

Catch-U-DNA

1st REVIEW MEETING

***WP6 integrated acoustic detection
of ctDNAs in serum***

Jobst Technologies GmbH

Capturing non-Amplified Tumor Circulating DNA with Ultrasound Hydrodynamics

WP6 (Jobst, Forth, AWS, UoC)

Integrated acoustic detection of ctDNAs in serum

➤ Objectives

- 1. Develop/optimize/characterize sensor array compatible with passive/active microfluidics maximizing sensitivity**
- 2. Integrate the measurement electronics, fluidics control, and assembled chips interface into one PC connective instrument.**

Task 6.1 Microfluidics and assembly (JOBST, FORTH, M1-M15)

Task 6.2 Fluidics control (JOBST, M10-M24)

Task 6.3 Instrument (AWS, JOBST, FORTH, M16-M30)

Achievements - Task 6.1 Microfluidics and assembly Benchmarks elaboration

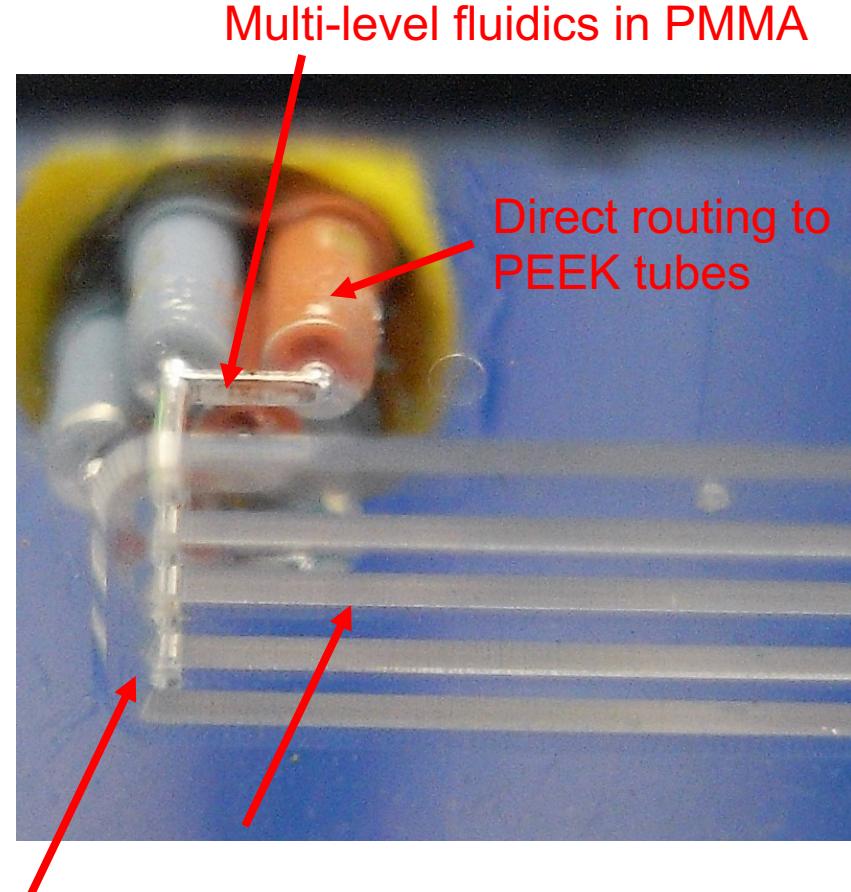
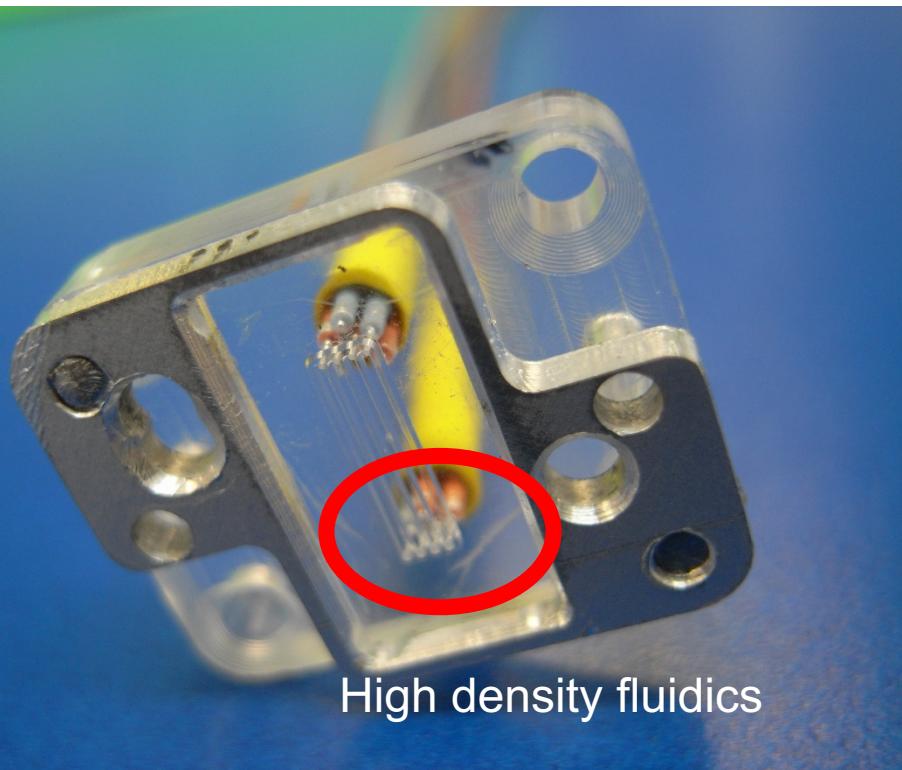
1. To enable high shear rate to allow for
 - Maximizing binding rate
 - Detaching of loosely bound materials
 - Regeneration of chip surface (detaching Avidin)
Applying **up to 100pN drag force** to a 200nm Liposome
2. To enable a fluid resting period to allow for
 - “threshold contact times for bond formation”
At around **10ms no-flow condition** for bond formation in a consecutive process
3. Not to break the 10microns thin sensing windows of the Quartz chip
 - Burst pressure calculations yield
A pressure limit of 1.0bar

$$Th = \sqrt{\frac{1.1 * (P) * (R)^2 * SF}{MR}}$$

Material	Modulus of rupture [psi]	K	Pressure [bar]	Diameter [mm]	Safety Factor SF	Thickness [mm]
Fused silica (SiO ₂)	7.000	1,1	1	0,4	1	0,010

