

Journal of Enzyme Inhibition and Medicinal Chemistry



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ISSN: (Print) (Online) Journal homepage: www.tandfonline.com/journals/ienz20

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To cite this article: Jingbo Qin, Jie Liu, Chunxiao Wu, Jianwen Xu, Bowen Tang, Kaiqiang Guo, Xiaohui Chen, Weihao Liu, Tong Wu, Hu Zhou, Meijuan Fang & Zhen Wu (2020) Synthesis and biological evaluation of (3/4-(pyrimidin-2-ylamino)benzoyl)-based hydrazine-1-carboxamide/carbothioamide derivatives as novel RXRα antagonists, Journal of Enzyme Inhibition and Medicinal Chemistry, 35:1, 880-896, DOI: 10.1080/14756366.2020.1740692

To link to this article: https://doi.org/10.1080/14756366.2020.1740692

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Synthesis and biological evaluation of (3/4-(pyrimidin-2-ylamino)benzoyl)-based hydrazine-1-carboxamide/carbothioamide derivatives as novel RXR α antagonists

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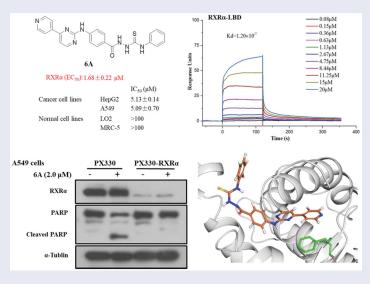
Fujian Provincial Key Laboratory of Innovative Drug Target Research and State Key Laboratory of Cellular Stress Biology, School of Pharmaceutical Sciences, Xiamen University, Xiamen, China

ABSTRACT

Abnormal alterations in the expression and biological function of retinoid X receptor alpha (RXR α) have a key role in the development of cancer. Potential modulators of RXR α as anticancer agents are explored in growing numbers of studies. A series of (4/3-(pyrimidin-2-ylamino)benzoyl)hydrazine-1-carboxamide/carbothioamide derivatives are synthesised and evaluated for anticancer activity as RXR α antagonists in this study. Among all synthesised compounds, **6A** shows strong antagonist activity (half maximal effective concentration (EC₅₀) = 1.68 \pm 0.22 μ M), potent anti-proliferative activity against human cancer cell lines HepG2 and A549 cells (50% inhibition of cell viability (IC₅₀) values < 10 μ M), and low cytotoxic property in normal cells such as LO2 and MRC-5 cells (IC₅₀ values > 100 μ M). Further bioassays indicate that **6A** inhibits 9-cis-RA-induced activity in a dose-dependent manner, and selectively binds to RXR α -=LBD with submicromolar affinity (Kd = 1.20 × 10⁻⁷ M). **6A** induces time-and dose-dependent cleavage of poly ADP-ribose polymerase, and significantly stimulates caspase-3 activity, leading to RXR α -dependent apoptosis. Finally, molecular docking studies predict the binding modes for RXR α -LBD and **6A**.

GRAPHIC ABSTRACT

Compound **6A** with low cytotoxic property in normal cells acts as a selective RXR alpha ligand to promote TNF alpha-mediated apoptosis of cancer cells.



ARTICLE HISTORY

Received 11 November 2019 Revised 7 February 2020 Accepted 1 March 2020

KEYWORDS

Hydrazine carbothioamide; RXRα antagonist; selectivity; anticancer activity; apoptosis

1. Introduction

Retinoid X receptor alpha (RXR α) is a ligand-dependent transcription factor of the nuclear receptor superfamily, and it is composed of A/B region (modulating N-terminal domain or AF-1 domain), DNA-binding domain (DBD), hinge region, and ligand-binding

domain (LBD)^{1–3}. RXR α forms dimers with itself or other nuclear receptors such as the thyroid hormone receptor (TR), retinoic acid receptor (RAR), vitamin D receptor (VDR), liver X receptor (LXR), and peroxisome-proliferator-activated receptor (PPAR), to exert its transcriptional activities in the nucleus and act biologically in the

CONTACT Meijuan Fang fangmj@xmu.edu.cn; Zhen Wu wuzhen@xmu.edu.cn; Hu Zhou huzhou@xmu.edu.cn Fujian Provincial Key Laboratory of Innovative Drug Target Research and State Key Laboratory of Cellular Stress Biology, School of Pharmaceutical Sciences, Xiamen University, Xiamen, 361102, China

Supplemental data for this article can be accessed here.

^{*}These authors contributed equally to this work.

Target compounds

Figure 1. Design and modification strategies of target compounds.

cytoplasm^{4–7}. It's worth noting that RXR α dimers are activated by endogenous ligand binding to its LBD, and then regulates gene expression in various biological processes^{8,9}. Besides, dysregulations of $RXR\alpha$ expression and function are implicated in the initiation and development of many cancers^{1,10–12}. So exogenous $RXR\alpha$ modulators may obstruct the development of various cancer and diseases. Experiments have shown that agonist can activate $\mbox{RXR}\alpha$ and upregulated the expression of CDKN1A (a target gene of RXR), resulting in p21WAF1/CIP1-dependent cell apoptosis and suppressing cell proliferation 13,14, while synthetic antagonists can promote mitochondria-mediated apoptosis and inflammation 10,15,16 , or induce TNF α -dependent apoptosis through suppressing PI3K/AKT-mediated cell survival pathway 17,18. In a word, previous studies have suggested a potential role for RXRα in cancer initiation and cancer therapy.

In recent years, increasing RXRα modulators including many agonists and few antagonists were identified based on virtual screening and structure-based design¹⁹⁻²¹. Among them, only Bexarotene (an RXR full agonist) has been clinically used for the treatment of cancers such as Cutaneous T cell lymphoma²². However, synthetic RXRα ligands reported are associated with unacceptable toxicity or undesirable side effects²³. For example, Bexarotene has adverse effects such as elevation of blood triglyceride, weight gain, and hepatomegaly^{24,25}. So, the current challenge is to develop a variety of novel RXR α modulator with more potent, high specific and low cytotoxic properties.

In this work, to explored novel RXRα antagonists, a series of new (3/4-(pyrimidin-2-ylamino)benzoyl)-based hydrazine-1-carboxamide/carbothioamide derivatives were designed based on the structures of compounds NSC-640358 and 24 (Figure 1). First, we fused 4-substituted pyrimidine (replacing 3-phenyl-1H-pyrazole moiety in NSC-640358) to 2-benzoylhydrazine-1-carboxamide (as a substitute for (E)-N'-benzylideneacetohydrazide moiety in compound 24) to build the scaffold structure of target compounds. Second, to obtain potent RXR α antagonists, we modified the two moiety structures such as introducing the aryl group to the C-4 position (R¹) of pyrimidine ring, introducing the alkyl/aryl/aralkyl

group to the nitrogen atom of carboxamide (R²), and synthesising carboxamide or carbothioamide derivatives (X). Next, we synthesised target compounds and evaluated their biological activities such as in vitro antiproliferative activity and modulating RXR α activity. Then, we attempt to establish the preliminary structure – activity relationships, and the selected compound 6 A as a new high potent RXRa antagonist was further investigated for the RXR α -dependent apoptosis induction including inhibition of RXR α transactivation, physical binding of **6A** to RXRα-LBD, and induction of cell apoptosis, and checking the level of cleaved poly ADPribose polymerase (c-PARP) and caspase-3. At last, a molecular docking study was performed to explore the binding nature of 6A to the ligand-binding pocket (LBP) of RXRα with antagonistic conformation (PDB: 3A9E).

2. Results and discussion

2.1. Chemistry

The synthetic strategy of target compounds (Series A andB) is depicted in Scheme 1. Commercially available ketones 1a-1e were reacted with N,N-dimethylformamide dimethyl acetal (DMF-DMA) in dry toluene at 110 °C for 18 h to give corresponding compounds 3a-3e. Ethyl 4-aminobenzoate (2a) and ethyl 3-aminobenzoate (2b) were refluxed with cyanamide in the presence of concentrated HCl for 24 h in ethanol, and then treated with ammonium nitrate to afford 4-guanidino benzoic acid ethyl ester nitrate (4a) and 3-guanidino benzoic acid ethyl ester nitrate (4b), respectively. Reactions of 4a with 3-(dimethylamino)-1-arylprop-2-en-1-ones (3a-3e) in the presence of sodium hydroxide were performed under reflux conditions in ethanol for 48 h to produce 4-((4-arylpyrimidin-2-yl)amino)benzoic acids, followed by refluxing with hydrazine hydrate in ethanol to get corresponding aroylhydrzides 5a-5e. Finally, target compounds of series A were prepared via reactions between aroylhydrzides 5a-5e with different isocyanates or isothiocyanates. On the other hand, target compounds of series B were synthesised with 4b instead of 4a in a similar method.

Series A: 6a-6t and 6A-6J Series B: 7a-7f and 7A-7D

Scheme 1. Synthesis of target compounds of Series A and B. Reaction conditions: (a) DMF-DMA, 110 °C; (b) Cyanamide, concentrated HCl, EtOH, reflux, 24 h and NH₄NO₃, 0 °C; (c) NaOH, EtOH, reflux; (d) Hydrazine monohydrate, EtOH, reflux; and (e) EtOH, reflux.

Table 1. In vitro antiproliferative activities of 2–(4-((-pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide derivatives (Series A) on two selected cancer cell lines.^a

	R1	R2	Х	IC50[μM]	
Compounds				HepG2	A549
6a	4-Pyridyl	n-Butyl	0	5.32 ± 0.07	21.69 ± 1.33
6b	4-Pyridyl	t- Butyl	0	5.39 ± 0.11	18.09 ± 1.25
6c	4-Pyridyl	Cyclohexyl	0	2.17 ± 0.08	8.39 ± 0.92
6d	4-Pyridyl	Cyclopentyl	0	2.71 ± 0.13	16.26 ± 1.21
6e	4-Pyridyl	p-Tolyl	0	1.28 ± 0.09	11.27 ± 1.05
6f	4-Pyridyl	2,4-difluorophenyl	0	8.10 ± 0.12	26.38 ± 1.42
6g	4-Pyridyl	3-chloro-4-methylphenyl	0	>40	>40
6ĥ	4-Pyridyl	Phenethyl	0	3.89 ± 0.04	13.11 ± 1.11
6i	3-Pyridyl	n-Butyl	0	>40	38.93 ± 1.59
6j	3-Pyridyl	Cyclopentyl	0	5.73 ± 0.26	17.09 ± 1.23
6k	3-Pyridyl	p-Tolyl	0	>40	8.385 ± 0.92
6l	3-Pyridyl	Phenethyl	0	5.48 ± 0.21	23.71 ± 1.37
6m	2-Pryidyl	n- Butyl (0	>40	>40
6n	2-Pryidyl	t-Butyl	0	15.71 ± 0.11	>40
60	2-Pryidyl	Cyclopentyl	0	>40	>40
6p	2-Pryidyl	m-Tolyl	0	23.31 ± 0.07	>40
6q	2-Pryidyl	3,5-dimethylphenyl	0	20.32 ± 0.14	>40
6r	4-Chlorophenyl	t- Butyl	0	4.48 ± 0.09	19.49 ± 1.29
6s	4-Chlorophenyl	Ethyl	0	20.34 ± 0.34	>40
6t	4-Chlorophenyl	Cyclohexyl	0	13.08 ± 0.46	>40
6A	4-Pyridyl	Phenyl	S	5.13 ± 0.14	5.09 ± 0.70
6B	4-Pyridyl	Benzyl	S	0.61 ± 0.07	15.95 ± 1.20
6C	3-Pyridyl	Benzyl	S	1.47 ± 0.42	17.57 ± 1.24
6D	2-Pryidyl	Phenyl	S	6.74 ± 0.05	38.51 ± 1.58
6E	2-Pryidyl	Benzyl	S	4.59 ± 0.11	>40
6F	Phenyl	Cyclohexyl	S	>40	30.79 ± 1.48
6G	Phenyl	Phenyl	S	>40	20.68 ± 1.31
6H	Phenyl	Benzyl	S	>40	22.27 ± 1.34
6l	4-Chlorophenyl	Phenyl	S	17.24 ± 0.07	28.32 ± 1.45
6J	4-Chlorophenyl	Benzyl	S	1.29 ± 0.09	12.76 ± 1.10
Sorafenib	. ,	•		16.89 ± 1.22	>40

 $^{\mathrm{a}}\mathrm{Data}$ are means $\pm\,\mathrm{SD}$ of triplicate experiments.

