Unix II and Bedtools

Bioinformatics Applications (PLPTH813)

Sanzhen Liu

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Outline

- more Unix apps: sort, find, awk, sed, wget
- data transfer
- bedtools

sort - sort lines of text files

```
sort -k 2n fruit.txt
cat fruit.txt
                            banana 5
orange 8
apple 6
                            apple
peach 12
                            orange 8
banana 5
                            peach 12
                            sort -k 2nr fruit.txt
sort fruit.txt
                            peach 12
apple 6
banana 5
                            orange 8
orange 8
                            apple 6
peach 12
                            banana 5
sort -k 2 fruit.txt
                            sort -k 1,2 fruit.txt
peach 12
                            apple
banana 5
                            banana 5
apple 6
                            orange 8
orange 8
                            peach 12
```

find - search for files in a directory hierarchy

Finding files >10M find . -size +10M # Finding files <10M</pre> find . -size -10M # find a file find -name "fruit.txt" # find a file in the current directory find -maxdepth 1 -name "fruit.txt"

find [pathnames] [conditions]

find - II

```
# find files containing a specific word in its name
find -name "fruit*"
# find files whose name are not "fruit.txt"
find -not -name "fruit.txt"
# find files modified within 30 minutes
find . -mmin -30
# find files modified within 1 day
find \cdot -mtime -1
# find files accessed within 1 hour.
find . -amin -60
```

awk - I

• **awk**: a programming language designed for text processing and typically used as a data extraction and reporting tool.

```
cat fruit.txt
orange
         8
apple 6
peach 12
         5
banana
awk '{print $1}' fruit.txt # output first field
orange
apple
peach
banana
```

awk - II

```
awk 'BEGIN {start_action} {action} END {stop_action}' filename

# add up values in a column
awk 'BEGIN {sum=0} {sum=sum+$2} END {print sum}' fruit.txt
31

# print lines that satisfying certain conditions
awk '{if ($2 > 10) print }' fruit.txt
peach 12
```

awk - III:

```
# NF - Number of fileds variable:
awk '{print NF}' fruit.txt
2
 NR - number of records variable:
awk 'END {print NR}' fruit.txt
4
# length of strings
awk '{print length($1)}' fruit.txt
tolower(string)
toupper(string)
```

sed - a stream editor used for modifying files in unix

```
fruit.txt
                                            orange 8
sed 's/apple/strawberry/' fruit.txt
                                            apple 6
orange 8
                                           peach 12
strawberry 6
                                           banana 5
peach 12
banana 5
sed 's/apple/strawberry/g' fruit.txt
orange 8
strawberry 6
peach 12
banana 5
```

sed - II

```
sed 's/apple/{&}/' fruit.txt
orange 8
{apple} 6
peach 12
banana 5
sed '/12/ s/peach/kiwi/' fruit.txt
orange 8
apple 6
kiwi 12
banana 5
```

fruit.txt

orange 8
apple 6
peach 12
banana 5

wget

```
wget <url link to a file>
wget <a ftp link>
```

example:

wget http://129.130.89.83/tmp/public/sequence.cost.png

scp

scp user@hostname:directory/remotefile localfile

scp <eid>@beocat.cis.ksu.edu:<path/files> .

Cyberduck



SFTP (SSH File Transfer Protocol)		
Server: URL:	beocat.cis.ksu.edu sftp://liu3zhen@beocat.cis.ksu.edu	Port: 22
Username:	liu3zhen	
Password:	•••••	
SSH Private Key:	Anonymous Login None	•
Add to Keychain	? Cancel	Connect

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BED format

The first three required BED fields are:

chrom - the chromosome
chromStart - the starting position of the feature; The first base is numbered 0.
chromEnd - the ending position of the feature in the chromosome or scaffold.
The chromEnd base is not included in the display of the feature.
For example, the first 100 bases of a chromosome are defined as
chromStart=0, chromEnd=100, and span the bases numbered 0-99.

The additional optional BED fields are:
name - Defines the name of the BED line.
score - A score between 0 and 1000
strand - Defines the strand - either '+' or '-'.
...