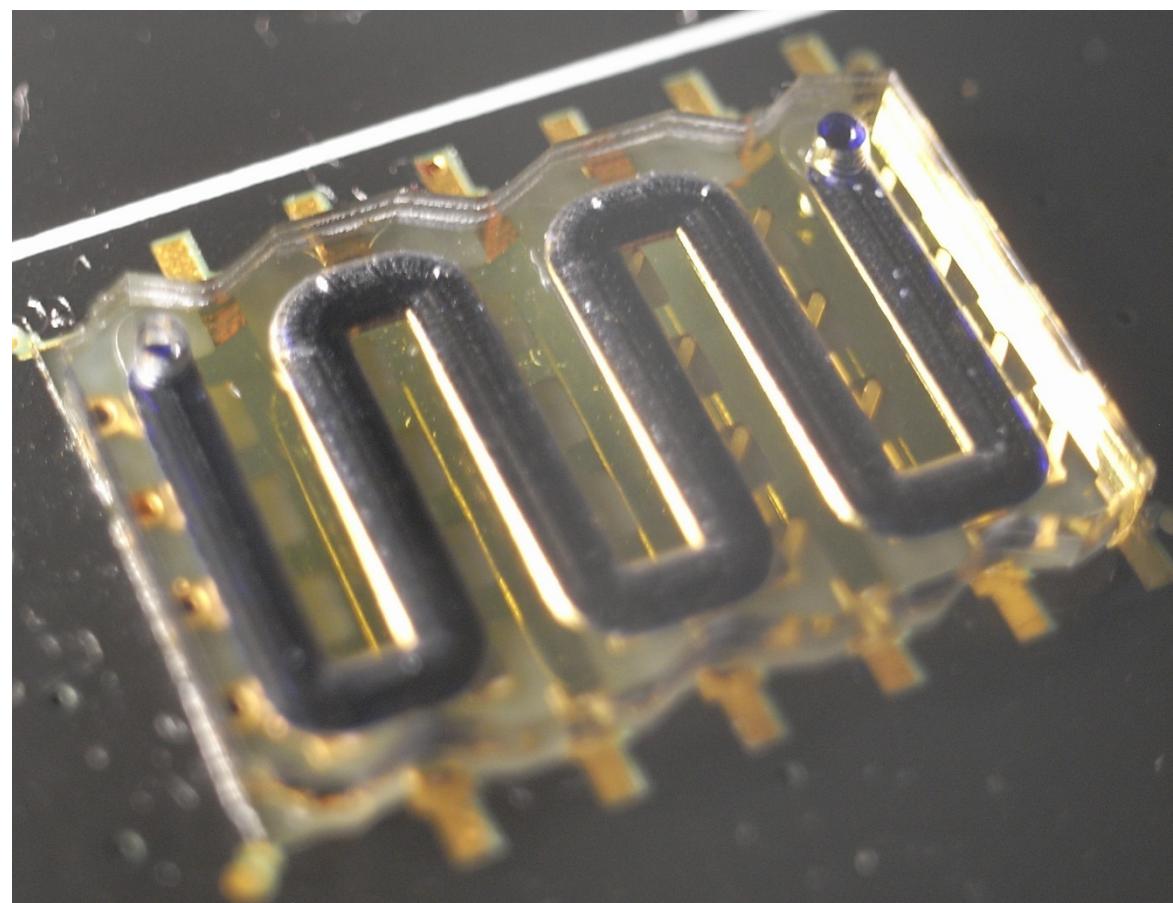
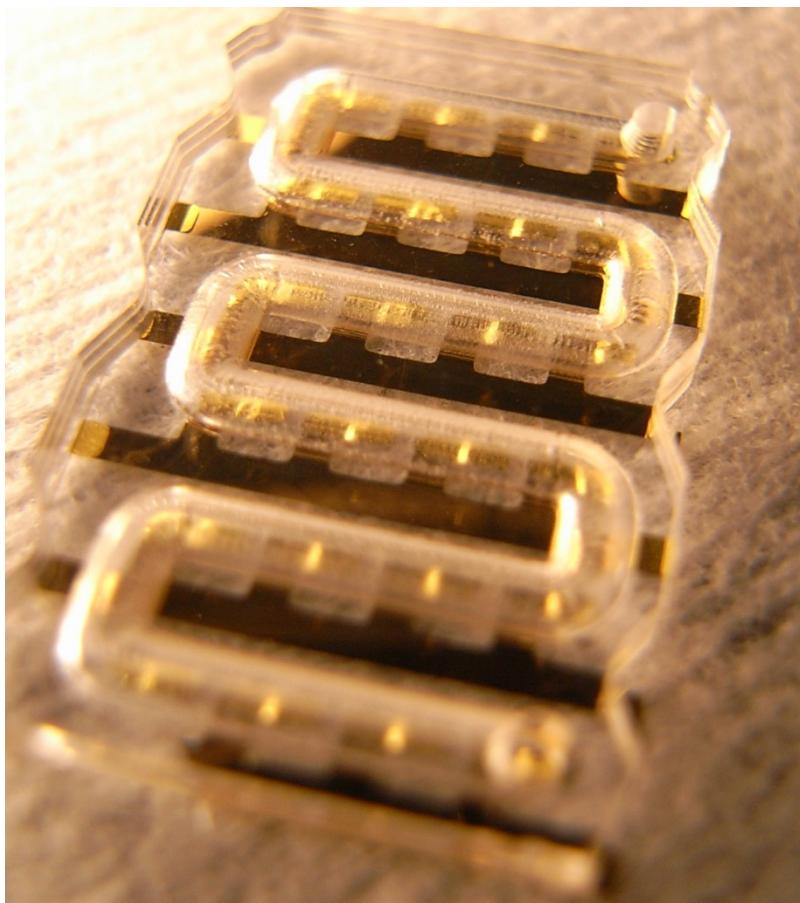
Achievements - Task 6.1 Microfluidics and assembly