2.2. Biology activity

2.2.1. 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay and structure activity relationships (SARs) of the anti-proliferative activity

To explore the antitumor activity of the target compounds, all synthesised target compounds were first assayed for *in vitro*

antiproliferative activities against two human cancer cell lines (HepG2 and A549) by MTT method with Sorafenib as a reference. The concentrations of compounds required for 50% inhibition of cell viability (IC_{50}) were determined and listed in Tables 1 and 2. According to these data, the preliminary SARs of these novel 2–(3/4-((-pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide

derivatives were summarised in Figure 2: (1) First, the R¹ substituent is crucial to the antitumor activities. As shown in Table 1, urea derivatives with 4-pyridyl at the R¹-position showed stronger antiproliferative activity than those with 3-pyridyl and 2-pyridyl (6a vs. 6i and 6m, 6d vs. 6j and 6o, 6e vs. 6k, 6h vs. 6l, and 6b vs. 6n). The synthesised thiourea derivatives **6A-6J** possessed a similar SAR, with the 4-pyridyl group to the C-4 position (R¹) of pyrimidine ring being the better substitution. 4-Pyridyl substitution at R¹ position was also better than 4-chlorophenyl and phenyl substitutions (6b vs. 6t, 6A vs. 6l, 6A vs. 6G, and 6B vs. 6H). (2) For the substituent R^2 of compounds (**6a** \sim **6h**) which contained 4-pyridyl group at R¹ position, it was found that both compounds **6a-6d** bearing alkyl groups (n-Butyl, t-Butyl, Cyclohexyl, and Cyclopentyl) and 6h having an aralkyl group (phenethyl) generally had good anti-proliferative properties, while the R² analogues **6e** (p-Tolyl), **6f** (2,4-Difluorophenyl), and 6g (3-Chloro-4-methylphenyl) exhibited different cytotoxicity due to their different substituted phenyl ring at the R²-position. A comparison of the activities of compounds 6e and 6f shows that the p-Tolyl substitution is better than 2,4-Difluorophenyl, but when 3-Chloro-4-methylphenyl was introduced into the molecule, the biological activity of 6g was significantly reduced. Among the synthesised thiourea derivatives

Table 2. In vitro antiproliferative activities of 2–(3-((-pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide derivatives (Series B) on two selected cancer cell lines.

$$\mathbb{R}^1 \xrightarrow[N]{N} \mathbb{N} \xrightarrow[H]{N} \mathbb$$

				IC50[μM]	
Compounds	R1	R2	Χ	HepG2	A549
7a	4-Pyridyl	t-Buty	0	14.37 ± 0.26	31.63 ± 1.50
7b	4-Pyridyl	Cyclopentyl	0	>40	30.24 ± 1.48
7c	3-Pyridyl	t-Butyl	0	31.45 ± 0.44	>40
7d	3-Pyridyl	n-Butyl	0	>40	>40
7e	3-Pyridyl	2,4-dimethoxyphenyl	0	>40	>40
7f	2-Pyridyl	n-Butyl	0	>40	30.59 ± 1.48
7A	4-Pyridyl	Benzyl	S	14.96 ± 0.22	34.43 ± 1.53
7B	3-Pyridyl	Benzyl	S	17.86 ± 0.28	>40
7C	2-Pyridyl	Phenyl	S	17.45 ± 0.34	>40
7D	2-Pyridyl	Benzyl	S	20.23 ± 0.08	>40

^aData are means ± SD of triplicate experiments.

(6A-6J), compounds with one aralkyl group (benzyl) at R²-position displayed the better cytotoxicity against HepG2 cells than those with one aryl group (phenyl) such as 6B vs. 6A, 6E vs. 6D, and 6J vs 61. (3) The various scaffold had a certain influence on antitumor activity. As shown in Tables 1 and 2, replacement of the 4-(pyrimidin-2-ylamino)benzohydrazide scaffold with the 3-(pyrimidin-2-ylamino)benzohydrazide tended to decrease the activity (7a vs. 6b, 7b vs. 6d, 7d vs. 6i, and 7f vs. 6m).

2.2.2. Identification of novel RXRα modulators from get compounds

 $RXR\alpha$ is an attractive target for the treatment of cancer and we'd like to discover new potential modulators of RXR α as anticancer agents. So, all target compounds were evaluated for their effect on regulating RXRα transcriptional function using a luciferase transcription assay in HEK293T cells with the Gal4- RXRa/LBD chimaera and Gal4 reporter system. When tested at 1.0 μM concentration with absence 9-cis-RA in this system, no compound exhibited increase transcriptional activity, suggesting no agonist effect for them. However, 11 compounds (6c, 6e, 6k, 6A, 6B, 6C, 6D, 6E, 6F, 6G, and 6H) of series A showed good antagonist effect on 9cis-RA-induced RXR α transactivation with an inhibition of >50% (Figure 3), while target compounds of series B exhibited little antagonist effect on 9-cis-RA-induced reporter activity, indicating 4-(pyrimidin-2-ylamino)benzohydrazide scaffold being crucial to the transcriptional activity of RXRa. Among these candidate compounds, compound 6A showed the strongest inhibition activity 9cis-RA-induced Gal4-DBD-RXRα-LBD transactivation at the tested concentration (1.0 µM). To confirm the antagonist effect of these 11 potential antagonists of RXRα, we next used a retinoid X receptor response element (RXRE)-luciferase reporter-based assay to evaluate their half maximal effective concentration (EC₅₀) values of the inhibition on 9-cis-RA-induced RXRE-luciferase reporter activation. As shown in Table 3, many of them (6k, 6A, 6B, 6C, 6G, and **6H**) showed good antagonism with EC₅₀ less than 10.0 μ M, while the EC₅₀ values of several compounds (6c, 6e, 6D, and 6E) could not be calculated. Furthermore, we evaluated the cytoxic effect of the six potent RXR α antagonists (EC₅₀ $< 10.0 \,\mu\text{M}$) on two normal cell lines to determine their safety (Table 3). Among the six tested compounds, 6A showed low cytotoxic property to both LO2 and MRC-5 with both IC₅₀ values more than 100 μ M.

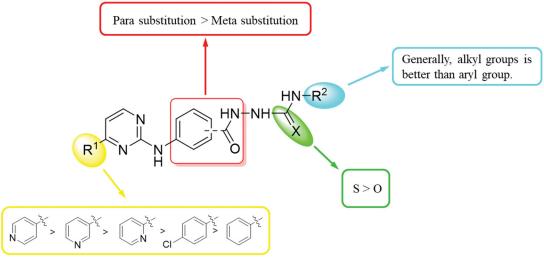
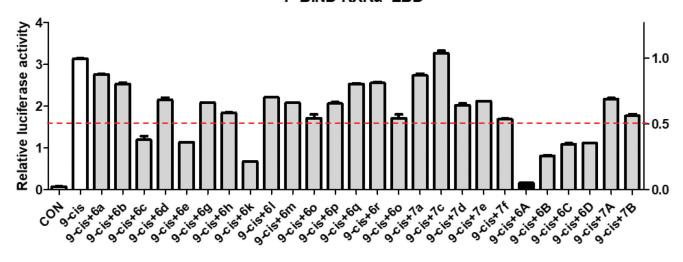


Figure 2. Summarised SARs of synthesised compounds (Series A and B) in Scheme 1.

P-BIND-RXRα -LBD



P-BIND-RXRα -LBD

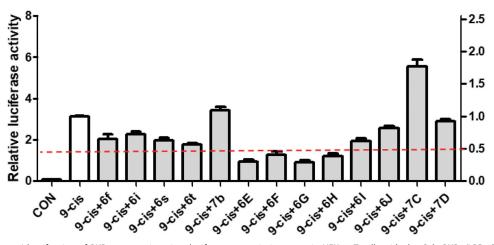


Figure 3. Identification of RXRα antagonists via a luciferase transcription assay in HEK293T cells with the Gal4-RXRα/LBD chimaera and Gal4 reporter system. The inhibition of target compounds at $1 \mu M$ on 9-cis-RA activating Gal4-DBD-RXRα-LBD transcriptional activity in HEK293T cells transfected with p-BIND-RXRα-LBD (50 ng) and pG5-luc (50 ng) were tested. Luciferase activities were measured 12 h after treatment using the dual-luciferase assay system kit and Renilla luciferase values were normalised to firefly luciferase activity and plotted as relative luciferase activity.

Table 3. The antagonist effect on RXR α (EC₅₀) and cytotoxic effect on two normal cell lines (IC₅₀) of the selected compounds.^a

, 50	,			
	EC ₅₀ (μM)	IC ₅₀ (μM)		
Compounds	RXRα	LO2	MRC-5	
6с	ND	ND	ND	
6e	ND	ND	ND	
6k	1.05 ± 0.18	24.94 ± 1.40	>100	
6A	1.68 ± 0.22	>100	>100	
6B	7.23 ± 0.32	14.27 ± 1.16	>100	
6C	1.75 ± 0.35	32.09 ± 1.51	>100	
6D	ND	ND	ND	
6E	ND	ND	ND	
6F	27.69 ± 1.35	ND	ND	
6G	0.56 ± 0.14	21.1 ± 1.32	>100	
6H	0.54 ± 0.06	16.89 ± 1.22	>100	

^aData are means ± SD of triplicate experiments.

2.2.3. 6A binds to RXRa and acts as an antagonist

Compound **6A** displayed low cytotoxic property, potent antiproliferative ($IC_{50} < 5.30 \,\mu\text{M}$) and high antagonistic ($EC_{50} = 1.68 \pm 0.22 \,\mu\text{M}$) effects, so compound **6A** was selected for further biological study. Compound **6A** was assessed in a dose-dependent reporter gene assay. As shown in Figure 4(A,B), compound **6A**

showed the dose-dependent effect on inhibiting the 9-cis-RA-induced Gal4-DBD-RXR α -LBD transactivation and 9-cis-RA-induced RXRE-luciferase reporter activation, and it at 2.0 μ M displayed a similar inhibitory effect when compared with UVI3003 (1.0 μ M), a known RXR α classical antagonist. In order to confirm compound **6A** as RXR α ligand, we then performed a surface plasmon resonance (SPR) technology-based experiment to directly measure the physical binding of compound **6A** to RXR α -LBD protein. As shown in Figure 4(C), we found that 6 A dose dependently bound to RXR α -LBD with a Kd value of 1.20×10^{-7} M. Furthermore, we measured the physical binding of compound **6A** to other nuclear receptors (ER α -LBD, PPAR γ -LBD, and TR3-LBD). The results showed that compound **6A** could not bound to these tested nuclear receptors well (Figure 3(D–F)). This implied that compound **6A** was able to selectively interact with RXR α .

2.2.4. 6A induces RXRα-dependent apoptosis of cancer cells

Previous studies showed RXR α antagonists with anticancer activity could induce apoptotic cell death. So, we studied whether the inhibitory effect of **6A** on the growth of cancer cells was caused by cellular apoptosis. First, we examined the expression of c-PARP (an apoptosis-related protein) in several cancer cell lines treated

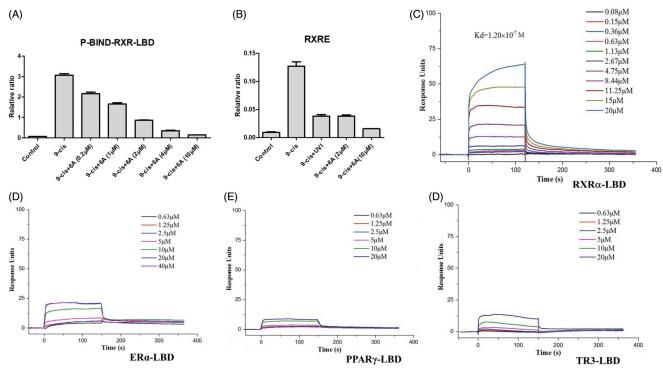


Figure 4. The antagonist effect of 6A by reporter gene assay (A,B) and the physical binding of 6A to RXRα-LBD (C), ERα-LBD (D), PPARγ-LBD (E), and TR3-LBD (F) by SPR assay. (A) Dose-dependent effect of 6A on inhibiting RXRα transactivation. HEK293T cells transfected with p-BIND-RXRα-LBD and pG5-luc were treated with indicated concentrations of 6A in the presence of 9-cis-RA (0.10 μM). (B) Inhibition of 9-cis-RA-induced RXRE-luciferase reporter activation by 6A. HEK293T cells transfected with RXRE (50 ng), full-length RXRα (50 ng), Renilla (1 ng) were treated with 9-cis-RA in the presence of indicated concentrations of 6A or UVI3003 (1 μM). (C) Surface plasmon resonance of 6A binding to RXRα-LBD. D. Surface plasmon resonance of 6A binding to PPARγ-LBD. F. Surface plasmon resonance of 6A binding to TR3-LBD. (C–F) Gradient concentrations of 6A were respectively injected through chips immobilised with RXRα-LBD, ERα-LBD, PPARγ-LBD, and TR3-LBD.

