

Research Paper Title:

PT-AML: Machine learning framework to identified personalized treatments for acute myeloid leukemia

Journal: *TBD*

Authors: Raghvendra Mall^{1#}, Siddhi Jani², ...

Keywords: []

SC: Deep Learning; Machine Learning; AML; Drug Sensitivity; Precision Medicine; Mutations; Cell State; Mechanism of Action

Abstract: [Checklist: Statements are factually accurate; provides a concise summary that is interesting and attracts attention; provides key background, purpose of article, and key findings from the study]

ABSTRACT EXAMPLE:

First line: State the topic; should be related to the title

Resistance to cell death is a leading hallmark of cancer. Therapies aimed at activating cell death are therefore of high interest to improve treatment and understanding the induction of cell death during cancers is a key strategy to identify therapies.

Second and third sentences: Expand on the critical background information relating to the topic, including why it is a relevant

Fourth sentence and beyond: These sentences should mirror the subsections from your results outline

.

Last sentence: These results demonstrate...

SC:

Research Article Outline:

PT-AML: Machine learning framework to identified personalize treatments for acute myeloid leukemia

[Checklist: Statements are factually accurate, key references from Kanneganti lab and Others ref lists are included]

1. **Introduction** **[Checklist:** limit to 3 paragraphs; describe the key concepts; provide relevant background information in the context of our lab's interests; attract the readers attention and inform about the purpose of the article and what you aim to achieve]

SC:

- 2.
- 3.
- 4.
- 5.

6. **Discussion**
-

Figures

[Checklist: concise title and legend; include labels and enough detail to be understood without the text; fonts and colors follow lab template, and the formatting is correct; no image duplications]

Figure Checklist

First authors and coauthors are responsible for ensuring that: 1) data are reproducible and there are no image duplications; 2) data have been independently verified by 2 people other than the first author (provide their names here).

See the example figure on the next page for clarifications. Please carefully check each item and sign off on the completion of this checklist before bringing figures to Thiru or Rebecca. Finalized figures must be approved by Thiru before you begin writing the manuscript.

Many of these points also apply for review figures. Particularly, for review figure color scheme ideas, refer to the colors used in this poster from InvivoGen:

https://www.invivogen.com/sites/default/files/invivogen/resources/documents/2016-poster_tlr-nlr-invivogen_0.pdf

- ¹ Figure # is at the bottom right of each figure in Arial font, size 14, bold (ex: **Figure 1**)
- ² Figure title is at the top of the page in Arial font, size 11, bold (ex: **Figure title**)
- ³ Figure panels are denoted by bold, uppercase letters in Arial font, size 14 (ex: **A**)
- ⁴ All text within the figure panels is in Arial font, size 11, not bold (ex: Media)
- ⁵ Western blot images include labels for the lanes along the top of the image; other labels are also at the top
- ⁶ Lane and border thickness are set to 1 pt with dotted lines on western blots set to 0.5 pt
- ⁷ No shadows or shadow lines are present
- ⁸ Terminology and labeling are used consistently throughout all figures (ex: *Casp11^{-/-}*)
- ⁹ Colors should be consistent throughout all figures (wild type, black; knock-out, red)
- ¹⁰ Red and green are not included on the same graph (due to red-green colorblindness)
- ¹¹ All microscopy images include the scale bar, which is defined in the figure legend
- ¹² Arrows can be used in microscopy images to callout cell death or other noteworthy staining; these arrows should be triangles (ex:)
- ¹³ All blots and gels have molecular weights (or base pair size for DNA/RNA) for all proteins or DNA/RNA species indicated
- ¹⁴ All blots must have an accompanying raw blot file (see example in Figure 2)
- ¹⁵ **There are no image duplications** (no duplicated blots, microscopy images, etc.) and data are reproducible and have been validated by 2 additional people as listed above

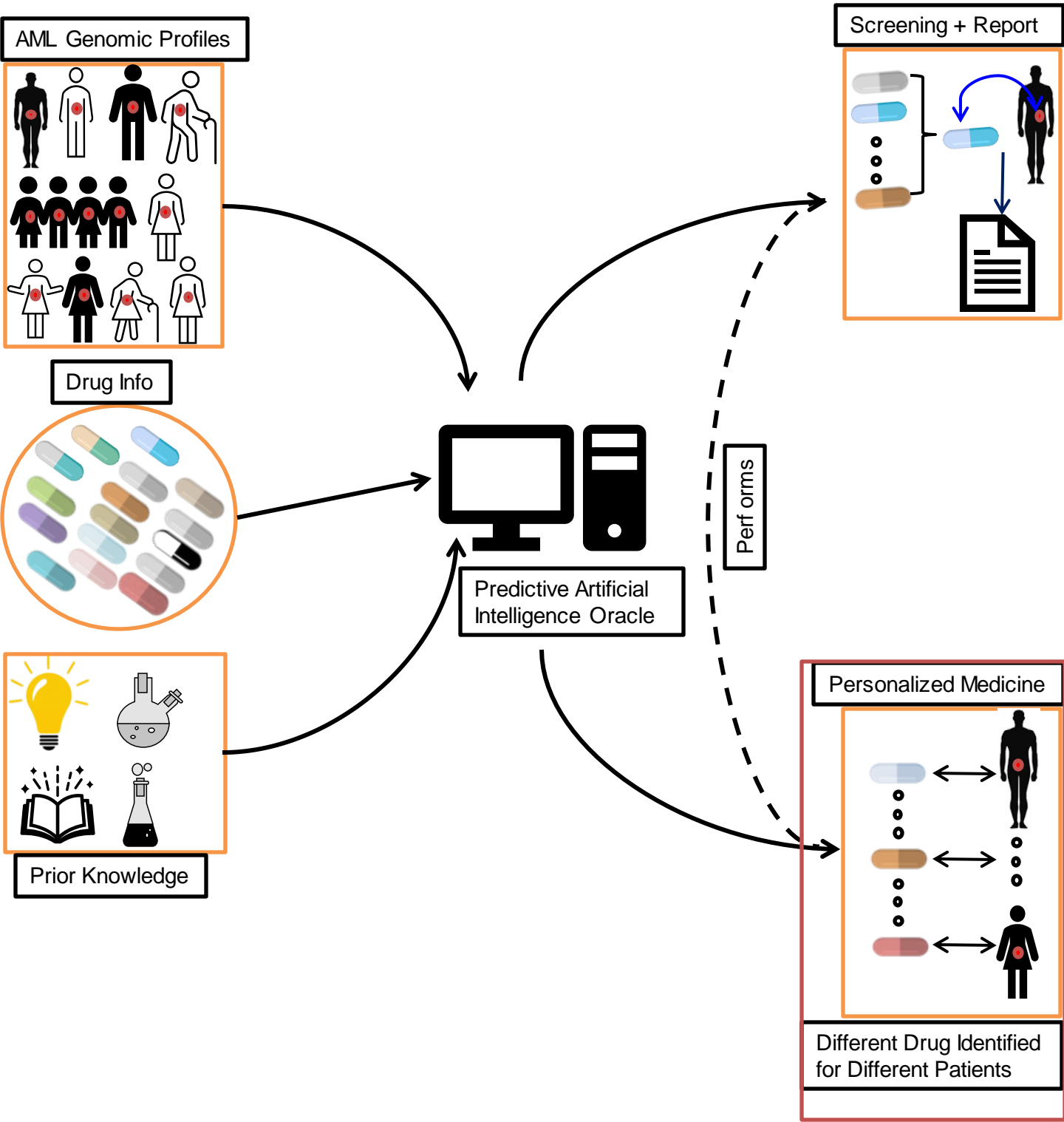
I confirm that my figures meet the above criteria, along with any other journal-specific criteria.

