American Society of Hematology
2021 L Street NW, Suite 900,
Washington, DC 20036
Phone: 202-776-0544 | Fax 202-776-0545
editorial@hematology.org

Description of a novel subtype of acute myeloid leukemia defined by recurrent *CBFB* insertions

Tracking no: BLD-2022-017874R1

Georgina Ryland (Centre for Cancer Research, Australia) Masayuki Umeda (St. Jude Children's Research Hospital, United States) Linda Holmfeldt (The Beijer Laboratory, Sweden) Soren Lehmann (Karolinska Insitute, Sweden) Morten Herlin (Aarhus University Hospital, Denmark) Jing Ma (St Jude Children's Research Hospital, United States) Mahsa Khanlari (St. Jude Children's Research Hospital, United States) Jeffrey Rubnitz (St. Jude Children's Research Hospital, United States) Rhonda Ries (Fred Hutchinson Cancer Research Center, United States) Hansen Kosasih (Murdoch Children's Research Institute, Australia) Paul Ekert (Murdoch Children's Research Institute, Australia) Hwee Ngee Goh (Peter MacCallum Cancer Centre, Australia) Ing Tiong (Peter MacCallum Cancer Centre, Australia) Sean Grimmond (University of Melbourne, Australia) Claudia Haferlach (MLL Munich Leukemia Laboratory, Germany) Ryan Day (Washington University St Louis, United States) Timothy Ley (Washington University School of Medicine, United States) Soheil Meshinchi (Fred Hutchinson Cancer Research Center, United States) Xiaotu Ma (St Jude Children's Research Hospital, United States) Piers Blombery (University of Melbourne, Australia) Jeffery Klco (St Jude Children's Research Hospital, United States)

Abstract:

Conflict of interest: COI declared - see note

COI notes: P.G.E. receives an annual payment related to the Walter and Eliza Hall Institute distribution of royalties scheme. P.G.E. consults for Illumina. The other authors declare no competing financial interests.

Preprint server: No;

Author contributions and disclosures: G.R., M.U., P.B., and J.M.K. conceived of the project; G.R., M.U., L.H., S.L., M.K.H., J.M., M.K., J.E.R., H.J.K., P.G.E., H.G., I.S.T., S.M.G., C.H., R.B.D., T.J.L., S.M., X.M., P.B., and J.M.K. identified cases, and provided genomic data and clinical information; G.R., M.U., J.M., and X.M. analyzed genomic data and collated clinical information; G.R., M.U., P.B., and J.M.K. wrote the first version of the manuscript; all authors reviewed and approved the final version of the manuscript.

Non-author contributions and disclosures: No;

Agreement to Share Publication-Related Data and Data Sharing Statement: Publicly available data used in this study is listed in the data supplement available with the online version of this article. For all other data please contact to corresponding authors.

Clinical trial registration information (if any):

Description of a novel subtype of acute myeloid leukemia defined by recurrent *CBFB* insertions

Georgina L. Ryland[1-3]*, Masayuki Umeda[4]*, Linda Holmfeldt[5,6], Sören Lehmann[7], Morten Krogh Herlin[8,9], Jing Ma[4], Mahsa Khanlari[4], Jeffrey E. Rubnitz[10], Rhonda E. Ries[11], Hansen J. Kosasih[12], Paul G. Ekert[3,12,13,14,15], Hwee Ngee Goh[1], Ing S. Tiong[1], Sean M. Grimmond[2], Claudia Haferlach[16], Ryan B. Day[17], Timothy J. Ley[17], Soheil Meshinchi[11], Xiaotu Ma[18], Piers Blombery[1,3,19]*, and Jeffery M. Klco[4]*