VCF format

```
##fileformat=VCFv4.2
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=file:///seq/references/1000GenomesPilot-NCBI36.fasta
##contig=<ID=20,length=62435964,assembly=B36,md5=f126cdf8a6e0c7f379d618ff66beb2da,species="Homo sar
##phasing=partial
##INFO=<ID=NS, Number=1, Type=Integer, Description="Number of Samples With Data">
##INFO=<ID=DP, Number=1, Type=Integer, Description="Total Depth">
##INFO=<ID=AF, Number=A, Type=Float, Description="Allele Frequency">
##INFO=<ID=AA, Number=1, Type=String, Description="Ancestral Allele">
##INFO=<ID=DB, Number=0, Type=Flag, Description="dbSNP membership, build 129">
##INFO=<ID=H2, Number=0, Type=Flag, Description="HapMap2 membership">
##FILTER=<ID=q10,Description="Quality below 10">
##FILTER=<ID=s50, Description="Less than 50% of samples have data">
##FORMAT=<ID=GT, Number=1, Type=String, Description="Genotype">
##FORMAT=<ID=GQ, Number=1, Type=Integer, Description="Genotype Quality">
##FORMAT=<ID=DP, Number=1, Type=Integer, Description="Read Depth">
##FORMAT=<ID=HQ, Number=2, Type=Integer, Description="Haplotype Quality">
#CHROM POS
                                         QUAL FILTER INFO
               TD
                         REF
                                ALT
                                                                                        FORMAT
20
       14370
               rs6054257 G
                                Α
                                             PASS
                                                     NS=3;DP=14;AF=0.5;DB;H2
                                                                                        GT:GQ:DP:HQ C
                                         29
20
       17330
                                         3
                                              q10
                                                     NS=3;DP=11;AF=0.017
                                                                                        GT:GQ:DP:HQ C
       1110696 rs6040355 A
                                             PASS
                                                     NS=2;DP=10;AF=0.333,0.667;AA=T;DB GT:GQ:DP:HQ 1
20
                                G,T
                                         67
20
       1230237 .
                                         47
                                             PASS
                                                     NS=3;DP=13;AA=T
                                                                                        GT:GQ:DP:HQ C
                                G,GTCT
20
       1234567 microsat1 GTC
                                         50
                                            PASS
                                                     NS=3;DP=9;AA=G
                                                                                        GT:GQ:DP
```

coverage

bedtools coverage -abam \$bam -b \$bed

```
Bed input: Interval_1 1 16452 10 -3.84 ...
```

Output:

```
1 2 3 4
Interval_1 1 16452 10 -3.84 5432 16302 16451 0.9909
```

- 1. Read number
- 2. Coverage (bp)
- 3. Interval length
- 4. Coverage (%)

annotate and intersect

```
Input 1: #Chr chrStart chrEnd Len
```

10 10006596 10023047 16451

Input 2: #CHR POS REF ALT

10 64 C G

bedtools annotate -i Input1 -files Input2 -both

bedtools intersect -wo -a Input1 -b Input2

* -wo: write the original A and B entries plus the number of basepairs of overlap between the two features.

closest

```
# find the closest, non-overlapping gene for each interval where # both experiments had a peak # -io ignores overlapping intervals and returns only the closest, # non-overlapping interval (in this case, genes)
```

bedtools closest -a both.bed -b genes.bed -io

slop & complement

```
# Step 1. Add 500 bp up and downstream of each probe bedtools slop -i probes.bed -b 500
```

```
# Step 2. Get a BED file of all regions not covered by the probes (+500 bp up/down) bedtools complement -i p.500bp.bed -g hg18.genome
```

window

#Report all genes that are within 10000 bp upstream or downstream of xxx.

```
bedtools window -a xxx.bed -b genes.bed -w 10000
```

Report all genes that are within 10000 bp upstream or 5000 bp downstream of xxx.

```
bedtools window -a xxx.bed -b genes.bed -l 10000 -r 5000
```

#Report all SNPs that are within 5000 bp upstream or 1000 bp downstream of genes. Define upstream and downstream based on strand.

```
bedtools window -a genes.bed -b snps.bed -l 5000 -r 1000 -sw
```

merge

Merge overlapping repetitive elements into a single entry.

```
bedtools merge -i example.bed
```

Merge overlapping repetitive elements into a single entry, returning the number of entries merged.

```
bedtools merge -i example.bed -n
```

Merge nearby (within 1000 bp) repetitive elements into a single entry.

```
bedtools merge -i example.bed -d 1000
```

Random

```
#Generate random sequences from the genome
bedtools random [options] -g <genome>
# typical options:
-1
       the length of the interval to generate
       the number of intervals to generate
-n
The genome file that is supplied takes the form:
      <chromName><TAB><chromSize>
    For example, Human (hg19):
    chr1
           249250621
    chr2 243199373
    ...
```

chr18_gl000207_random 4262

flanking

```
head -n2 genes.bed
chr1 134212701 134230065 Nuak2 8 +
chr1 134212701 134230065 Nuak2 7 +

bedtools flank -i genes.bed -g ref.chromsizes -1 2000 -r 0 -s >
prom.bed

This will give you the upstream regions based on strand as follows:
chr1 134210701 134212701 Nuak2 8 +
chr1 134210701 134212701 Nuak2 7 +
```

bedtools getfasta -fi ref.fa -bed prom.bed -fo prom.fa

- -I The number of base pairs that a flank should start from orig. start coordinate.
- -r The number of base pairs that a flank should end from orig. end coordinate
- -s Define -l and -r based on strand.
 e.g. if used, -l 500 for a negative-stranded feature,
 it will start the flank 500 bp downstream. Default = false.