Name

Date

PT-AML: Machine learning framework to identified personalized treatments for acute myeloid leukemia

Graphical Abstract



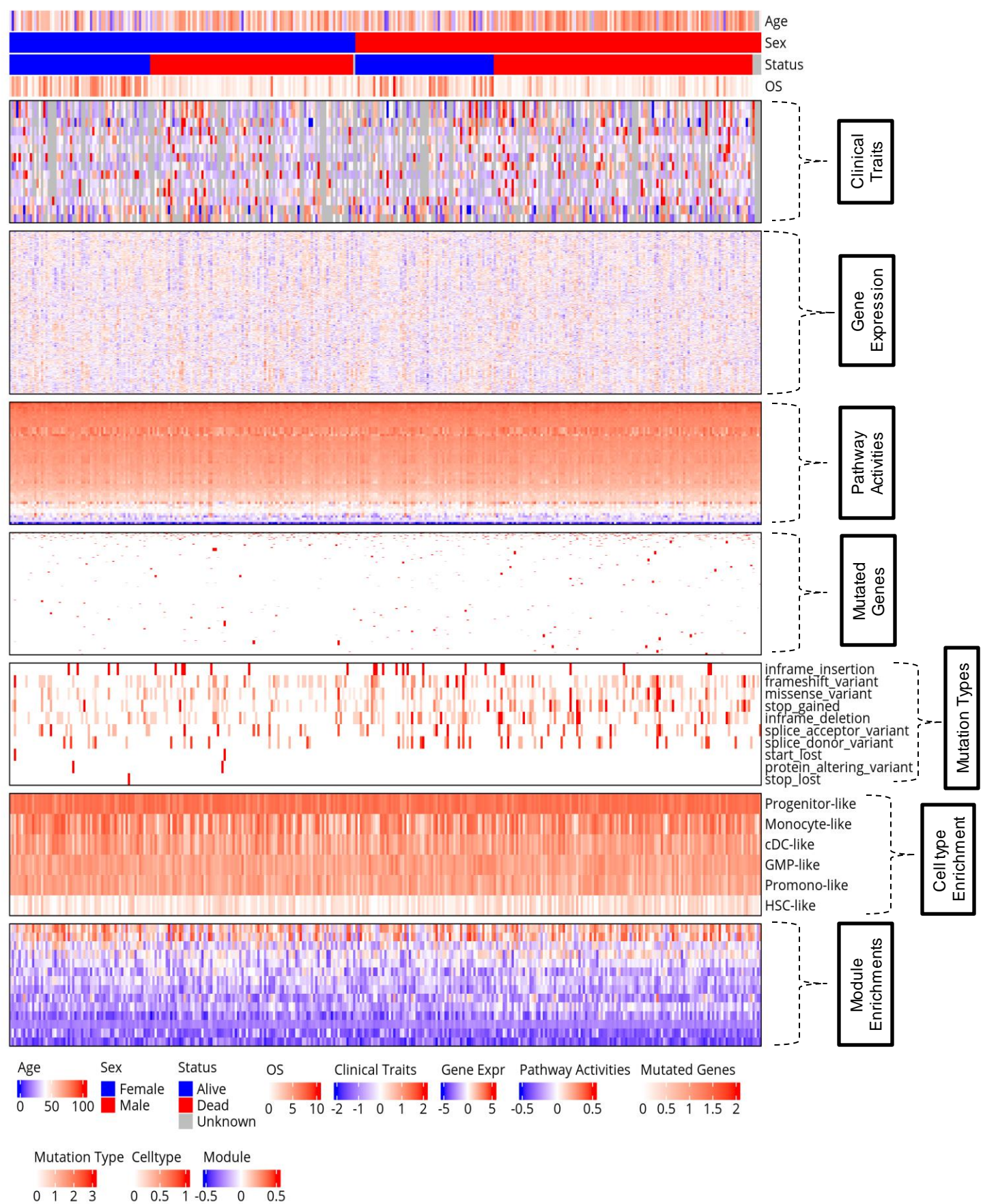


Figure 1

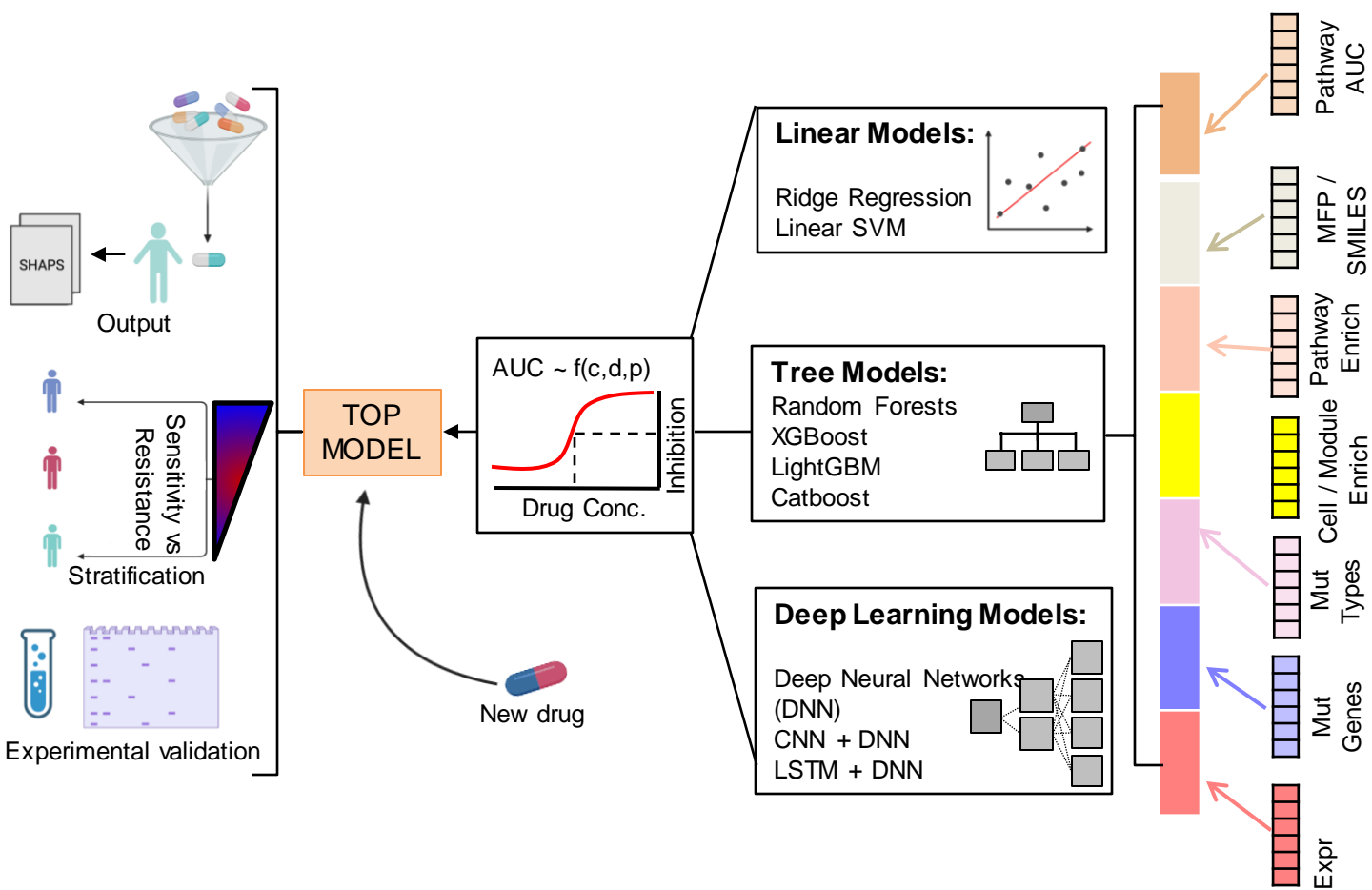
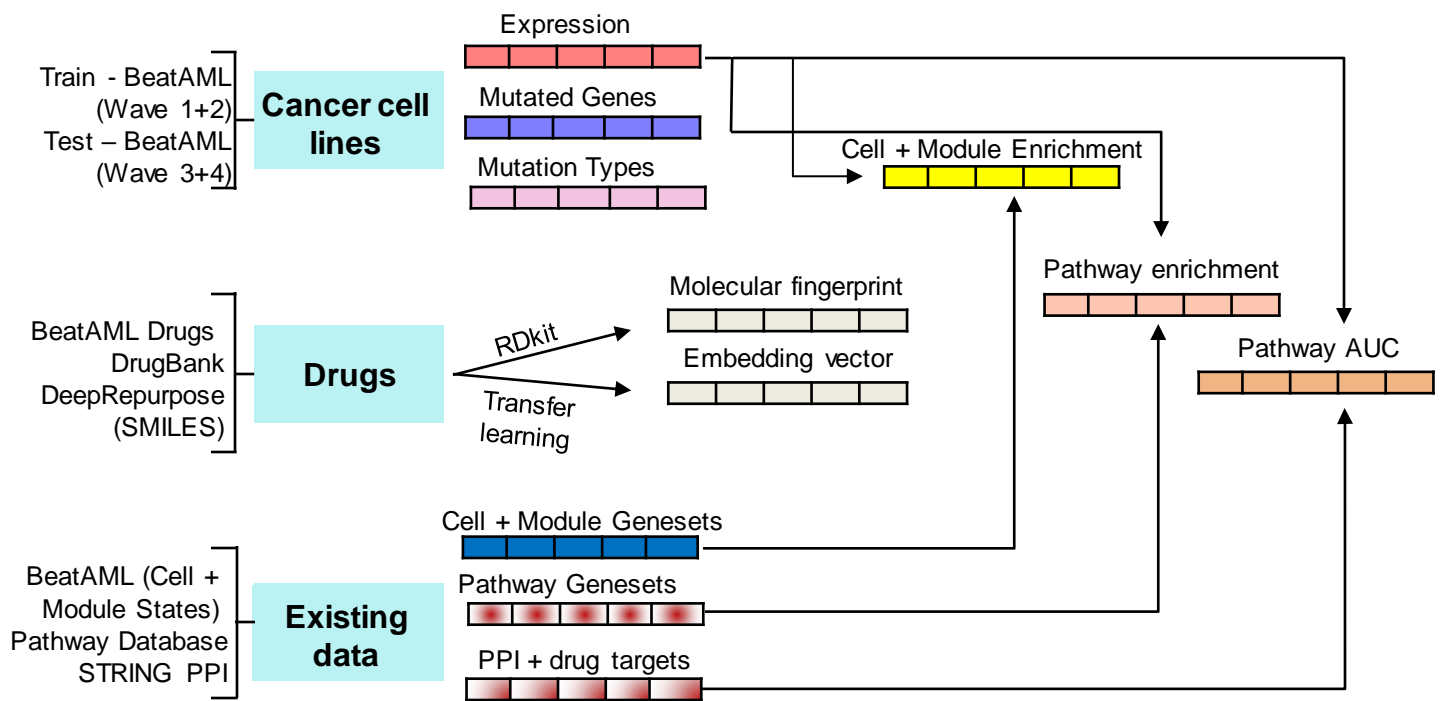
