

# Quality Control for scRNA and scATAC seq data

Load and Merge the data

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```
library(tibble)
library(tidyr)
library(dplyr)
library(rtracklayer)
library(dplyr)
library(Seurat)
library(Signac)
library(EnsDb.Hsapiens.v86)
library(ggplot2)
library(cowplot)
library(simspec)
library(cowplot)
library(AnnotationHub)
```

```
# load function from local files
```

```
# load function from local files
```

```
load(here::here("data","reference","annotations.rdata")) # load the annotations
```

# 1. Read the Raw Data

In this section, we will read the raw data from the cellrange-arc output files. We have 4 samples: PSZ-6, TSC-tube, TSC-edge, and CTRL. We will rename the samples as PSZ, TSC-tube, TSC-edge, and TSC-outside, respectively.

```
options(Seurat.assay.version = "v3")

counts.psz <- Read10X_h5(here::here("data", "raw-data", "cellrange-arc-out",
                                     "PSZ-6", "filtered_feature_bc_matrix.h5"))
seurat.psz <- CreateSeuratObject(counts = counts.psz$`Gene Expression`,
                                assay = "RNA",
                                project = "PSZ-6")
fragments_path <- here::here("data", "raw-data", "cellrange-arc-out",
                              "PSZ-6", "atac_fragments.tsv.gz")
seurat.psz[['ATAC']] <- CreateChromatinAssay(counts = counts.psz$`Peaks`,
                                             annotation = annotations,
                                             fragments = fragments_path,
                                             sep = c(":", "-"),
                                             genome = 'hg38')

counts.tube <- Read10X_h5(here::here("data", "raw-data", "cellrange-arc-out",
                                     "TSC-tube", "filtered_feature_bc_matrix.h5"))
seurat.tube <- CreateSeuratObject(counts = counts.tube$`Gene Expression`,
                                assay = "RNA",
                                project = "TSC-tube")
fragments_path <- here::here("data", "raw-data", "cellrange-arc-out",
                              "TSC-tube", "atac_fragments.tsv.gz")
seurat.tube[['ATAC']] <- CreateChromatinAssay(counts = counts.tube$`Peaks`,
                                             annotation = annotations,
                                             fragments = fragments_path,
                                             sep = c(":", "-"),
                                             genome = 'hg38')

counts.edge <- Read10X_h5(here::here("data", "raw-data", "cellrange-arc-out",
                                     "TSC-edge", "filtered_feature_bc_matrix.h5"))
seurat.edge <- CreateSeuratObject(counts = counts.edge$`Gene Expression`,
                                assay = "RNA",
                                project = "TSC-edge")
fragments_path <- here::here("data", "raw-data", "cellrange-arc-out",
                              "TSC-edge", "atac_fragments.tsv.gz")
seurat.edge[['ATAC']] <- CreateChromatinAssay(counts = counts.edge$`Peaks`,
                                             annotation = annotations,
                                             fragments = fragments_path,
                                             sep = c(":", "-"),
                                             genome = 'hg38')

counts.outside <- Read10X_h5(here::here("data", "raw-data", "cellrange-arc-out",
                                     "CTRL", "filtered_feature_bc_matrix.h5"))
seurat.outside <- CreateSeuratObject(counts = counts.outside$`Gene Expression`,
                                assay = "RNA",
                                project = "TSC-outside")
fragments_path <- here::here("data", "raw-data", "cellrange-arc-out",
                              "CTRL", "atac_fragments.tsv.gz")
seurat.outside[['ATAC']] <- CreateChromatinAssay(counts = counts.outside$`Peaks`,
```

```

annotation = annotations,
fragments = fragments_path,
sep = c(":", "-"),
genome = 'hg38')

```

## 2. Merge the samples

```

# Add unique identifiers to the cell names for each Seurat object
seurat.psz <- RenameCells(seurat.psz,
                          new.names = paste("psz", Cells(seurat.psz), sep = "_"))
seurat.tube <- RenameCells(seurat.tube,
                           new.names = paste("tube", Cells(seurat.tube), sep = "_"))
seurat.edge <- RenameCells(seurat.edge,
                           new.names = paste("edge", Cells(seurat.edge), sep = "_"))
seurat.outside <- RenameCells(seurat.outside,
                              new.names = paste("outside",
                                                  Cells(seurat.outside), sep = "_"))

