# $\mathbf{1}^{st}$ QTB PBR Hack'a'thing

# Soldering for and by beginners.

# March 2–4, 2016

# **Contents**

1	PBF	PBR Hack'a'thing Projects		
	1.1	Gas Flux: Gas'o'meter	5	
	1.2	Light Flux: Spectrometer	7	
	1.3	Liquid Flux: Continuous Culture & Turbidostat	8	
	1.4	The Server	10	
	1.5	Heat Flux: Water Bath Thermostat	11	
	1.6	The Kaiten Eppi: Automated Sampling Device	12	
	1.7	Single Cell Biology: Microfluidic Device	13	
2	Program			
	2.1	Day 1 <12:00 : Building Bioreactors	14	
	2.2	Day 1 >13:00 : Hack'a'thing I		
	2.3	Day 2 : Hack'a'thing II		
	2.4	Day 3: Hack'a'thing III	14	
	2.5	Participants & Teams	15	
3 Outlook:		look: 2 <sup>nd</sup> QTB PBR Hack'a'thing	15	
	3.1	Growth Dynamics: Photobioreactors in Research	15	
	3.2	Single Cell Dynamics: Microfluidic Devices		
	3.3	Omics: Sterile and Automated Sampling Devices		

# 1 PBR Hack'a'thing Projects

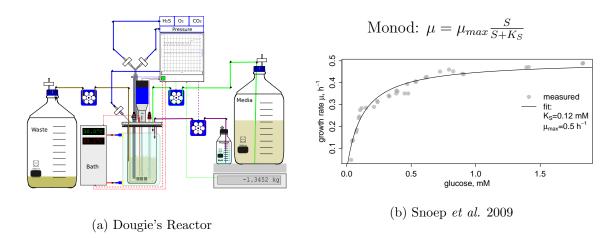


Figure 1: Bioreactors

$$\frac{dX}{dt} = (\mu_{ab} - \phi)X$$

$$\frac{dS}{dt} = \phi(S_{in} - S) - (\mu_{ab} + \mu_{cd})X$$

$$\frac{datp}{dt} = (n_{cd}\mu_{cd} - n_{ab}\mu_{ab} - \mu_m)\frac{C_c}{V_c} - \mu_{ab}atp$$

$$\frac{dG}{dt} = k_L a(G_{in}^* - G) - n_g \mu_{cd}X$$

$$adp = a_{tot} - atp$$

$$\mu_{ab} \equiv f(S, atp)$$

$$\mu_{cd} \equiv f(S, G, adp)$$

$$\mu_m \equiv f(S, ROS)n_{cd}, n_{ab} \equiv f(S)$$

Figure 2: Growth in Continuous Culture: liquid and gas flux equations

### Resources - Materials, Designs and Ideas

#### **DIY Bioreactors:**

- Open source DIY bioreactor and its code on github
- Github project for Arduino-controlled bioreactor
- Onlinebioreactor: Arduino wiring schematics and source code for a temperature and pH measurements of a bioreactor
- An Algae Bioreactor from Recycled Water Bottles
- Google Group: DIY Bioreactor and Bioreactor controller
- The Jun lab's Mother Machine
- Pelling lab's CO2 incubator for mammalian cell culture
- Biomonstaaar: PBR instructable, and spacegambit project proposal
- Openbioreacteurs: DIY algae reactors

#### 3D Printed Labware and Bioreactors:

- Tsuda et al. PLoS ONE 2015: 3D Printed 'Plug and Play' Millifluidic Devices
- Lücking et al. Eng. Life Sci. 2015: 3D printed labware, incl. "well plate with different baffle geometries, shake flask cap with built-in luer connections, and filter holder for an in-house developed membrane reactor system"; also see paper by Büchs group on baffle geometries
- Baden *et al.* PLoS ONE 2015: Open Labware: 3-D Printing Your Own Lab Equipment

#### **Designs & Instructions:**

- Arduino Playground: Wiki for Arduino-based Designs
- Instructables: Diverse DIY Designs
- Thingiverse: 3D Printer Designs

### Shops:

- Conrad: online and offline shop, DE
- Electrow: online shop, China

 $\bullet\,$  Adafruit: online shop, USA

 $\bullet\,$  Fasttech: online shop, China

### 1.1 Gas Flux: Gas'o'meter

**Project:** Extend existing setup (co2meter's  $O_2$  and  $CO_2$  sensors with Sainsmart's Arduino Mega+Touch screen) by our brand new digital mass flow meter Aalborg XFM; write calibration routines for all sensors; build water trap and tubing to connect to the PSI or our DIY reactor and increase gas transfer (smaller bubbles) and decrease overall gas flow so that we can measure cellular activity. Perhaps add valve control to measure multiple reactors in series.