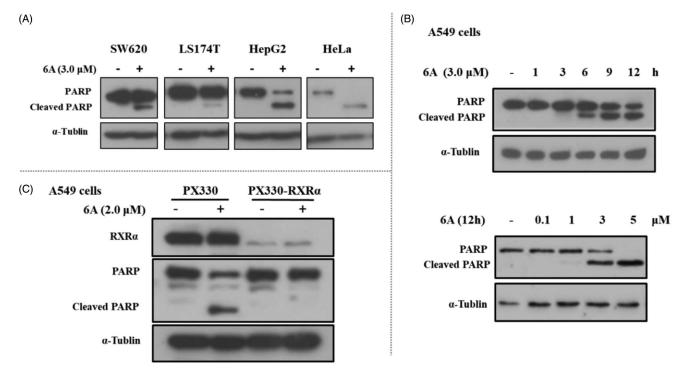


Figure 5. 6A-induced PARP cleavage in cancer cells. (A) SW620, LS174T, HepG2, and Hela cells were treated with 6A for 12 h and cell lysates prepared were analysed by Western blotting. (B) A549 cells were time- and dose- dependently treated with 6A for 12 h and cell lysates prepared were analysed by Western blotting for PARP cleavage. (C) A549 and RXRα-/- A549 cells were treated with 6A for 12 h and cell lysates prepared were analysed by Western blotting for PARP cleavage.

with or without **6A**. As shown in Figure 5(A), the c-PARP increased markedly in HepG2 and Hela cells after treatment with **6A** at $3.0\,\mu\text{M}$ for $12\,\text{h}$, whereas **6A** at the same concentration has a weak

effect on PARP cleavage in SW620 and LS174T cells. A time- and dose- dependent increase of c-PARP, was also observed in A549 cells after treatment with **6A** (Figure 5(B)). To address the role of

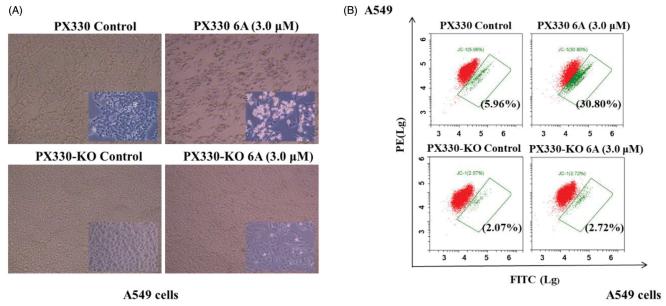


Figure 6. 6A-induced apoptotic effect. (A) The images of A549 and RXR α -/- A549 cells after treatment with or without 6A for 12 h; (B) Flow cytometry analysis of A549 and RXR α -/- A549 cells after treatment with or without 6A for 12 h.

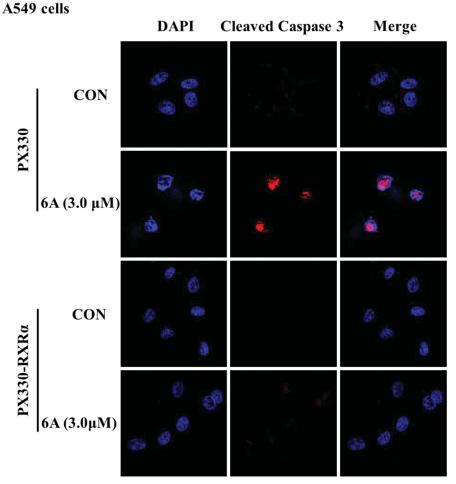


Figure 7. The effect of 6A on caspase-3 activation. A549 and RXR α -/- A549 cells were treated with or without 6A for 12 h, then stained with cleaved-caspase 3 anti-body and DAPI and assessed by immunofluorescence.

RXR α , we then evaluated the effect of **6A** on cleaved PARP in RXR α knockout A549 cells (PX330-KO). Comparison of the obvious effect of **6A** in normal A549 cells (PX330), **6A** showed little effect on inducing PARP cleavage in RXR α knockout A549 cells (PX330-

KO; Figure 5(C)). Thus, **6A** could induce RXR α -dependent PARP cleavage in cancer cells.

To further confirm the role of RXR α in the apoptosis induction by **6A**, we evaluated the apoptosis effect of compound **6A** in

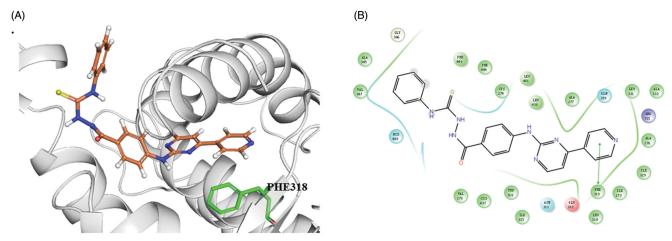


Figure 8. Predicted binding modes of 6A with RXR\(\alpha\)-LBP under antagonistic conformation (PDB ID: 3A9E). (A) Close-up view depiction (3 D model) of the superposition of RXR\alpha LBP bound to 6A. The RXR\alpha binding site is presented with a grey ribbon. The 6A is shown in stick with orange carbon, blue nitrogen, and yellow sulphur. The figure was prepared using PyMol (http://www.pymol.org/). (B) Two-dimensional diagram of interactions between compound 6A and RXR α . The π - π interactions are displayed as green solid line. Colour lines around 6A stand for the binding pocket and the residues in colours nearby established the pocket. The green colour denotes the hydrophobic nature of amino acids, the red colour denotes the acid amino acids, the purple denotes the alkalinity of amino acids, the cyan denotes the polar amino acids, and the grey points of ligand atoms denotes the solvent accessibility.

A549 and RXR α knockout A549 cells. As observed in Figure 6(A), **6A**-treated A549 cells exhibited obvious apoptosis characteristics with the accumulation of the nuclear convolution and fragmentation in membrane-enclosed bodies, but little apoptotic bodies were detected in treated RXRα knockout A549 cells. Meanwhile, Figure 6(B) showed that when A549 cells were incubated with 6A at $10.0\,\mu\text{M}$, the percentage of apoptotic cells was strikingly elevated to 30.8% from the control group (5.96%). However, the percentages of apoptotic cells in RXRa knockout A549 cells without and with 6A treatment were 2.07 and 2.72%, respectively. These results demonstrated that apoptosis cells increased significantly after the treatment of 6A in A549 cells, and the apoptosis induction of **6A** was mainly through RXRα-dependent pathways. It indicates the expression of $RXR\alpha$ plays a role in the apoptosis induction by 6A.

Finally, to find out whether the apoptotic induction of 6A is dependent on caspase pathway, we next used immunofluorescence assay to evaluate the effect of 6A on the activation of caspase-3 a major executioner caspase known to cleave PARP. As shown in Figure 7, 6A treatment of A549 cells resulted in the strong activation of caspase-3 and nuclear shrinkage compared with control cells, whereas there is no significant difference between treated and untreated RXR-/- A549 cells. This result was consistent with c-PARP. It suggests that compound 6A induces cell death in A549 cells via caspase-dependent pathway and RXRa dependent pathway.

2.3. Molecular docking studies

Based on the antagonist effect of 6A on RXR α , the molecular docking of **6A** with antagonistic conformation of RXR α (PDB: 3A9E) was performed using Schrodinger 2015.1 Glide protocol to explore their binding mode. The docking study showed that 6A was well accommodated into LBP of RXR α (see Figure 8(A) and Supplementary movie). The docking results predict the binding features of **6A** with RXR α . First, the right 4-(pyridin-4-yl)pyrimidine portion of compound 6A occupies the sub-pocket formed by hydrophobic aromatic amino acid residues LEU438, LEU331, ALA276, PHE318, and TRP310, the pyridine ring forming π - π interaction with PHE318. Second, the hydrazine-1-carbothioamide

structure locates at the polar region involving HID440. Third, the left phenyl group of Compound 6A is directed to the protein-solvent interface and makes hydrophobic interactions with surrounding lipophilic amino acids such as VAL347 and PHE443. These findings may provide a direction for the further modifications of this new class of RXR α regulator.

3. Conclusion

In summary, 2-(4-(pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide/carbothioamide (Series A) and 2-(3-(pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide/carbothioamide (Series B) derivatives were first synthesised and evaluated as potential antitumour agents targeting RXRα. Compounds 6c, 6e, 6h, 6A, and 6J exhibited potent antiproliferative activity with IC50 values of lower than 15 µM against two human tumour cell lines, while compounds 6k, 6A, 6B, 6C, 6G, and 6H were identified as good antagonists of RXR α with EC₅₀ values of lower than 10.0 μ M at RXRE-luciferase reporter-based assay. Compound 6A with the low cytotoxity of normal cells was identified as the most potent anticancer agent targeting RXRa. Moreover, it was found that the representative compound **6A** effectively induced RXRα-dependent apoptosis of cancer cells in a time- and concentration- dependent manner. SPR technology-based experiment confirmed that compound 6A could selectively bind to RXRα-LBD with a Kd value of 1.20×10^{-7} M, and molecular docking analysis showed that compound **6A** could bind to the LBP of RXRα like the classical RXRα antagonists. Taken together, this class of synthetic compounds may be served as a good starting point to design and synthesise new RXR α antagonists, and further studies of compound **6A** on activity in vivo would be considered in the future.

4. Experimental section

4.1. Chemistry

All reagents and solvents were purchased from commercial sources and were used without further purification. Oxygen or water sensitive reactions, which required the use of nitrogen atmosphere. All reactions were magnetically stirred and monitored by thin-layer chromatography (TLC) on (Qingdao Haiyang Chemical, China) silica gel 60 F-254 by fluorescence. ¹H-NMR and ¹³C-NMR spectra were obtained using a Bruker AV2 600 Ultra shield spectrometer at 600 and 150 MHz, respectively. Chemical shifts were given in ppm (δ) relative to tetramethylsilane as the internal standard. Spin multiplicities were described as s (singlet), d (doublet), dd (double doublet), t (triplet), br (broad signal), and orm (multiplet). High-resolution mass spectra (HRMS) data were acquired using a quadrupole-orbitrap (Q-Exactive) mass spectrometer (Thermo Scientific, Hemel Hempstead, UK) equipped with a heated electrospray ionisation source (HESI-II). Melting points were measured on SGW X-4 micro-melting point spectrometer and were uncorrected. The high-performance liquid chromatography (HPLC) analysis was performed on an Agilent Technologies 1100 Series HPLC system (Palo Alto, CA). The mobile phase was (A) methanol and (B) water in a linear gradient mode as follows: (A) from 5 to 100% and (B) from 95 to 0% during 0-30 min. The flow rate was 1 ml/min, and the detection wavelengths were 254 and 280 nm.

4.1.1. General procedure for the synthesis of compounds 3a-3e

To a solution of appropriate 1a-1e (0.041 mol) in toluene (50 ml), DMF-DMA (0.082 mol) was added, the reaction mixture was stirred at 110 °C for 18 h. Then, the mixture was cooled to room temperature, the solvent was removed by reduced pressure distillation. Then 20 ml ethyl acetate was added. The resulting solution was stirred at room temperature for 1 h, and filtered under suction, washed with (ethyl acetate: n-hexane = 3:1) to afford the compounds 3a-3e.

4.1.1.1. (E)-3-(dimethylamino)-1-(pyridin-4-yl)prop-2-en-1-one (3a). According to the general procedure, compound **3a** was obtained by using 4-acetylpyridine, yellow solid, 4.2 g, yield: 58.3%. Melting point (mp): 84–85 °C. ¹H NMR (600 MHz, DMSO-d₆) δ 8.67–8.69 (m, 2H), 7.79 (d, J= 12.1 Hz, 1H), 7.76 (d, J= 5.5 Hz, 2H), 5.84 (d, J= 12.1 Hz, 1H), 3.18 (s, 3H), 2.95 (s, 3H); ¹³C NMR (150 MHz, DMSO-d₆) δ 155.7, 150.6 (2C), 147.2, 121.5 (2C), 91.2, 45.2, 37.8 (2C).

4.1.1.2. (E)-3-(dimethylamino)-1-(pyridin-3-yl)prop-2-en-1-one (3b). According to the general procedure, compound **3b** was obtained by using 3-acetylpyridine, yellow solid, 3.2 g, yield: 49.5%. mp: 83–85 °C. ¹H NMR (600 MHz, DMSO-d₆) δ 9.07 (d, J= 1.7 Hz, 1H), 8.66 (dd, J= 4.8, 1.7 Hz, 1H), 8.22 (td, J= 8.0, 1.7 Hz, 1H), 7.77 (d, J= 12.1 Hz, 1H), 7.47–7.51 (m, 1H), 5.87 (d, J= 12.1 Hz, 1H), 3.17 (s, 3H), 2.94 (s, 3H); ¹³C NMR (150 MHz, DMSO-d₆) δ 155.1, 151.8, 149.0, 135.6, 135.1, 123.9, 91.4, 45.1, 37.7 (2C).

4.1.1.3. (E)-3-(dimethylamino)-1-(pyridin-2-yl)prop-2-en-1-one (3c). According to the general procedure, compound **3c** was obtained by using 2-acetylpyridine, Yellow solid, 3.5 g, yield: 53%. mp: 78–79 °C. 1 H NMR (600 MHz, DMSO-d₆) δ ppm 8.62–8.64 (m, 1 H) 7.99 (d, J=7.7 Hz, 1 H) 7.91 (td, J=7.7, 1.7 Hz, 1 H) 7.80 (d, J=12.0 Hz, 1 H) 7.50 (m, 1 H) 6.38 (d, J=12.0 Hz, 1 H) 3.18 (s, 3 H) 2.92 (s, 3 H); 13 C NMR (150 MHz, DMSO-d₆) δ 156.2, 154.8, 148.9, 137.5, 126.1, 121.6, 90.5, 45.1, 37.6 (2C).

