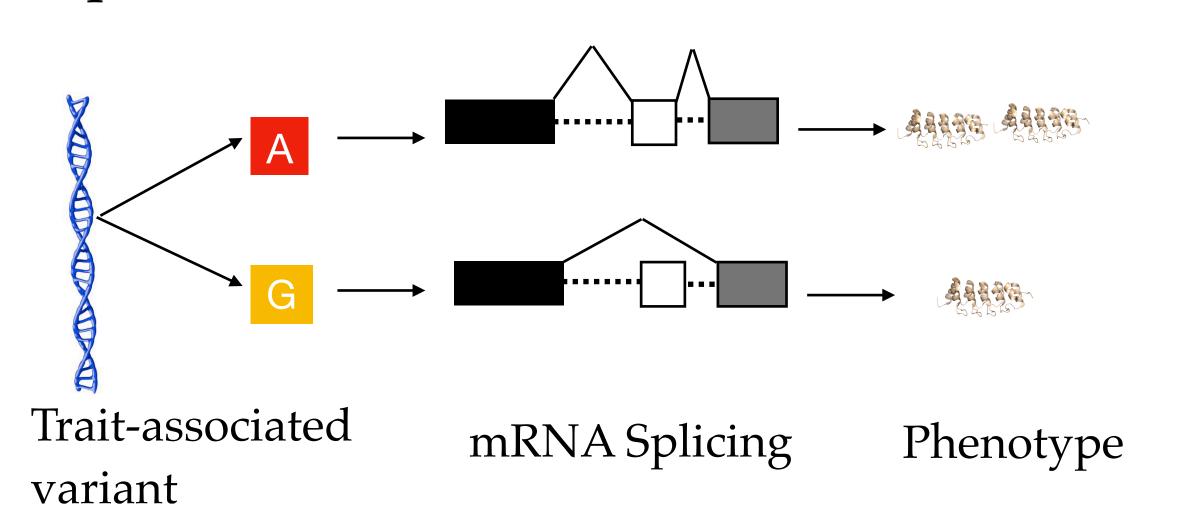


Elucidating the role of mRNA splicing in disease pathogenesis Ankeeta Shah¹, Yang I Li^{2,3}

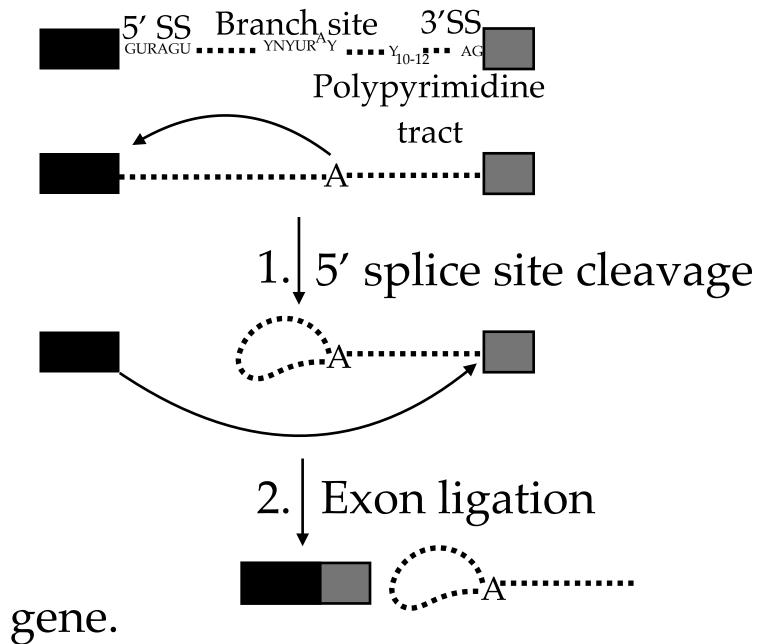
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75% of disease-associated genetic variants may disrupt gene regulation in a manner independent of promoters and enhancers*.