Capabilities for reusable flow cell – to be used for evaluation work



Achievements - Task 6.1 Microfluidics and assembly

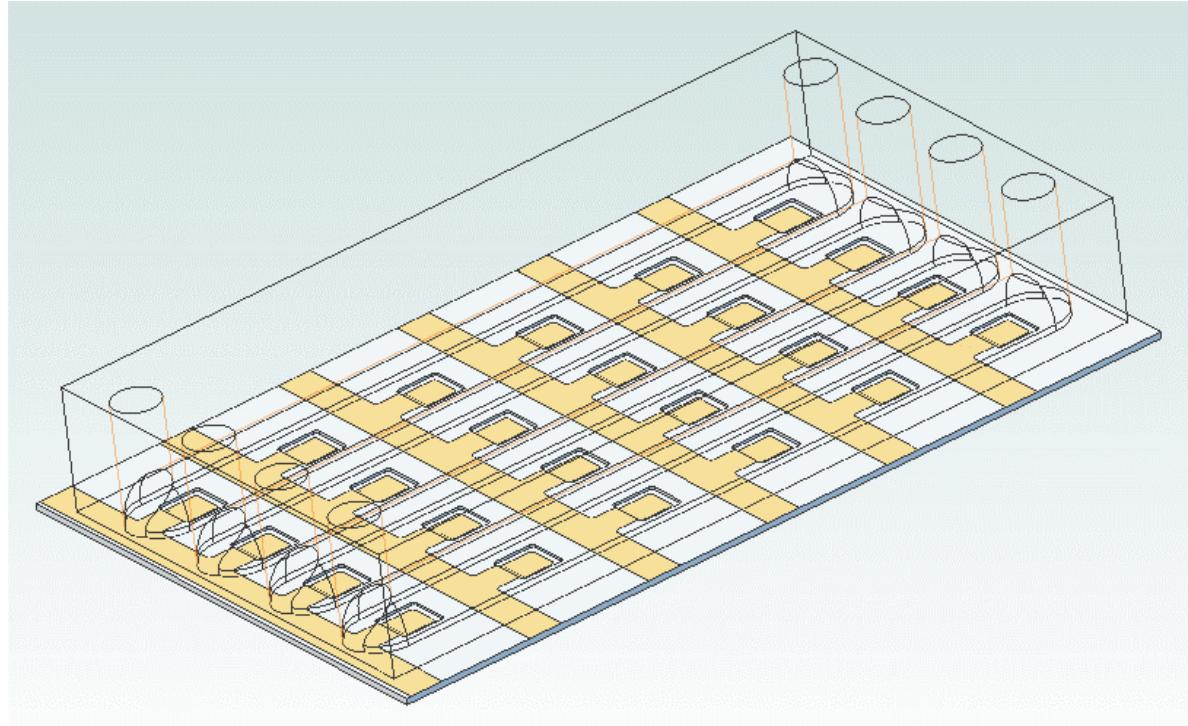
A flow cell permanently bonded to a 24x chip of AWS
(flow cells machined in PMMA and bonded with UV curing underfiller)



Achievements - Task 6.1 Microfluidics and assembly

But pressure drop adds up for each sensing site covered.

⇒ Make parallel channels with separate in- and outlets



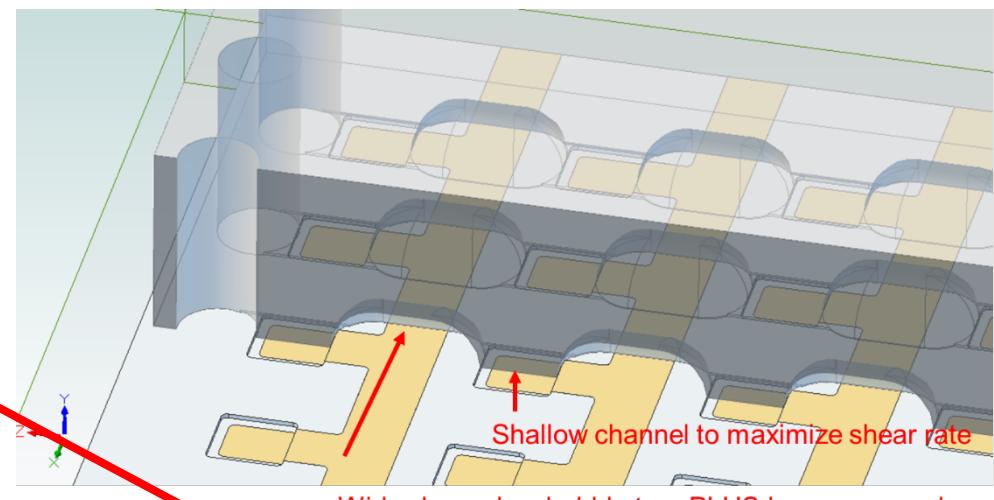
Achievements - Task 6.1 Microfluidics and assembly