4.1.1.4. (E)-3-(dimethylamino)-1-phenylprop-2-en-1-one (3d). According to the general procedure, compound **3d** was obtained by using acetophenone, Yellow solid, 2.6 g, yield: 60%. mp: $95-96\,^{\circ}\text{C.}^{-1}\text{H}$ NMR (600 MHz, DMSO-d₆) δ 7.87–7.89 (m, 2 H) 7.71

(d, J = 12.3 Hz, 1 H), 7.46–7.50 (m, 1 H), 7.41–7.45 (m, 2 H), 5.82 (d, J = 12.3 Hz, 1 H), 3.14 (s, 3 H), 2.91 (s, 3 H); ¹³C NMR (150 MHz, DMSO-d₆) δ 154.6, 140.7, 131.2, 128.6 (2C), 127.6 (2C), 91.4, 45.0, 37.6 (2C).

4.1.1.5. (E)-1-(4-chlorophenyl)-3-(dimethylamino)prop-2-en-1-one (3e). According to the general procedure, compound 3e was obtained by using 4-chloroacetophenone, yellow solid, 2.6 g, yield: 56%. mp: $101-102\,^{\circ}$ C. 1 H NMR (600 MHz, DMSO-d₆) δ 7.89–7.94 (m, 2H), 7.73 (d, J=12.1 Hz, 1H), 7.49–7.51 (m, 1H), 7.47–7.48 (m, 1H), 5.82 (d, J=12.1 Hz, 1H), 3.15 (s, 3H), 2.92 (s, 3H); 13 C NMR (150 MHz, DMSO-d₆) δ 184.7, 155.0, 139.4, 136.0, 129.5 (2C), 128.6 (2C), 91.1, 45.0, 37.6.

4.1.2. 4-Guanidinobenzoate ethyl ester nitrate (4a) and 3-guanidinobenzoate ethyl ester nitrate (4A)

4.1.2.1. 4-guanidinobenzoate ethyl ester nitrate (4a). To a solution of ethyl p-aminobenzoate (2a; 5.0 g, 0.030 mol) in ethanol (20 ml), cyanamide (50.0% w/w, 7.6 g, 0.091 mol) was added, Concentrated hydrochloric acid (3.8 ml, 0.045 mol) was added dropwise with stirring at 0 °C. After the completion of the dropwise addition, the temperature was raised to reflux for 24 h. The reaction mixture was concentrated under reduced pressure. The residue was diluted with 50 ml of water, and then an aqueous solution of ammonium nitrate (4.8 g, 0.061 mol) was added dropwise at 0 °C. After the completion of the dropwise addition, the mixture was kept for 1 h, then filtered to give 4a.

White solid, 6.3 g, yield: 78%. mp: $175-176\,^{\circ}\text{C}$. ^{1}H NMR (600 MHz, DMSO-d₆): δ 9.96 (br s, 1H), 8.00 (d, $J=8.6\,\text{Hz}$, 2H), 7.65 (s, 3H), 7.36 (d, $J=8.6\,\text{Hz}$, 2H), 4.32 (q, $J=7.1\,\text{Hz}$, 2H), 1.30–1.34 (m, 3H); ^{13}C NMR (150 MHz, DMSO-d₆): δ 165.6, 155.8, 140.9, 131.2(2C), 127.2, 123.4(2C), 61.2, 14.6.

4.1.2.2. 3-guanidinobenzoate ethyl ester nitrate (4A). According to the synthetic procedure of **4a**, compound **4A** was obtained by using ethyl 3-aminobenzoate (**2b**).

White solid, 6.0 g, yield: 74%. mp: $177-178\,^{\circ}\text{C}$. ¹H NMR (600 MHz, DMSO-d₆): δ 7.85 (m, 1H), 7.78 (t, $J=1.8\,\text{Hz}$, 1H), 7.58–7.62 (m, 1H), 7.51–7.56 (m, 4H), 4.34 (q, $J=7.0\,\text{Hz}$, 2H), 1.33 (t, $J=7.0\,\text{Hz}$, 3H); ¹³C NMR (150 MHz, DMSO-d₆): δ 165.6, 156.3, 136.5, 131.7, 130.7, 129.6, 127.3, 125.3, 61.5, 14.6.

4.1.3. General procedure for the synthesis of compounds 5a-5e

To a solution of (E)-3-(dimethylamino)-1-(pyridin-4-yl)prop-2-en-1-one (1.3 g, 7.4 mmol) and ethyl 4-guanidinobenzoate nitrate (4a;2.0 g, 7.4 mmol), sodium hydroxide (355 mg, 8.9 mmol) was added, and the mixture was heated to reflux for 48 h. After the completion of the reaction, the mixture was concentrated and purified by column chromatography using appropriate mixtures of dichloromethane and methanol (MeOH) to afford ethyl 4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoate, then to a solution of appropriate compound. To a solution of ethyl 4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoate (1.00 eq) in ethanol, hydrazine hydrate (1.00 eq) was added, and the reaction mixture was stirred at 87 °C for 20 h. The reaction mixture was cooled to room temperature, filtered and the concentrated filter cake was dried to obtain 5a. Yield: 84.3%.

Compounds **5b–5e** were synthesised following the general procedure as described above with the yield of 52–68%.



4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzohydrazide (5a). Yellow solid 1.1 g, yield: 58%. mp: 298–300 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.11 (s, 1H), 9.62 (s, 1H), 8.81 (dd, J = 4.4, 1.6 Hz, 2H), 8.72 (d, J = 5.1 Hz, 1H), 8.11 (dd, J = 4.4, 1.6 Hz, 2H), 7.92 (dt, J = 8.0, 1.8 Hz, 2H), 7.83 (dt, J = 8.0, 1.8 Hz, 2H), 7.60 (d, $J=5.1\,\mathrm{Hz},\ 1\mathrm{H}),\ 4.46$ (br s, 2H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.3, 162.0, 160.5, 160.4, 151.1, 144.2(2C), 143.4, 128.2 (2C), 126.5, 121.4 (2C), 118.4 (2C), 109.7.

4.1.3.2. 4-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzohydrazide (5b). White solid, 1.0 g, yield: 53%. mp: 285-287 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.05 (s, 1H), 9.61 (s, 1H), 9.36–9.37 (m, 1H), 8.75 (m, 1H), 8.66 (d, J = 5.1 Hz, 1H), 8.52 (dt, J = 8.01.8 Hz, 1H), 7.90 (dt, J = 8.0, 1.8 Hz, 2H), 7.82 (dt, J = 8.0, 1.8 Hz, 2H), 7.59–7.63 (m, 1 H), 7.58 (d, J = 5.1 Hz, 1H), 4.45 (br s, 2H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.3, 162.2, 160.4, 159.9, 152.1, 148.7, 143.5, 134.9, 132.6, 128.2, 126.4, 124.4 (2C), 118.3 (2C), 109.4.

4.1.3.3. 4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzohydrazide (5c). White solid, 0.8 g, yield: 68%. mp: 286–288 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.06 (s, 1H), 9.62 (s, 1H), 8.76–8.78 (m, 1 H), 8.70 (d, J = 5.1 Hz, 1H), 8.43 (d, J = 7.9 Hz, 1H), 8.07 (td, J = 7.9, 1.8 Hz, 1H), 7.92 (dt, J = 8.0, 1.8 Hz, 2H), 7.84 (dt, J = 8.0, 1.8 Hz, 2H), 7.79 (d, J = 5.1 Hz, 1H), 7.59 (m, 1H), 4.46 (br s, 2 H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.3, 163.4, 160.3, 160.1, 154.0, 150.1, 143.6, 138.1, 128.2, 126.3, 126.3, 121.6 (2C), 118.3 (2C), 109.1.

4.1.3.4. 4-((4-phenylpyrimidin-2-yl)amino)benzohydrazide White solid, 0.6 g, yield: 67%. mp: 303–305 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 9.98 (s, 1H), 9.60 (s, 1 H), 8.59–8.63 (m, 1H), 8.18–8.20 (m, 2H), 7.93 (d, J = 8.8 Hz, 2H), 7.82 (d, J = 8.8 Hz, 2H), 7.56–7.59 (m, 3H), 7.49 (d, $J = 5.1 \,\text{Hz}$, 1H), 4.44 (s, 2H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.3, 164.2, 160.4, 159.6, 143.7, 137.0, 131.5 (2C), 129.4 (2C), 128.2, 127.4(2C), 126.2, 118.2 (2C), 109.1

4.1.3.5. 4-((4-(4-chlorophenyl)pyrimidin-2-yl)amino)benzohydrazide (5e). Yellow solid, 1.1 g, yield: 58%. mp: 300-302 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 9.98 (s, 1 H), 9.60 (s, 1H), 8.61 (d, J = 5.1 Hz, 1H), 8.18-8.20 (m, 2H), 7.93 (d, J=8.8 Hz, 2H), 7.82 (d, J=8.8 Hz, 2H), 7.57–7.58 (m, 2H), 7.49 (d, J = 5.1 Hz, 1H), 4.44 (s, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.3, 163.0, 160.4, 159.8, 143.6, 136.3, 135.8, 129.5(2C), 129.2 (2C), 128.2 (2C), 126.3, 118.3 (2C), 109.0.

4.1.4. General procedure for the synthesis of compounds 5A-5C According to the synthesis of compounds 5a-5c, compounds 5A-5C were synthesised with 4A instead of 4a in a similar method.

3-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzohydrazide 4.1.4.1. (5A). Yellow solid, 0.5 g, yield: 63%. mp: 302–304°C. ¹H NMR (600 MHz, DMSO-d₆) δ 9.98 (s, 1H), 9.73 (s, 1H), 8.79–8.80 (m, 1H), 8.79–8.79 (m, 1H), 8.69 (d, J = 5.1 Hz, 1H), 8.45 (s, 1H), 8.13–8.17 (m, 2H), 7.87-7.91 (m, 1H), 7.58 (d, J = 5.1 Hz, 1H), 7.41-7.44 (m, 1H), 7.38-7.41 (m, 1H), 4.54 (br s, 2H); ¹³C NMR (150 MHz, DMSO d_6) δ 166.8, 161.8, 160.7, 160.4, 151.1 (2C), 144.2, 140.9, 134.5, 128.9, 122.0, 121.4 (2C), 120.2, 118.8, 109.2.

4.1.4.2. 3-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzohydrazide (5B). Yellow solid, 1.2 q, yield: 79%. mp: 306–308 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 9.92 (s, 1H), 9.72 (s, 1H), 9.38 (dd, J = 2.2, 0.7 Hz, 1H), 8.74 (dd, J = 4.7, 1.6, Hz, 1H), 8.64 (d, J = 5.1 Hz, 1H), 8.57 (dt, J = 8.1, 1.9 Hz, 1H), 8.41 (s, 1H), 7.89–7.92 (m, 1H), 7.60 (m, 1H), 7.55 (d, J = 5.1 Hz, 1H), 7.41–7.45 (m, 1H), 7.34–7.40 (m, 1H), 4.49–4.54 (m, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.8, 162.1, 160.6, 159.9, 152.0, 148.7, 141.0, 134.9, 134.5, 132.6, 128.9, 124.4, 121.9, 120.1, 118.7, 108.9.

3-((4-(Pyridine-2-yl)pyrimidin-2-yl)amino)benzohydrazide 4.1.4.3. (5C). Yellow solid, 1.1 g, yield: 68%. Mp: 311–312°C. ¹H NMR (600 MHz, DMSO-d₆): δ 9.92 (s, 1H), 9.72 (s, 1H), 8.76 (td, J = 4.6, 1.1 Hz, 1H), 8.67 (d, J = 5.1 Hz, 1H), 8.51 (d, J = 7.7 Hz, 1H), 8.48 (s, 1H), 8.05 (dt, J = 7.7, 1.7, Hz, 1H), 7.90 (td, J = 7.7, 1.7 Hz, 1H), 7.76 (d, J = 5.1 Hz, 1H), 7.58 (ddd, J = 7.7, 4.6, 1.1 Hz, 1H), 7.41-7.43 (m, J = 5.1 Hz, 1H), 7.58 (ddd, J = 7.7, 4.6, 1.1 Hz, 1H), 7.41-7.43 (m, J = 5.1 Hz, 1H), 7.58 (ddd, J = 7.7, 4.6, 1.1 Hz, 1H), 7.41-7.43 (m, J = 5.1 Hz, 1H)1H), 7.37-7.41 (m, 1H), 4.52 (br s, 2H); ¹³C NMR (150 MHz, DMSO d_6): δ 166.9, 163.3, 160.5, 160.1, 154.0, 150.1, 141.1, 138.1, 134.5, 128.9, 126.2, 121.8, 121.7, 120.0, 118.6, 108.7.

4.1.5. General procedure for the synthesis of target compounds 6a-6t and 6A-6J (Series A)

Compounds 5a-5e (1.00 eq) were separately used to react with an appropriate isocyanate or isothiocyanate (1.00 eq) in ethanol at 70 °C for 4h, then cooled to room temperature, filtered to get crude products of compounds 6a-6t and 6A-6J, which were further purified by silica gel column chromatography (DCM:MeOH = 50:1).