1. Department of Pathology, Peter MacCallum Cancer Centre, Melbourne, Victoria, Australia
2. Centre for Cancer Research, University of Melbourne, Parkville, Victoria, Australia
3. Sir Peter MacCallum Department of Oncology, University of Melbourne, Parkville, Victoria, Australia
4. Department of Pathology, St. Jude Children's Research Hospital, Memphis, Tennessee, United States
5. Department of Immunology, Genetics, and Pathology, Science for Life Laboratory, Uppsala University, Uppsala, Sweden
6. The Beijer Laboratory, Uppsala, Sweden
7. Department of Hematology, Karolinska University Hospital, Stockholm
8. Department of Clinical Genetics, Aarhus University Hospital, Aarhus, Denmark
9. Department of Pediatrics and Adolescent Medicine, Aarhus University Hospital, Aarhus, Denmark
10. Department of Oncology, St. Jude Children's Research Hospital, Memphis, Tennessee, United States
11. Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, Washington, United States
12. Murdoch Children's Research Institute, Parkville, Victoria, Australia
13. Children's Cancer Institute, Lowy Cancer Research Centre, UNSW Sydney, Sydney, New South Wales, Australia
14. Cancer Immunology Program, Peter MacCallum Cancer Centre, Parkville, Victoria, Australia
15. Discipline of Paediatrics and Child Health, School of Clinical Medicine, UNSW Medicine & Health, UNSW Sydney, Sydney, New South Wales, Australia
16. Munich Leukemia Laboratory, Munich, Germany
17. Division of Oncology, Department of Medicine, Washington University School of Medicine, St. Louis, Missouri, United States
18. Department of Computational Biology, St. Jude Children's Research Hospital, Memphis, Tennessee, United States
19. Clinical Haematology, Peter MacCallum Cancer Centre and Royal Melbourne Hospital, Melbourne, Victoria, Australia

*These authors contributed equally to the manuscript

Correspondence:

Jeffery M. Klco, Mail Stop 342, Room D4047B, St. Jude Children's Research Hospital, 262 Danny Thomas Place, Memphis, TN 38105; e-mail: jeffery.klco@stjude.org

Piers Blombery, Department of Pathology, Peter MacCallum Cancer Centre, 305 Grattan Street, Melbourne, Victoria, Australia 3000; e-mail: piers.blombery@petermac.org

Word count: 1127/1200

Number of Figures: 1

Number of Tables: 1

Supplemental Data: Yes

Reference Count: 14/25

Molecular analysis of pediatric and adult acute myeloid leukemia (AML) is routinely used to identify subtype-defining driver structural variants and mutations which may provide important information for risk stratification. For example, the core-binding factor (CBF) AML subgroup defined by t(8;21) *RUNX1::RUNX1T1* or inv(16)/t(16;16) *CBFB::MYH11* are associated with favorable outcomes[1,2]. Despite the extensive genomic characterization of pediatric and adult AML, there still remains an important proportion of previously unclassified cases where new driver lesions are still being identified, including those that can influence patient management due to their association with outcomes. This includes the recent identification of tandem duplications in *UBTF* in pediatric AML[3,4] and structural variants that dysregulate *BCL11B* in lineage ambiguous acute leukemia[5]. Herein we describe an additional new subtype of AML characterized by a recurrent insertion mutation in *CBFB*.

We initially re-analyzed a cohort of 553 pediatric AML transcriptomes from our previous study[3] and identified two patients (PARANT:SJAML040573, PARUTH:SJAML040605) with similar gene expression profiles to *CBFB::MYH11* AML but without a *CBFB::MYH11* fusion or other known leukemic driver by conventional testing, and without finding a *CBFB::MYH11* fusion by manual inspection of RNA sequence data (Supplemental Figure 1, Table 1). However, in both of these patients, we identified a somatic nine base-pair insertion in exon 3 of *CBFB* (NM_022845.3). The core-binding factor β (CBFβ) protein encoded by *CBFB* forms the non-DNA binding regulatory subunit of a heterodimeric transcription factor complex with a DNA-binding CBFα subunit (RUNX1, RUNX2, or RUNX3). Interestingly, both *CBFB* mutations were predicted to lead to the same amino acid change, substituting aspartic acid at position 87 (D87) for glycine, aspartic acid, serine, and tyrosine [p.(Asp87delinsGlyAspSerTyr); GDSY] within the N-terminal RUNX-binding domain[6] (Figure 1A).