⇒ And localize pressure drop to sensing site only

Just yields 50pN

At the burst limit

Just 25µm flow cell

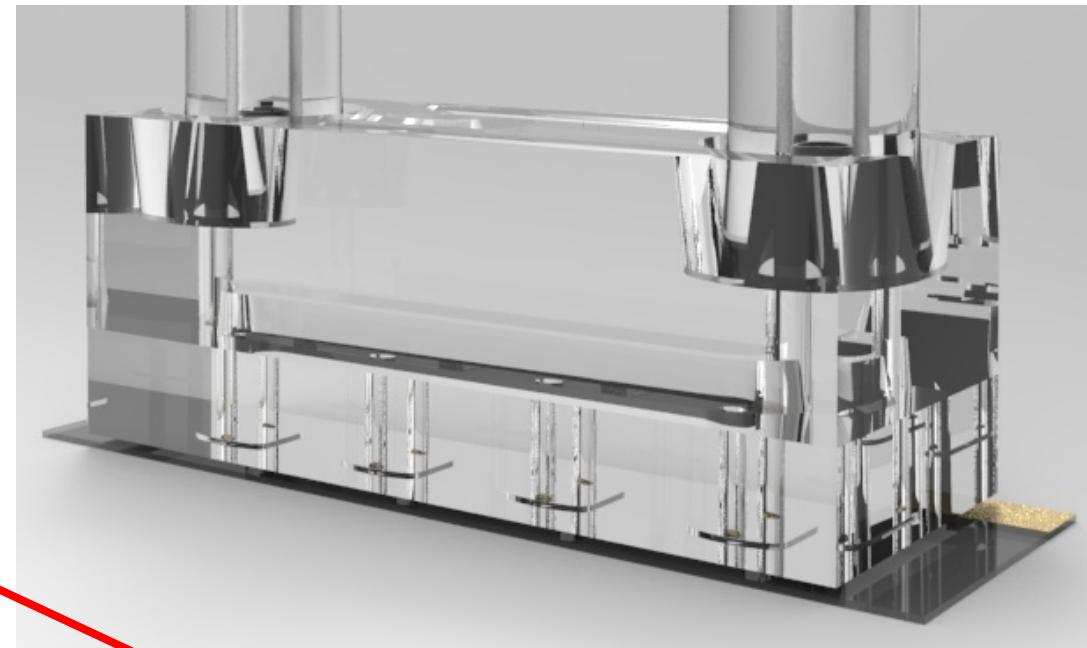


Scenario channel per 6 sensors

η [mPa.s]	1	SUM	Pressure drop calc for laminar flow only		$\tau = \frac{4\eta Q}{R^2} / \pi$	$T = \frac{6\eta Q}{h^2 W}$	Sphere diameter r_{um}
v. [$\mu\text{l}/\text{min}$]	1.650		dp [mbar]	dp [bar]			
conn. Tube			1.019,9	1,0			
in via	0,4	d/a [μm]					
Flow cell S1 6x	6	25					
out via	0,4	225					
conn. Tube	50	500					
		b [μm]	dp [mbar]	dp [bar]	% pD	v [mm/s]	Re
			8,964	0,009	0,9%	140,1	70,0
			1,749	0,002	0,2%	691,6	2,24
			998,514	0,999	97,9%	1.100,0	24,59
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Achievements - Task 6.1 Microfluidics and assembly

⇒ Flow cell for each single sensing site



Yields 100pN

Savely < burst limit

Scenario channel per 1 sensors

n [mPa.s] 1
v. [µl/min] 3.300

		dp [mbar]	dp [bar]
SUM		375,7	0,4

If value set:
width of rectangle
- otherwise tube

Pressure drop calc for laminar flow only!

Geometry	
round	$\tau = \frac{4\eta Q}{R^3}/\text{pi.}$
rect.	$\tau = \frac{6\eta Q}{h^2 W}$

Sphere diameter [µm]
0,2

Stokes drag [pN]

$$F_d = 6\pi \eta R v$$

	I [mm]	d/a [µm]	b [µm]	dp [mbar]	dp [bar]	% pD	v [mm/s]	Re
conn. Tube	50	500		17,927	0,018	4,8%	280,1	140,1
in via	0,4	225		3,497	0,003	0,9%	1.383,3	311,2
Flow cell S1	1	25	1000	332,838	0,333	88,6%	2.200,0	107,3
out via	0,4	225		3,497	0,003	0,9%	1.383,3	311,2
conn. Tube	50	500		17,927	0,018	4,8%	280,1	140,1

	dp [mbar]	dp [bar]	% pD	v [mm/s]	Re	Shear rate [s-1]
	375,7	0,4				
				280,1	140,1	4,48
				1.383,3	311,2	49,18
				2.200,0	107,3	528,00
				1.383,3	311,2	49,18
				280,1	140,1	4,48

