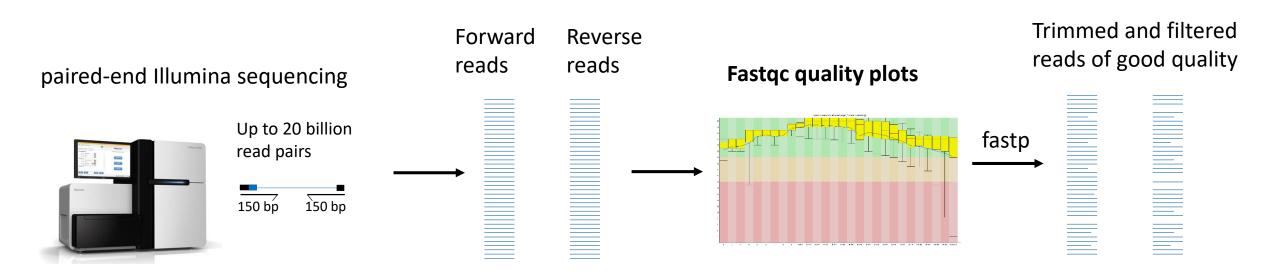
# Mapping to a reference genome

### 1. Quality check and trimming raw reads



### 2. Alignment to the reference genome

reference bam individual 1 bam individual 2

#### Tools to align short read data

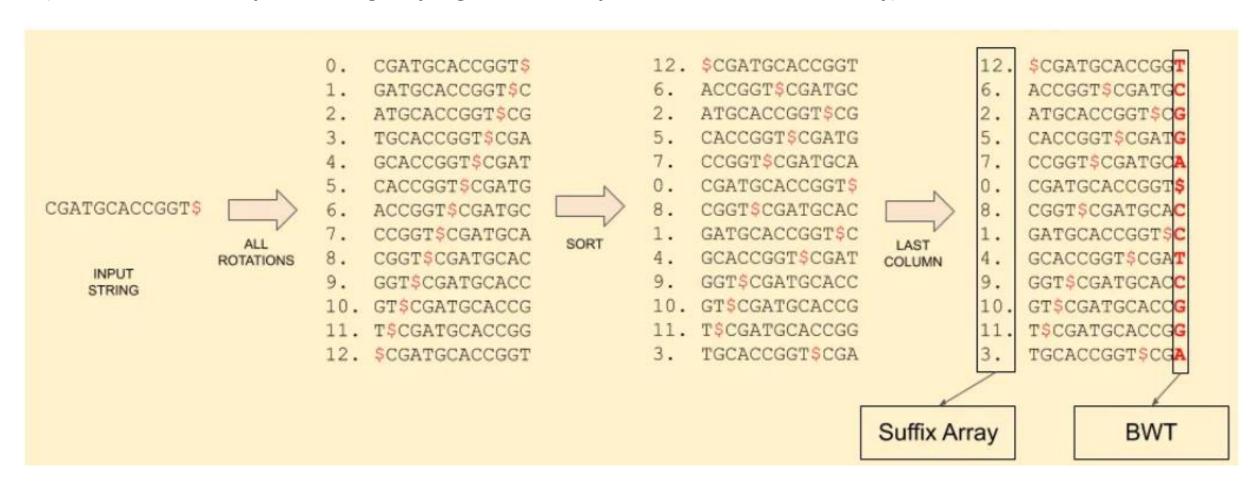
reference read —

**bwa-mem2** (Burroughs-Wheeler alignment – maximum exact matches)

**Bowtie2** 

#### **Burroughs Wheeler Transform**

(also used in file compression, e.g. bzip, figures from https://medium.com/@mr-easy)



#### **Burroughs Wheeler Alignment**

\$CGATGCACCGGT ACCGGT SCGATGC ATGCACCGGT\$CG CACCGGT\$CGATG CCGGT\$CGATGCA CGATGCACCGGT\$ CGGT\$CGATGCAC GATGCACCGGTSC GCACCGGT\$CGAT COLUMN GGT\$CGATGCACC GT\$CGATGCACCG TSCGATGCACCGG TGCACCGGTSCGA Suffix Array **BWT** 

