Potentially Hazardous Biological Agents Risk Assessment Form (6A)

Required for research involving microorganisms, rDNA, fresh/frozen tissue (including primary cell lines, human and other primate established cell lines and tissue cultures), blood, blood products and body fluids.

SRC/IACUC/IBC approval required before experimentation.

Student's Name(s) Yujia Li

Title of Project Intracellular Trafficking of Ajuba in Human Cells

To be completed by the QUALIFIED SCIENTIST/DESIGNATED SUPERVISOR in collaboration with the student researcher(s). All questions are applicable and must be answered; additional page(s) may be attached.

SECTION 1: PROJECT ASSESSMENT

1. Identify potentially hazardous biological agents to be used in this experiment. Include the source, quantity and the biosafety level risk group of each microorganism.

Ethidium bromide-contained in safety cabinet and disposed of in appropriate biohazard weste.

E coli bacterial cells used for molecular cloring. HeLa II cells for transfection. None other required specific biosafety labelling or equip-

- Describe the site of experimentation including the level of biological containment. Laboratory 913N in Hunter College. North building. Laboratory possesses all required chemical biosafety cabinets and is inspected and approved by Hunter College Office of Environmental Health and Safety. Hunter College
- Describe the procedures that will be used to minimize risk (personal protective equipment, hood type, etc.). Latex gloves, lab coat for personal protective equipment. Cell culture will be performed using a sterile biosafety cabinets class II and the cells will be incubated in adequate CO2 incubator
- What final biosafety level do you recommend for this project given the risk assessment you conducted? First level of biosafety is adequate for these manipulations- the lab is BSL-1 level facility (recombinant DNA use).
- 5. Describe the method of disposal of all cultured materials and other potentially hazardous biological agents. All biologicals and chemical will be disposed in a contained biohazard waste handled by College professionals.

SECTION 2: TRAINING

- 1. What training will the student receive for this project?
 - Experimental design and data interpretation on human cell biology. Experimentation using cell biology and molecular biology protocols
- Experience/training of Designated Supervisor as it relates to the student's area of research (if applicable).
 - Over 15 years of experience training students at all levels, including 13 years at Hunter College, with 4 Ph.D. students and many undergraduate and high school students.

SECTION 3: For ALL MICROORGANISMS, CELL LINES and TISSUES - To be completed by the QUALIFIED SCIENTIST or DESIGNATED SUPERVISOR - Check the appropriate box(es) below:

- Experimentation on the microorganisms/cell lines/tissues used in this study will NOT be conducted at a Regulated Research Institution, but will be conducted at a (check one) __BSL-1 or __BSL-2 laboratory. This study has been reviewed by the local SRC and the procedures have been approved prior to experimentation.
- Experimentation on the microorganisms/cell lines/tissues used in this study will be conducted at a Regulated Research Institution and was approved by the appropriate institutional board prior to experimentation; institutional approval forms are attached. Date of IACUC/IBC approval May 30th 2017 (IRB-Institute Origin of cell lines: ATCC (American Tissue Culture Collection)
- Experimentation on the microorganisms/cell lines/tissues used in this study will be conducted at a Regulated Research Institution, which does not require pre-approval for this type of study. The SRC has reviewed that the student received appropriate training and the project complies with Intel ISEF rules.

CERTIFICATION – To be SIGNED by the QUALIFIED SCIENTIST or DESIGNATED SUPERVISOR

The QS/DS has seen this project's research plan and supporting documentation and acknowledges the accuracy of the information provided above. This study has been approved as a (check one) BSL-1/ BSL-2 study, and will be conducted in an appropriate laboratory.

Diego Loayza	Diego Loayza	Digitally signed by Diego Loeyza Date: 2019 07 10 13 04:35 -04'00'
QS/DS Printed Name	Signature	
07/10/19		
Date of review (mm/dd/yy)		

SECTION 4: CERTIFICATION	 To be completed by 	the LOCAL or AFFILIATED FAIR SRC
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The SKC has seen this pro	jects research plan and supportin	g documentation and acknowledges	the accuracy	of the information	n provided above
			, ,	_	provided above.

SRC Printed Name

Date of review (mm/dd/yy)