99

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Sphere diameter [µm]
0,2

Stokes drag [pN]

$$F_d = 6\pi \eta R v$$

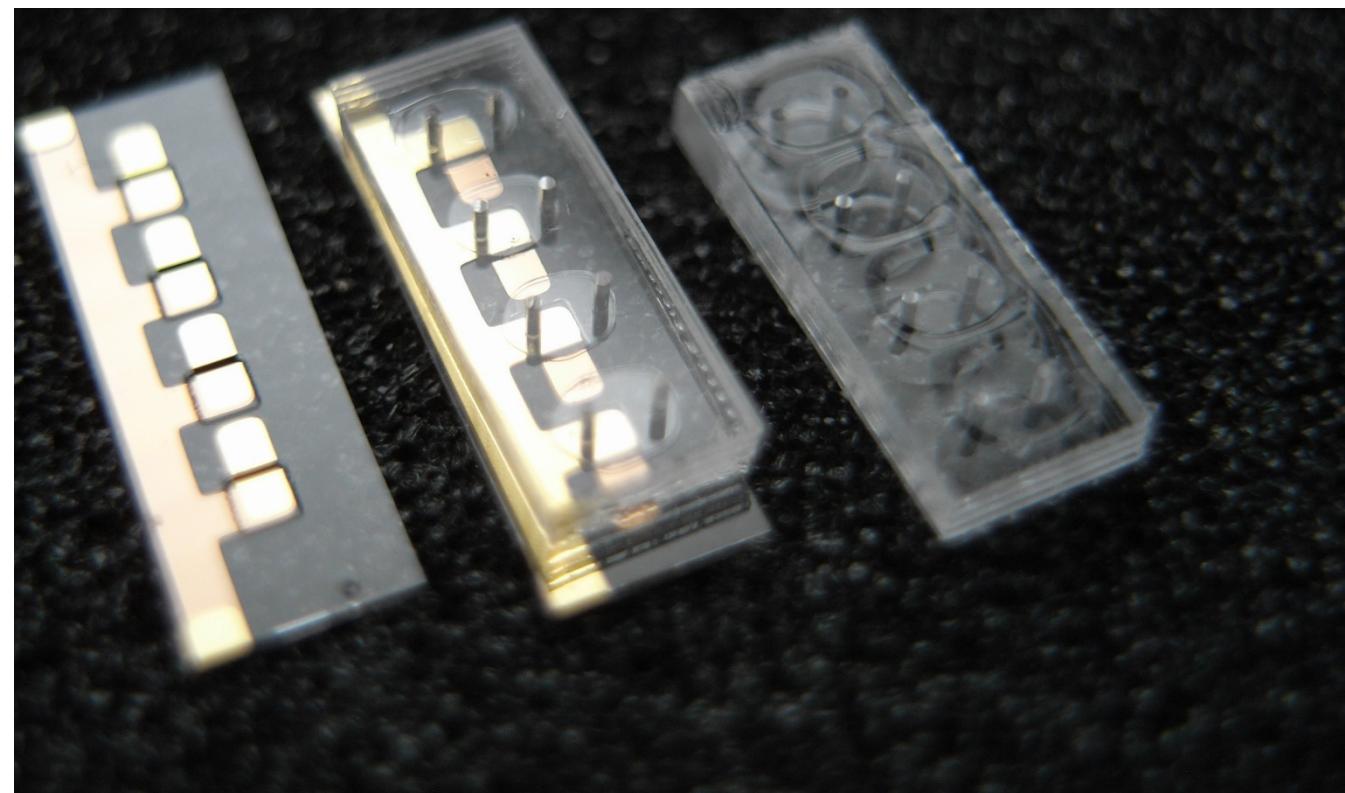
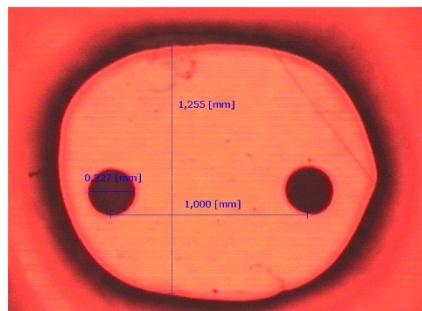
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Achievements - Task 6.1 Microfluidics and assembly

⇒ Flow cell for each single sensing site

First prototype
made

25µm flow cell
realized by
dispensing of
adhesive rubber
to flow cell body



Achievements - Task 6.1 Microfluidics and assembly

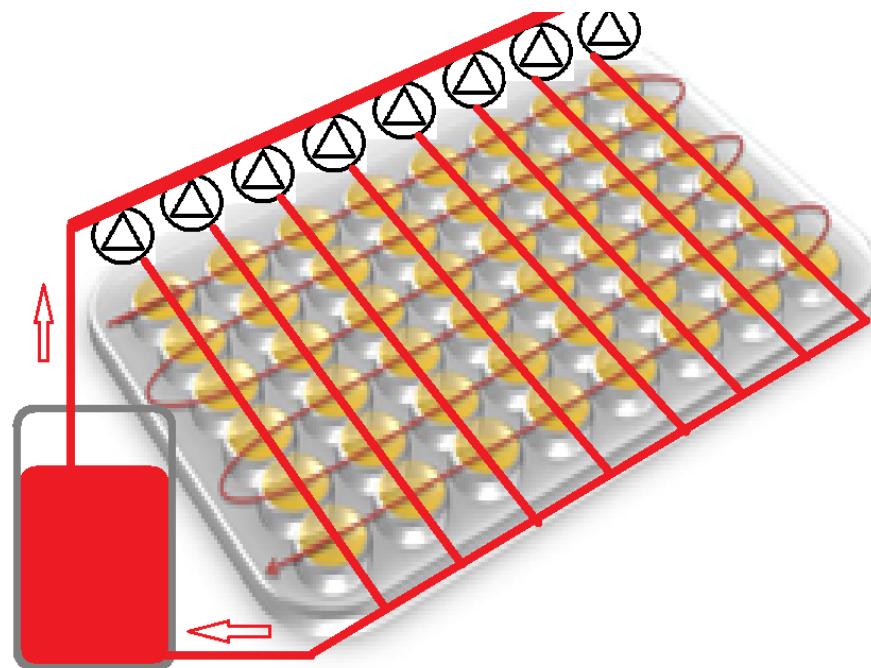
Conclusion:

- Required benchmarks elaborated
- Flow cells for AWS 24x and 4x chips designed
- Manufactured
- And assembled
- Successfully
- Performance follows expectations

Achievements - Task 6.2 Fluidics control

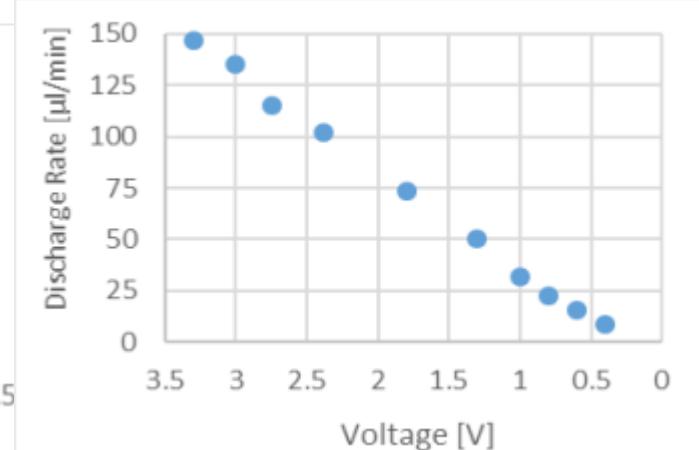
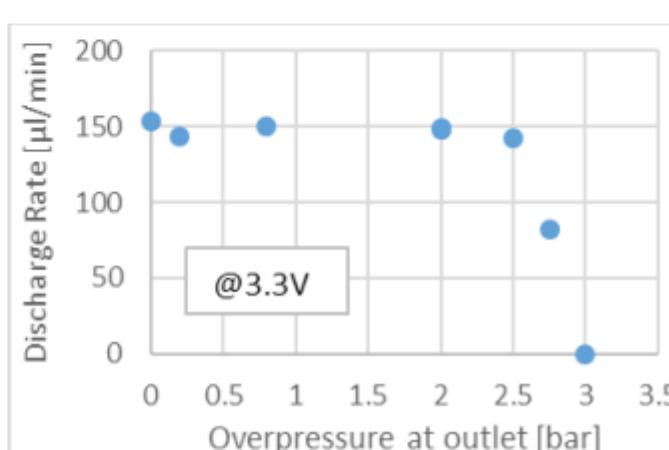
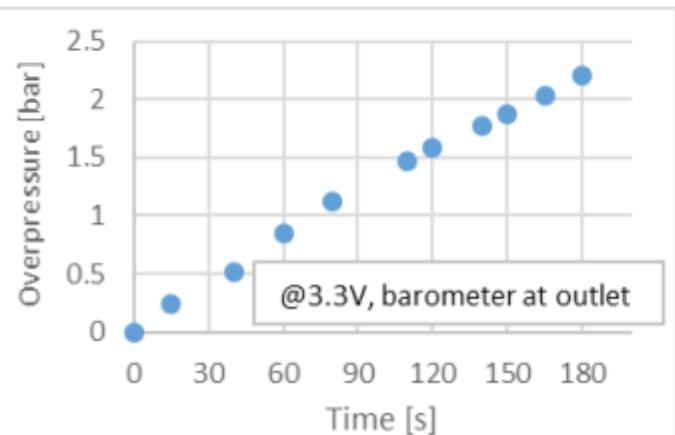
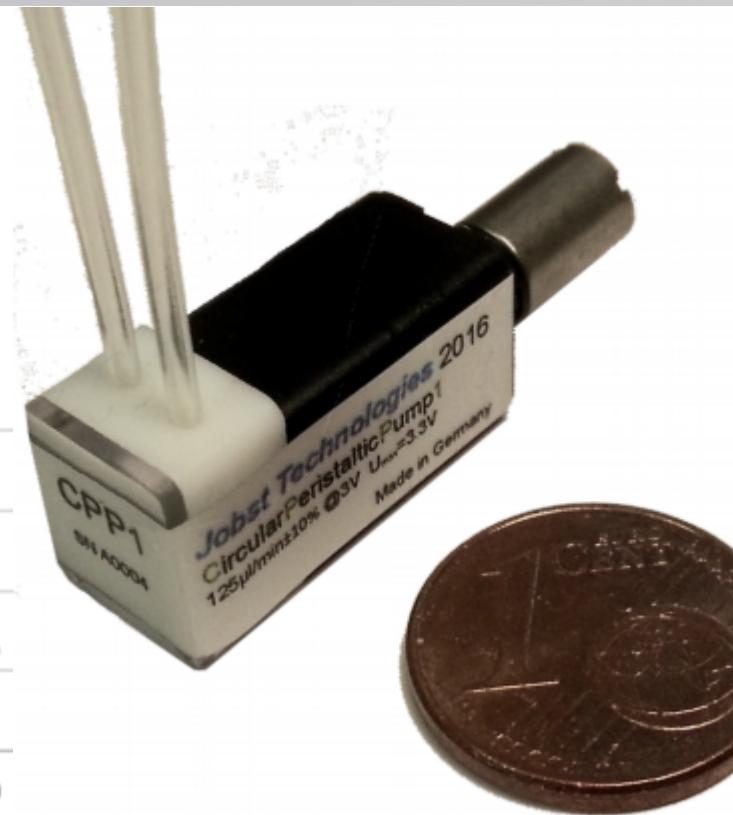
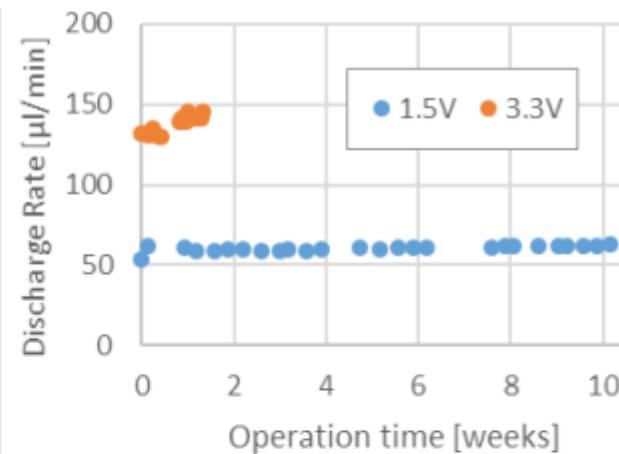
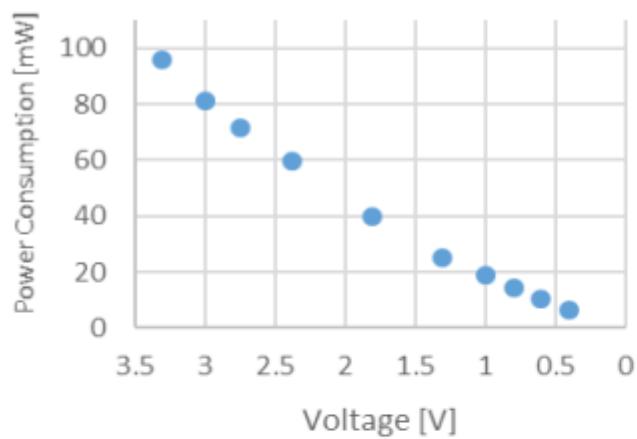
To achieve 100pN drag a total flow rate of 80ml/min is diverted into the 24 flow cells