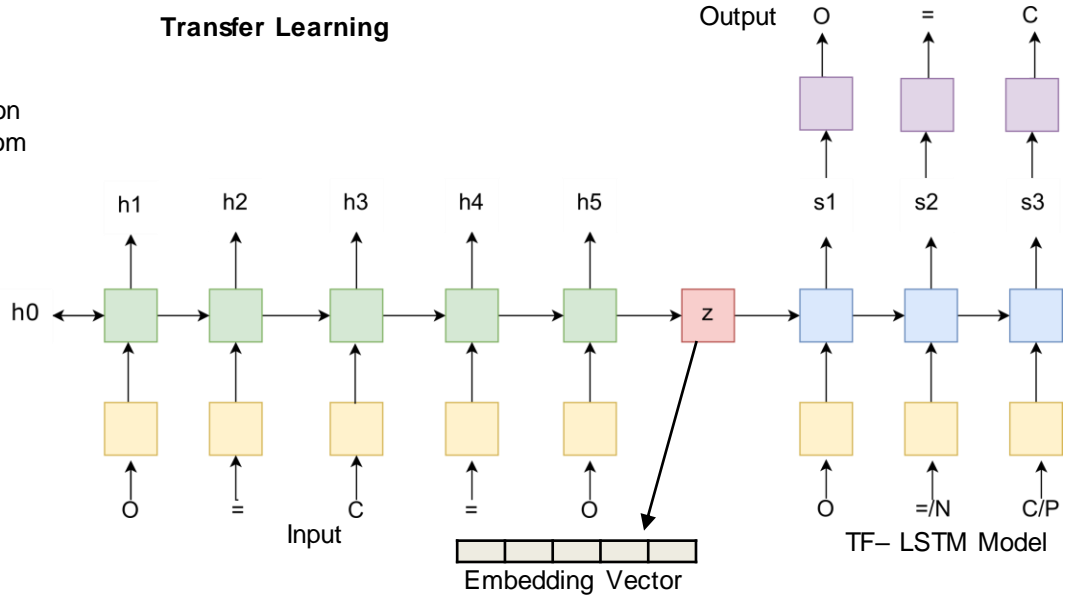
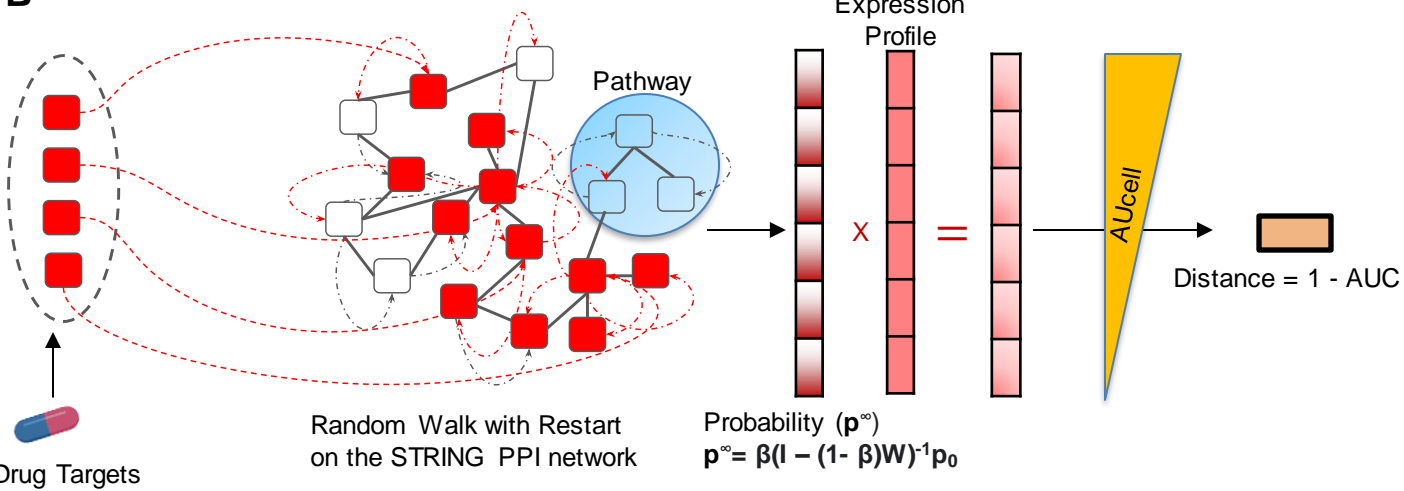


Figure 2

A**Transfer Learning**DeepRepurpose:

- TF-LSTM model trained on 2.5 million compounds from PubChem + MOSES.
- 96.7% reconstructed compounds are valid.
- μ_{error} per sample is 0.001