# Now merge the Seurat objects
seurat <- merge(seurat.psz, y = list(seurat.tube, seurat.edge, seurat.outside))

# remake the peaks link for the
peaks <- reduce(unlist(as(c(seurat.psz@assays$ATAC@ranges,
                           seurat.tube@assays$ATAC@ranges,
                           seurat.edge@assays$ATAC@ranges,
                           seurat.outside@assays$ATAC@ranges),
                           "GRangesList"))))

peakwidths <- width(peaks)
peaks <- peaks[peakwidths < 10000 & peakwidths > 20]

counts_atac_merged <- FeatureMatrix(seurat@assays$ATAC@fragments,
                                    features = peaks,
                                    cells = colnames(seurat))
seurat[['ATAC']] <- CreateChromatinAssay(counts_atac_merged,
                                         fragments = seurat@assays$ATAC@fragments,
                                         annotation = seurat@assays$ATAC@annotation,
                                         sep = c(":", "-"),
                                         genome = "hg38")

save(seurat, file = here::here("data", "processed-data", "seurat-merged_2025-03-15.rdata"))
# save(seurat, file = here::here("data", "processed-data", "seurat-merged_2025-03-14.rdata"))

```

## sessionInfo()

```
## R version 4.4.0 (2024-04-24)
## Platform: aarch64-apple-darwin20
## Running under: macOS Sonoma 14.3.1
##
## Matrix products: default
## BLAS:   /Library/Frameworks/R.framework/Versions/4.4-arm64/Resources/lib/libRblas.0.dylib
## LAPACK: /Library/Frameworks/R.framework/Versions/4.4-arm64/Resources/lib/libRlapack.dylib; LAPACK v
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## time zone: America/New_York
## tzcode source: internal
##
## attached base packages:
## [1] stats4      stats      graphics  grDevices  utils      datasets  methods
## [8] base
##
## other attached packages:
## [1] AnnotationHub_3.12.0      BiocFileCache_2.12.0
## [3] dbplyr_2.5.0             simspec_0.0.0.9000
## [5] cowplot_1.1.3            EnsDb.Hsapiens.v86_2.99.0
## [7] ensemblDb_2.28.1         AnnotationFilter_1.28.0
## [9] GenomicFeatures_1.56.0   AnnotationDbi_1.66.0
## [11] Biobase_2.64.0           Signac_1.14.0
## [13] Seurat_5.2.1             SeuratObject_5.0.2
## [15] sp_2.2-0                 rtracklayer_1.64.0
## [17] GenomicRanges_1.56.2     GenomeInfoDb_1.40.1
## [19] IRanges_2.38.1           S4Vectors_0.42.1
## [21] BiocGenerics_0.50.0      knitr_1.49
## [23] lubridate_1.9.4          forcats_1.0.0
## [25] stringr_1.5.1            dplyr_1.1.4
## [27] purrr_1.0.4              readr_2.1.5
## [29] tidyr_1.3.1              tibble_3.2.1
## [31] ggplot2_3.5.1            tidyverse_2.0.0
##
## loaded via a namespace (and not attached):
## [1] RcppAnnoy_0.0.22         splines_4.4.0
## [3] later_1.4.1             BiocIO_1.14.0