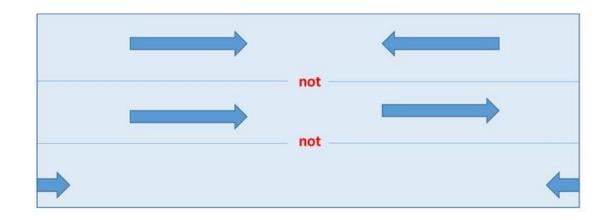
CGATGCACCGGT\$

Read: GCA

	Re	Read:			C	4	Read:			GCA			Read:			<b>G</b> CA		
12.	0.	\$				T	0.	\$				T	0.	\$ .			T	
6.	1.	A				C	1.	Α				C	1.	Α.			C	
2.	2.	A				G	2	Α				G	2.	Α.			G	
5.	3.	C				G	3.	CI	1			G	3.	CA			G	
7.	4.	C				A	4.	С				A	4.	С.			A	
0.	5.	C				\$	5.	C				\$	1/5.	С.			\$	
8.	6.	C				C	6.	C				C	16/4	С.			C	
1.	7.	G				C	7.	G				C	1/	G.			C	
4.	8.	G				T	8.	G				T	8.	GCA	*		T	
9.	9.	G			٠	C	9.	G				C	9.	G.			C	
10	10.	G				G	10.	G				G	10.	G.			G	
11	11.	T				G	11.	T				G	11.	т.			G	
3.	12.	T	٠			A	12.	T		٠		A	12.	т.		٠	A	

#### Aligning reads is complicated by:

- Imperfect match to the reference due to mutations or sequencing errors, or errors in the reference genome
- Multiple positions where the read could match (repeated regions)
- Low quality of the read
- With paired-end reads: only one read maps or the other read maps on a different chromosome



Alignment tools such as bwamem2 are able to handle all of these complications and give information on the mapping quality.

## Regions that are repeated in the genome make it very difficult to map reads

Reference genome



Read to be aligned to the reference



## Regions that are repeated in the genome make it very difficult to map reads

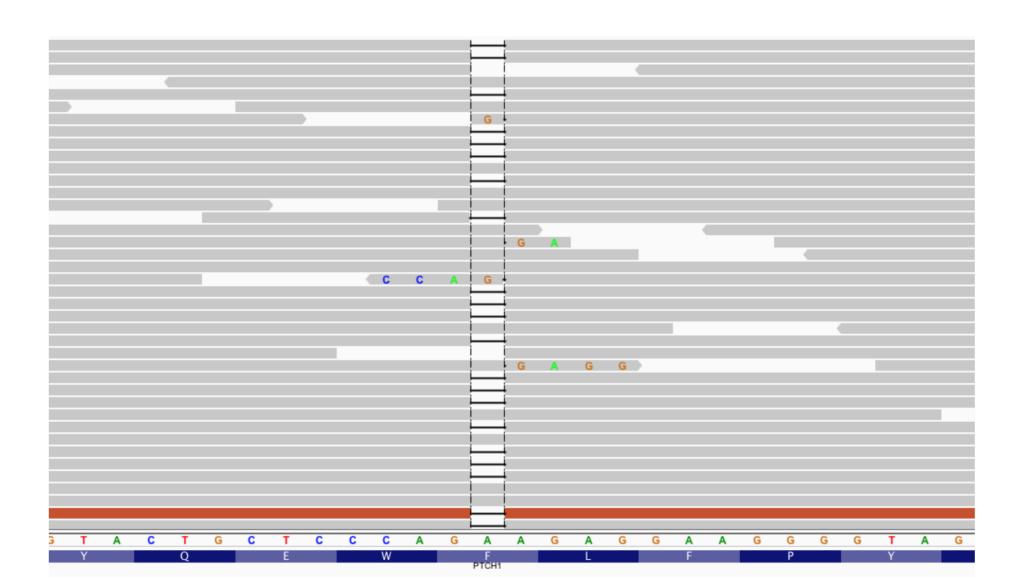
Reference genome



Read to be aligned to the reference



#### Indels: deletions and insertions



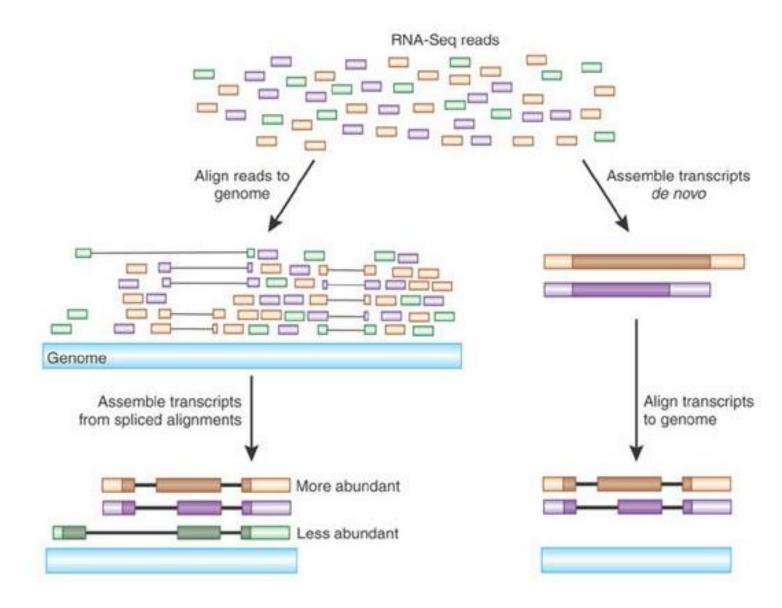
#### Aligning other type of data: RNA data

Tools:

e.g.