- 1. **build** water trap, tubing path from reactor, and casing for sensors and Arduino; **build** improved gassing system (glas blowers!) to allow lower flow
- 2. **code** sensor calibration routines via touch-screen (use PSI gas mixing system)
- 3. **build** & **code** interface to Aalborg XFM digital mass flow meter: connect the Aalborg's RS 485 interface to Arduino hardware serial Tx3/Rx3, and Ground
- 4. **build** & **code** valve control to measure several reactors; connect via Arduino software serial connections; perhaps attach to PSI Multicultivator

#### **Resources:**

- Sensor manuals in code/manuals/offgas/ at the PBR git:
   Manual-CM-0201-UV-Flux-Oxygen.pdf, Manual-GSS-Sensors.pdf, and
   A\_XFM\_Manual\_TD0701M[...].pdf
- Code in code/offgas/arduino/ at the PBR git

- Existing setup: available
- Aalborg XFM, with RS 485 interface: available
- Valve system for gas tubing, controllable via serial interface: obtain

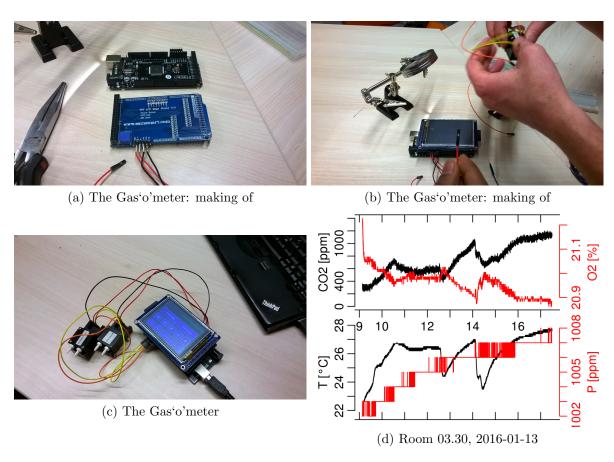


Figure 3: Gas Measurement Module.

### 1.2 Light Flux: Spectrometer

**Project:** Simple spectrometric measuring tool based on AvaSpec-Mini2048l-V25

- 1. Basic: Connect to Rasperry Pi, using drivers provides by Avantes; **code** simple interface with display and/or recording functions
- 2. Advanced: use LED for absorbance, reflectance, or fluorescence measurements; build light paths and perhaps a reactor probe for online recording

#### **Resources:**

- AvaSpec-Mini data sheet in code/manuals/light/ at the PBR git
- The AS5216 microprocessor board a lib file for Raspberry Pi model 1 B+ is available in code/light/

- AvaSpec-Mini2048l-V25, Minispectrometer: available Mini spectrometer, 2048 Large pixels, grating-MN0600-0.50 (350-885nm), OSC, 25µm slit, USB2 interface, AvaSoft-Basic
- Fiber optic cables, VIS/NIR: 1 m, 200 μm VIS/NIR and 1m, 600 μm: available SMA terminations, metal protection sleeves
- Raspberry Pi Version 1 Model B+: obtain
- LED system: use PSI LEDs or obtain
- Reactor probe: **build** together with fine mechanics or glas blowers



Figure 4: Spectrometer Module.

