# Class07 Machine Learning 1

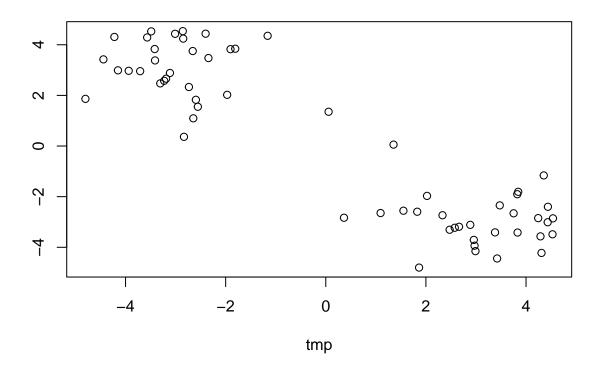
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#Clustering methods Find groups (aka) clusters in my data #K-means clustering

Make up some data to test with.

```
#Make up some data w/ 2 clear groups
tmp <- c( rnorm(30, mean = 3), rnorm(30, mean = -3) )
x <- cbind(tmp, rev(tmp))
plot(x)</pre>
```



The 'kmeans()' function does k-means clustering. >Q1. How many points are in each cluster?

```
k <- kmeans(x, centers=4, nstart=20)</pre>
## K-means clustering with 4 clusters of sizes 23, 23, 7, 7
## Cluster means:
##
          tmp
## 1 -3.172893 3.567209
## 2 3.567209 -3.172893
## 3 1.506122 -2.182678
## 4 -2.182678 1.506122
##
## Clustering vector:
## [1] 2 2 3 2 2 2 2 2 2 2 2 2 2 3 3 2 3 2 2 3 2 2 2 3 3 2 2 2 3 2 1 4 1 1 1 4 1 1
## Within cluster sum of squares by cluster:
## [1] 30.144489 30.144489 8.859244 8.859244
## (between_SS / total_SS = 93.6 %)
## Available components:
##
## [1] "cluster"
                    "centers"
                                   "totss"
                                                 "withinss"
                                                               "tot.withinss"
## [6] "betweenss"
                                                 "ifault"
                    "size"
                                   "iter"
```

We can use the dollar syntax to get at the results (components of the returned list).

What 'component' pf your result object details: -cluster size? -cluster assignment/membership? -cluster center?

### k\$size

## [1] 23 23 7 7

### k\$cluster

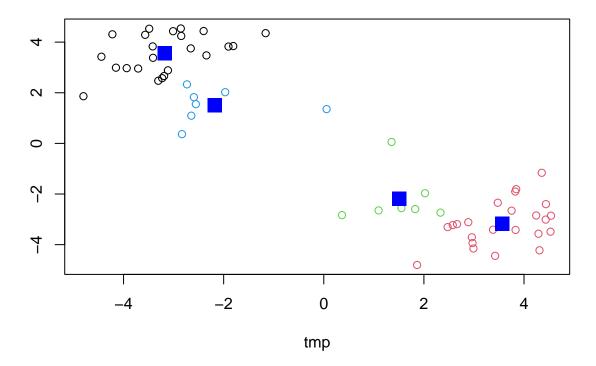
```
## [1] 2 2 3 2 2 2 2 2 2 2 2 2 2 3 3 2 3 2 2 2 2 2 3 2 2 2 3 2 1 4 1 1 1 4 1 1
```

#### k\$centers

```
##
          tmp
## 1 -3.172893 3.567209
## 2 3.567209 -3.172893
## 3 1.506122 -2.182678
## 4 -2.182678 1.506122
```

Q3. Plot x colored by the kmeans cluster assignment and add cluster centers as blue points.

```
plot(x, col=k$cluster)
points(k$centers, col="blue", pch=15, cex=2)
```



# ##Hierarchical Clustering

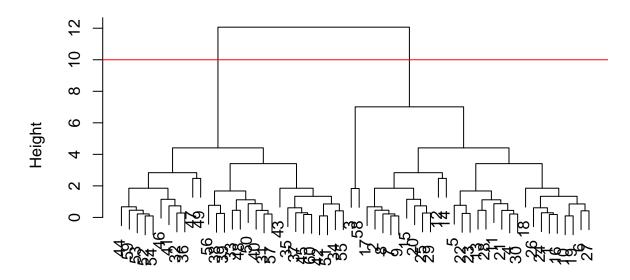
The hclust() function needs a distance matrix as input nor our original data. For this we use the 'dist(' function

```
hc <- hclust(dist(x))
hc

##
## Call:
## hclust(d = dist(x))
##
## Cluster method : complete
## Distance : euclidean
## Number of objects: 60

plot(hc)
abline(h=10, col="red")</pre>
```