4.1.5.1. N-butyl-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide (6a). Yellow solid (94.52%), 0.056 g, yield: 58%, mp: 213–215 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.16 (s, 1H), 9.96 (br s, 1H), 8.82 (dd, J = 4.4, 1.4 Hz, 2H), 8.73 (d, J = 5.1 Hz, 1H), 8.13 (dd, J = 4.4, 1.4 Hz, 2H), 7.94 (d, J = 9.0 Hz, 2H), 7.89 (d, J = 9.0 Hz, 2H), 7.74 (s, 1H), 7.62 (d, J = 5.1 Hz, 1H), 6.45 (br s, 1H), 3.03 (q, J = 6.7 Hz, 2H), 1.38 (quin, J = 7.3 Hz, 2H), 1.28 (sxt, J = 7.3 Hz, 2H), 0.87 (t, J = 7.3 Hz, 3H); ¹³C NMR (150 MHz, DMSO d_6): δ 166.5, 161.9, 160.5, 159.0, 150.9 (2C), 144.4, 143.9 (2C), 128.9 (2C), 125.8, 121.5 (2C), 118.3 (2C), 109.8, 39.3, 32.5, 19.9, 14.2; HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ $[M+H]^+$ 406.1986, found 406.1985.

4.1.5.2. N-(tert-butyl)-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (6b). Yellow solid (99.24%), 0.054 g, yield: 60%, mp: 171–172 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.16 (s, 1H), 9.95 (s, 1H), 8.80 (dd, J = 4.4, 1.4 Hz, 2H), 8.73 (d, J = 5.1 Hz, 1H), 8.11 (dd, J = 4.4, 1.4 Hz, 2H), 7.93 (d, J = 9.0 Hz, 2H), 7.88 (d, J = 9.0 Hz, 2H), 7.61 (s, 1H), 7.60 (d, J = 5.1 Hz, 1H)6.08 (br s, 1H), 1.27 (s, 9H); 13 C NMR (150 MHz, DMSO- d_6): δ 166.3, 162.0, 160.5, 157.8, 151.1(2C), 144.2, 143.9 (2C), 128.7 (2C), 125.7, 121.4 (2C), 118.3 (2C), 109.8, 49.9, 29.6 (3 C); HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ [M+H]⁺ 406.1986, found 406.1983, calcd for $C_{21}H_{23}N_7O_2Na^+$ [M + Na]⁺ 428.1805, found 428.1803.

4.1.5.3. N-cyclohexyl-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (6c). White solid (93.67%), 0.055 g, yield: 58%, mp: 181–183 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.15 (s, 1H), 9.96 (s, 1H), 8.80 (dd, J = 4.4, 1.4 Hz, 2H), 8.72 (d, J = 5.1 Hz, 1H), 8.10 (dd, J = 4.4, 1.4 Hz, 2H), 7.92 (d, J = 9.0 Hz, 2H), 7.88 (d, J = 9.0 Hz, 2H), 7.69 (s, 1H), 7.60 (d, J = 5.1 Hz, 1H), 6.26 (d, J = 8.0 Hz, 1H), 3.38–3.43 (m, 1H), 1.75 (dd, J = 9.3, 3.8 Hz, 2H), 1.64

(dd, J= 9.3, 3.8 Hz, 2H), 1.53 (dd, J= 9.3, 3.8 Hz, 1H), 1.22–1.30 (m, 2H), 1.11–1.20 (m, 3H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.4, 162.0, 160.4, 158.2, 151.1(2C), 144.2, 143.9 (2C), 128.8 (2C), 125.7, 121.4 (2C), 118.3(2C), 109.8, 48.6, 33.5 (2C), 25.7, 25.1 (2C); HRMS (ESI, m/z): calcd for $C_{23}H_{26}N_7O_2^+$ [M + H]⁺ 432.2142, found 432.2141, calcd for $C_{23}H_{25}N_7O_2Na^+$ [M + Na]⁺ 454.1962, found 454.1960.

4.1.5.4. N-cyclopentyl-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)-benzoyl) hydrazine-1-carboxamide (6d). Yellow solid (95.45%), 0.054 g, yield: 57%, mp: 238–239 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.15 (s, 1H), 9.95 (s, 1H), 8.80 (dd, J = 4.4, 1.4 Hz Hz, 2H), 8.72 (d, J = 5.1 Hz, 1H), 8.11 (dd, J = 4.4, 1.4 Hz, 2H), 7.92–7.95 (m, 2H), 7.88 (d, J = 9.0 Hz, 2H), 7.67 (s, 1H), 7.60 (d, J = 5.1 Hz, 1H), 6.36 (d, J = 7.5 Hz, 1H), 3.90 (m, 1H), 1.76–1.83 (m, 2H), 1.58–1.65 (m, 2H), 1.45–1.53 (m, 2H), 1.33–1.41 (m, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.4, 162.0, 160.4, 158.5, 151.1(2C), 144.2, 143.9 (2C), 128.8 (2C), 125.8, 121.4 (2C), 118.3 (2C), 109.8, 51.6, 33.1 (2C), 23.7 (2C); HRMS (ESI, m/z): calcd for $C_{22}H_{24}N_7O_2^+$ [M + H] $^+$ 418.1986, found 418.1984, calcd for $C_{22}H_{23}N_7O_2Na^+$ [M + Na] $^+$ 440.1805, found 440.1804.

4.1.5.5. 2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl)-N-(ptolyl)hydrazine-1-carboxamide (6e). Yellow solid (99.58%), 0.061 g, yield: 69%, mp: 230–231 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.17 (s, 1H), 10.14 (s, 1H), 8.80 (dd, J = 4.4, 1.4 Hz, 2H), 8.76 (br s, 1H), 8.72 (d, J = 5.1 Hz, 1H), 8.11 (dd, J = 4.4, 1.4 Hz, 2H), 8.07 (s, 1H), 7.95 (d, J = 9.0 Hz, 2H), 7.93 (d, J = 9.0 Hz, 2H), 7.60 (d, J = 5.1 Hz, 1H), 7.35 (d, J = 8.2 Hz, 2H), 7.07 (d, J = 8.2 Hz, 2H), 2.23 (s, 3H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.7, 162.0, 160.5, 156.3, 151.1 (2C), 144.2, 144.1 (2C), 137.5, 131.2, 129.5 (2C), 128.9 (2C), 125.6, 121.4 (2C), 119.1(2C), 118.3 (2C), 109.8, 20.8; HRMS (ESI, m/z): calcd for $C_{24}H_{22}N_7O_2^+$ [M + H] $^+$ 440.1829, found 440.1824, calcd for $C_{24}H_{21}N_7O_2Na^+$ [M + Na] $^+$ 462.1649, found 462.1642.

4.1.5.6. N-(2,4-difluorophenyl)-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (6f). Yellow solid (93.67%), 0.065 g, yield: 71%, mp: 252–254 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.23 (br s, 1H), 10.17 (s, 1H), 8.79 (dd, J=4.4, 1.4 Hz, 2H), 8.72 (d, J=5.1 Hz, 1H), 8.65 (br s, 1H), 8.42 (s, 1H), 8.11 (dd, J=4.4, 1.4 Hz, 2H), 7.95 (d, J=9.0 Hz, 2H), 7.90 (d, J=9.0 Hz, 2H), 7.61 (d, J=5.1 Hz, 1H), 7.26–7.31 (m, 1H), 6.99–7.08 (m, 1H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.6, 162.0, 160.5, 160.4, 156.1, 151.1(2C), 144.2, 144.1(2C), 128.9 (2C), 125.4, 124.2, 121.4, 118.4 (2C), 111.6, 111.4, 109.8, 104.5, 104.3, 104.3, 104.1; HRMS (ESI, m/z): calcd for C₂₃H₁₈F₂N₇O₂+ [M+H]+ 462.1485, found 462.1476, calcd for C₂₃H₁₇F₂N₇O₂Na+ [M+Na]+ 484.1304, found 484.1296.

4.1.5.7. N-(3-chloro-4-methylphenyl)-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino) benzoyl)hydrazine-1-carboxamide (6g). Yellow solid (99.12%), 0.063 g, yield: 64%, mp: 209–210 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.18 (s, 1H), 10.15 (s, 1H), 8.97 (br s, 1H), 8.81 (dd, J= 4.4, 1.4 Hz, 2H), 8.73 (d, J= 5.1 Hz, 1H), 8.23 (br s, 1H), 8.12 (dd, J= 4.4, 1.4 Hz, 2H), 7.96 (d, J= 9.0 Hz, 2H), 7.93 (d, J= 9.0 Hz, 2H), 7.68 (d, J= 1.4 Hz, 1H), 7.62 (d, J= 5.1 Hz, 1H), 7.29 (br s, 1H), 7.22 (d, J= 8.2 Hz, 1H), 2.25 (s, 3H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.7, 162.0, 160.5, 160.4, 151.0(2C), 144.3, 144.1 (2C), 139.4, 133.4, 131.5, 128.9 (2C), 128.7, 125.6 (2C), 121.5 (2C), 118.3 (2C), 109.8, 19.2; HRMS (ESI, m/z): calcd for $C_{24}H_{21}CIN_7O_2^+$ [M + H] $^+$ 474.1440, found 474.1440, calcd for $C_{24}H_{20}CIN_7O_2Na^+$ [M + Na] $^+$ 496.1259, found 496.1259.

4.1.5.8. N-phenethyl-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)-benzoyl)hydrazine-1-carboxamide (6h). Yellow solid (96.95%), 0.072 g, yield: 75%, mp: 240–241 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.17 (s, 1H), 9.99 (s, 1H), 8.81 (dd, J = 4.4, 1.4 Hz, 2H), 8.74 (d, J = 5.1 Hz, 1H), 8.12 (dd, J = 4.4, 1.4 Hz, 2H), 7.94 (d, J = 9.0 Hz, 2H), 7.91 (d, J = 9.0 Hz, 2H), 7.86 (s, 1H), 7.62 (d, J = 5.1 Hz, 1H), 7.29 (t, J = 7.6 Hz, 3H), 7.20–7.24 (m, 3H), 7.17–7.20 (m, 1H), 6.51 (br s, 1H), 3.27 (d, J = 7.3 Hz, 2H), 2.72 (t, J = 7.3 Hz, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.5, 162.0, 160.5, 158.9, 151.1(2C), 144.2, 144.0 (2C), 140.1, 129.1 (2C), 128.9 (2C), 128.8, 128.8, 126.5, 125.8, 121.4 (2C), 118.3 (2C), 109.8, 41.4, 36.5; HRMS (ESI, m/z): calcd for $C_{25}H_{24}N_7O_2^+$ [M + H] $^+$ 454.1986, found 454.1979, calcd for $C_{25}H_{23}N_7NaO_2^+$ [M + Na] $^+$ 476.1805, found 476.1799.

4.1.5.9. N-butyl-2-(4-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide (6i). Yellow solid (99.63%), 0.076 g, yield: 81%, mp: 179–181 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.10 (s, 1H), 9.96 (br s, 1H), 9.36 (d, J = 2.0 Hz, 1H), 8.75 (dd, J = 4.7, 2.0, Hz, 1H), 8.67 (d, J = 5.1 Hz, 1H), 8.53 (dt, J = 8.0, 2.0 Hz, 1H), 7.93 (d, J = 9.0 Hz, 2H), 7.89 (d, J = 9.0 Hz, 2H), 7.73 (s, 1H), 7.61 (dd, 1H)J = 8.0, 4.7, Hz, 1H, 7.59 (d, J = 5.1 Hz, 1H), 6.45 (br s, 1H), 3.03–3.04 (m, 2H), 1.38 (quin, J = 7.3 Hz, 2H), 1.28 (sxt, J = 7.3 Hz, 2H), 0.87 (t, J = 7.3 Hz, 3H); ¹³C NMR (150 MHz, DMSO- d₆): δ 166.5, 162.2, 160.3, 160.0, 159.0, 152.1, 148.6, 144.0, 134.9, 132.5, 128.9 (2C), 125.7, 124.5, 118.2 (2C), 109.5, 39.3, 32.5, 19.9, 14.2; HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ $[M + H]^{+}$ 406.1986, found 406.1980.

4.1.5.10. N-cyclopentyl-2-(4-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)-benzoyl) hydrazine-1-carboxamide (6j). Yellow solid (95.47%), 0.071 g, yield: 75%, mp: 159–161 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.11 (s, 1H), 9.94 (s, 1H), 9.37 (d, J= 1.6 Hz, 1H), 8.75 (dd, J= 4.7, 1.6, Hz, 1H), 8.68 (d, J= 5.1 Hz, 1H), 8.53 (d, J= 7.5 Hz, 1H), 7.92–7.95 (d, J= 9.0 Hz, 2H), 7.88 (d, J= 9.0 Hz, 2H), 7.66 (s, 1H), 7.61–7.63 (m, 1H), 7.59 (d, J= 5.1 Hz, 1H), 6.35 (d, J= 7.5 Hz, 1H), 3.91 (m, 1H), 1.76–1.83 (m, 2H), 1.58–1.65 (m, 2H), 1.45–1.53 (m, 2H), 1.33–1.41 (m, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.4, 162.2, 160.4, 159.9, 158.5, 152.1, 148.7, 144.0, 134.9, 132.5, 128.8 (2C), 125.7, 124.5, 118.2 (2C), 109.5, 51.5, 33.1 (2C), 23.7 (2C); HRMS (ESI, m/z): calcd for $C_{22}H_{23}N_7NaO_2^+$ [M+H] $^+$ 418.1986, found 418.1980, calcd for $C_{22}H_{23}N_7NaO_2^+$ [M+Na] $^+$ 440.1805, found 440.1799.