Given these findings, we hypothesized that *CBFB* mutations may be recurrent in AML and a defining feature of a novel subtype. Through a combination of published data, clinical sequencing, and screening driver-negative AML cohorts from independent sources (see Supplemental Methods), we identified an additional 16 cases with *CBFB* insertions involving D87, including 15 AML and one B/myeloid mixed phenotype acute leukemia (MPAL), for a total of 18 cases (Table 1). These additional cases also lacked a *CBFB::MYH11* fusion or other known leukemic driver alterations (Supplemental Table 1). *CBFB* mutations were confirmed in both DNA and RNA sequencing when available, or Sanger sequencing, and were confirmed to be somatic in 11 of 11 cases where matched germline data was available (Supplemental Table 1, Supplemental Methods, Supplemental Figure 2). Remarkably, we identified ten different nucleotide insertions at codon 87 in these 18 cases; nine out of ten were predicted to encode for the same in-frame GDSY amino acid change (p.(Asp87delinsGlyAspSerTyr)), with the other nucleotide insertion leading to a GDTY amino acid change (p.(Asp87delinsGlyAspThrTyr)) (Figure 1A). This highly stereotyped change at the protein level (i.e. GDXY) strongly implies a functional relevance.

We next integrated eight of these additional cases into our transcriptome cohort and observed a tight cluster of cases with *CBFB* insertions adjacent to the *CBFB::MYH11* cluster (Figure 1B). Gene set enrichment analysis (GSEA) confirmed broad similarities between AML with *CBFB* insertions and CBF AMLs (Figure 1C, Supplemental Figure 3, Supplemental Table 2). However, *CBFB* insertion

cases showed uniquely high expression of *BCL2L14*, *MEIS1*, and *HOXA* cluster genes, demonstrating a more stem-like signature compared to *CBFB::MYH11* AML.

We also examined the cooperating mutations in ten *CBFB* insertion cases from RNA sequencing data and integrated these findings with mutational information from other studies (Figure 1D, Supplemental Table 1, Supplemental Table 3). Recurrent mutations were detected in *BCORL1* (7/18, 39%), *FLT3* (7/18, 39%), *NRAS* (6/18, 33%), *ETV6* (5/18, 28%), *KDM6A* (4/18, 22%) and *NF1* (2/18 11%). *FLT3* tyrosine kinase domain (TKD) mutations were most common although internal tandem duplications and mutations outside the TKD were also observed. Overall, these mutations are different from the mutational spectrum previously reported for CBF leukemias[7], most notably the absence of *KIT* mutations and a higher frequency of *FLT3*-TKD and *BCORL1* mutations. Recurrent chromosomal alterations in the *CBFB* insertion group included trisomy 6 and trisomy 22, whereas trisomy 8 was not observed (Figure 1D, Supplemental Table 1). Additionally, the *CBFB* mutation was conserved at both diagnosis and relapse in two cases profiled at both time points (AML075 and 115225). Collectively these data suggest that *CBFB* insertions are a subtype-defining lesion, and we have provisionally termed this group *CBFB*-GDXY.

Like CBF AML, AMLs with *CBFB*-GDXY mutations were observed in both children and adults but were enriched in adolescent and young adult age groups (median age 16.5 years, range 9 - 27). Overall, this mutation in AML cohorts was rare, including 3/188 in the TARGET pediatric AML cohort, 5/1048 in the pediatric AAML1031 cohort and 1/350 in the Clinseq-AML Swedish adult cohort, while more than 2000 cases from multiple large cohorts comprised primarily of adult AMLs did not harbor a *CBFB* insertion (see Supplemental Data for a description of cohorts screened). Additionally, unlike the typical myelomonocytic morphology with abnormal eosinophils (FAB M4 Eo) observed for *CBFB::MYH11* AML, *CBFB*-GDXY AML had less mature morphologies (FAB M0: n=1, FAB M1: n=4, or FAB M2: n=4), consistent with the stem-related expression profiles, where morphologic reports were available. However, an increase in eosinophils was still observed (Figure 1E). Like *RUNX1::RUNX1T1* AML[8], we noted that *CBFB*-GDXY AML may express *CD19* (Supplemental Figure 4, Supplemental Table 1). Further supporting this observation is the identification of the GDXY insertion in one case of B/myeloid mixed phenotype acute leukemia.