# **Cluster Dendrogram**



dist(x)
hclust (\*, "complete")

To get our cluster membership vector we must cut our tree. For this we use 'cutree()'.

```
cutree(hc, h=10)
```

We can cut by a given height (h=) or into a given number of k groups w/k=.

```
cutree(hc, k=2)
```

#Principal Component Analysis

##PCA of UK food data

Let's read the data about the stuff people in the UK eat and drink:

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url, row.names=1)</pre>
```

Look at the first bit of file:

## head(x)

##		England	Wales	Scotland	N.Ireland
##	Cheese	105	103	103	66
##	Carcass_meat	245	227	242	267
##	Other_meat	685	803	750	586
##	Fish	147	160	122	93
##	Fats_and_oils	193	235	184	209
##	Sugars	156	175	147	139

How many columns in dataset:

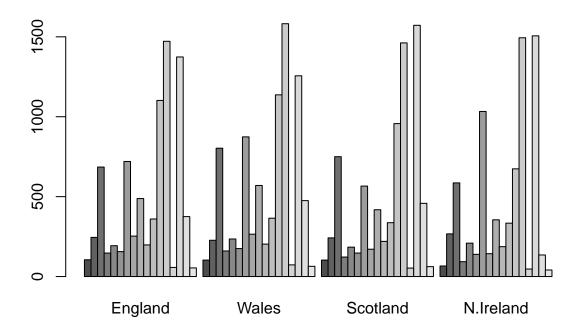
ncol(x)

### ## [1] 4

Q1. How many rows and columns are in your new data frame named x? What R functions could you use to answer this questions? 4 columns and 17 rows Q2. Which approach to solving the 'row-names problem' mentioned above do you prefer and why? Is one approach more robust than another under certain circumstances? Using the codeblock and running it multiple times eventually led to the removal of all the columns, so it's not my preferred approach as I feel like it's error-prone.

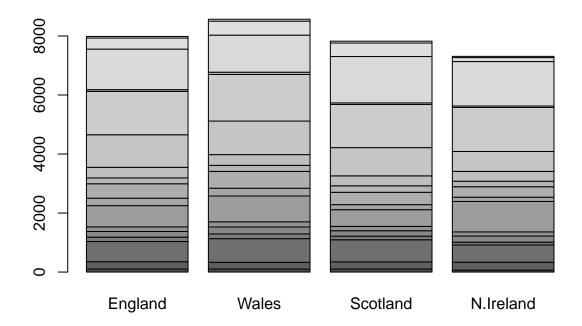
We can make some plots to try to understand this data a bit more. for example barplots:

barplot(as.matrix(x), beside=TRUE)



Q3: Changing what optional argument in the above barplot() function results in the following plot? Changing the 'beside' function from TRUE to FALSE would result in a stacked plot.

barplot(as.matrix(x), beside=FALSE)



## #PCA

The main base R function for PCA is 'prcomp()'.

```
PCA <- prcomp(t(x))
summary(PCA)</pre>
```

```
## Importance of components:

## PC1 PC2 PC3 PC4

## Standard deviation 324.1502 212.7478 73.87622 4.189e-14

## Proportion of Variance 0.6744 0.2905 0.03503 0.000e+00

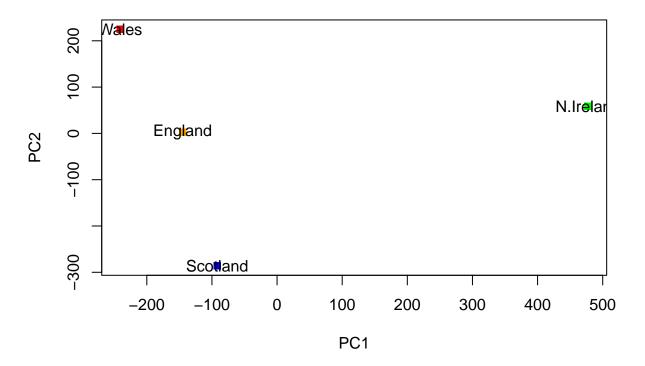
## Cumulative Proportion 0.6744 0.9650 1.00000 1.000e+00
```

What's in this returned PCA object?

### attributes(PCA)