Split into 8 lines feeding 6 flow cells each => 10ml/min per pump



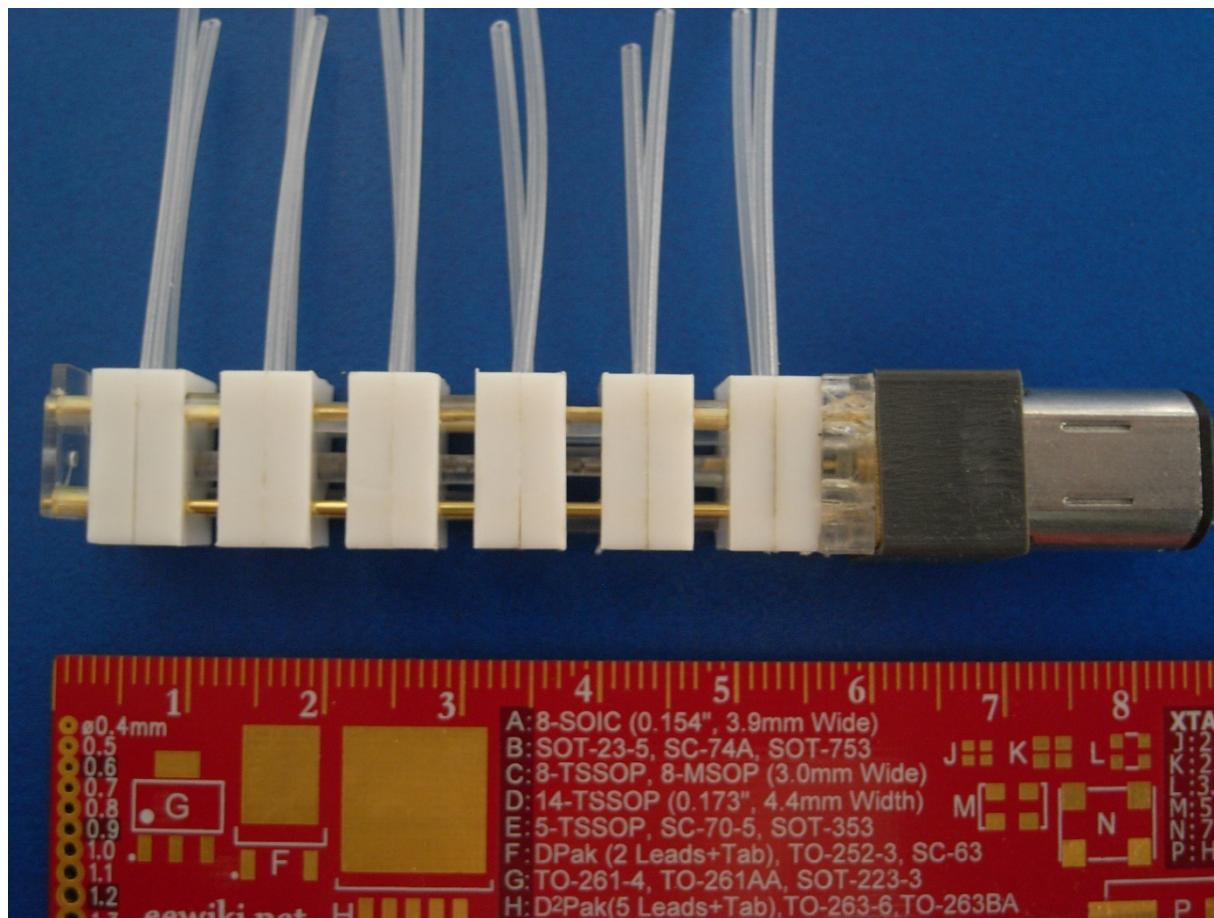
Achievements - Task 6.2 Fluidics control - pumps

JOBST Micro-pumps performance



Achievements - Task 6.2 Fluidics control - pumps

The first version of a compact 6 channel multi-head pump



Deviations

- WRT to DOW:
- **Deliverable D6.1** Report on electrical and fluidics assembly providing high signal quality and compatibility with bio-functionalization (M15)
- **D6.2** Report on sensitivity increase scaling by local and temporal shear rate engineering (M24)
- **D6.3** Final instrument prototype (M30)