This cohort is small and collected from different sources with varied treatment protocols, precluding a definitive assessment of the impact of this mutation on outcomes. However, 8 out of 17 patients (where data was available) had either relapsed or refractory disease after initial treatment, whereas patients with *RUNX1::RUNX1T1* or *CBFB::MYH11* AMLs commonly have a good outcome and these AMLs are considered favorable risk. To investigate the treatment response in one patient, we designed a *CBFB* mutation-specific ultra-deep NGS assay (see Supplemental Methods) for longitudinal tracking of measurable residual disease (MRD). This 25-year-old patient (SBJ00860) was treated with cytarabine and idarubicin induction (7+3), followed by high-dose cytarabine consolidation. MRD assessment after each cycle of chemotherapy showed detectable but decreasing *CBFB* insertion variant allele frequency, becoming undetectable after the last cycle of therapy and remaining undetectable at nine months of follow-up (Figure 1F).

In summary, we have reported a novel subtype of AML characterized by recurrent in-frame insertion mutations in *CBFB* leading to a GDXY amino acid sequence change at position D87. Molecular characterization demonstrated transcriptional similarity to CBF AML, while also highlighting an enrichment of *FLT3*-TKD mutations, lack of *KIT* mutations, and stemness-related gene expression signature. Recognition of this subtype and further study in clinical trials, as well as investigation of the underlying leukemogenic mechanism of the *CBFB* insertion, will be important to understand the full clinical relevance of this novel entity.

Acknowledgements

The authors wish to thank all patients and their families from all participating institutions for their contribution of the biological specimens used in this study. This work was supported by grants from the Wilson Centre for Blood Cancer Genomics and the Snowdome Foundation, as well as the American Lebanese and Syrian Associated Charities of St. Jude Children's Research Hospital. The authors also gratefully acknowledge the Haematology Tissue Bank (Peter MacCallum Cancer Centre/Royal Melbourne Hospital) for assistance with sample collection and the Genomics Core Facility and Genomics Platform Group (University of Melbourne Centre for Cancer Research) for sequencing and analysis support, as well as the Biorepository and the Hartwell Center for Bioinformatics and Biotechnology at St. Jude Children's Research Center (P30 CA021765, Cancer Center Support Grant). Tumor samples and coded data were supplied by the Children's Cancer Centre Biobank at the Murdoch Children's Research Institute and The Royal Children's Hospital (mcri.edu.au/research/projects/childrens-cancer-centre-biobank). Establishment and running of the Children's Cancer Centre is made possible through generous support by Cancer In Kids @ RCH (www.cika.org.au), The Royal Children's Hospital Foundation and the Murdoch Children's Research Institute. P.G.E. acknowledges the support of the Samuel Nissen Charitable Foundation and the Leukemia and Lymphoma Society USA Specialized Center of Research (SCOR) Project Grant "Apoptosis Controllers". J.M.K. holds a Career Award for Medical Scientists from the Burroughs Wellcome Fund and is a previous recipient of the V Foundation Scholar Award (Pediatric).

This research content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

Contribution:

G.R., M.U., P.B., and J.M.K. conceived of the project; G.R., M.U., L.H., S.L., M.K.H., J.M., M.K., J.E.R., H.J.K., P.G.E., H.G., I.S.T., S.M.G., C.H., R.B.D., T.J.L., S.M., X.M., P.B., and J.M.K. identified cases, and provided genomic data and clinical information; G.R., M.U., J.M., and X.M. analyzed genomic data and collated clinical information; G.R., M.U., P.B., and J.M.K. wrote the first version of the manuscript; all authors reviewed and approved the final version of the manuscript.