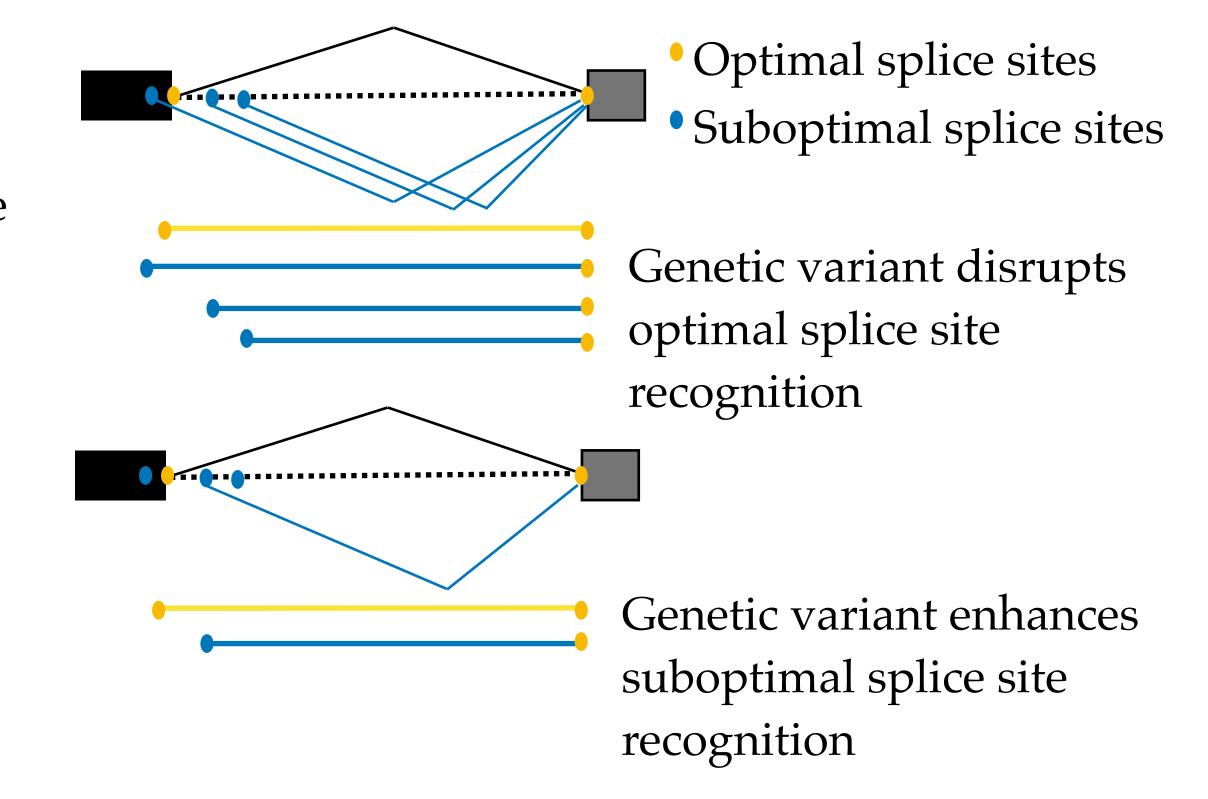


mRNA splicing can create multiple transcripts from the same gene. However, the function of many of these alternative transcripts is unknown (i.e. a result of splicing errors/noisy splicing, Pickrell et al., PLoS Genetics, 2010). *Chun et al., Nature Genetics, 2017.

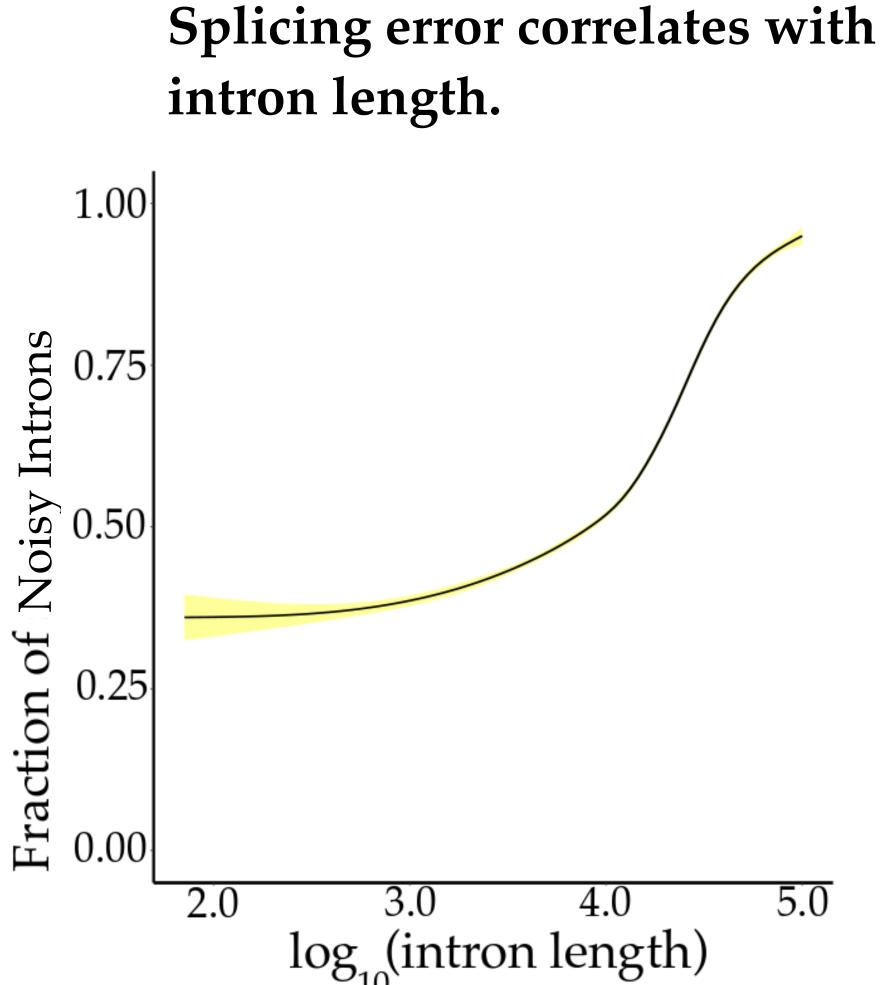
mRNA splicing involves two transesterification reactions.

