$\mathbf{1}^{st}$ QTB PBR Hack'a'thing

Soldering for and by beginners.

March 2–4, 2016

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1 PBR Hack'a'thing Projects

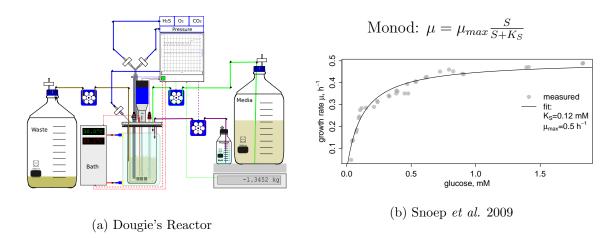


Figure 1: Bioreactors

$$\frac{dX}{dt} = (\mu_{ab} - \phi)X$$

$$\frac{dS}{dt} = \phi(S_{in} - S) - (\mu_{ab} + \mu_{cd})X$$

$$\frac{datp}{dt} = (n_{cd}\mu_{cd} - n_{ab}\mu_{ab} - \mu_m)\frac{C_c}{V_c} - \mu_{ab}atp$$

$$\frac{dG}{dt} = k_L a(G_{in}^* - G) - n_g \mu_{cd}X$$

$$adp = a_{tot} - atp$$

$$\mu_{ab} \equiv f(S, atp)$$

$$\mu_{cd} \equiv f(S, G, adp)$$

$$\mu_m \equiv f(S, ROS)n_{cd}, n_{ab} \equiv f(S)$$

Figure 2: Growth in Continuous Culture: liquid and gas flux equations

Resources - Material and Ideas

Designs & Instructions:

• Arduino Playground: Wiki for Arduino-based Designs

• Instructables: Diverse DIY Designs

• Thingiverse: 3D Printer Designs

Shops:

• Conrad: online and offline shop, DE

• Elecrow: online shop, China

• Adafruit: online shop, USA

• Fasttech: online shop, China

1.1 Gas Flux: Gas'o'meter

Project: Extend existing setup (co2meter's O_2 and CO_2 sensors with Sainsmart's Arduino Mega+Touch screen) by our brand new digital mass flow meter Aalborg XFM; write calibration routines for all sensors; build water trap and tubing to connect to the PSI or our DIY reactor and increase gas transfer (smaller bubbles) and decrease overall gas flow so that we can measure cellular activity. Perhaps add valve control to measure multiple reactors in series.

- 1. **build** water trap, tubing path from reactor, and casing for sensors and Arduino; **build** improved gassing system (glas blowers!) to allow lower flow
- 2. **code** sensor calibration routines via touch-screen (use PSI gas mixing system)
- 3. **build** & **code** interface to Aalborg XFM digital mass flow meter: connect the Aalborg's RS 485 interface to Arduino hardware serial Tx3/Rx3, and Ground
- 4. **build** & **code** valve control to measure several reactors; connect via Arduino software serial connections; perhaps attach to PSI Multicultivator

Resources:

- Sensor manuals in code/manuals/offgas/ at the PBR git:
 Manual-CM-0201-UV-Flux-Oxygen.pdf, Manual-GSS-Sensors.pdf, and
 A_XFM_Manual_TD0701M[...].pdf
- Code in code/offgas/arduino/ at the PBR git

- Existing setup: available
- Aalborg XFM, with RS 485 interface: available
- Valve system for gas tubing, controllable via serial interface: obtain

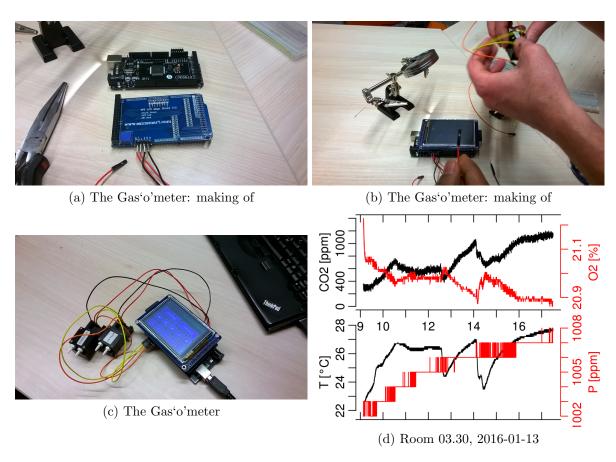


Figure 3: Gas Measurement Module.