Conflict-of-interest disclosure: P.G.E. receives an annual payment related to the Walter and Eliza Hall Institute distribution of royalties scheme. P.G.E. consults for Illumina. The other authors declare no competing financial interests.

References

1. Byrd JC, Mrózek K, Dodge RK, et al. Pretreatment cytogenetic abnormalities are predictive of induction success, cumulative incidence of relapse, and overall survival in adult patients with de novo acute myeloid leukemia: results from Cancer and Leukemia Group B (CALGB 8461). *Blood*. Dec 15 2002;100(13):4325-36. doi:10.1182/blood-2002-03-0772
2. Schlenk RF, Benner A, Krauter J, et al. Individual patient data-based meta-analysis of patients aged 16 to 60 years with core binding factor acute myeloid leukemia: a survey of the German Acute Myeloid Leukemia Intergroup. *J Clin Oncol*. Sep 15 2004;22(18):3741-50. doi:10.1200/jco.2004.03.012
3. Umeda M, Ma J, Huang BJ, et al. Integrated Genomic Analysis Identifies UBTF Tandem Duplications as a Recurrent Lesion in Pediatric Acute Myeloid Leukemia. *Blood Cancer Discov*. May 5 2022;3(3):194-207. doi:10.1158/2643-3230.BCD-21-0160
4. Stratmann S, Yones SA, Mayrhofer M, et al. Genomic characterization of relapsed acute myeloid leukemia reveals novel putative therapeutic targets. *Blood Advances*. 2021;5(3):900-912. doi:10.1182/bloodadvances.2020003709
5. Montefiori LE, Bendig S, Gu Z, et al. Enhancer Hijacking Drives Oncogenic BCL11B Expression in Lineage-Ambiguous Stem Cell Leukemia. *Cancer Discov*. Nov 2021;11(11):2846-2867. doi:10.1158/2159-8290.Cd-21-0145
6. de Bruijn MFTR, Speck NA. Core-binding factors in hematopoiesis and immune function. *Oncogene*. 2004/05/01 2004;23(24):4238-4248. doi:10.1038/sj.onc.1207763
7. Faber ZJ, Chen X, Gedman AL, et al. The genomic landscape of core-binding factor acute myeloid leukemias. *Nature Genetics*. 2016/12/01 2016;48(12):1551-1556. doi:10.1038/ng.3709
8. Walter K, Cockerill PN, Barlow R, et al. Aberrant expression of CD19 in AML with t(8;21) involves a poised chromatin structure and PAX5. *Oncogene*. May 20 2010;29(20):2927-37. doi:10.1038/onc.2010.56
9. Bolouri H, Farrar JE, Triche T, Jr., et al. The molecular landscape of pediatric acute myeloid leukemia reveals recurrent structural alterations and age-specific mutational interactions. *Nat Med*. Jan 2018;24(1):103-112. doi:10.1038/nm.4439
10. Ma X, Liu Y, Liu Y, et al. Pan-cancer genome and transcriptome analyses of 1,699 paediatric leukaemias and solid tumours. *Nature*. 2018/03/01 2018;555(7696):371-376. doi:10.1038/nature25795
11. Alexander TB, Gu Z, Iacobucci I, et al. The genetic basis and cell of origin of mixed phenotype acute leukaemia. *Nature*. Oct 2018;562(7727):373-379. doi:10.1038/s41586-018-0436-0
12. Fornerod M, Ma J, Noort S, et al. Integrative Genomic Analysis of Pediatric Myeloid-Related Acute Leukemias Identifies Novel Subtypes and Prognostic Indicators. *Blood Cancer Discov*. Nov 2021;2(6):586-599. doi:10.1158/2643-3230.Bcd-21-0049
13. Stratmann S, Yones SA, Garbulowski M, et al. Transcriptomic analysis reveals proinflammatory signatures associated with acute myeloid leukemia progression. *Blood Advances*. 2022;6(1):152-164. doi:10.1182/bloodadvances.2021004962
14. Petti AA, Khan SM, Xu Z, et al. Genetic and Transcriptional Contributions to Relapse in Normal Karyotype Acute Myeloid Leukemia. *Blood Cancer Discovery*. 2022;3(1):32-49. doi:10.1158/2643-3230.Bcd-21-0050

Figures

Figure 1. A novel subtype of AML characterized by *CBFB*-GDXY mutations.