**B****Pathway Distance Estimation****Figure 3**

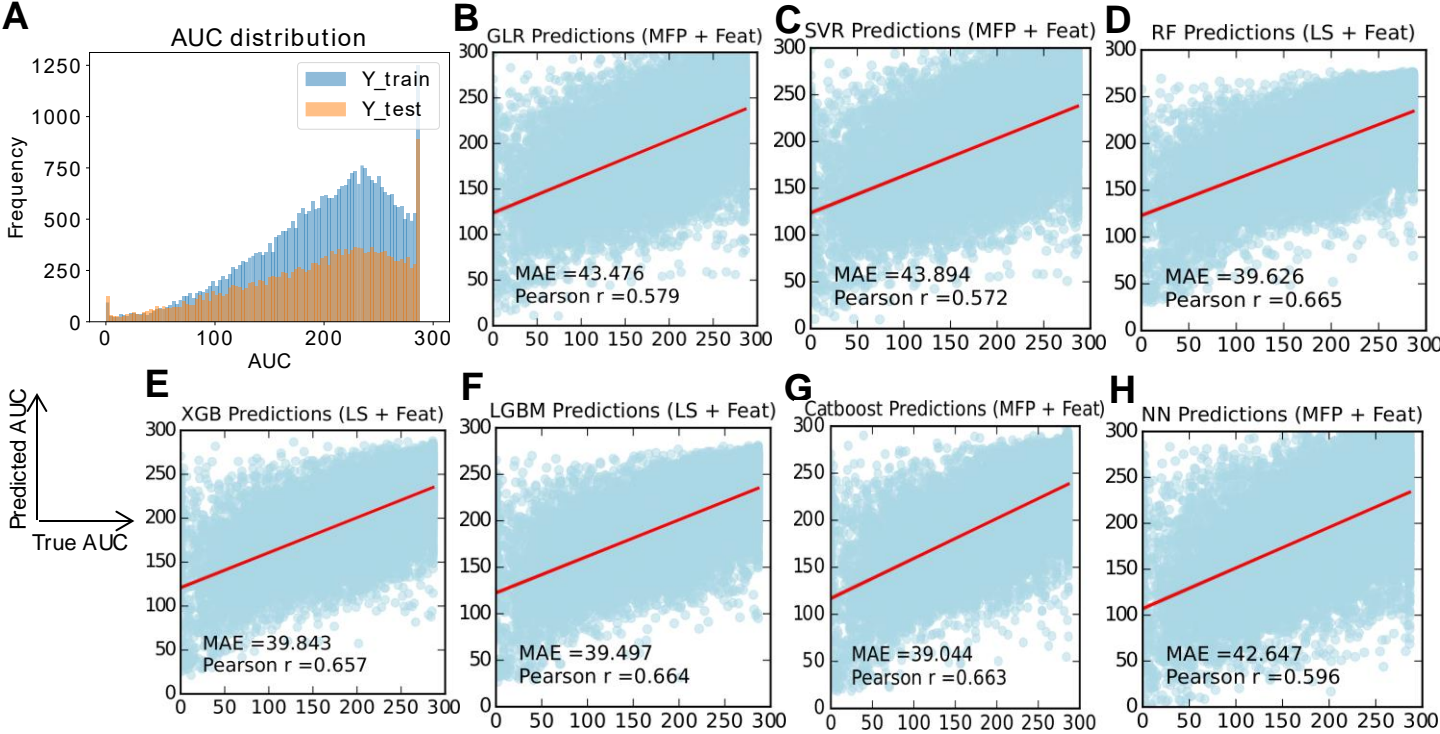
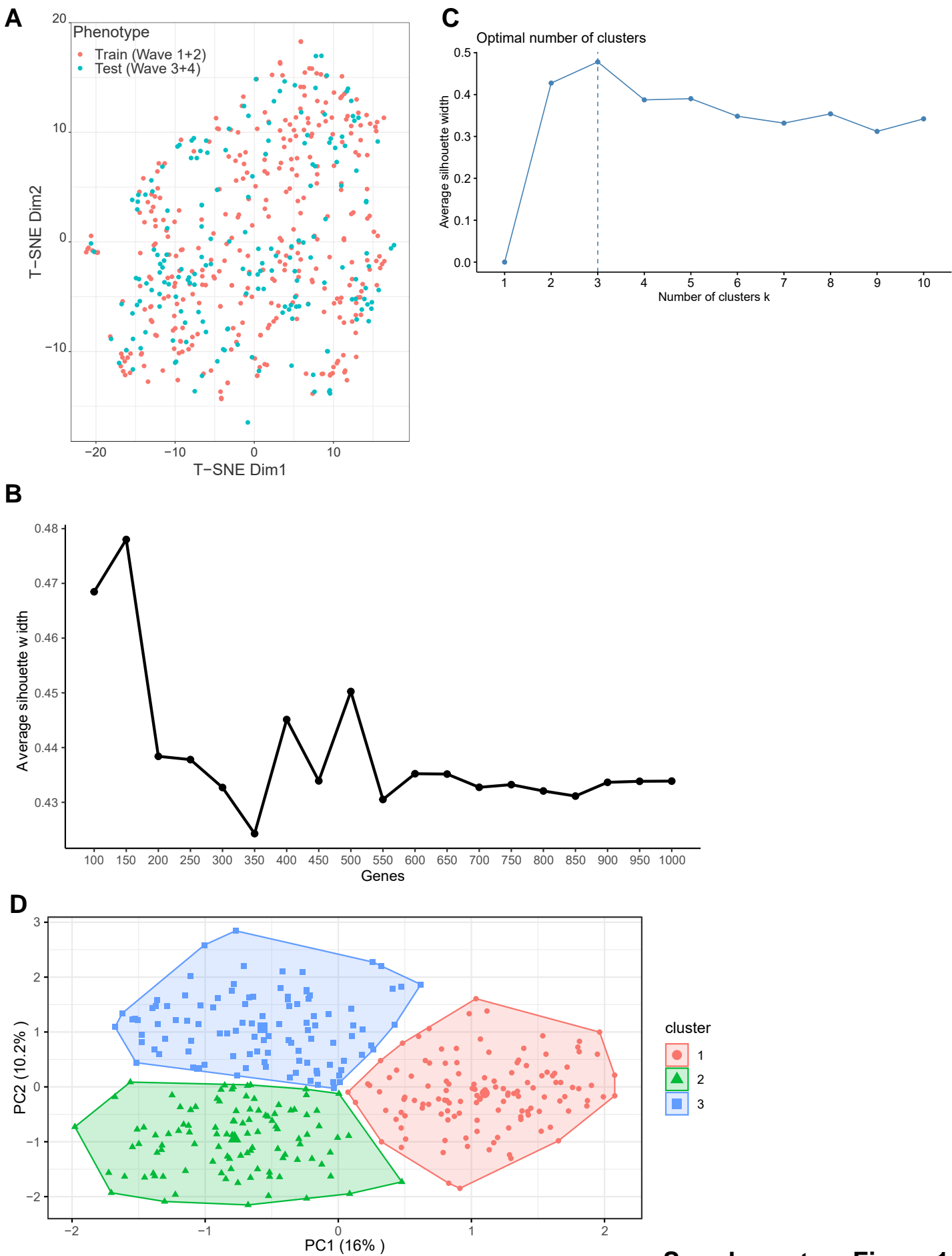
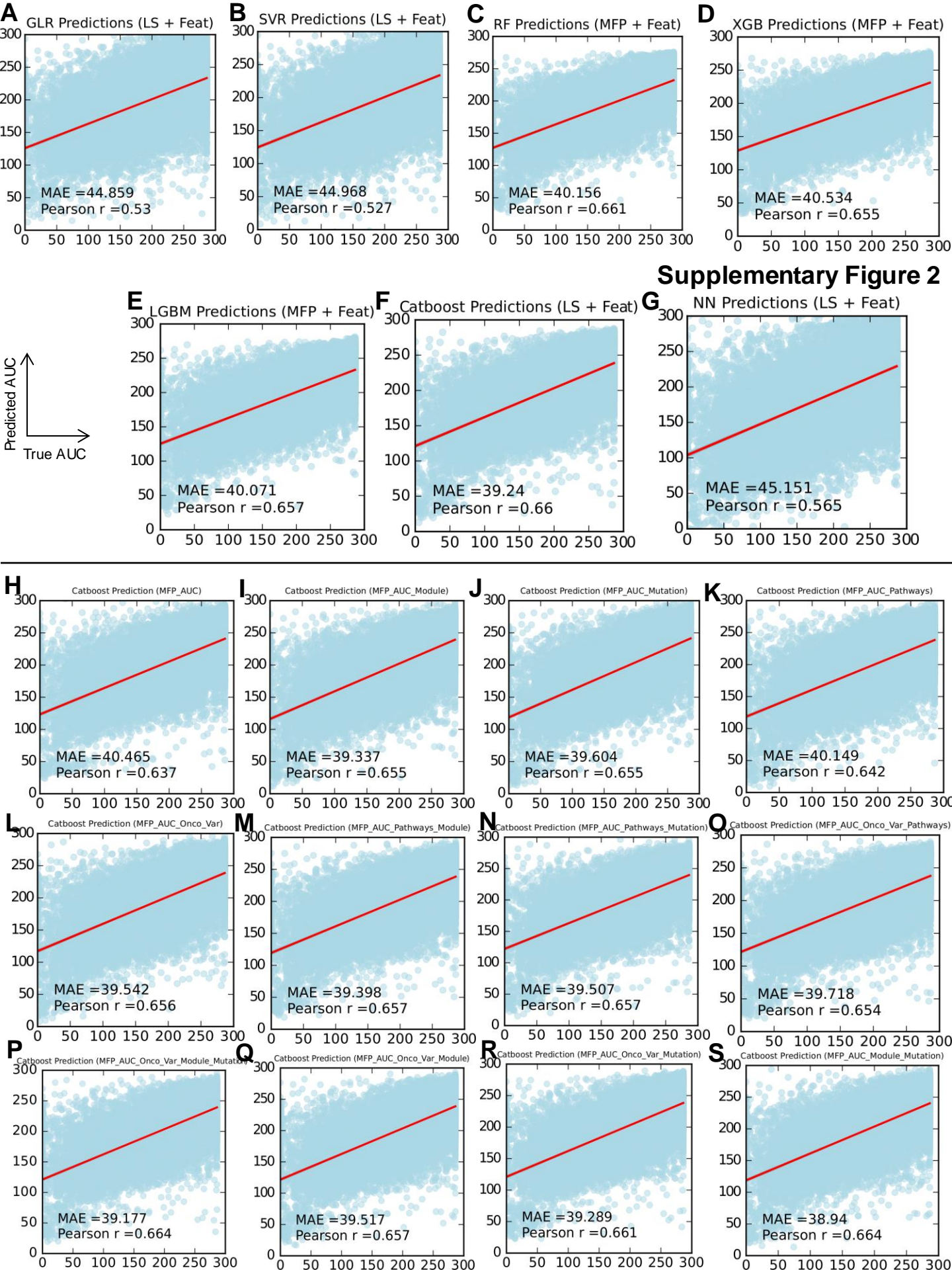
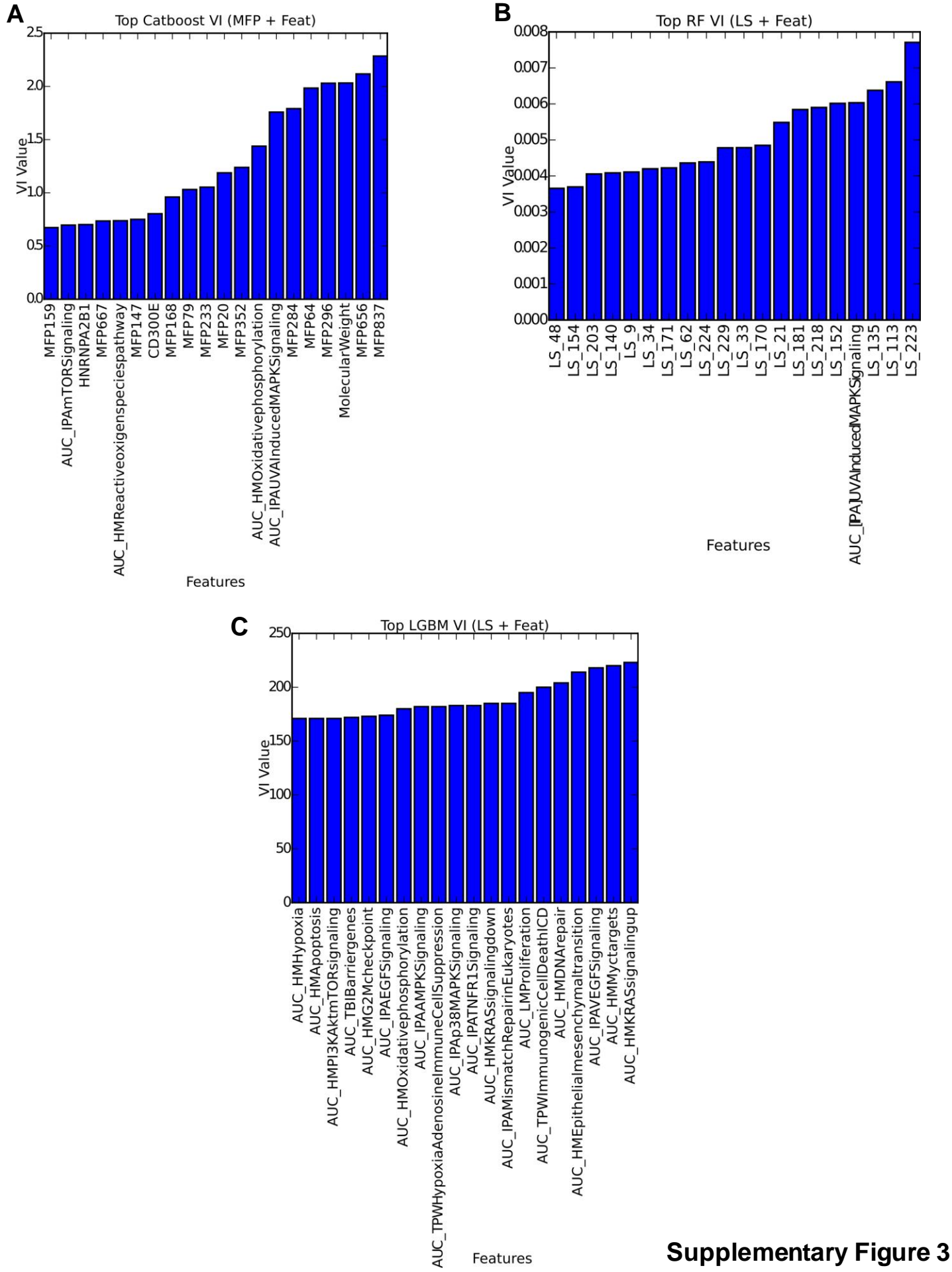


