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## **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see Authors & Referees and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section

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1 01	an statistical analyses, commit that the following items are present in the ligare regend, table regend, main text, or internous section.
n/a	Confirmed
	The exact sample size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated

Our web collection on statistics for biologists contains articles on many of the points above.

## Software and code

Policy information about availability of computer code

Data collection

No software was used to collect data.

Data analysis

DNA barcodes obtained using Sanger sequencing at SourceBioscience. High-throughput data obtained using Illumina HiSeq-3000 at the Center for Genome Research and Biocomputing, Oregon State University. All software used for phylogenetic analysis is available online

- DNA barcodes (assembly): Mega v.5.0, Mega v.6.0, Geneious v.9.1.2
- DNA barcodes (alignment and phylogenetic inference): MAFFT v.7.2.1, jModelTest 2, RAXML v.8, FastTree 2, MrBayes v.3.2.
- High-throughput data (assembly): YASRA, PRICE, SSPACE, BLASTN, MUSCLE, PHYLIP, SPades.
- -High-throughput data (alignment and phylogenetic inference): MAFFT, PRANK, GBlocks, jModelTest 2, RaxML, FastTree 2, Astral-II, MrBaves v.3.2.
- Time-calibrated phylogenies: MAFFT, Gblocks, RevBayes v.1.0.4, Tracer v.1.6.0, treePL, BAMM (including set\_priors.R script),

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Passport data of all herbarium specimens included in the molecular studies presented in this paper is available as supplementary file. Additional records and

information of the collections included in this study and of specimens added subsequently are available through the website of the project (https://herbaria.plants.ox.ac.uk/bol/ipomoea). All data used for phylogenetic analysis is available in public repositories: DNA barcode sequences are available through GenBank and genome assemblies are available through the Oxford Repository Archive. Illumina raw reads are available through the Sequence Read Archive.

The reference copy of the document with all sections, see <u>nature com/documents/fir reporting summary flat pdf</u> Ecological, evolutionary & environmental sciences study design  all studies must disclose on these points even when the disclosure is negative.  Study description  Monographic study of the genus Ipomoea (morning glories) integrating the morphological study of herbarium specimens with the use of high-throughput DNA sequencing (flyth-Seigl and DNA barcodes for phylogenetic analysis.  Research sample  All Ipomoea specimens from 47 herbaria worldwide used for morphological study of herbarium specimens with the use of high-throughput DNA sequencing (flyth-Seigl and DNA barcodes for phylogenetic analysis.  Sampling strategy  Sampling strategy aimed at obtaining, when possible, multiple specimens from each putative species studied. When available, material collection in the last 50 years was preferred as this is more suitable to produce good sequencing results.  Data collection  Samples for DNA analysis collected from herbarium material by P.M.R., J.R.I.W., B.R.M.W. and A.S. Morphological study of the specimens conducted by J.R.I.W.  Timing and spatial scale  Project developed between 2012 and 2019 with continuous sampling of herbarium material.  Data exclusions  No data was excluded from the study.  Methodology is extensively described and all data files, including raw sequence data, are made available through publicly accessible repositories.  Reporting for specific materials, systems and methods used in many studies. Here, indicate whether each material, yetern or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.  Materials & experimental systems  Methods    Involved in the study	Field-specific	creporting		
Study description	Please select the one below			
Study description  Monographic study of the genus Ipomoea (morning glories) integrating the morphological study of herbarium specimens with the use of high-throughput DNA sequencing (Hyb-Seq) and DNA barcodes for phylogenetic analysis.  All Ipomoea specimens from 47 herbaria worldwide used for morphological study. 385 samples representing c. 210 species sequenced to obtain their whole childroplast genome and 605 putative single copy nuclear coding DNA regions using Hyb-Seq. 1,560 specimens sequenced to obtain their whole childroplast genome and 605 putative single copy nuclear coding DNA regions using Hyb-Seq. 1,560 specimens sequenced for at least one DNA barcode using Sanger sequencing.  Sampling strategy  Sampling strategy aimed at obtaining, when possible, multiple specimens from each putative species studied. When available, material collected in the last 50 years was preferred as this is more suitable to produce good sequencing results.  Data collection  Samples for DNA analysis collected from herbarium material by P.M.R., J.R.I.W., B.R.M.W. and A.S. Morphological study of the specimens conducted by J.R.I.W.  Timing and spatial scale  Project developed between 2012 and 2019 with continuous sampling of herbarium material.  Data exclusions  No data was excluded from the study.  Methodology is extensively described and all data files, including raw sequence data, are made available through publicly accessible repositories.  Methodos  We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, yetem or method listed is relevant to your study. If you are not sure if a list trem applies to your research, read the appropriate section before selecting a response.  Materials & experimental systems  Methods    Involved in the study   Involved in the st	.,			
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Eukaryotic cell lines  Flow cytometry  Palaeontology  MRI-based neuroimaging				
Palaeontology MRI-based neuroimaging				
Animals and other organisms		—		

Human research participants

Clinical data