(A) Graphical representation of *CBFB* exons 1-6 (NM_022845.3) showing the location (arrowhead) and sequence of the 9 bp insertion mutations (highlighted in red) relative to the wild type complementary DNA sequence. The number of patients with each mutation is shown in the circles. The predicted protein sequence of the *CBFB* mutations is also shown in red.

(B) Uniform Manifold Approximation and Projection (UMAP) of expression profiles of the pediatric AML cohort (AML, n=561; cord blood CD34+ control, n=5) performed with the top 133 most variably expressed genes. Dots are colored by the molecular feature of the sample.

(C) Gene Set Enrichment Analysis (GSEA) between AML with *CBFB* insertions and non-CBF AML (left) using gene sets derived from differentially expressed genes in *CBFB::MYH11* AML or *RUNX1::RUNX1T1* AML against non-CBF AML. Volcano plots (right) of genes differentially expressed between AML with *CBFB* insertions and *CBFB::MYH11* or *RUNX1::RUNX1T1*.

(D) Mutational landscape of *CBFB*-GDXY AML (n=18) in this study (mutations detected at diagnosis are shown) and *CBFB::MYH11* AML (n=65) collected in the previous study[3]. Pre-selected 75 genes frequently mutated in AML were subjected to mutation calling from RNAseq data. Eight *FLT3* mutations were detected in seven patients and are categorized as internal tandem duplications (ITD), mutations in the tyrosine kinase domain (TKD), or mutations outside the TKD (other domains).

(E) (i) Giemsa stained peripheral blood showed blasts with myeloid and monoblastic features; arrowhead marks single slender Auer rod (original x1000 magnification). (ii) Giemsa stained bone marrow aspirate smears showed immature myeloid elements with granules, blasts, and eosinophils (original x1000 magnification); arrowhead marks salmon-colored granules in the cytoplasm of the myeloid cell (inset). (iii) Hematoxylin and eosin-stained bone marrow biopsy (original magnification x500) showed a hypercellular marrow almost completely replaced by a diffuse infiltrate of medium-sized blasts with increased eosinophils in the background.

(F) Measurable residual disease assessment of the *CBFB* c.259_260insGGGACTCCT mutation by ultra-deep next generation sequencing in SBJ00860.

Tables

Table 1. Clinical characteristics of the patients with acute leukemia harboring a *CBFB*-GDXY mutation.