Figure 4

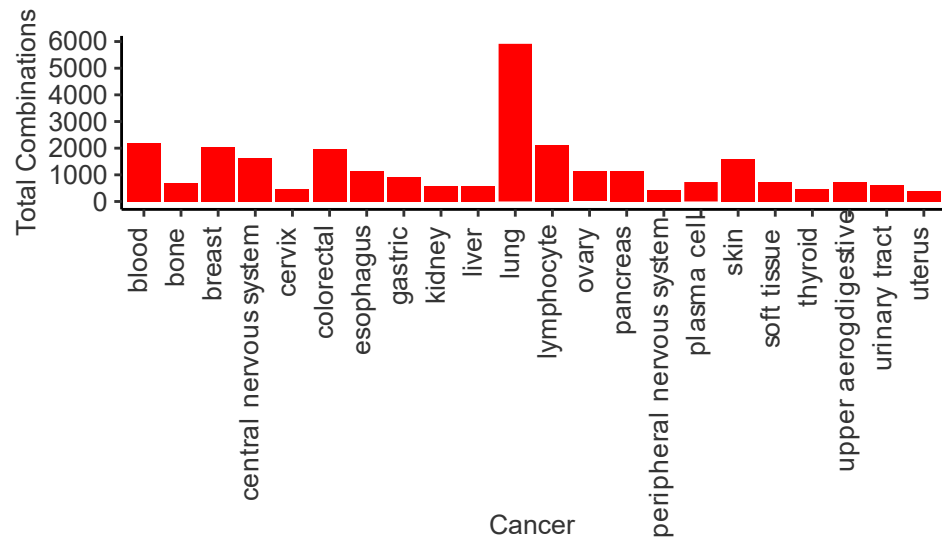
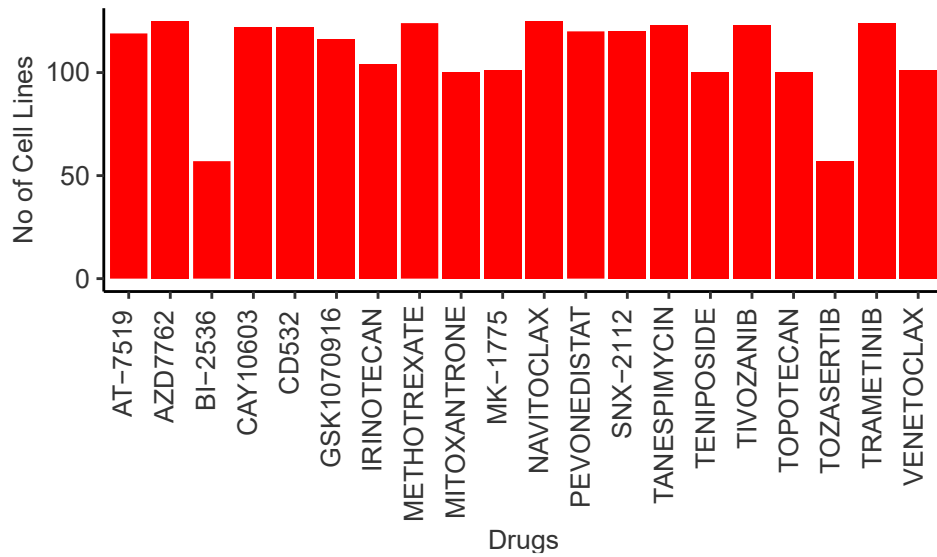