## [5] filelock_1.0.3          bitops_1.0-9
## [7] polyclip_1.10-7         XML_3.99-0.18
## [9] fastDummies_1.7.5       lifecycle_1.0.4
## [11] rprojroot_2.0.4         hdf5r_1.3.12
## [13] globals_0.16.3          lattice_0.22-6
## [15] MASS_7.3-64             magrittr_2.0.3
## [17] plotly_4.10.4           rmarkdown_2.29
## [19] yaml_2.3.10             httpuv_1.6.15
## [21] sctransform_0.4.1       spam_2.11-1
## [23] spatstat.sparse_3.1-0   reticulate_1.40.0
## [25] pbapply_1.7-2           DBI_1.2.3
## [27] RColorBrewer_1.1-3      abind_1.4-8
## [29] zlibbioc_1.50.0         Rtsne_0.17
```

## [31]	RCurl_1.98-1.16	rappdirs_0.3.3
## [33]	GenomeInfoDbData_1.2.12	ggrepel_0.9.6
## [35]	irlba_2.3.5.1	listenv_0.9.1
## [37]	spatstat.utils_3.1-2	goftest_1.2-3
## [39]	RSpectra_0.16-2	spatstat.random_3.3-2
## [41]	fitdistrplus_1.2-2	parallelly_1.42.0
## [43]	codetools_0.2-20	DelayedArray_0.30.1
## [45]	RcppRoll_0.3.1	tidyselect_1.2.1
## [47]	UCSC.utils_1.0.0	farver_2.1.2
## [49]	matrixStats_1.5.0	spatstat.explore_3.3-4
## [51]	GenomicAlignments_1.40.0	jsonlite_1.9.0
## [53]	progressr_0.15.1	ggridges_0.5.6
## [55]	survival_3.8-3	tools_4.4.0
## [57]	ica_1.0-3	Rcpp_1.0.14
## [59]	glue_1.8.0	gridExtra_2.3
## [61]	SparseArray_1.4.8	here_1.0.1
## [63]	xfun_0.51	MatrixGenerics_1.16.0
## [65]	withr_3.0.2	BiocManager_1.30.25
## [67]	fastmap_1.2.0	digest_0.6.37
## [69]	timechange_0.3.0	R6_2.6.1
## [71]	mime_0.12	colorspace_2.1-1
## [73]	scattermore_1.2	tensor_1.5
## [75]	spatstat.data_3.1-4	RSQLite_2.3.9
## [77]	generics_0.1.3	data.table_1.16.4
## [79]	httr_1.4.7	htmlwidgets_1.6.4
## [81]	S4Arrays_1.4.1	uwot_0.2.2
## [83]	pkgconfig_2.0.3	gtable_0.3.6
## [85]	blob_1.2.4	lmtest_0.9-40
## [87]	XVector_0.44.0	htmltools_0.5.8.1
## [89]	dotCall64_1.2	ProtGenerics_1.36.0
## [91]	scales_1.3.0	png_0.1-8
## [93]	spatstat.univar_3.1-1	rstudioapi_0.17.1
## [95]	tzdb_0.4.0	reshape2_1.4.4
## [97]	rjson_0.2.23	nlme_3.1-167
## [99]	curl_6.2.1	cachem_1.1.0
## [101]	zoo_1.8-12	BiocVersion_3.19.1
## [103]	KernSmooth_2.23-26	parallel_4.4.0
## [105]	miniUI_0.1.1.1	restfulr_0.0.15
## [107]	pillar_1.10.1	grid_4.4.0
## [109]	vctrs_0.6.5	RANN_2.6.2
## [111]	promises_1.3.2	xtable_1.8-4
## [113]	cluster_2.1.8	evaluate_1.0.3
## [115]	cli_3.6.4	compiler_4.4.0
## [117]	Rsamtools_2.20.0	rlang_1.1.5
## [119]	crayon_1.5.3	future.apply_1.11.3
## [121]	plyr_1.8.9	stringi_1.8.4
## [123]	viridisLite_0.4.2	deldir_2.0-4
## [125]	BiocParallel_1.38.0	munsell_0.5.1
## [127]	Biostrings_2.72.1	lazyeval_0.2.2
## [129]	spatstat.geom_3.3-5	Matrix_1.7-2
## [131]	RcppHNSW_0.6.0	hms_1.1.3
## [133]	patchwork_1.3.0	bit64_4.6.0-1
## [135]	future_1.34.0	KEGGREST_1.44.1
## [137]	shiny_1.10.0	SummarizedExperiment_1.34.0

```
## [139] ROCR_1.0-11          memoise_2.0.1
## [141] igraph_2.1.4             fastmatch_1.1-6
## [143] bit_4.5.0.1
```