1.2 Light Flux: Spectrometer

Project: Simple spectrometric measuring tool based on AvaSpec-Mini2048l-V25

- 1. Basic: Connect to Rasperry Pi, using drivers provides by Avantes; **code** simple interface with display and/or recording functions
- 2. Advanced: use LED for absorbance, reflectance, or fluorescence measurements; build light paths and perhaps a reactor probe for online recording

Resources:

- AvaSpec-Mini data sheet in code/manuals/light/ at the PBR git
- The AS5216 microprocessor board a lib file for Raspberry Pi model 1 B+ is available in code/light/

- AvaSpec-Mini2048l-V25, Minispectrometer: available Mini spectrometer, 2048 Large pixels, grating-MN0600-0.50 (350-885nm), OSC, 25µm slit, USB2 interface, AvaSoft-Basic
- Fiber optic cables, VIS/NIR: 1 m, 200 μm VIS/NIR and 1m, 600 μm: available SMA terminations, metal protection sleeves
- Raspberry Pi Version 1 Model B+: obtain
- LED system: use PSI LEDs or obtain
- Reactor probe: **build** together with fine mechanics or glas blowers



Figure 4: Spectrometer Module.

1.3 Liquid Flux: Continuous Culture & Turbidostat

Project: build a module consisting of media and waste bottles, a reactor vessel, peristaltic pump(s), and a scale; pump and scale are controlled *via* serial interfaces from an Arduino+Touchscreen. The flow rate is controlled *via* the pump motor speed and recorded *via* the scale; the flow rate is recorded or can be set after a setup-specific (tubing) calibration routine

- 1. **build** a simple reactor vessel (Schott bottles) with liquid media flow, from media bottles through reactor vessel and out to waste bottle
- 2. **code**: calibration routine for the weight sensor module
- 3. **code**: analog control of peristaltic motor speed and recording of weight loss and/or gain to record mass flow (g/min)
- 4. **code**: routine to calibrate pump speed to weight loss/gain for a specific setup; store calibration on SD card, which allows to also set pump speed in g/min, or if provided with a culture volume, as culture dilution rate (h^{-1})
- 5. build & code: combine with 1.2 to make turbidostatic control
- 6. build: add gassing system of project 1.1 to make a first simple bioreactor

Resources:

- Arduino tutorials for Adafruit peristaltic pump and general DC motor control and Adafruit motor shield
- Arduino library for Electrow weight sensor kit 3 kg
- HX711 24-bit analog-to-digital converter (ADC) for load cells

Materials:

- Bottles, screw caps with inlet/outlet openings, and tubing: available & obtain!
- Scale ordered

fancy: Mettler Toledo, PBK785-3XS/f

cheap: Elecrow Weight Sensor kit 3kg for Arduino \leftarrow ordered

• Peristaltic pumps - ordered

fancy: Longer Pump LP-BT100-2J, DG-2(10)

cheap: Ismatec Ecoline VC-MS/CA4-12

cheaper: Welco WPM or cheapest: Adafruit \leftarrow ordered

• Toshiba TA7291P Bridge Driver - ordered

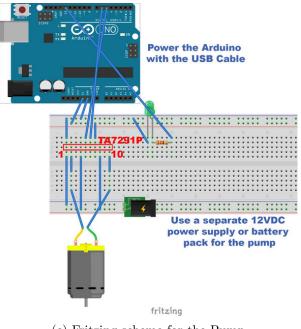
• Sainsmart Arduino Mega + Touchscreen - ordered



(a) Elecrow Scale Module, 3 kg



(b) Adafruit Peristaltic Pump



(c) Fritzing scheme for the Pump

Figure 5: Continuous Culture Module. The electrow scale module is based on the HX711 load-cell amplifier (24-bit analog- to-digital converter) and connected to 5V, Gnd and two analog pins of the Arduino and comes with an Arduino library. An Adafruit peristaltic pump (12 v) is run by a "geared down DC motor" and requires a Toshiba TA7291P Bridge Driver (0-20V 1A; 2A peak) to control pump direction and speed via Arduino's PWM pins, see the instructable tutorial.

1.4 The Server

Project: a master software running on a (detachable) linux desktop that synchronizes and speaks via a comon interface to all Arduino and Raspberry Pi modules; the modules themselves can interpret get, set and act impulses (use arguments only when absolutely necessary).

During an initialization the server may inquire what an attached module provides (*via* data IDs and SI units, meaningful time resolution) and handle it automatically. Variable higher order control or processing logics can be built using defined data and control IDs.

- 1. **build** combine of gas (1.1), liquid (1.3) and light (1.2) modules into a bioreactor
- 2. **code**: master program to synchronize and record data from the three modules
- 3. code: combine e.g. 1.2 & 1.3 to implement turbidostat control

- setTime(time_t t): sets the current master time to all modules
- get(..., time_t t): get all values, currently available (with a time stamp), or from a previous time t
- act(..., time_t t): act (switch on and off, set to a specific value), now or at future time t

1.5 Heat Flux: Water Bath Thermostat

Project: build a water bath for growth vessels, control T, read-out energy required for maintaining constant T and estimate the amount of heat withdrawn or administered

1.