**STAR** 

HISAT2



## PacBio mapping

Karin Näsvall



Sequel IIe Instrument

Data Transfer/SMRT Link Server

Generate Sequencing Data

#### Sequel lle System output files

```
<your_specified_output_directory>/r64009_20200825_221039/1_A01/
|-- m64009_200825_222052.baz2bam_1.log
|-- m64009_200825_222052.ccs.log
|-- m64009_200825_222052.ccs_reports.json
|-- m64009_200825_222052.ccs_reports.txt
|-- m64009_200825_222052.consensusreadset.xml
|-- m64009_200825_222052.reads.bam
|-- m64009_200825_222052.reads.bam.pbi
|-- m64009_200825_222052.sts.xml
|-- m64009_200825_222052.transferdone
|-- m64009_200825_222052.zmw_metrics.json.gz
```



#### **SMRT Link GUI**

Access using a web browser

#### Automatic HiFi reads generation (Export Reads)

hifi\_reads.fastq.gz- FASTQ file containing HiFi Reads hifi\_reads.fasta.gz- FASTA file containing HiFi Reads hifi\_reads.bam- BAM file containing HiFi Reads

HiFi reads QV > 20

https://pacbiofileformats.readthedocs.io/en/13.0/

https://pacbiofileformats.readthedocs.io/en/13.0/BAM.html#hifi-reads

- \$ samtools view —H /pacbio/m84093\_240426\_124306\_s1.ccs.bc2033.rmdup.bam |head =n2
- @HD VN:1.6 S0:coordinate pb:5.0.0
- @RG ID:f56a67e5/0--0 PL:PACBIO
  DS:READTYPE=CCS;Ipd:CodecV1=ip;PulseWidth:CodecV1=pw;B
  INDINGKIT=102-739-100;SEQUENCINGKIT=102-118800;BASECALLERVERSION=5.0;FRAMERATEHZ=100.000000;Barco
  deFile=/lustre/scratch123/tol/resources/barcodes/PacBi
  o\_ULI\_adapter.fasta;BarcodeHash=cf95303a081e62fbaedf29
  888b16fdb7;BarcodeCount=1;BarcodeMode=Symmetric;Barcod
  eQuality=Score\_LB:TRAC-2-8589
  PU:m84093\_240426\_124306\_s1\_SM:Meier\_Genomes13637824
  PM:REVIO\_BC:AAGCAGTGGTATCAACGCAGAGTACT\_CM:R/P1-C1/5.025M

https://pacbiofileformats.readthedocs.io/en/13.0/BAM.html#hifi-reads

- \$ samtools view /pacbio/m84093\_240426\_124306\_s1.ccs.bc2033.bam |head -n2
- ----

```
cat m84093_240426_124306_s1.ccs.bc2033.stats
-
A = 7108650862 (34.9%), C = 3058767823 (15.0%), G = 3061827434 (15.0%),
T = 7127816130 (35.0%), CpG = 1003156910 (4.9%)
sum = 20357062249, n = 2218131, mean = 9177.57438537219, largest = 30301, smallest = 135
```

### PacBio mapping – Minimap2

Minimap2 <a href="https://github.com/lh3/minimap2">https://github.com/lh3/minimap2</a>

Uses **minimizers** – short sequences of length –k [default 15 bases]

- 1. Extracts minimizers from the reference (target) and index them
- 2. Match each minimizer in the query sequence against the reference set of minimizers
- 3. Sorts the position of each minimizer after position
- Make a chain of the minimizers
- 5. Repeat for all query sequences

Minimap2 -ax map-hifi target.fa query.fa > output.sam

Align PacBio high-fidelity (HiFi) reads to a reference genome (-k19 -w19 -U50,500 - g10k -A1 -B4 -O6,26 -E2,1 -s200)

- Preprocessing
  - Filtering adapters (blastn)
- Alignment
  - MINIMAP2
- Alignment post-processing
  - Statistics



https://pipelines.tol.sanger.ac.uk/readmapping