

1.1-section-result

February 14, 2025

1 Section 1 - Quantifying the association of a feature with an outcome

- We frequently want to know if a **feature** is **associated** with an **outcome**. For example, we might want to determine:
 - Is the presence of a specific cell stress condition (feature) associated with the formation of stress granules (outcome)?
 - Is the amino acid sequence of a protein (feature) associated with its propensity to misfold (outcome)?
- This type of analysis is invaluable for checking for correlations between features and outcomes in complex data
- In this section we will explore using contingency tables, odds ratios, and Fisher's Exact Test to test for association

1.1 Example 1.1

Application 1.1: We hypothesize that *E. coli* proteins that contain native entanglements are more likely to misfold than proteins without entanglements.

- To proceed, we need information about which proteins in *E. coli* contain entanglements and which proteins in *E. coli* misfold.

1.1.1 What is an entanglement? How can we tell if a protein is entangled?

- Entanglements are a structural motif in proteins formed by two segments: a loop (closed by a native contact) and a thread (**Figures 1.1.1 & 1.1.2**).

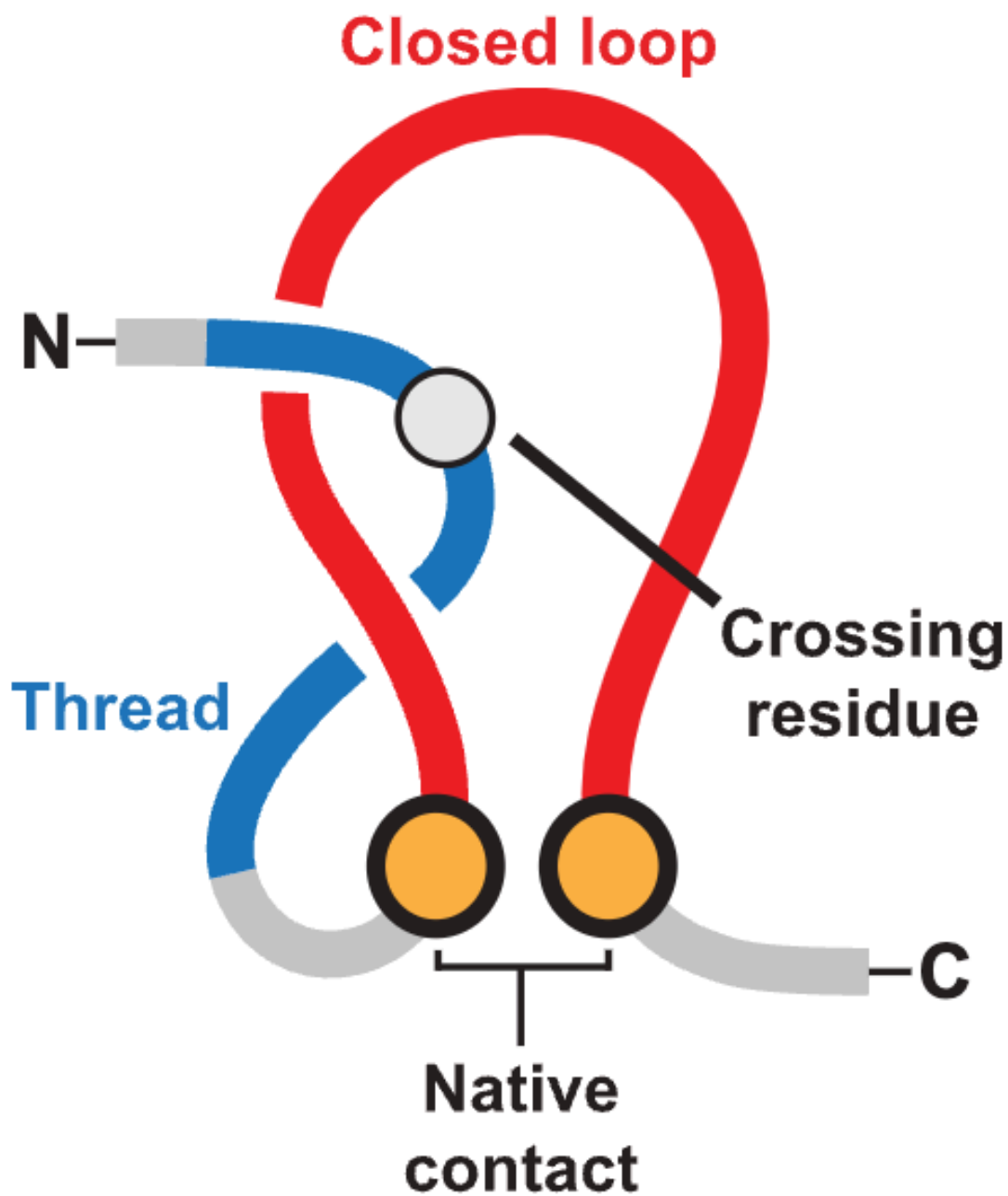


Figure 1.1.1. General structure of a non-covalent lasso entanglement. The threading segment (blue) passes through a loop (red) that is closed by a native contact (yellow). *J. Mol. Biol.* 436 (2024) 168487.

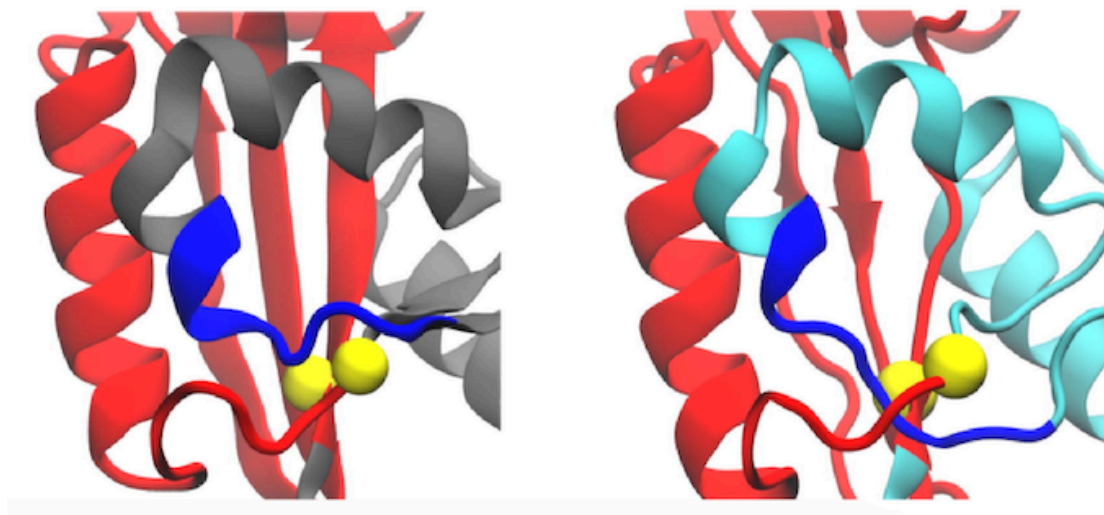


Figure 1.1.2. 3D structures of oligoribonuclease without (left) and with (right) an entanglement. The threading segment (blue) passes through a loop (red) that is closed by a native contact (yellow). *J. Mol. Biol.* 436 (2024) 168487.

- Some proteins contain entanglements in their native state (*i.e.*, native entanglements)
- Some proteins can gain or lose entanglements during misfolding
- For the current hypothesis, we are concerned with native entanglements; this information can be obtained by analyzing either experimental structures or predicted structures
- Our analysis will use data on entanglements computed from experimental structures of *E. coli* proteins

1.1.2 How can we tell if a protein misfolds?

- The structural proteomics technique *limited proteolysis mass spectrometry* (LiP-MS) profiles changes in protein structures across the proteome in response to perturbations (**Figure 1.1.3**)

