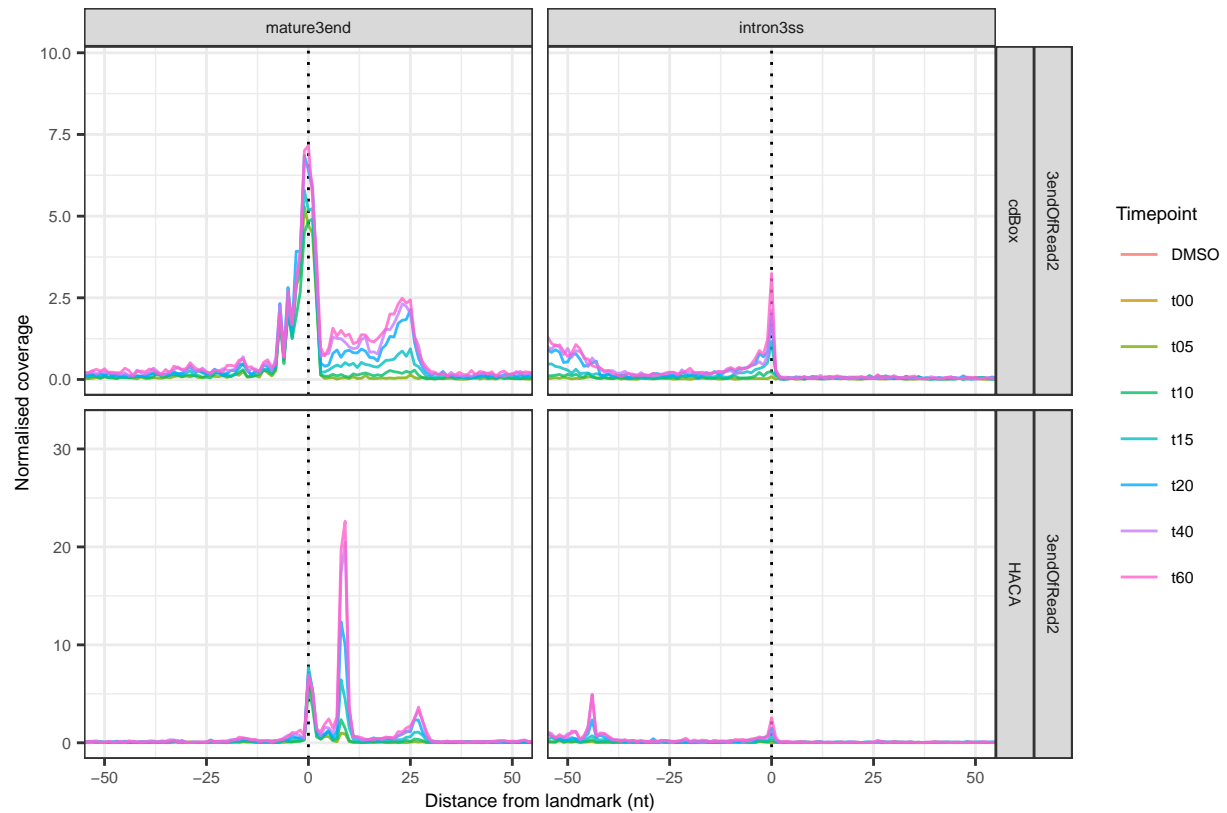
```
figure_data %>%
```

```
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "3endOfRead2" &
         !(Timepoint %in% c("t00", "DMSO"))) %>%
```

```
  ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
                        size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
                                                    col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
                                                size = 0.5) + geom_blank(data = dummy_data, aes(x = DistToLandmark,
                                                    y = sum_count_norm_annotation_number, col = Timepoint), stat = "summary",
                                                fun = "mean", alpha = 0.8, size = 1) + facet_grid(Read ~ snoRNAType ~ region, scales = "free_y") + theme_bw() + coord_cartesian(xlim = c(-50,
50)) + xlab("Distance from landmark (nt)") + ylab("Normalised coverage") +
  labs(subtitle = "Fig 6. d - Time course; 3'CLIP") + theme(text = element_text(size = 7,
family = "Arial"))
```

```
## Warning: Ignoring unknown parameters: alpha, size
```


Fig 6. d – Time course; 3'CLIP



#Supplementary Figure 6 c - e coverage as above over snoRNAs, except showing cross-link sites also showing cross-link sites at 5' end, and upstream 5'SS for snoRNA containing introns