4.1.5.11. 2-(4-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzoyl)-N-(ptolyl)hydrazine-1-carboxamide (6k). Yellow solid (96.10%), 0.056 g, yield:59%, mp: 209–210 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.13 (s, 2H), 9.37 (d, J = 1.6 Hz, 1H), 8.76 (dd, J = 4.7, 1.6 Hz, 1H), 8.74 (br s, 1H), 8.68 (d, J = 5.1 Hz, 1H), 8.53 (dt, J = 8.0, 1.6 Hz, 1H), 8.07 (s, 1H), 7.95 (d, J = 9.0 Hz, 2H), 7.92 (d, J = 9.0 Hz, 2H), 7.61–7.63 (m, 1H), 7.60 (d, J = 5.1 Hz, 1H), 7.36 (d, J = 8.2 Hz, 2H), 7.07 (d, J = 8.2 Hz, 2H), 2.24 (s, 3H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.6, 162.2, 160.4, 160.0, 152.1, 148.7, 144.2, 137.6, 135.0, 132.5, 131.1, 129.6, 129.5 (2C), 128.9 (2C), 125.5, 124.5, 121.5 (2C), 118.3 (2C), 109.5, 20.8; HRMS (ESI, m/z): calcd for $C_{24}H_{21}N_7NaO_2^+$ [M + H]⁺ 440.1829, found 440.1823, calcd for $C_{24}H_{21}N_7NaO_2^+$ [M + Na]⁺ 462.1649, found 462.1643.

4.1.5.12. N-phenethyl-2-(4-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)-benzoyl) hydrazine-1-carboxamide (6l). Yellow solid (95.57%), 0.054 g, yield: 56%, mp: 177–178 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.09 (s, 1H), 9.99 (s, 1H), 9.35 (d, J = 2.0 Hz, 1H), 8.74 (dd, J = 4.7, 1.4 Hz, 1H), 8.66 (d, J = 5.1 Hz, 1H), 8.52 (dt, J = 8.0, 1.4 Hz,

1H), 7.93 (d, $J = 9.0 \,\text{Hz}$, 2H), 7.88–7.90 (d, $J = 9.0 \,\text{Hz}$, 2H), 7.86 (s, 1H), 7.61 (dd, J = 8.0, 4.7 Hz, 1H), 7.58 (d, J = 5.1 Hz, 1H), 7.26-7.30 (m, 2H), 7.20-7.23 (m, 2H), 7.17-7.19 (m, 1H), 6.53 (br s, 1H), 3.22-3.29 (m, 2H), 2.71 (t, $J = 7.3 \,\text{Hz}$, 2H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.6, 162.2, 160.3, 160.0, 159.0, 152.1, 148.6, 144.1, 140.0, 135.0, 132.5, 129.1 (2C), 128.9 (2C), 128.8 (2C), 126.5, 125.6, 124.5, 118.2 (2C), 109.5, 41.4, 36.4; HRMS (ESI, m/z): calcd for $C_{25}H_{24}N_7O_2^+$ [M + H]⁺ 454.1986, found 454.1981.

4.1.5.13. N-(tert-butyl)-2-(4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (6m). Yellow solid (98.73%), 0.076 g, yield: 79%, mp: 213–214 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.10 (s, 1H), 9.95 (s, 1H), 8.76 (s, 1H), 8.71 (d, $J = 4.9 \, \text{Hz}$, 1H), 8.43 (d, $J = 7.8 \,\mathrm{Hz}$, 1H), 8.07 (td, J = 7.8, 1.6 Hz, 1H), 7.94 (d, J = 8.6 Hz, 2H), 7.88 (d, J = 8.6 Hz, 2H), 7.80 (d, J = 4.9 Hz, 1H), 7.58-7.61 (m, 2H), 6.08 (br s, 1H), 1.27 (s, 9H); 13C NMR (150 MHz, DMSO-d₆): δ 166.3, 163.4, 160.2, 160.1, 157.8, 153.9, 150.2, 144.1, 138.1, 128.7 (2C), 126.3, 125.6, 121.6, 118.2 (2C), 109.2, 49.9, 29.6 (3 C); HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ $[M+H]^+$ 406.1986, found 406.1982.

4.1.5.14. N-butyl-2-(4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide (6n). White solid (96.55%), 0.072 g, yield: 78%, mp: 228–229 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.10 (s, 1H), 9.96 (s, 1H), 8.77 (s, 1H), 8.71 (d, J = 4.9 Hz, 1H), 8.43 (d, J = 7.8 Hz, 1H), 8.07 (td, J = 7.8, 1.6 Hz, 1H), 7.95 (d, J = 8.6 Hz, 2H), 7.90 (d, $J = 8.6 \,\text{Hz}$, 2H), 7.80 (d, $J = 4.9 \,\text{Hz}$, 1H), 7.74 (s, 1H), 7.57-7.62 (m, 1H), 6.44 (br s, 1H), 3.03 (m, 2H), 1.39 (quin, J = 7.3 Hz, 2H), 1.28 (sxt, J = 7.3 Hz, 2H), 0.88 (t, J = 7.3 Hz, 3H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.6, 163.4, 160.2, 160.1, 159.1, 153.9, 150.1, 144.1, 138.1, 128.9 (2C), 126.3, 125.6, 121.6, 118.2 (2C), 109.2, 32.4, 19.9, 14.2; HRMS (ESI, m/z): calcd for C₂₁H₂₄N₇O₂ $[M+H]^+$ 406.1986, found 406.1985, calcd for $C_{21}H_{23}N_7O_2Na^+$ $[M + Na]^+$ 428.1805, found 428.1806.

4.1.5.15. N-cyclopentyl-2-(4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (60). White solid (95.44%), 0.063 g, yield: 73%, mp: 156–158 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.10 (s, 1H), 9.94 (s, 1H), 8.77 (d, $J = 4.0 \,\text{Hz}$, 1H), 8.71 (d, J = 4.9 Hz, 1H), 8.43 (d, J = 7.8 Hz, 1H), 8.07 (td, J = 7.8, 1.6 Hz, 1H), 7.94–7.97 (d, $J = 8.6 \,\text{Hz}$, 2H), 7.90 (d, $J = 8.6 \,\text{Hz}$, 2H), 7.80 (d, J = 4.9 Hz, 1H), 7.66 (s, 1H), 7.59 (m, 1H), 6.35 (d, J = 7.5 Hz, 1H), 3.91 (m, 1H), 1.76-1.84 (m, 2H), 1.58-1.66 (m, 2H), 1.46-1.54 (m, 2H), 1.34–1.41 (m, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.4, 163.4, 160.2, 160.1, 158.5, 153.9, 150.2, 144.1, 138.1, 128.8 (2C), 126.3, 125.6, 121.6, 118.2 (2C), 109.2, 51.5, 33.1 (2C), 23.7 (2C); HRMS (ESI, m/z): calcd for $C_{22}H_{24}N_7O_2^+$ $[M+H]^+$ 418.1986, found 418.1980.

4.1.5.16. 2-(4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl)-N-(mtolyl) hydrazine-1-carboxamide (6p). Yellow solid (97.54%), 0.054 g, yield: 67%, mp: 229–230 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.13 (s, 1H), 8.77 (d, J = 4.0 Hz, 2H), 8.72 (d, J = 5.1 Hz, 1H), 8.44 (d, J = 7.7 Hz, 1H), 8.11 (s, 1H), 8.07 (dt, J = 7.7, 1.6 Hz, 1H), 7.98 (d, J = 7.7, 1.6 Hz, 1Hz), 7.98 (d, J = 7.7, 1.6 Hz), 7.98 (d, JJ = 8.6 Hz, 2H), 7.93 (d, J = 8.6 Hz, 2H), 7.81 (d, J = 5.1 Hz, 1H), 7.58–7.61 (m, 1H), 7.31 (s, 1H), 7.29 (d, $J = 7.7 \,\mathrm{Hz}$, 1H), 7.14 (t, J = 7.7 Hz, 1H), 6.78 (d, J = 7.7 Hz, 1H), 2.26 (s, 3H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.6, 163.4, 160.3, 160.1, 156.3, 153.9, 150.2, 144.2, 140.1, 138.2, 138.1, 128.9 (2C), 128.9, 126.3, 125.5, 123.1, 121.6, 119.5, 118.2 (2C), 116.1, 109.2, 21.7; HRMS (ESI, m/z): calcd for $C_{24}H_{22}N_7O_2^+$ [M+H]⁺ 440.1829, found 440.1826, calcd for $C_{24}H_{21}N_7NaO_2^+$ [M + Na]⁺ 462.1649, found 462.1646.

4.1.5.17. N-(3,5-dimethylphenyl)-2-(4-((4-(pyridin-2-yl)pyrimidin-2yl)amino) benzoyl)hydrazine-1-carboxamide (6q). Yellow solid (98.69%), 0.073 g, yield: 79%, mp: 245–246 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.13 (s, 1H), 10.12 (s, 1H), 8.77 (d, $J = 4.0 \,\text{Hz}$, 1H), 8.72 (d, J = 5.1 Hz, 1H), 8.68 (br s, 1H), 8.44 (d, J = 7.8 Hz, 1H), 8.06-8.10 (m, 2H), 7.98 (d, J=8.6 Hz, 2H), 7.93 (d, J=8.6 Hz, 2H), 7.81 (d, J = 5.1 Hz, 1H), 7.58–7.61 (m, 1H), 7.11 (s, 2H), 6.60 (s, 1H), 2.22 (s, 6H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.6, 163.4, 160.2, 160.1, 156.2, 153.9, 150.2, 144.2, 140.0, 138.1, 138.0 (2C), 128.9 (2C), 126.3, 125.5, 123.9 (2C), 121.6, 118.2 (2C), 116.7, 109.2, 21.6 (2C); HRMS (ESI, m/z): calcd for $C_{25}H_{24}N_7O_2^+$ [M+H]⁺ 454.1986, found 454.1983.

4.1.5.18. N-(tert-butyl)-2-(4-((4-(4-chlorophenyl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (6r). White solid (92.63%), 0.065 g, yield:76%, mp: 170–172 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.05 (s, 1H), 9.93 (s, 1H), 8.63 (d, J = 5.1 Hz, 1H), 8.21 (d, J = 8.6 Hz, 2H), 7.92 (d, J = 8.8 Hz, 2H), 7.86 (d, J = 8.8 Hz, 2H), 7.64 $(d, J = 8.6 \,Hz, 2H), 7.59 (s, 1H), 7.51 (d, J = 5.1 \,Hz, 1H), 6.07 (br s, 1H)$ 1H), 1.26 (s, 9H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.3, 163.0, 160.3, 159.8, 157.8, 144.1, 136.3, 135.8, 129.5 (2C), 129. 2(2C), 128.7 (2C), 125.5, 118.2 (2C), 109.1, 49.9, 29.6 (3 C); HRMS (ESI, m/z): calcd for $C_{22}H_{24}CIN_6O_2^+$ $[M+H]^+$ 439.1644, found 439.1641, calcd for $C_{22}H_{23}CIN_6NaO_2^+$ [M + Na]⁺ 461.1463, found 461.1459.

4.1.5.19. 2-(4-((4-(4-chlorophenyl)pyrimidin-2-yl)amino)benzoyl)-Nethylhydrazine-1-carboxamide (6s). White solid (97.87%), 0.077 g, yield: 82%, mp: 241–242 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.06 (s, 1H), 9.95 (s, 1H), 8.64 (d, J = 5.1 Hz, 1H), 8.22 (d, J = 8.6 Hz, 2H), 7.93 (d, $J = 8.8 \,\text{Hz}$, 2H), 7.88 (d, $J = 8.8 \,\text{Hz}$, 2H), 7.76 (s, 1H), 7.62–7.68 (m, 2H), 7.52 (d, J = 5.1 Hz, 1H), 6.45 (br s, 1H), 3.01–3.11 (m, 2H), 1.02 (t, $J = 7.0 \,\text{Hz}$, 3H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.6, 163.0, 160.3, 159.8, 158.9, 144.1, 136.3, 135.7, 129.5 (2C), 129.2 (2C), 128.9 (2C), 125.5, 118.2 (2C), 109.1, 34.5, 16.0; HRMS (ESI, m/z): calcd for $C_{20}H_{20}CIN_6O_2^+$ $[M+H]^+$ 411.1331, found 411.1330, calcd for $C_{20}H_{19}CIN_6NaO_2^+$ $[M + Na]^+$ 433.1150, found 433.1149.