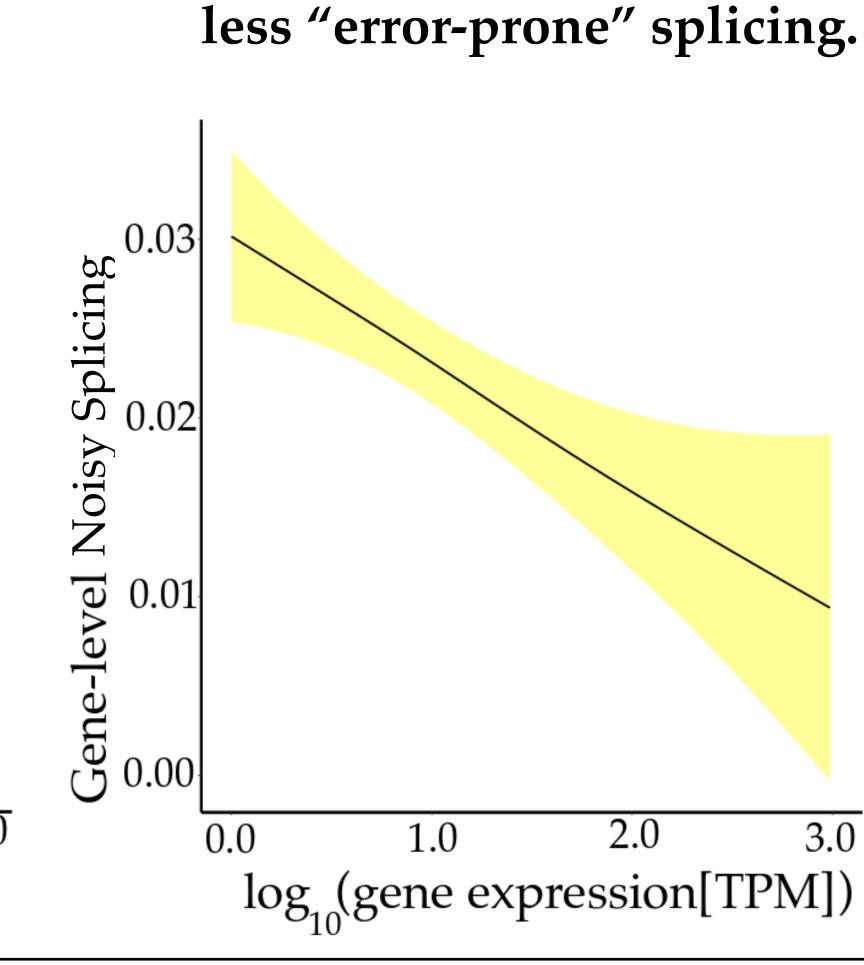


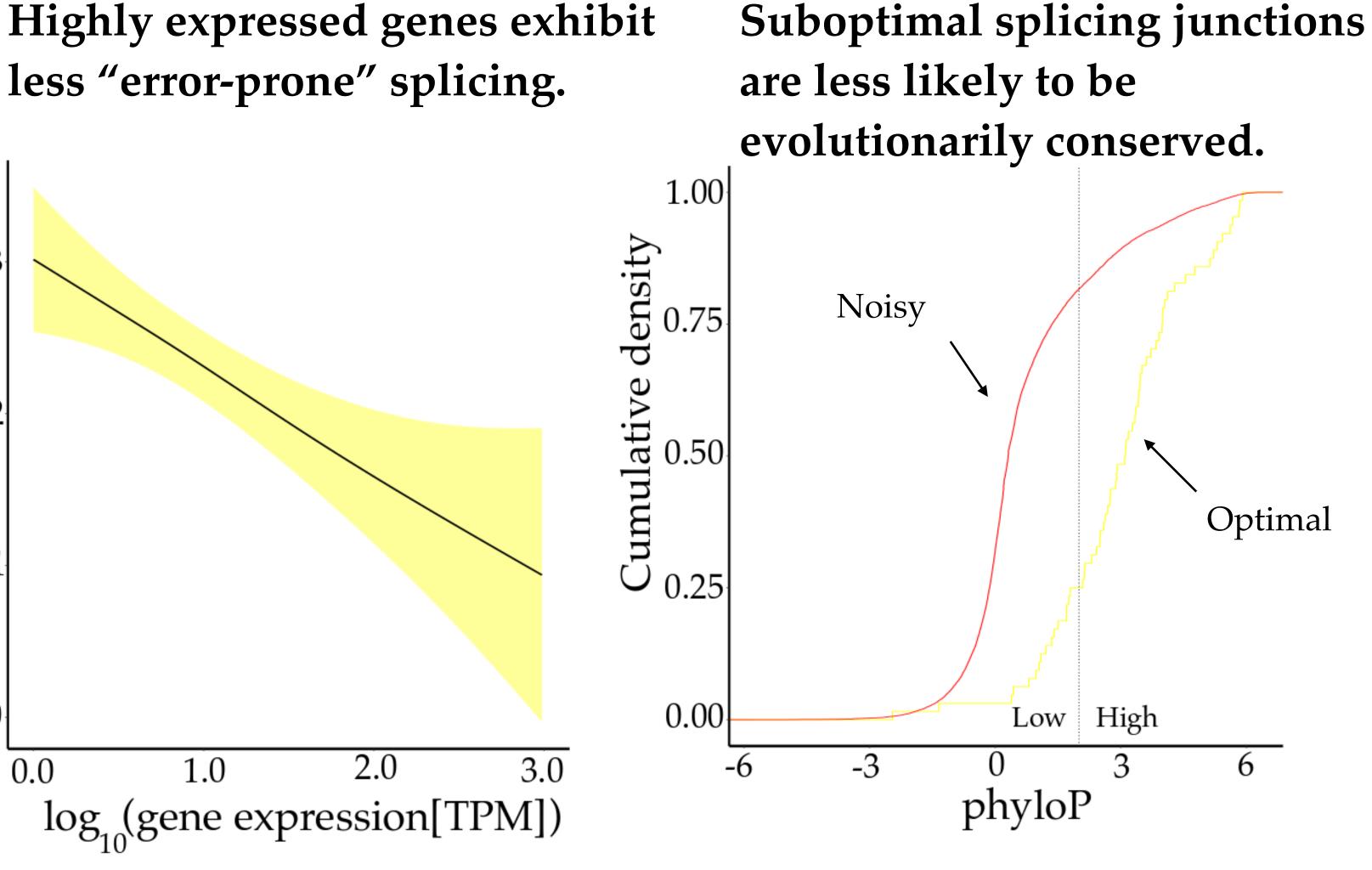
Hypothesis: genetic variants that increase risk of disease through splicing may do so by reducing splicing accuracy.



494,296 splicing junctions extracted from Yoruba **GEUVADIS*** lymphoblastoid cells lines (LCLs) 204,159 "noisy" splicing junctions** 290, 137 optimal splicing junctions *Lappalainen et al., Nature, 2013 **Noisy splicing events called using an adaptation of LeafCutter (Li et al., *Nature Genetics*, 2018)







Splicing quantitative trait loci (sQTLs) can be explained by SNPs that (a) disrupt recognition of splice sites or (b) enhance recognition of "noisy" splice sites.

individual i T/T $Y_i = X_i \beta + \epsilon_i$

Genotype for

splicing nosiness) for individual i Noisy sQTL eQTL sQTL Yoruba

2,553

Aggregated "noisy"

event usage (i.e.

LCLs

(N = 89)

FastQTL, Ongen et al., Bioinformatics, 2016

2,096

T/G G/G G allele at rs6808944 increases use of a single cryptic SS

 $0.12 \mid \beta = -0.218073$ 0.04T/G G/G

T/G

G/G

0.8 + p < 2.05264e-18, $\beta = 1.89595$

Noisy sQTL

0.6

n. 0.4

0.2

0.50 0.25 T/G G/GFunctional transcript Individual noisy introns $0.6 \mid \beta = 1.51729 *$

p < 1.62597e-27, $\beta = -1.79843$

sQTL

1.00

0.75

T/G G/GNon-functional transcript

178

Error

Effect size

Conclusions and Future Directions

Having gained intuition about how noisy splicing can explain how some SNPs act to contribute to variation in splicing, we will test the hypothesis that SNPs that increase disease risk through splicing likely do so by reducing splicing accuracy (e.g. produce nonfunctional mRNA transcripts) by performing transcriptome-wide association studies (TWAS) for a number of diseases.

pair, resulting in the inclusion of a pseudo-exon.

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