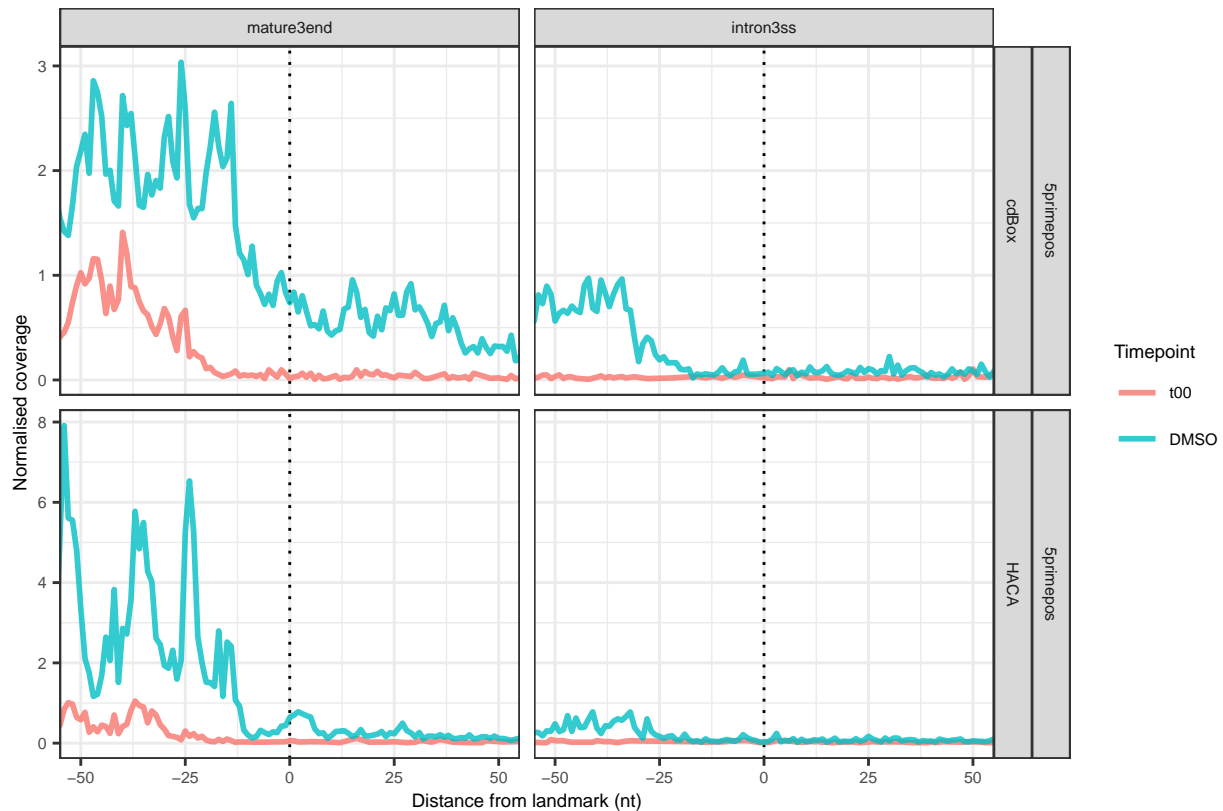
supplementary fig 6d

Supplementary Figure 6 cross-link

figure_data %>%

```
filter(snoRNAType %in% c("HACA", "cdBox") & Read == "5primepos" &
  (Timepoint %in% c("t00", "DMSO"))) %>%
ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
  size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
  col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
  size = 1) + facet_grid(Read + snoRNAType ~ region, scales = "free_y") +
  theme_bw() + coord_cartesian(xlim = c(-50, 50)) + xlab("Distance from landmark (nt)") +
  ylab("Normalised coverage") + labs(subtitle = "Sup Fig. 6 d - Controls; cross-link") +
  theme(text = element_text(size = 7, family = "Arial"))
```