### 1.3 Liquid Flux: Continuous Culture & Turbidostat

**Project:** build a module consisting of media and waste bottles, a reactor vessel, peristaltic pump(s), and a scale; pump and scale are controlled *via* serial interfaces from an Arduino+Touchscreen. The flow rate is controlled *via* the pump motor speed and recorded *via* the scale; the flow rate is recorded or can be set after a setup-specific (tubing) calibration routine

- 1. **build** a simple reactor vessel (Schott bottles) with liquid media flow, from media bottles through reactor vessel and out to waste bottle
- 2. **code**: calibration routine for the weight sensor module
- 3. **code**: analog control of peristaltic motor speed and recording of weight loss and/or gain to record mass flow (g/min)
- 4. **code**: routine to calibrate pump speed to weight loss/gain for a specific setup; store calibration on SD card, which allows to also set pump speed in g/min, or if provided with a culture volume, as culture dilution rate  $(h^{-1})$
- 5. build & code: combine with 1.2 to make turbidostatic control
- 6. build: add gassing system of project 1.1 to make a first simple bioreactor

#### **Resources:**

- Arduino tutorials for Adafruit peristaltic pump and general DC motor control and Adafruit motor shield
- Arduino library for Electrow weight sensor kit 3 kg
- HX711 24-bit analog-to-digital converter (ADC) for load cells

#### **Materials:**

- Bottles, screw caps with inlet/outlet openings, and tubing: available & obtain!
- Scale ordered

fancy: Mettler Toledo, PBK785-3XS/f

cheap: Elecrow Weight Sensor kit 3kg for Arduino  $\leftarrow$  ordered

• Peristaltic pumps - ordered

fancy: Longer Pump LP-BT100-2J, DG-2(10)

cheap: Ismatec Ecoline VC-MS/CA4-12

cheaper: Welco WPM or cheapest: Adafruit  $\leftarrow$  ordered

• Toshiba TA7291P Bridge Driver - ordered

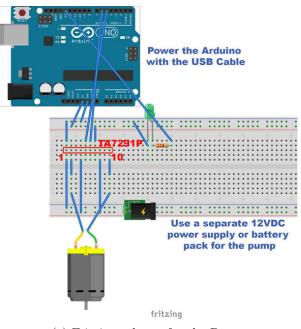
#### • Sainsmart Arduino Mega + Touchscreen - ordered



(a) Elecrow Scale Module, 3 kg



(b) Adafruit Peristaltic Pump



(c) Fritzing scheme for the Pump

Figure 5: Continuous Culture Module. The electrow scale module is based on the HX711 load-cell amplifier (24-bit analog- to-digital converter) and connected to 5V, Gnd and two analog pins of the Arduino and comes with an Arduino library. An Adafruit peristaltic pump (12 v) is run by a "geared down DC motor" and requires a Toshiba TA7291P Bridge Driver (0-20V 1A; 2A peak) to control pump direction and speed *via* Arduino's PWM pins, see the *instructable* tutorial.

#### 1.4 The Server

**Project:** a master software running on a (detachable) linux desktop that synchronizes and speaks via a comon interface to all Arduino and Raspberry Pi modules; the modules themselves can interpret get, set and act impulses (use arguments only when absolutely necessary).

During an initialization the server may inquire what an attached module provides (*via* data IDs and SI units, meaningful time resolution) and handle it automatically. Variable higher order control or processing logics can be built using defined data and control IDs.

- 1. **build** combine of gas (1.1), liquid (1.3) and light (1.2) modules into a bioreactor
- 2. **code**: master program to synchronize and record data from the three modules
- 3. code: combine e.g. 1.2 & 1.3 to implement turbidostat control

- setTime(time\_t t): sets the current master time to all modules
- get(..., time\_t t): get all values, currently available (with a time stamp), or from a previous time t
- act(..., time\_t t): act (switch on and off, set to a specific value), now or at future time t

### 1.5 Heat Flux: Water Bath Thermostat

**Project:** build a water bath for growth vessels, control T, read-out energy required for maintaining constant T and estimate the amount of heat withdrawn or administered

1.