Supplementary Figure 1





Supplementary Figure 3

A**B**

| Methods | Features Used | CV (MAE) | Test (MAE) | CV (RMSE) | Test (RMSE) | CV (r2) | Test (r2) | CV Pearson (r) | Test Pearson (r) | CV Spearman (r) | Test Spearman (r) |
|--------------------------------|--------------------|-------------------------|---------------|-------------------------|---------------|------------------------|--------------|------------------------|------------------|------------------------|-------------------|
| Ridge Regressor | LS + Cell Line | 30.224 +/- 1.972 | 44.859 | 40.342 +/- 2.906 | 60.005 | 0.555 +/- 0.052 | 0.281 | 0.744 +/- 0.034 | 0.53 | 0.742 +/- 0.032 | 0.556 |
| Ridge Regressor | MFP + Cell Line | 29.925 +/- 1.917 | 43.476 | 40.103 +/- 2.891 | 57.257 | 0.559 +/- 0.052 | 0.336 | 0.747 +/- 0.034 | 0.579 | 0.745 +/- 0.033 | 0.578 |
| Linear SVR | LS + Cell Line | 30.125 +/- 1.955 | 44.968 | 40.249 +/- 2.898 | 60.366 | 0.556 +/- 0.051 | 0.278 | 0.745 +/- 0.034 | 0.527 | 0.743 +/- 0.032 | 0.557 |
| Linear SVR | MFP + Cell Line | 29.934 +/- 1.922 | 43.894 | 40.088 +/- 2.894 | 57.731 | 0.559 +/- 0.052 | 0.328 | 0.747 +/- 0.034 | 0.572 | 0.745 +/- 0.033 | 0.571 |
| RF | LS + Cell Line | 21.824 +/- 0.973 | 39.626 | 29.865 +/- 1.514 | 52.077 | 0.787 +/- 0.021 | 0.442 | 0.887 +/- 0.012 | 0.665 | 0.886 +/- 0.009 | 0.656 |
| RF | MFP + Cell Line | 20.749 +/- 1.221 | 40.156 | 28.287 +/- 1.834 | 52.528 | 0.817 +/- 0.021 | 0.437 | 0.904 +/- 0.012 | 0.661 | 0.9 +/- 0.01 | 0.649 |
| XGBoost | LS + Cell Line | 24.288 +/- 1.212 | 39.843 | 31.939 +/- 1.755 | 52.339 | 0.728 +/- 0.038 | 0.432 | 0.853 +/- 0.022 | 0.657 | 0.835 +/- 0.022 | 0.65 |
| XGBoost | MFP + Cell Line | 22.937 +/- 1.009 | 40.534 | 30.101 +/- 1.445 | 52.876 | 0.782 +/- 0.037 | 0.43 | 0.884 +/- 0.21 | 0.655 | 0.867 +/- 0.022 | 0.642 |
| LightGBM | LS + Cell Line | 21.529 +/- 1.119 | 39.497 | 28.748 +/- 1.63 | 52.091 | 0.786 +/- 0.029 | 0.441 | 0.887 +/- 0.016 | 0.664 | 0.872 +/- 0.016 | 0.655 |
| LightGBM | MFP + Cell Line | 23.232 +/- 1.079 | 40.071 | 31.123 +/- 1.651 | 52.554 | 0.752 +/- 0.037 | 0.432 | 0.867 +/- 0.021 | 0.657 | 0.854 +/- 0.021 | 0.644 |
| Catboost | LS + Cell Line | 26.35 +/- 1.559 | 39.24 | 37.256 +/- 2.51 | 52.449 | 0.623 +/- 0.044 | 0.436 | 0.789 +/- 0.028 | 0.66 | 0.789 +/- 0.024 | 0.655 |
| Catboost | MFP + Cell Line | 21.02 +/- 1.48 | 39.044 | 31.919 +/- 2.432 | 52.032 | 0.725 +/- 0.036 | 0.439 | 0.851 +/- 0.021 | 0.663 | 0.85 +/- 0.017 | 0.652 |
| DNN | LS + Cell Line | 23.511 +/- 1.431 | 45.151 | 32.091 +/- 2.1 | 58.69 | 0.718 +/- 0.032 | 0.32 | 0.847 +/- 0.019 | 0.565 | 0.836 +/- 0.018 | 0.552 |
| DNN | MFP + Cell Line | 18.189 +/- 1.05 | 42.647 | 25.422 +/- 1.582 | 56.177 | 0.824 +/- 0.018 | 0.356 | 0.908 +/- 0.01 | 0.596 | 0.899 +/- 0.01 | 0.587 |
| Graph Attention Network + FFNN | SMILES + Cell Line | | | | | | | | | | |
| CNN + FFNN | SMILES + Cell Line | | | | | | | | | | |
| LSTM + FFNN | SMILES + Cell Line | | | | | | | | | | |