Deviations

- WRT to DOW:
- **Milestone MS11** First generation chip assembly operational (M15)
- **MS12** Sensitivity by shear rate engineering >100 fold (M24)
- **MS13** Demonstration of electronics control unit (M30)

Future plans - Task 6.1 Microfluidics and assembly

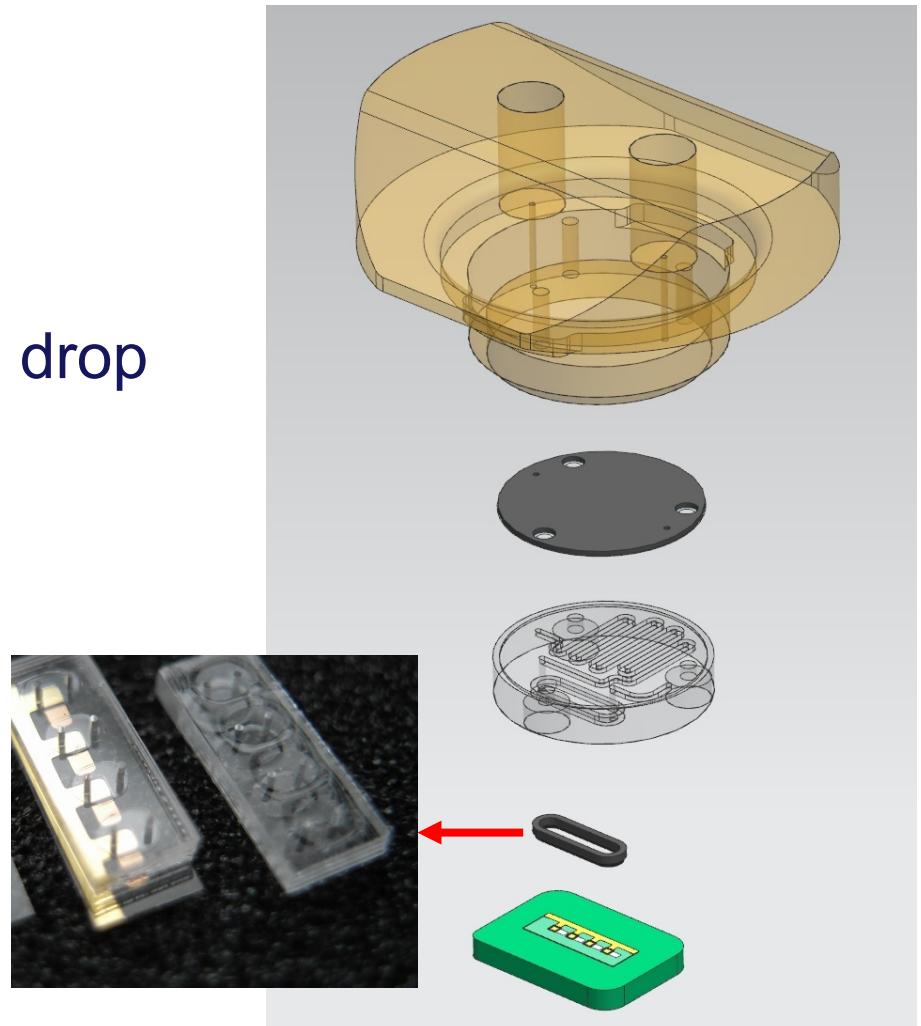
Realize integrated

Flow channel + seal manifold

For:

- Optimized shear @ low pressure drop
- Air bubble tolerant design
- Sealing
- Internal volume
- Thermal equilibration

Consider DNA electro-sorption



Future plans - Task 6.2 Fluidics control

- Get full process flow from sampling to readout elaborated
- Benchmark multi-channel pump
- Design pump heads delivering flow only for <50% of rotation
 - To enable >10ms resting period

Backup content...

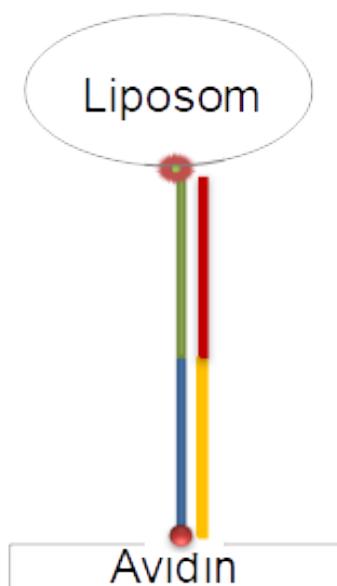
T 6.1 Microfluidics and assembly Shear force assisted performance improvement

Kinds of binding: **unspecific binding** of DNA, micro-particles:

- Minimizing the surface to volume ratio of the fluidics minimizes analyte loss due to unspecific binding to walls.
- Realize drag forces in the range of 1-100 pN to detach unspecific bound microspheres^a

a) M. Javanmard, F. Babrzadeh, R.W. Davis, "Microfluidic force spectroscopy for characterization of biomolecular interactions with piconewton resolution" Appl. Phys. Lett. 97, 173704

T 6.1 Microfluidics and assembly Shear force assisted performance improvement



Threshold contact times for bond formation:
10 milliseconds for antibody coated 4.5 microns micro-spheres^{b)}

Shear stress resistance:

sensor surface- avidin (adsorption on Au) +

avidin/ biotin@5' DNA +++

cholesterol@3' DNA / liposom ?

Method b: Direct immobilization to a Avidin-modified surface

b)..A. Pierres, A. Benoliel, C. Zhu, P. Bongrand, "Diffusion of Microspheres in Shear Flow Near a Wall: Use to Measure Binding Rates between Attached Molecules" Biophysical Journal 2001, 81, 25– 42