4.1.5.20. 2-(4-((4-(4-chlorophenyl)pyrimidin-2-yl)amino)benzoyl)-Ncyclohexyl-hydrazine-1-carboxamide (6t). White solid (97.94%), 0.046 g, yield: 48%, mp: $165-167\,^{\circ}\text{C}$. ¹H NMR (600 MHz, DMSO-d₆): δ 10.06 (s, 1H), 9.94 (s, 1H), 8.64 (d, J = 5.1 Hz, 1H), 8.20–8.23 (m, 2H), 7.92 (d, $J = 8.8 \,\text{Hz}$, 2H), 7.88 (d, $J = 8.8 \,\text{Hz}$, 2H), 7.69 (s, 1H), 7.63–7.66 (m, 2H), 7.52 (d, $J = 5.1 \,\text{Hz}$, 1H), 6.24 (d, $J = 8.0 \,\text{Hz}$, 1H), 3.38-3.47 (m, 1H), 1.73-1.78 (m, 2H), 1.66 (m, 2H), 1.51-1.56 (m, 1H), 1.22-1.31 (m, 2H), 1.10-1.21 (m, 3H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.4, 163.0, 160.3, 159.8, 158.1, 144.1, 136.3, 135.8, 129.5 (2C), 129.2 (2C), 128.8 (2C), 125.6, 118.1 (2C), 109.1, 48.6, 33.5 (2C), 25.7, 25.1 (2C); HRMS (ESI, m/z): calcd for $C_{24}H_{26}CIN_6O_2^+$ $[M+H]^+$ 465.1800, found 465.1799, calcd for $C_{24}H_{25}CIN_6NaO_2^+$ $[M + Na]^+$ 487.1620, found 487.1619.

4.1.5.21. N-phenyl-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6A). Yellow solid (97.57%), 0.054 g, yield: 66%, mp: 267–269 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.40 (s, 1H), 10.19 (s, 1H), 9.79 (br s, 1H), 9.68 (br s, 1H), 8.80 (dd, J = 4.4, 1.4 Hz, 2H), 8.74 (d, J = 4.9 Hz, 1H), 8.11 (dd, J = 4.4,1.4 Hz, 2H), 7.96 (br s, 4H), 7.63 (d, J = 4.9 Hz, 1H), 7.46 (br s, 2H), 7.33 (t, J = 7.7 Hz, 2H), 7.16 (t, J = 7.7 Hz, 1H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.1, 162.0, 160.5, 160.4, 151.1(2C), 144.2, 139.8 (2C), 129.3 (2C), 128.4 (2C), 128.2, 126.4 (2C), 125.5(2C), 121.4, 118.4, 118.1 (2C), 109.8; HRMS (ESI, m/z): calcd for $C_{23}H_{20}N_7OS^+$ $[M+H]^+$

442.1445, found 442.1440, calcd for $C_{23}H_{19}N_7NaOS^+$ [M + Na]⁺ 464.1264, found 464.1261.

4.1.5.22. N-benzyl-2-(4-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6B). Yellow solid (98.33%), 0.065 g, yield: 73%, mp: 240–241 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.27 (s, 1H), 10.18 (s, 1H), 9.42 (s, 1H), 8.80 (dd, J = 4.4, 1.4 Hz, 2H), 8.73 (d, J = 5.1 Hz, 1H), 8.65 (br s, 1H), 8.11 (dd, J = 4.4, 1.4 Hz, 2H), 7.91–7.97 (m, 4H), 7.62 (d, J = 5.1 Hz, 1H), 7.29–7.33 (m, 4H), 7.20–7.24 (m, 1H), 4.75 (d, J = 6.0 Hz, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.1, 162.0, 160.5, 160.4, 151.1(2C), 144.1, 140.0 (2C), 129.2 (2C), 128.5 (2C), 127.5, 127.0 (2C), 125.5, 121.4(2C), 118.1(2C), 109.8, 47.2; HRMS (ESI, m/z): calcd for $C_{24}H_{21}N_7OSNa^+$ [M + H]⁺ 456.1601, found 478.1415.

4.1.5.23. N-benzyl-2-(4-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6C). Yellow solid (98.60%), 0.072 g, yield: 75%, mp: 159–160 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.27 (s, 1H), 10.12 (s, 1H), 9.42 (s, 1H), 9.35–9.38 (s, 1H), 8.75 (dd, J= 4.9, 1.8 Hz, 1H), 8.68 (d, J= 5.1 Hz, 1H), 8.66 (d, J= 4.9 Hz, 1H), 8.52 (td, J= 8.0, 1.8 Hz, 1H), 7.92–7.96 (br s, 4H), 7.61–7.63 (m, 1H), 7.60 (d, J= 5.1 Hz, 1H), 7.28–7.34 (m, 4H), 7.20–7.24 (m, 1H), 4.75 (d, J= 6.0 Hz, 2H); 13 C NMR (150 MHz, DMSO- d₆): δ 166.2, 162.2, 160.3, 160.0, 152.1, 148.7, 144.2, 140.0, 134.9, 132.5, 129.2 (2C), 128.5 (2C), 127.5, 127.0 (2C), 125.4, 124.5, 118.1 (2C), 109.5, 47.2; HRMS (ESI, m/ z): calcd for C₂₄H₂₁N₇NaOS⁺ [M + H]⁺ 456.1601, found 478.1419.

4.1.5.24. N-phenyl-2-(4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6D). Yellow solid (99.24%), 0.046 g, yield: 48%, mp: 200–202 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.39 (s, 1H), 10.13 (s, 1H), 9.80 (br s, 1H), 9.68 (s, 1H), 8.76–8.78 (d, J= 4.0 Hz, 1H), 8.72 (d, J= 5.1 Hz, 1H), 8.43 (d, J= 7.7 Hz, 1H), 8.07 (td, J= 7.7, 1.8 Hz, 1H), 7.98 (s, 4H), 7.81 (d, J= 5.1 Hz, 1H), 7.59 (m, 1H), 7.46 (d, J= 1.8 Hz, 2H), 7.34 (t, J= 7.7 Hz, 2H), 7.13–7.19 (m, 1H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.2, 163.4, 160.2, 160.1, 153.9, 150.2, 144.3, 139.8, 138.1, 129.3 (2C), 128.4 (2C), 126.5, 126.3 (2C), 125.4, 121.6, 118.0 (2C), 109.3, 40.5; HRMS (ESI, m/z): calcd for $C_{23}H_{20}N_7OS^+$ [M + H] $^+$ 442.1445, found 442.1444, calcd for $C_{23}H_{19}N_7OSNa^+$ [M + Na] $^+$ 464.1264, found 464.1265.

4.1.6.25. N-benzyl-2-(4-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6E). White solid (99.14%), 0.045 g, yield: 67%, mp: 222–223 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.27 (s, 1H), 10.12 (s, 1H), 9.42 (s, 1H), 8.76–8.78 (m, 1H), 8.71 (d, J = 4.9 Hz, 1H), 8.65 (br s, 1H), 8.42 (d, J = 7.7 Hz, 1H), 8.07 (td, J = 7.7, 1.8 Hz, 1H), 7.92–7.98 (br s, 4H), 7.80 (d, J = 4.9 Hz, 1H), 7.59 (m, 1H), 7.28–7.34 (m, 4H), 7.20–7.24 (m, 1H), 4.75 (m, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.2, 163.4, 160.2, 160.2, 153.9, 150.2, 144.3, 140.0, 138.1, 129.3 (2C), 128.5 (2C), 127.5, 127.0 (2C), 126.3, 125.3, 121.6, 118.0 (2C), 109.3, 70.2, 47.2; HRMS (ESI, m/z): calcd for $C_{24}H_{22}N_7OS^+$ [M + H] $^+$ 456.1601, found 456.1599, calcd for $C_{24}H_{21}N_7NaOS^+$ [M + Na] $^+$ 478.1421, found 478.1419.

4.1.6.26. N-cyclohexyl-2-(4-((4-phenylpyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6F). White solid (98.99%), 0.032 g, yield:59%, mp: 230–232 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.10 (br s, 1H), 10.05 (s, 1H), 9.16 (br s, 1H), 8.63 (d, J=5.1 Hz, 1H), 8.20–8.21 (m, 1H), 8.18–8.20 (m, 1H), 7.96 (d, J=8.8 Hz, 2H), 7.92 (d, J=8.8 Hz, 2H), 7.68 (d, J=3.8 Hz, 1H), 7.56–7.60 (m, 3H), 7.51 (d, J=5.1 Hz, 1H), 4.15 (br s, 1H), 1.79 (br s, 2H), 1.64–1.72 (m,

2H), 1.57 (d, J=13.0 Hz, 1H), 1.28–1.33 (m, 2H), 1.22–1.27 (m, 2H), 1.01–1.12 (m, 1H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.0, 164.2, 160.3, 159.6, 144.4, 137.0, 131.5 (2C), 129.5 (2C), 129.2, 127.4 (2C), 125.2, 118.0 (2C), 109.3, 53.4, 32.3 (2C), 25.6, 25.4 (2C); HRMS (ESI, m/z): calcd for C₂₄H₂₇N₆OS⁺ [M + H]⁺ 447.1962, found 447.1959, calcd for C₂₄H₂₆N₆NaOS⁺ [M + Na]⁺ 469.1781, found 469.1780.

4.1.5.27. N-phenyl-2-(4-((4-phenylpyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6G). White solid (97.80%), 0.034 g, yield: 50%, mp: 210–212 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.38 (br s, 1H), 10.06 (s, 1H), 9.79 (br s, 1H), 9.67 (br s, 1H), 8.63 (d, J = 5.3 Hz, 1H), 8.19–8.21 (m, 2H), 7.98 (d, J = 8.8 Hz, 2H), 7.95 (d, J = 8.8 Hz, 2H), 7.57–7.59 (m, 3H), 7.51 (d, J = 5.3 Hz, 1H), 7.43–7.49 (m, 2H), 7.33 (t, J = 7.8 Hz, 2H), 7.13–7.19 (m, 1H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.2, 164.2, 160.3, 159.7, 144.5, 139.8, 137.0, 131.5 (2C), 129.5 (2C), 129.3 (2C), 128.4, 127.4 (2C), 126.5 (2C), 125.5, 125.3, 118.0 (2C), 109.3; HRMS (ESI, m/z): calcd for C₂₄H₂₁N₆OS⁺[M+H]⁺ 441.1492, found 441.1490, calcd for C₂₄H₂₀N₆NaOS⁺[M+Na]⁺ 463.1312, found 463.1309.

4.1.5.28. N-benzyl-2-(4-((4-phenylpyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (6H). White solid (99.08%), 0.054 g, yield: 62%, mp: 289–291 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.26 (s, 1H), 10.05 (s, 1H), 9.41 (s, 1H), 8.65 (br s, 1H), 8.62 (d, J=5.1 Hz, 1H), 8.17–8.22 (m, 2H), 7.97 (d, J=8.8 Hz, 2H), 7.93 (d, J=8.8 Hz, 2H), 7.57 (m, 3H), 7.51 (dd, J=5.1, 1.6 Hz, 1H), 7.28–7.34 (m, 4H), 7.22 (d, J=5.5 Hz, 1H), 4.75 (d, J=5.5 Hz, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.2, 164.2, 160.3, 159.7, 144.4, 140.0, 136.9, 131.6 (2C), 129.5 (2C), 129.2, 128.5 (2C), 127.5, 127.4 (2C), 127.0 (3 C), 125.2, 117.9 (2C), 109.3, 47.2; HRMS (ESI, m/z): calcd for $C_{25}H_{23}N_6OS^+$ [M+H]⁺ 455.1649, found 455.1644, calcd for $C_{25}H_{22}N_6NaOS^+$ [M+Na]⁺ 477.1468, found 477.1465.

4.1.5.29. 2-(4-((4-(4-chlorophenyl)pyrimidin-2-yl)amino)benzoyl)-N-phenylhydrazine-1-carbothioamide (6l). White solid (98.95%), 0.045 g, yield: 48%, mp: 275–277 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.38 (s, 1H), 10.09 (s, 1H), 9.79 (br s, 1H), 9.68 (s, 1H), 8.65 ((d, J=5.1, 1H), 8.22 (d, J=8.6 Hz, 2H), 7.95 (br s, 4H), 7.65 (d, J=8.6 Hz, 2H), 7.52 (d, J=5.1 Hz, 1H), 7.46 (br s, 2H), 7.33 (m, 2H), 7.13–7.19 (m, 1H). 13 C NMR (150 MHz, DMSO-d₆): δ 163.0, 160.3, 159.9, 144.3, 139.8, 136.3, 135.8, 129.5 (2C), 129.3 (2C), 129.2 (2C), 128.4 (2C), 126.5 (2C), 125.5, 125.4, 118.0 (2C), 109.2; HRMS (ESI, m/z): calcd for C₂₄H₂₀ClN₆OS⁺ [M+H]⁺ 475.1102, found 475.1100, calcd for C₂₄H₁₉ClN₆NaOS⁺ [M+Na]⁺ 497.0922, found 497.0920.