Identifier	*CBFB*-GDXY mutation (*CBFB* NM_022845.3)	Age/Sex	Diagnosis	FAB category	Karyotype	Cooperating gene mutations	Risk group	Treatment protocol	Outcome	Reference
PARANT:SJAML040573	c.259_260insGAGATTCCT p.(Asp87delinsGlyAspSerTyr)	17M	AML	M2	46,XY	*ETV6*, *KRAS*, *NF1*	Standard	AAML0531	Alive/CR	9,10
PARCEC	c.259_260insGAGACTCCT p.(Asp87delinsGlyAspSerTyr)	23F	AML	Unknown	46,XX	*NRAS*	Standard	AAML0531	Refractory/Dead	9,10
PARUTH:SJAML040605	c.259_260insGGGACTCCT p.(Asp87delinsGlyAspSerTyr)	9M	AML	Unknown	46,XY.nuc ish CBFBx3	*FLT3*, *NRAS*, *BCORL1*	Standard	AAML0531	Relapse/Dead	9
PAVLBB	c.259_260insGTGACTCCT p.(Asp87delinsGlyAspSerTyr)	15M	AML	Unknown	46,XY		Standard	AAML1031	Refractory/Dead	Unpublished
PAWIHN	c.259_260insGGGATTCCT p.(Asp87delinsGlyAspSerTyr)	17F	AML	Unknown	46,XX,i(7)(p10)	*KDM6A*	Standard	AAML1031	Alive/CR	Unpublished
PAWZIX	c.259_260insGGGACTCCT p.(Asp87delinsGlyAspSerTyr)	13F	AML	Unknown	46,XX	*ETV6*, *NRAS*	Standard	AAML1031	Relapse/Alive/CR2	Unpublished
PAXCCW	c.259_260insGGGACTCCT p.(Asp87delinsGlyAspSerTyr)	12F	AML	Unknown	47,XX,+6	*NRAS*	Standard	AAML1031	Alive/CR	Unpublished
PAXDVZ	c.259_260insGGGATTCCT p.(Asp87delinsGlyAspSerTyr)	20M	AML	Unknown	46,XY	*BCORL1*, *KDM6A*, *NRAS*	Standard	AAML1031	Refractory/Dead	Unpublished
SJMPAL017975	c.259_260insGAGACAGTT p.(Asp87delinsGlyAspSerTyr)	18M	B/M MPAL	Unknown	51,XY,+Y,+4,+6,+13,+22	*FLT3*, *ASXL1*, *BCORL1*	Unknown	Unknown	Alive/CR	11
SJAML016545	c.259_260insGAGACTCGT p.(Asp87delinsGlyAspSerTyr)	16M	AML	M2	47,XY,+22	*FLT3*	Intermediate	AML02	Alive/CR	12
SJAML031769	c.259_260insGGGATTCCT p.(Asp87delinsGlyAspSerTyr)	12M	AML	M2	47,XY,+6	*NRAS*, *FLT3*	Intermediate	AML16	Alive/CR	Unpublished
SJAML033048	c.259_260insGGGATTCCT p.(Asp87delinsGlyAspSerTyr)	14F	AML	M2	46,XX	*BCORL1*	Intermediate	AML16	Alive/CR	Unpublished
SBJ00860	c.259_260insGGGACTCCT p.(Asp87delinsGlyAspSerTyr)	25M	AML	M1		*BCORL1*, *ETV6*, *KMT2D*	Intermediate	Induction: 7 + 3 Cytarabine / idarubicin Consolidation: HiDAC x3 cycles	Alive/CR	Unpublished
AML075	c.259_260insGGGATTCGT p.(Asp87delinsGlyAspSerTyr)	10M	AML	M0-NOS	46,XY,inv(9)(q11q12)	*NF1*, *KDM6A*, *WT1*	Intermediate	NOPHO-AML-93	Relapse/Dead	4,13
AMLNOS004	c.259_260insGCGATTCCT p.(Asp87delinsGlyAspSerTyr)	15M	AML	M1	47,XY,+6	*FLT3*, *BCORL1*, *KDM6A*	Standard	NOPHO AML 2004	Alive/CR	Unpublished
ALG201115	c.259_260insGAGATTCCT p.(Asp87delinsGlyAspSerTyr)	27M	AML	M1	46,XY		Intermediate	VP2010-2012	Relapse/Alive/CR2	Unpublished
MLL_75644	c.259_260insGGGATTCCT p.(Asp87delinsGlyAspSerTyr)	17M	AML	M1	46,XY	*FLT3*, *BCORL1*, *ETV6*	Intermediate	Unknown	Unknown	Unpublished
115225	c.259_260insGAGATACCT p.(Asp87delinsGlyAspThrTyr)	22M	AML	Indeterminate	46,XY	*FLT3*, *ETV6*, *WT1*, *DNMT3A*	Unknown	Induction: 7 + 3 Cytarabine / daunorubicin, with concurrent midostaurin (vs. placebo) Consolidation: high dose	Relapse/Alive/CR2	14

			cytarabine x3 cycles with concurrent midostaurin (vs placebo) + 12 mth maintenance midostaurin (vs placebo)

Full information is found in Supplemental Table 1.

Figure 1