```
## $names
## [1] "sdev" "rotation" "center" "scale" "x"
##
## $class
## [1] "prcomp"
```

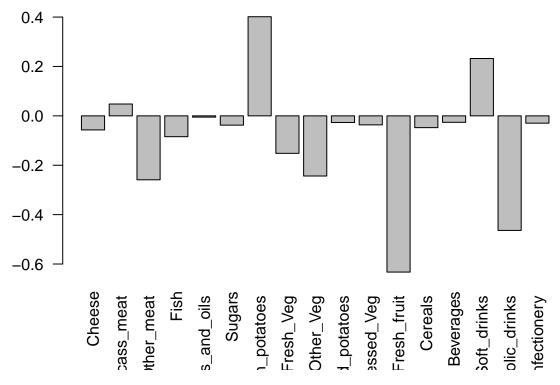
```
plot(PCA$x[,1:2], col=c("orange", "red", "blue", "green"), pch=15)
text(PCA$x[,1], PCA$x[,2], labels=colnames(x))
```



Q7. Complete the code to generate a plot of PC1 vs PC2. T Q8. Customize your plot so that the colors of the country names match the colors in our UK and Ireland map and table at start of this document.

We can look at how the variables contribute to our new PCs by examining the 'PCA\$rotation' component of our results.

barplot(PCA\$rotation[,1], las=2)



```
url2 <- "https://tinyurl.com/expression-CSV"
rna.data <- read.csv(url2, row.names=1)
head(rna.data)</pre>
```

```
##
                   wt3
                        wt4 wt5 ko1 ko2 ko3 ko4 ko5
## gene1
          439 458
                   408
                        429 420
                                 90
                                     88
                                         86
                                              90
## gene2
          219 200
                   204
                        210 187 427 423 434 433 426
## gene3 1006 989 1030 1017 973 252 237 238 226 210
                        856 760 849 856 835 885 894
## gene4
          783 792
## gene5
          181 249
                   204
                        244 225 277 305 272 270 279
## gene6 460 502
                   491
                        491 493 612 594 577 618 638
```

nrow(rna.data)

## [1] 100

ncol(rna.data)

## [1] 10

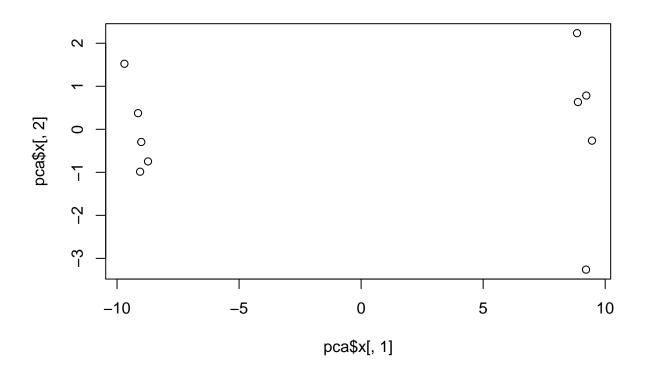
Q10: How many genes and samples are in this data set? 100 genes; 10 samples Let's do PCA of htis dataset; first take the transpose as that's what hte prcomp() function wants.

```
pca <- prcomp(t(rna.data), scale=TRUE)
summary(pca)</pre>
```

```
## Importance of components:
                             PC1
                                    PC2
                                            PC3
                                                    PC4
                                                             PC5
                                                                     PC6
##
                                                                             PC7
## Standard deviation
                          9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111
## Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642
## Cumulative Proportion 0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
                              PC8
                                      PC9
                                               PC10
## Standard deviation
                          0.62065 0.60342 3.348e-15
## Proportion of Variance 0.00385 0.00364 0.000e+00
## Cumulative Proportion 0.99636 1.00000 1.000e+00
```

We can make our score plot (aka PCA plot) from the 'pca\$x'.

```
plot(pca$x[,1], pca$x[,2])
```



Make a color vector by wt and ko

```
rep("red", 5)
```

```
## [1] "red" "red" "red" "red" "red"
```

```
rep("blue", 5)

## [1] "blue" "blue" "blue" "blue"

colvec <- c(rep("red", 5), rep("blue", 5))
plot(pca$x[,1], pca$x[,2], col=colvec, pch=15)</pre>
```