Sup Fig. 6 d – Controls; cross-link



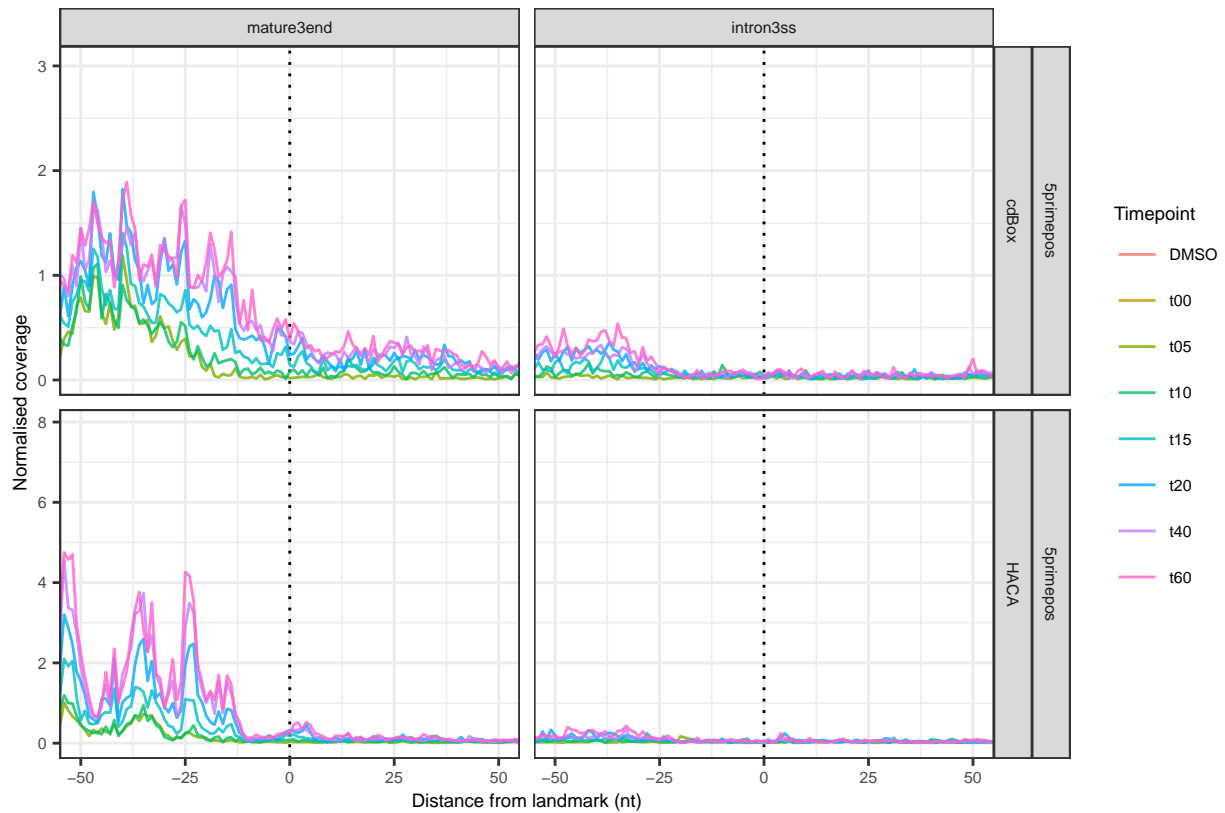
supplementary fig 6 e

```
# invisible data to keep yaxis same
dummy_data <- figure_data %>%
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "5primepos" &
    (Timepoint %in% c("t00", "DMSO")))

# cross-link
figure_data %>%
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "5primepos" &
    !(Timepoint %in% c("t00", "DMSO"))) %>%
  ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
    size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
    col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
    size = 0.5) + geom_blank(data = dummy_data, aes(x = DistToLandmark,
    y = sum_count_norm_annotation_number, col = Timepoint), stat = "summary",
    fun = "mean", alpha = 0.8, size = 1) + facet_grid(Read +
    snoRNAType ~ region, scales = "free_y") + theme_bw() + coord_cartesian(xlim = c(-50,
    50)) + xlab("Distance from landmark (nt)") + ylab("Normalised coverage") +
    labs(subtitle = "Sup Fig. 6e Time course; cross-link") +
    theme(text = element_text(size = 7, family = "Arial"))
```

Warning: Ignoring unknown parameters: alpha, size

Sup Fig. 6e Time course; cross-link



supplementary fig 6 c

```
supplementary_figure_data <- snoRNA_and_host_introns_wrangled_sum_for_graph %>%
  filter(snoRNAType %in% c("cdBox", "HACA") & snoRNALocation ==
    "intronic" & Timepoint != "negative" & Protein == "RBM7" &
    region %in% c("mature5end", "intron5ss"))

# supplementary figure 6 c

supplementary_figure_data %>%
  filter(snoRNAType %in% c("HACA", "cdBox") & Read == "5primepos") %>%
  ggplot() + geom_vline(xintercept = 0, linetype = "dotted",
    size = 0.5) + geom_line(aes(x = DistToLandmark, y = sum_count_norm_annotation_number,
    col = Timepoint), stat = "summary", fun = "mean", alpha = 0.8,
    size = 1) + facet_grid(Read + snoRNAType ~ region, scales = "free_y") +
  theme_bw() + coord_cartesian(xlim = c(-50, 50), ylim = c(0,
    5)) + xlab("Distance from landmark (nt)") + ylab("Normalised coverage") +
  labs(subtitle = "Sup Fig6 C - Controls & Time course; cross-link") +
  theme(text = element_text(size = 7, family = "Arial"))
```

Sup Fig6 C – Controls & Time course; cross-link