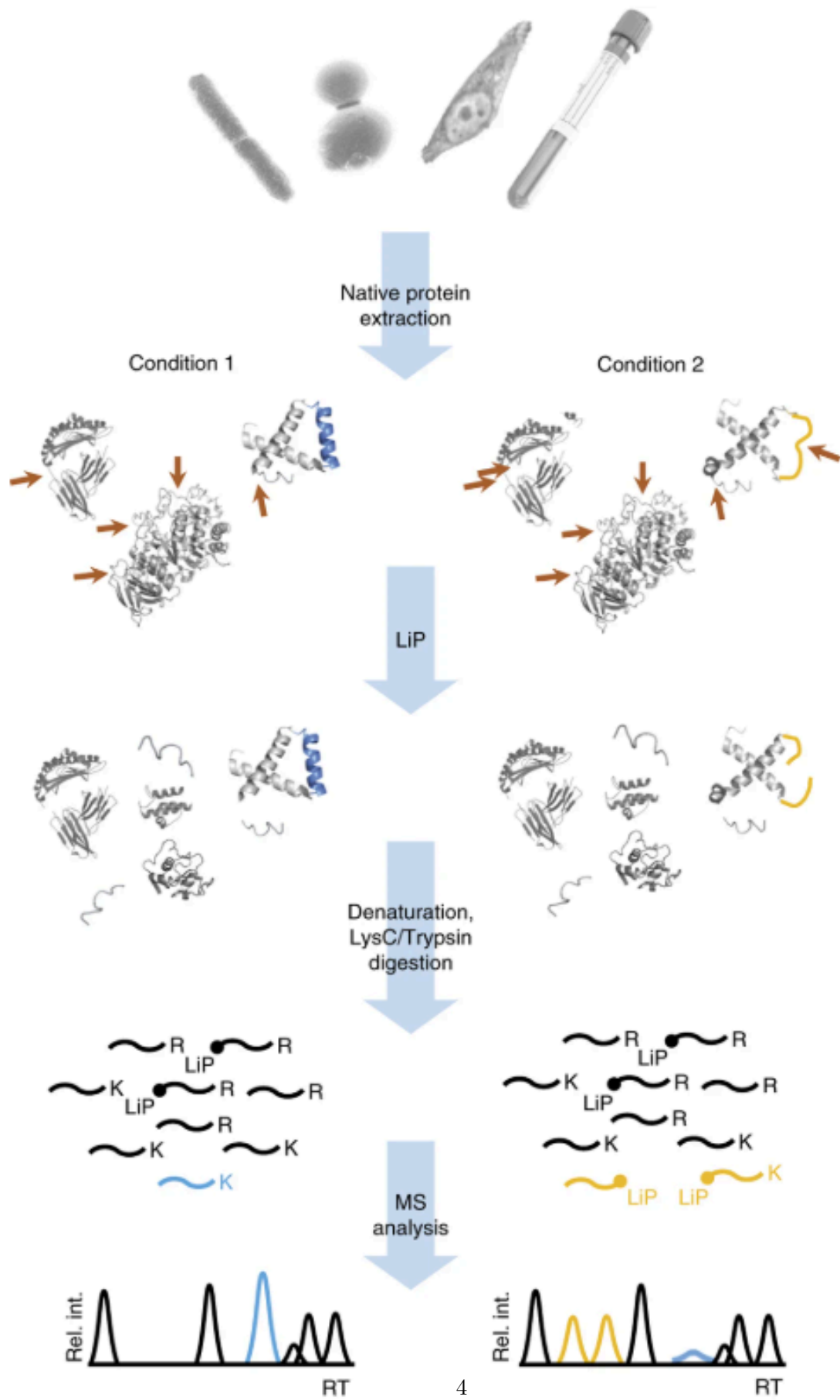


Figure 1.1.3. *Schematic of a LiP-MS experiment. When studying misfolding, one sample will be treated with guanidinium chloride to induce unfolding before a dilution jump is used to stimulate refolding; the other sample is not treated with guanidinium chloride, preserving protein native states. Nature Protocols volume 12, p. 2391–2410 (2017).*

- LiP-MS compares differences in protein structures between two samples
- In the case at hand, a protein is considered to misfold if there is a significant change in its limited proteolysis digestion pattern between a guanidinium chloride-unfolded/refolded and an untreated sample containing natively folded protein
- We will use LiP-MS data from *E. coli* to match the *E. coli* entanglement data

1.1.3 We have our data - what now?

- Now that we have identified relevant data to test our hypothesis, let's dive into some code.

1.2 Testing our hypothesis in Python

1.2.1 Step 0 - Load libraries

- We first need to make sure we have access to all of the functions etc. that we need for this analysis - let's load some libraries

```
[1]: import numpy as np
import pandas as pd
from scipy.stats import fisher_exact
import matplotlib.pyplot as plt
```

1.2.2 Step 1 - Load the data

- After loading the libraries, we now need to load the data into memory

```
[2]: # "data1" is a pandas DataFrame object
data_path = "/home/jovyan/data-store/data/iplant/home/shared/NCEMS/
↳BPS-training-2025/"
data1 = pd.read_csv(data_path + "NativeEntanglements_and_SigCuts_EXP_buffC.
↳csv")
```

