

1 Introduction: Nanoparticles in medicine

1.1 Polymeric colloids

- 1.1.1 Functionalization for protein binding
- 1.1.2 Polymerization consequences (initiator, co-monomer, surfactants...)

1.2 Liposomal nanocarriers (formation for amphiphilic lipids)

- 1.2.1 phospholipid bilayer (typical molecules HSPC, DPPC, cholesterol, PEG...)
- 1.2.2 polydispersity control (extrusion, paper w/ Zoltan)
- 1.2.3 Drug carrier, SSLs (stealth function, bilayer stability, filling)

1.3 Physicochemical characterization

- 1.3.1 Dimensional metrology and traceability
- 1.3.2 Characterization tools
 - 1.3.2.1 Single-particle (AFM, TEM, SEM, TSEM...)
 - 1.3.2.2 Ensemble (DLS, DCS, SAXS)

2 Theoretical Background

2.1 Interaction of light and matter

2.2 Small Angle X-ray Scattering (SAXS)

- 2.2.1 Physical Process
- 2.2.2 Evaluation of the scattering intensity (Form factor * $S(q)$, electron density, number of colloids)
 - 2.2.2.1 What is q ?
 - 2.2.2.2 Modelling
 - 2.2.2.2.1 Sphere (Gudrun polymeric colloids)
 - 2.2.2.2.2 Core-shell (interface effects???)
 - 2.2.2.2.3 Vesicle model (5 gaussian) ????
 - 2.2.2.2.4 Inclusion of background ??? ($a+b*q^{-4}$)
 - 2.2.2.3 Guinier approach (deviation when using few points, polydispersity...)
- 2.2.3 Contrast variation (solvent variation, ASAXS...)
 - 2.2.3.1 Isoscattering point
 - 2.2.3.1.1 Polydispersity and ellipticity smearing (simulation, calculation)
 - 2.2.3.2 Basic functions approach
 - 2.2.3.2.1 Shape factor
 - 2.2.3.2.2 Guinier rule
 - 2.2.3.2.3 $I(0)$ for polydisperse systems

3 Experimental setup

3.1 BESSY II

3.2 FCM Beamline

- 3.2.1 Transmission measurements (calibrated diodes)

3.3 SAXS

- 3.3.1 Pilatus detector (high dynamic range, noise free...)

- 3.3.2 HZB SAXS setup

- 3.3.2.1 distance calibration (10^{-4} uncertainty)

- 3.3.3 Radial integration and error propagation

- 3.3.4 Absolute intensity calibration

- 3.3.4.1 Flux monitor (thin diode)

- 3.3.4.2 Detector efficiency (pilatus, thin diode)

3.4 Continuous contrast variation

- 3.4.1 Filling of capillaries (galden at bottom, reference layer)

- 3.4.1.1 Capillary homogeneity (Hilgenberg)

- 3.4.2 Calibration of solvent density and finding of main axis

- 3.4.3 Limitations

- 3.4.3.1 Density range (sucrose, fructose, iodixanol)

- 3.4.3.2 Background, induced aggregation (heavy salts)

- 3.4.3.3 Comparison to SANS / RSoXS in polymeric Nps (H. Abe, 2006)

4 Contrast variation in SAXS with the density gradient technique (1st paper)

4.1 Materials and methods

- 4.1.1 Particle and chemicals

- 4.1.2 Diffusion time and calibration height (sucrose mass fraction formula)

4.2 Continuous contrast variation in SAXS on PS-PMMA nanoparticles

4.3 Model dependent evaluation

- 4.3.1 Core-shell form factor fit

4.4 Model-free approach to contrast variation data

- 4.4.1 Isoscattering point

- 4.4.1.1 Relative standard uncertainty

- 4.4.2 Guinier Region analysis

- 4.4.2.1 Average electron density

4.5 Summary

5 Simultaneous size and density determination of polymeric colloids (2nd paper)

5.1 Materials and methods

- 5.1.1 Particle and chemicals

- 5.1.2 Differential Centrifuge Sedimentation (DCS)

5.2 Technique validation for the determination of the particle size distribution

5.2.1 Inter-laboratory comparison of the mean particle size

5.2.2 Colloidal size distribution

5.3 Considerations about contrast variation data evaluation

5.3.1 Shape factor formalism (simulation, depending on number of curves)

5.3.1.1 Advantages / disadvantages

5.3.2 Isoscattering point formalism (simulation, dependent of many things)

5.3.2.1 Advantages / disadvantages

5.4 Determination of the particle physical density

5.4.1 Validation through comparison with DCS

5.4.1.1 Uncertainties (physical density inaccuracy, beam size)

5.4.2 Use for homogenous polymeric colloids (PMMA-COOH)

5.5 Summary

6 Continuous contrast variation applied to relevant relevant bio-materials (3rd paper)

6.1 Materials and methods

6.1.1 Caelyx: PEGylated liposomal doxorubicin

6.1.2 Iso-osmolar contrast agent: Iodixanol

6.1.3 Sterically Stabilized Liposomes (SSLs) of different sizes

6.1.4 Lipoproteins (HDL&LDL)

6.2 Traceable size determination of a liposomal drug

6.2.1 Isoscattering point approach

6.2.2 Shape factor calculation

6.3 Osmotic effects in liposomes

6.3.1 Application to drug-stabilized liposomes

6.3.2 Size dependency of the osmotic activity

6.4 Application to blood plasma components

6.4.1 HDL

6.4.2 LDL

6.4.3 Literature comparison

6.5 Protein-coated low-density nano-particles

6.5.1 Single-SAXS experiments (C. Minelli Paper)

6.5.2 Contrast variation (Isopoint subtraction)

6.6 Summary