Table 1: Performance comparison of different machine learning models

| Features Used | CV (MAE) | Test (MAE) | CV (RMSE) | Test (RMSE) | CV (r2) | Test (r2) | CV (Pearson r) | Test (Pearson r) | CV (Spearman r) | Test (Spearman r) |
|--|-------------------------|---------------|-------------------------|---------------|------------------------|--------------|------------------------|------------------|------------------------|-------------------|
| MFP + AUC | 27.458 +/- 1.207 | 40.465 | 38.85 +/- 1.856 | 54.114 | 0.591 +/- 0.054 | 0.406 | 0.768 +/- 0.035 | 0.637 | 0.774 +/- 0.028 | 0.626 |
| MFP + Onco +Var + AUC | 20.862 +/- 1.39 | 39.542 | 31.731 +/- 2.305 | 52.46 | 0.728 +/- 0.037 | 0.43 | 0.853 +/- 0.022 | 0.656 | 0.852 +/- 0.017 | 0.642 |
| MFP + Pathw ays + AUC | 19.556 +/- 1.005 | 40.149 | 30.409 +/- 1.866 | 53.287 | 0.751 +/- 0.03 | 0.412 | 0.866 +/- 0.017 | 0.642 | 0.867 +/- 0.014 | 0.628 |
| MFP + Modules + AUC | 20.996 +/- 1.183 | 39.337 | 31.81 +/- 2.001 | 52.509 | 0.728 +/- 0.033 | 0.429 | 0.853 +/- 0.019 | 0.655 | 0.854 +/- 0.015 | 0.644 |
| MFP + Mutations + AUC | 22.004 +/- 1.185 | 39.604 | 32.911 +/- 1.944 | 52.854 | 0.709 +/- 0.038 | 0.429 | 0.842 +/- 0.022 | 0.655 | 0.843 +/- 0.018 | 0.64 |
| MFP + AUC + Pathw ays + Modules | 25.35 +/- 1.324 | 39.398 | 36.136 +/- 2.137 | 52.482 | 0.646 +/- 0.044 | 0.432 | 0.803 +/- 0.027 | 0.657 | 0.804 +/- 0.023 | 0.647 |
| MFP + AUC + Pathw ays + Mutations | 25.59 +/- 1.377 | 39.507 | 36.397 +/- 2.138 | 52.792 | 0.641 +/- 0.044 | 0.431 | 0.8 +/- 0.027 | 0.657 | 0.801 +/- 0.023 | 0.648 |
| MFP + AUC + Pathw ays + Onco +Var | 24.765 +/- 1.473 | 39.718 | 35.574 +/- 2.351 | 52.744 | 0.658 +/- 0.042 | 0.427 | 0.81 +/- 0.026 | 0.654 | 0.81 +/- 0.021 | 0.643 |
| MFP + AUC + Modules + Mutations | 25.858 +/- 1.415 | 38.94 | 36.688 +/- 2.168 | 52.229 | 0.635 +/- 0.045 | 0.441 | 0.796 +/- 0.028 | 0.664 | 0.797 +/- 0.023 | 0.656 |
| MFP + AUC + Modules + Onco + Var | 24.768 +/- 1.405 | 39.517 | 35.575 +/- 2.239 | 52.653 | 0.657 +/- 0.042 | 0.432 | 0.81 +/- 0.026 | 0.657 | 0.809 +/- 0.021 | 0.648 |
| MFP + AUC + Mutations + Onco + Var | 24.749 +/- 1.477 | 39.289 | 35.578 +/- 2.396 | 52.387 | 0.657 +/- 0.044 | 0.437 | 0.81 +/- 0.027 | 0.661 | 0.81 +/- 0.022 | 0.651 |
| MFP + AUC + Mutations + Modules + Onco + Var | 24.743 +/- 1.51 | 39.177 | 35.559 +/- 2.445 | 52.323 | 0.657 +/- 0.043 | 0.44 | 0.81 +/- 0.026 | 0.664 | 0.81 +/- 0.021 | 0.655 |
| MFP + AUC + Mutations + Pathw ays + Onco + Var | 25.675 +/- 1.556 | 39.803 | 36.518 +/- 2.458 | 53.268 | 0.638 +/- 0.045 | 0.419 | 0.798 +/- 0.028 | 0.647 | 0.797 +/- 0.023 | 0.638 |
| MFP + AUC + Modules + Pathw ays + Onco + Var | 24.718 +/- 1.461 | 39.422 | 35.531 +/- 2.354 | 52.581 | 0.658 +/- 0.046 | 0.433 | 0.811 +/- 0.028 | 0.658 | 0.81 +/- 0.023 | 0.648 |
| MFP + AUC + Mutations + Modules + Pathw ays | 27.653 +/- 1.425 | 39.403 | 38.459 +/- 2.177 | 52.601 | 0.599 +/- 0.48 | 0.436 | 0.774 +/- 0.03 | 0.66 | 0.775 +/- 0.026 | 0.652 |

Supplementary Table 1: Ablation study of different feature sets used for the optimal model construction.

Background:

1. 699 cell lines from CCLE with 19,177 gene expression profiles. This information was downloaded from Cancer Dependency Map Public 21Q3. We filtered the original data consisting of 1,377 cell lines → 699 cell lines to keep only those cell lines with COSMIC ids to match the drug-response dataset from GDSC portal. We include 10 features related to cell line metadata including age, gender, type, name of cell line etc. for the cancer cell lines.
2. Genes of interest include genes which are part of several inflammasome/inflammatory cell death pathways including:
 - a) Reactome inflammasome, b) KEGG nod like signaling pathway, c) GO biological process inflammasome complex, d) Reactome pyroptosis, e) Necroptotic signaling pathway from GO, f) PANoptosis pathway, g) Immunogenic cell death pathway (ICR) → total of 170 (167 of which are present in the 19,177 genes)
3. Seven pathways considered for inflammatory cell death as mentioned above.
4. We got the mutation profile and copy number variation profile for the 170 genes of interest. The mutation profile and copy number variation was obtained from Harmonizome database from Mayan lab.
5. We removed genes which had no variation in expression, mutation or CNV across cell lines including: Mutation_ERBIN, Mutation_NLRP2B, Mutation_STMP1, Mutation_PYDC2, Mutation_CARD18, CNV_ERBIN, CNV_NLRP2B, CNV_STMP1
6. Total Features include:
 - a) Cell Line Features (10); b) Pathways (7); c) Expression (167); d) Mutation (162); e) CNV (164)
7. The dose response information is obtained from GDSC portal. It contains drug response for a particular cell line with prediction variables: IC50score and Z-score. In the GDSC portal, the Z-score is used to determine sensitive and resistant drugs with cut-offs of -2 and 2 respectively. We use the $-\log_{10}(\text{IC50score})$ as our y variable (term to predict). It contains 398 unique drugs and 989 cell lines.
8. The viability information is also obtained from GDSC portal. It contains cell viability at different dosage levels. **Currently not used.**

Methods:

1. The gene expression profile for each cell line is quantile normalized (19,177 genes), scaled and converted to z-scores.
2. T-sne plots are made for 699 cell lines with cancer types based on expression profiles as well as the features we have engineered.
3. A complexheatmap visualization of the different features used and how they look across the 699 cancer cell lines.

Background:

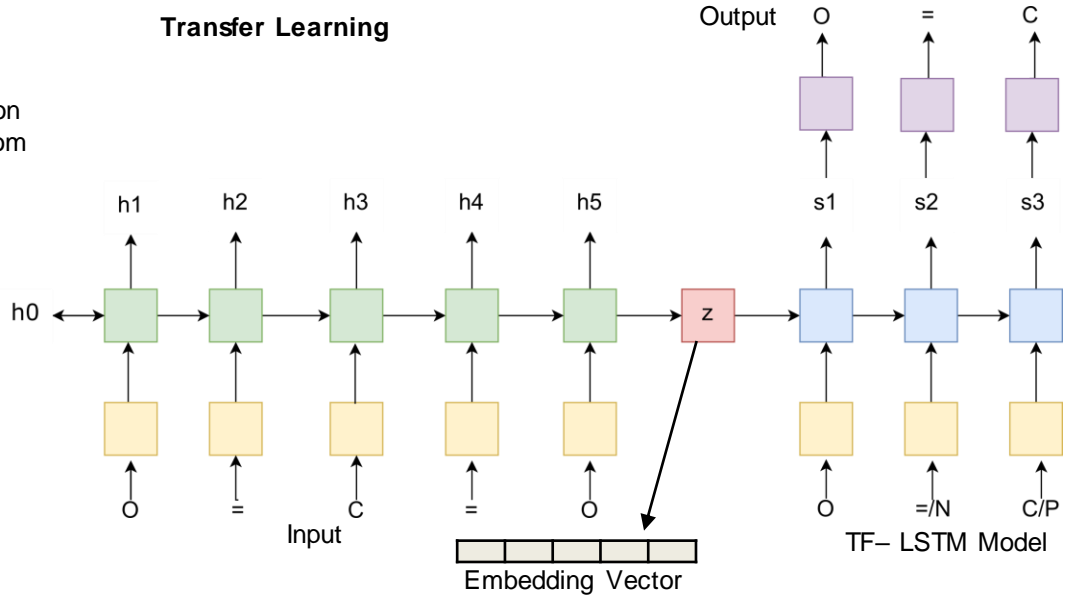
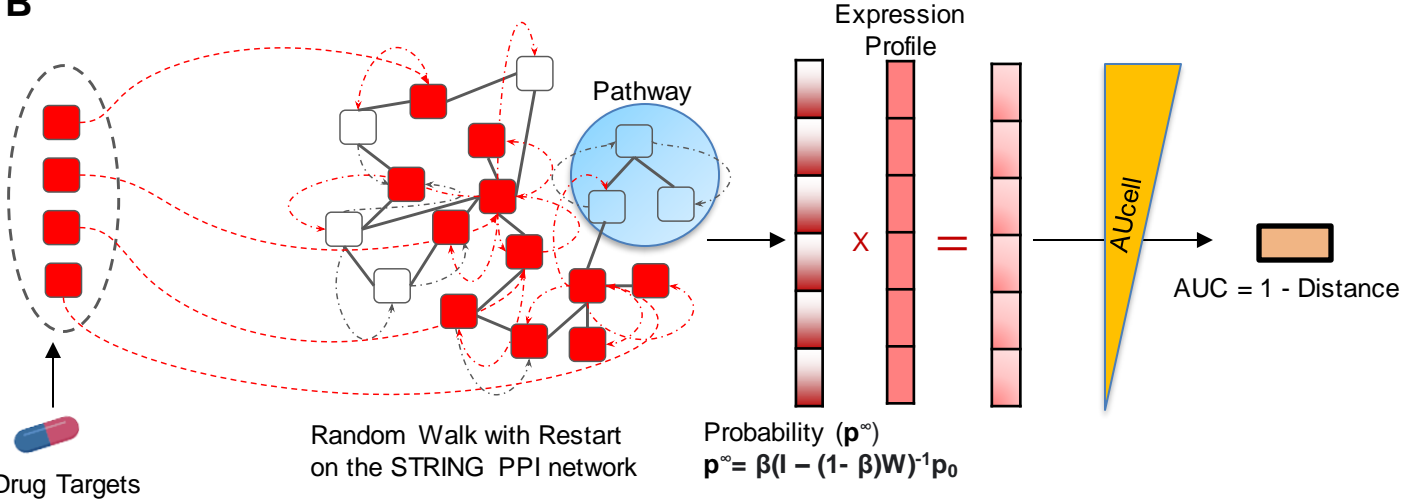
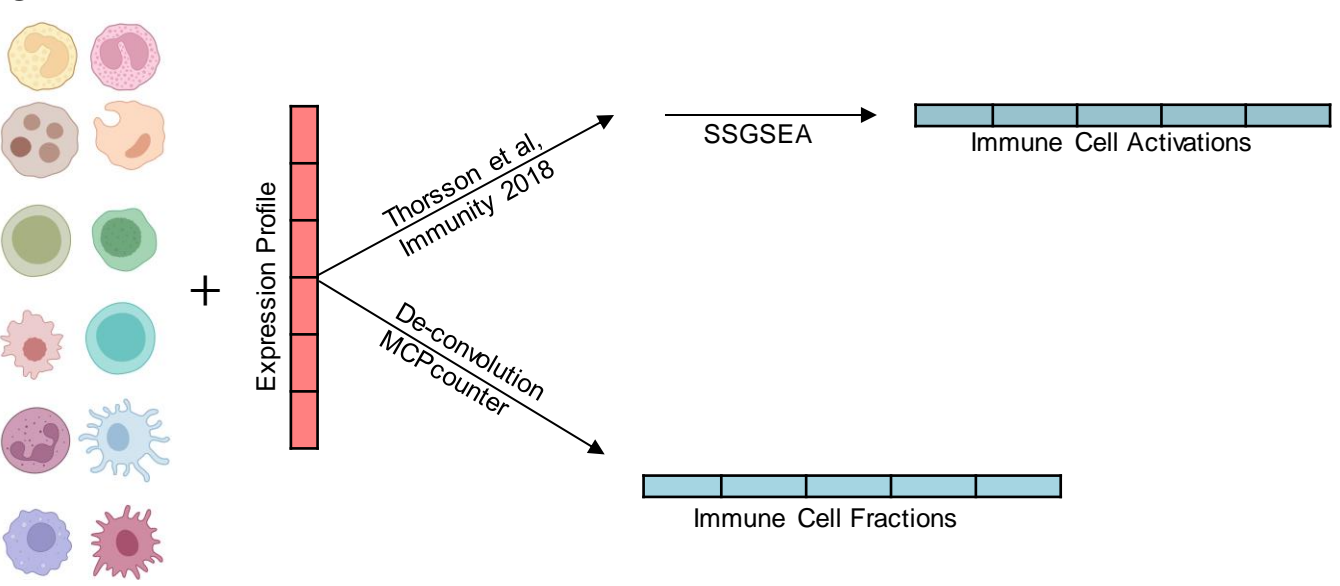
1. We obtained drug information for 399 drugs from GDSC portal and the cell line information for 699 cell lines from CCLE.
2. We curated and added each drug's target from resources like Drug Bank, its SMILES, Inchikey, Molecular weight, molecular formula representations from PubChem, ChEMBL and NCBI API.
3. There are a total of 373 unique drugs available from this dataset.
4. We find a total of 379,005 drug-cell line sensitivity profiles from GDSC portal (GDSC 1 & 2 combined).
5. After filtering, removing duplicates and combined drug plus cell line features, we end up with a total of 151,636 drug-cell line combination profiles consisting of 253 drugs and 693 cell lines.

Method:

1. We perform inner joins with drug profiles and cell line profiles to get drug-cell features used for the predictive models.
2. For each drug, we have its known targets. We perform a random walk with restart to get a random walk score for each drug using 'diffusr' package in R. It provides the probability of a drug impacting all the genes in our cell line's expression profile. This affinity is based on topology and doesn't consider individual gene's expression in a particular cell line. To get a cell line specific affinity, we multiply the random walk scores with corresponding genes' expression levels. Using this information and the gene set for each pathway, we estimate the distance of a drug from each of the 7 inflammasome related pathways using 'AUCell' package in R.
3. For each drug, we can estimate its molecular fingerprint representation using the RDKit package in python.
4. For each drug, we can also estimate its representation using a transfer-learning based approach. We pass the drug SMILES to a TF-LSTM autoencoder trained on over 2 million drug SMILES and obtain an embedding vector representation of the drug which can be fed to a machine learning algorithm.
5. All the cell line information (expression, mutation, copy number) + drug information (vector representation) + pathway enrichments (pathway activation based on expression) and distance of a pathway from a drug's known targets are used as features to predict the drug response.
6. We used a variety of machine learning methods including:
 1. Linear Regression
 2. Elastic Net
 3. SVM
 4. Random Forests
 5. Xgboost
 6. LightGBM
 7. Feed Forward Neural Networks (DNN)
 8. Graph Attention Network (GAT) + DNN
 9. Convolutional Neural Network (CNN) + DNN
 10. Long-Short Term Memory (LSTM) + DNN
7. We built different models on training set (565 cell lines) and test on a complete independent test set (128 cell lines on which no training is performed)
8. We highlight the variable importance (i.e. the top features) driving the prediction for each of these different machine learning models and performance is highlighted in Table.

A**Transfer Learning**DeepRepurpose:

- TF-LSTM model trained on 2.5 million compounds from PubChem + MOSES.
- 96.7% reconstructed compounds are valid.
- μ_{error} per sample is 0.001

**B****Pathway Distance Estimation****C****Immune Profiling****Unused Data**

Research Article Checklist should be placed after presentation

Research Article Checklist

First author and coauthors are responsible for ensuring that: 1) all statements are factually accurate; 2) references to seminal publications and Kanneganti lab publications are used consistently and wherever appropriate (refer to the reference lists); 3) the correct mouse source and information is provided.

- ☐ **Title:** informative, concise, and includes keywords
- ☐ **Author List:** all authors are included; names are spelled correctly
- ☐ **Keywords:** searchability; citability
- ☐ **Abstract:** Overall summary (provides key background, purpose, and findings)
 - Statements are factually accurate
 - Concise
 - Followed example
- ☐ **Outline or Full Article:**
 - Introduction: 3 paragraphs; key concepts and background
 - Statements are factually accurate
 - **References** (Key refs from Kanneganti lab and Others ref lists)
 - Discussion: 3 paragraphs; concise summary of info presented; contextualized; highlights any clinical relevance
 - Followed examples
- ☐ **Figures:**
 - Concise title and legend
 - Labels and enough detail to be understood without text
 - Followed lab template (correct formatting, no shadows, no typos)
 - No image duplications

I confirm that I have checked for the above points during my review and provided the necessary feedback to the first author.

Name

Date