1.2.3 Step 2 - Explore the data

- Let's explore the data quickly to get a better understanding of what we need to do

```
[3]: # first, print a quick summary
print ("Create a quick summary of the DataFrame:\n")
data1.info()

# second, print the first 10 rows of data1
print ("\nPrint the first 10 rows of the DataFrame:\n")
display(data1.head(10))
```

```
# third, count the number of unique gene identifiers in column "gene" of data1
print ("\nThe number of unique genes is:", len(data1["gene"].unique()))
```

Create a quick summary of the DataFrame:

```
<class 'pandas.core.frame.DataFrame'>
RangeIndex: 345 entries, 0 to 344
Data columns (total 4 columns):
#   Column          Non-Null Count  Dtype
---  -
0   buff            345 non-null    object
1   gene            345 non-null    object
2   NativeEnt       345 non-null    bool
3   NonRefoldable   345 non-null    bool
dtypes: bool(2), object(2)
memory usage: 6.2+ KB
```

Print the first 10 rows of the DataFrame:

	buff	gene	NativeEnt	NonRefoldable
0	C	P00350	True	True
1	C	P00370	True	True
2	C	P00448	True	True
3	C	P00509	True	True
4	C	P00561	True	False
5	C	P00579	False	False
6	C	P00864	True	True
7	C	P00934	True	True
8	C	P00954	False	True
9	C	P00957	True	True

The number of unique genes is: 345

- We can see from this summary that this data file contains 4 columns:
 - **buff**: the buffer condition for the experiment
 - **gene**: the unique gene identifier (there are no duplicates!)
 - **NativeEnt**: **True** if the protein has a native entanglement, **False** if it does not
 - **NonRefoldable**: **True** if the protein *did not* refold in LiP-MS experiment (i.e., misfolded), **False** if it *did* refold
- Now that we have a better understanding of the data, we are ready to run our analysis.

1.2.4 Step 3 - Run the analysis

```
[4]: # compute the values of {a, b, c, d} and construct the contingency table
a = len(data1[(data1["NativeEnt"] == True) & (data1["NonRefoldable"] == True_
↪)])
b = len(data1[(data1["NativeEnt"] == True) & (data1["NonRefoldable"] ==_
↪False)])
c = len(data1[(data1["NativeEnt"] == False) & (data1["NonRefoldable"] == True_
↪)])
d = len(data1[(data1["NativeEnt"] == False) & (data1["NonRefoldable"] ==_
↪False)])

# put values into a new format to enable a nice print statement & analysis
contingency_table = pd.DataFrame({"Protein Misfolded" : [a, c],
                                "Protein Not Misfolded": [b, d]},
                                index = ["Protein Entangled", "Protein Not_
↪Entangled"])

# print the output
print ("This is our contingency table:\n")

# create a table from our contingency_table using matplotlib
plt.clf()
fig, ax = plt.subplots(figsize = (5, 2))
ax.axis("tight")
ax.axis("off")
cell_text = contingency_table.reset_index().values.tolist()
col_labels = [""] + contingency_table.columns.tolist()
table = ax.table(cellText=cell_text, colLabels=col_labels, loc="center",_
↪cellLoc="center")
table.auto_set_font_size(False)
table.set_fontsize(14)
table.scale(2, 2)
plt.show()
```

This is our contingency table:

<Figure size 640x480 with 0 Axes>

	Protein Misfolded	Protein Not Misfolded
Protein Entangled	233	31
Protein Not Entangled	52	29

- We can complete this analysis by computing the odds ratio and p -value

```
[5]: # use the fisher_exact function from scipy.stats to compute the odds ratio and
      ↪p-value
odds_ratio, fisher_p_value = fisher_exact(contingency_table, alternative =
      ↪"two-sided")

# print the results of this analysis
print ("The odds ratio is:", "%.2f" %odds_ratio)
print ("The p-value is   :", "%.2e" %fisher_p_value)
```

The odds ratio is: 4.19

The p-value is : 2.93e-06

1.2.5 Step 4 - Interpret the results

- The odds ratio of 4.19 indicates that there is a **positive association** between entanglement and misfolding
- In other words, entanglement and misfolding tend to co-occur in the same protein, supporting our hypothesis
 - Odds ratios > 1 indicate positive association
 - Odds ratios $= 1$ indicate no association
 - Odds ratios < 1 indicate negative association
- We can also say that the **odds** of an entangled protein misfolding are 4.19 times greater than the odds of a non-entangled protein misfolding
- The p -value is «0.05, which is a common threshold for significance; in this instance, we conclude the result is significant