- Jacketed reactor vessel: build or obtain
- Julabo water bath, e.g. F25-ME
- Arduino and/or Raspberry Pi

1.6 The Kaiten Eppi: Automated Sampling Device

Projects: build sterile and automated sampling device; using a controllable syringe pump, sampling into the Kaiten Eppi (automated: pump sample into tubes, potentially pre-filled with chemicals, vortex, and transport them into liquid N_2 or other storage containers)

Materials:

• Sterile sampling device by HHU glas blowers: available

• Syringe pump: obtain

• Kaiten Eppi: build

• Sainsmart Arduino Mega & Touchscreen: obtain

1.7 Single Cell Biology: Microfluidic Device

Project: Basic microfluidics and live-cell imaging device; scratch growth chambers and liquid flow channels into microscope slide; attach 2-3 pumps; and control via arduino/screen

Resources:

• Tsuda et al. PLoS ONE 2015: 3D Printed 'Plug and Play' Millifluidic

Materials:

• Ilka's lab microscope: available

• Microscopy slides: available

• 2–3 peristaltic pumps for microfluidics: obtain

• Sainsmart's Arduino Mega + Touchscreen: obtain

2 Program

2.1 Day 1 <12:00 : Building Bioreactors

Talks, 30-60 min:

- Rob's DIY Reactor The Beginnings: NinjaPBR Arduino-controlled mini PBR
- Dougie's DIY Reactor 20 yrs Later
- Avantes Spectrometry: Spectrometry applications, incl. NIR for metabolite measurements and OD; software interface to Avantes spectrometers
- CellDeg Optimizing Photosynthetic Growth: Introduction to CellDeg's 2.5 k Euro algal growth setup (30 g/L cyano biomass in three days)

2.2 Day 1 > 13:00: Hack'a'thing I

- Introduction to the Gas'o'meter: connecting sensors with Arduino, making an autonomous measurement device via Sainsmart's Touch Screen
- Introduction to Rob's reactor: complete setup for photosynthetic growth
- Self-organizing into teams: lab hardware (tubing etc.), control hardware (soldering etc.), software and/or by by projects (1.1–1.7)

2.3 Day 2: Hack'a'thing II

- Hardware I: soldering, tubing
- Software I: probe/sensor/pump ⇔ arduino/raspi interfaces
- Visit HHU's fine mechanics and glas blower work-shops, place orders for stuff missing for above goals

2.4 Day 3: Hack'a'thing III

- Hardware II: Integrate projects 1.1,1.2&1.3 into a simple DIY reactor and/or with PSI FMT150 or Multicultivator
- Software II: arduino/raspi ⇔ master/server interface Standard data formats and interfaces
- Brain storming: relation of data and models and beer

2.5 Participants & Teams

$_{\mathrm{type}}$	name	e-mail	\mathbf{team}
	Sascha Wigger	sascha.wigger@student.htw-berlin.de	
	Nicolas Schmelling	nicolas.schmelling@uni-duesseldorf.de	
	Fiona Moejes	fionamoejes@hotmail.com	
	Stefano Magni	stefano.magni@uni-duesseldorf.de	
	Philipp Norf	philipp.norf@uni-duesseldorf.de	
	Elahe Radmaneshfar	radmaneshfar@gmail.com	
	Anna Matuszynska	anna.matuszynska@hhu.de	
	Suraj Sharma	suraj.sharma@uni-duesseldorf.de	
	Simon Schliesky	simon.schliesky@hhu.de	
	Antonella Succurro	succurro@hhu.de	
	Robert Lehmann	lehmann.rob@googlemail.com	
	Dougie Murray	dougie@ttck.keio.ac.jp	
	Oliver Ebenhöh	oliver.ebenhoeh@hhu.de	
	Rainer Machné	machne@hhu.de	

Table 1: should we have skill badges, https://www.adafruit.com/products/2857?q=skill%20badge&p=0& or 3D-printed awards, like the "red soldering iron", "the blue tube", "the green chip"?

3 Outlook: 2nd QTB PBR Hack'a'thing

3.1 Growth Dynamics: Photobioreactors in Research

Talks, 30-60 min:

Nir Keren, Hellingwerf, Jan Cerveny, Dougie Murray, something microfluidics?

3.2 Single Cell Dynamics: Microfluidic Devices

Integrate project 1.7 with the simple microscope in Ilka's lab, or a more advanced system (CAi?)

3.3 Omics: Sterile and Automated Sampling Devices

Proper sampling for high-throughput data acquisition (mass spectrometry, sequencing) - the Kaiten Eppi