- Jacketed reactor vessel: build or obtain
- Julabo water bath, e.g. F25-ME
- Arduino and/or Raspberry Pi

# 1.6 The Kaiten Eppi: Automated Sampling Device

**Projects:** build sterile and automated sampling device; using a controllable syringe pump, sampling into the Kaiten Eppi (automated: pump sample into tubes, potentially pre-filled with chemicals, vortex, and transport them into liquid  $N_2$  or other storage containers)

#### **Materials:**

• Sterile sampling device by HHU glas blowers: available

• Syringe pump: obtain

• Kaiten Eppi: build

• Sainsmart Arduino Mega & Touchscreen: obtain

# 1.7 Single Cell Biology: Microfluidic Device

**Project:** Basic microfluidics and live-cell imaging device; scratch growth chambers and liquid flow channels into microscope slide; attach 2-3 pumps; and control via arduino/screen

#### **Resources:**

• Tsuda et al. PLoS ONE 2015: 3D Printed 'Plug and Play' Millifluidic

#### **Materials:**

• Ilka's lab microscope: available

• Microscopy slides: available

• 2–3 peristaltic pumps for microfluidics: obtain

• Sainsmart's Arduino Mega + Touchscreen: obtain

# 2 Program

### 2.1 Day 1 <12:00 : Building Bioreactors

Talks, 30-60 min:

- Rob's DIY Reactor The Beginnings: NinjaPBR Arduino-controlled mini PBR
- Dougie's DIY Reactor 20 yrs Later
- Avantes Spectrometry: Spectrometry applications, incl. NIR for metabolite measurements and OD; software interface to Avantes spectrometers
- CellDeg Optimizing Photosynthetic Growth: Introduction to CellDeg's 2.5 k Euro algal growth setup (30 g/L cyano biomass in three days)

### 2.2 Day 1 > 13:00: Hack'a'thing I

- Introduction to the Gas'o'meter: connecting sensors with Arduino, making an autonomous measurement device via Sainsmart's Touch Screen
- Introduction to Rob's reactor: complete setup for photosynthetic growth
- Self-organizing into teams: lab hardware (tubing etc.), control hardware (soldering etc.), software and/or by by projects (1.1–1.7)

# 2.3 Day 2: Hack'a'thing II

- Hardware I: soldering, tubing
- Software I: probe/sensor/pump ⇔ arduino/raspi interfaces
- Visit HHU's fine mechanics and glas blower work-shops, place orders for stuff missing for above goals

# 2.4 Day 3: Hack'a'thing III

- Hardware II: Integrate projects 1.1,1.2&1.3 into a simple DIY reactor and/or with PSI FMT150 or Multicultivator
- Software II: arduino/raspi ⇔ master/server interface Standard data formats and interfaces
- Brain storming: relation of data and models and beer

### 2.5 Participants & Teams

$\mathbf{type}$	name	e-mail	$\mathbf{team}$
	Sascha Wigger	sascha.wigger@student.htw-berlin.de	
	Nicolas Schmelling	nicolas.schmelling@uni-duesseldorf.de	
	Fiona Moejes	fionamoejes@hotmail.com	
	Stefano Magni	stefano.magni@uni-duesseldorf.de	
	Philipp Norf	philipp.norf@uni-duesseldorf.de	
	Elahe Radmaneshfar	radmaneshfar@gmail.com	
	Anna Matuszynska	anna.matuszynska@hhu.de	
	Suraj Sharma	suraj.sharma@uni-duesseldorf.de	
	Simon Schliesky	simon.schliesky@hhu.de	
	Antonella Succurro	succurro@hhu.de	
	Robert Lehmann	lehmann.rob@googlemail.com	
	Dougie Murray	dougie@ttck.keio.ac.jp	
	Oliver Ebenhöh	oliver.ebenhoeh@hhu.de	
	Rainer Machné	machne@hhu.de	

Table 1: should we have skill badges, https://www.adafruit.com/products/2857?q=skill%20badge&p=0& or 3D-printed awards, like the "red soldering iron", "the blue tube", "the green chip"?

# 3 Outlook: 2<sup>nd</sup> QTB PBR Hack'a'thing

# 3.1 Growth Dynamics: Photobioreactors in Research

Talks, 30-60 min:

Nir Keren, Hellingwerf, Jan Cerveny, Dougie Murray, something microfluidics?

# 3.2 Single Cell Dynamics: Microfluidic Devices

Integrate project 1.7 with the simple microscope in Ilka's lab, or a more advanced system (CAi?)

# 3.3 Omics: Sterile and Automated Sampling Devices

Proper sampling for high-throughput data acquisition (mass spectrometry, sequencing) - the Kaiten Eppi