4.1.5.30. N-benzyl-2-(4-((4-(4-chlorophenyl)pyrimidin-2-yl)amino)-benzoyl) hydrazine-1-carbothioamide (6J). White solid (95.91%), 0.035 g, yield: 56%, mp: 235–237 °C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.26 (s, 1H), 10.08 (s, 1H), 9.42 (s, 1H), 8.63 (d, J = 5.1 Hz, 2H), 8.20 (d, J = 8.6 Hz, 2H), 7.91 – 7.95 (br s, 4H), 7.65 (d, J = 8.6 Hz, 2H), 7.52 (d, J = 5.1 Hz, 1H), 7.29–7.33 (m, 4H), 7.20– 7.24 (m, 1H), 4.75 (d, J = 6.0 Hz, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.2, 163.0, 160.3, 159.9, 144.3, 140.0, 136.3, 135.8, 129.5 (2C), 129.2 (2C), 128.5 (2C), 127.5 (2C), 127.0 (2C), 125.3, 118.0 (2C), 109.2, 47.2; HRMS (ESI, m/z): calcd for $C_{25}H_{22}$ CIN₆OS⁺ [M + H]⁺ 489.1259, found 489.1255, calcd for $C_{25}H_{21}$ CIN₆NaOS⁺ [M + Na]⁺ 511.1078, found 511.1075.

4.1.6. General procedure for the synthesis of compounds 7a-7f and 7A-7D (Series B)

Compounds **5A–5C** (1.00 eq) were separately used to react with an appropriate isocyanate or isothiocyanate (1.00 eq) in ethanol at



70 °C for 4h, then cooling to room temperature, filtered to get crude products of compounds 7a-7f and 7A-7D, which were further purified by silica gel column chromatography (DCM:MeOH).

N-(tert-butyl)-2-(3-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl) hydrazine-1-carboxamide (7a). Yellow solid (98.13%), 0.076 g, yield: 83%, mp: 161–162 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.09 (br s, 1H), 10.06 (s, 1H), 8.84 (d, J = 6.0 Hz, 2H), 8.75 (d, J = 5.1 Hz, 1H), 8.56 (s, 1H), 8.20 (d, J = 6.0 Hz, 2H), 7.94 (dd, J = 8.0, 1.4 Hz, 1H), 7.72 (s, 1H), 7.64 (d, J = 5.1 Hz, 1H), 7.54 (d, J = 7.7 Hz, 1H), 7.48 (t, J = 7.7 Hz, 1H), 6.04–6.20 (m, 1H), 1.33 (s, 9H); ¹³C NMR (150 MHz, DMSO- d_6): δ 167.0, 161.8, 160.7, 160.5, 157.7, 151.1, 144.2 (2C), 140.9, 133.9, 129.0, 122.5, 121.5 (2C), 120.7, 119.0, 109.3, 50.0, 29.6 (3 C); HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ $[M+H]^+$ 406.1986, found 406.1982, calcd for $C_{21}H_{23}N_7NaO_2^+$ $[M + Na]^+$ 428.1805, found 428.1801.

4.1.6.2. N-cyclopentyl-2-(3-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzovl) hydrazine-1-carboxamide (7b), Yellow solid (99,30%). 0.076 g, Yield: 79%, mp: 221–223 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.05 (s, 1H), 10.01 (s, 1H), 8.79 (d, $J = 6.0 \,\text{Hz}$, 2H), 8.70 (d, J = 5.1 Hz, 1H), 8.50 (s, 1H), 8.15 (d, J = 6.0 Hz, 2H), 7.91 (dd, J = 7.7, 1.5 Hz, 1H), 7.75 (s, 1H), 7.58 (d, J = 5.1 Hz, 1H), 7.51 (d, J = 7.7 Hz, 1H), 7.40-7.45 (J = 7.7 Hz, 1H), 6.36 (m, 1H), 3.93 (m, 1H), 1.78-1.85(m, 2H), 1.58-1.67 (m, 2H), 1.47-1.55 (m, 2H), 1.35-1.43 (m, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 167.2, 161.8, 160.6, 160.5, 158.4, 151.0, 144.2 (2C), 140.9, 133.9, 129.0, 122.5, 121.4 (2C), 120.8, 119.0, 109.3, 51.6, 33.1 (2C), 23.7 (2C); HRMS (ESI, m/z): calcd for $C_{22}H_{24}N_7O_2^+$ [M+H]⁺ 418.1986, found 418.1983, calcd for $C_{22}H_{13}N_7NaO^+$ [M + Na]⁺ 440.1805, found 440.1803.

4.1.6.3. *N-(tert-butyl)-2-(3-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)*benzoyl) hydrazine-1-carboxamide (7c). White solid (97.12%), 0.079 g, yield: 81%, mp: 206–207 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.04 (br s 1H), 9.94 (s, 1H), 9.38 (d, J = 1.6 Hz, 1H), 8.74 (dd, J = 4.8, 1.8 Hz, 1H), 8.64 (d, J = 5.1 Hz, 1H), 8.57 (dt, J = 7.9, 1.8 Hz, 1H), 8.45 (s, 1H), 7.93 (dd, J = 7.9, 1.8 Hz, 1H), 7.66 (s, 1H), 7.59 (dd, J = 7.9, 4.8 Hz, 1H), 7.56 (d, J = 5.1 Hz, 1H), 7.49 (d, J = 7.7 Hz, 1H), 7.40–7.43 (t, J = 7.7 Hz 1H), 6.08 (br s, 1H), 1.28 (s, 9H); ¹³C NMR (150 MHz, DMSO-d₆): δ 166.9, 162.1, 160.6, 159.9, 157.7, 152.0, 148.7, 141.0, 135.0, 133.8, 132.6, 128.9, 124.4, 122.4, 120.6, 119.0, 108.9, 49.9, 29.6 (3 C); HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ $[M+H]^+$ 406.1986, found 406.1982, calcd for $C_{21}H_{23}N_7NaO_2^ [M + Na]^+$ 428.1805, found 428.1801.

4.1.6.4. N-butyl-2-(3-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide (7d). Yellow solid (99.36%), 0.066 g, Yield: 79%, mp: 196–198 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.05 (s, 1H), 9.94 (s, 1H), 9.38 (d, J = 1.9 Hz, 1H), 8.74 (dd, J = 4.7, 1.4 Hz, 1H), 8.64 (d, J = 5.1 Hz, 1H), 8.57 (dt, J = 8.0, 1.9 Hz, 1H), 8.45 (s, 1H), 7.94 (dd, J = 8.0, 1.4 Hz, 1H), 7.82 (s, 1H), 7.59 (dd, J = 8.0, 4.7 Hz, 1H), 7.56 (d, J = 5.1 Hz, 1H), 7.52 (d, J = 7.7 Hz, 1H), 7.42 (t, J = 7.7 Hz, 1H), 6.42 (t, J = 7.7 Hz, 1H), 3.04 (q, J = 6.7 Hz, 2H), 1.39 (quin, J = 7.3 Hz, 2H), 1.28 (sxt, J = 7.3 Hz, 2H), 0.88 (t, J = 7.3 Hz, 3H); 13 C NMR (150 MHz, DMSO-d₆): δ 167.1, 162.1, 160.6, 159.9, 158.8, 152.0, 148.7, 141.0, 135.0, 133.9, 132.6, 128.8, 124.4, 122.5, 120.8, 119.1, 108.9, 39.4, 32.5, 19.9, 14.2; HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ [M+H]⁺ 406.1986, found 406.1980, calcd for $C_{21}H_{23}N_7NaO_2^+$ [M + Na]⁺ 428.1805, found 428.1802.

4.1.6.5. N-(2,4-dimethoxyphenyl)-2-(3-((4-(pyridin-3-yl)pyrimidin-2yl)amino) benzoyl)hydrazine-1-carboxamide (7e). White solid

(94.02%), 0.032 g, yield: 55%, mp: 231–232 °C. ¹H NMR (600 MHz, DMSO- d_6): δ 10.28 (br s, 1H), 9.97 (s, 1H), 9.38 (d, J = 1.8 Hz, 1H), 8.73 (dd, J = 4.7, 1.4 Hz, 1H), 8.64 (d, J = 5.1 Hz, 1H), 8.61 (s, 1H), 8.57 (dt, J = 8.0, 1.8 Hz, 1H), 8.48 (s, 1H), 8.03 (br s, 1H), 7.95 (dd, J = 8.0, 1.8 Hz, 1H), 8.48 (s, 1H), 8.03 (br s, 1H), 7.95 (dd, J = 8.0, 1.8 Hz, 1H), 8.48 (s, 1H), 8.03 (br s, 1H), 7.95 (dd, J = 8.0, 1.8 Hz, 1H), 8.48 (s, 1H), 8.03 (br s, 1H), 7.95 (dd, J = 8.0, 1.8 Hz, 1H), 8.48 (s, 1H), 8.03 (br s, 1H), 7.95 (dd, J = 8.0, 1.8 Hz, 1H), 8.03 (br s, 1H)1.4 Hz, 1H), 7.88 (d, J = 8.8 Hz, 1H), 7.57–7.59 (m, 1H), 7.55–7.57 (m, 1H), 7.49-7.52 (m, 1H), 7.43-7.47 (m, 1H), 6.62 (d, J=2.5 Hz, 1H), 6.48 (dd, J = 8.8, 2.5 Hz, 1H), 3.84 (s, 3H), 3.73 (s, 3H); ¹³C NMR (150 MHz, DMSO- d_6): δ 167.3, 162.1, 160.6, 160.0, 156.0, 155.4, 152.0, 149.6, 148.7, 141.1, 135.0, 133.7, 132.6, 129.0, 124.4, 122.6, 122.2, 120.6, 120.0, 118.9, 109.0, 104.6, 99.2, 56.3, 55.7; HRMS (ESI, m/z): calcd for $C_{25}H_{24}N_7O_4^+$ $[M+H]^+$ 486.1884, found 486.1883, calcd for $C_{25}H_{23}N_7NaO_4^+$ [M + Na]⁺ 508.1704, found 508.1703.

4.1.6.6. N-butyl-2-(3-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carboxamide (7f). Yellow solid (98.48%), 0.043 g, yield: 80%, mp: 183–184°C. 1 H NMR (600 MHz, DMSO-d₆): δ 10.05 (s, 1H), 9.94 (s, 1H), 8.74–8.77 (m, 1H), 8.68 (d, J = 4.9 Hz, 1H), 8.53 (s, 1H), 8.51 (d, J = 7.9 Hz, 1H), 8.04 (td, J = 7.9, 1.7 Hz, 1H), 7.92 (dd, J = 7.9, 1.7 Hz, 1H), 7.83 (s, 1H), 7.77 (d, J = 5.1 Hz, 1H), 7.58 (m, 1H), 7.52 (d, $J = 7.9 \,\text{Hz}$, 1H), 7.42 (t, $J = 7.9 \,\text{Hz}$, 1H), 6.43 (t, J = 5.1 Hz, 1H), 3.05 (q, J = 6.8 Hz, 2H), 1.39 (quin, J = 7.3 Hz, 2H), 1.29 (sxt, J = 7.3 Hz, 2H), 0.88 (t, J = 7.3 Hz, 3H); ¹³C NMR (150 MHz, DMSO-d₆): δ 167.2, 163.3, 160.5, 160.1, 158.8, 154.0, 150.1, 141.1, 138.1, 133.9, 128.9, 126.2, 122.4, 121.8, 120.7, 119.0, 108.7, 39.4, 32.5, 19.9, 14.2; HRMS (ESI, m/z): calcd for $C_{21}H_{24}N_7O_2^+$ [M + H]⁺ 406.1986, found 406.1980, calcd for $C_{21}H_{23}N_7NaO_2^+$ [M+Na]⁺ 428.1805, found 428.1802.

4.1.6.7. N-benzyl-2-(3-((4-(pyridin-4-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (7A). Yellow solid (98.66%), 0.042 g, yield: 53%, mp: 218-220 °C. ¹H NMR (600 MHz, DMSO- d₆): δ 10.40 (s, 1H), 10.01 (s, 1H), 9.51 (s, 1H), 8.79 (d, $J = 6.0 \,\mathrm{Hz}$, 2H), 8.69 (d, J = 5.1 Hz, 2H), 8.54 (br s, 1H), 8.15 (d, J = 6.0 Hz, 2H), 7.94 (dd, J = 1.2, 8.0 Hz, 1H), 7.57–7.61 (m, 2H), 7.44 (t, J = 7.8 Hz, 1H), 7.31-7.34 (m, 2H), 7.27- 7.31 (m, 2H), 7.19-7.23 (m, 1H), 4.76 (d, J = 5.8 Hz, 2H); ¹³C NMR (150 MHz, DMSO- d₆): δ 166.8, 161.8, 160.6, 160.5, 151.0, 144.2 (2C), 140.9, 139.9, 133.5, 128.8 (2C), 128.5, 127.5, 127.0 (2C), 122.8, 121.4, 121.3 (2C), 119.4, 109.3, 47.2; HRMS (ESI, m/ z): calcd for $C_{24}H_{22}N_7OS^+$ [M+H]⁺ 456.1601, found 456.1599, calcd for $C_{24}H_{21}N_7NaOS^+$ [M + Na]⁺ 478.1421, found 478.1418.

N-benzyl-2-(3-((4-(pyridin-3-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (7B). White solid (96.84%), 0.055 g, yield: 58%; mp: 212-213 °C . ¹H NMR (600 MHz, DMSO d_6): δ 10.38 (s, 1H), 9.95 (s, 1H), 9.49 (s, 1H), 9.38 (d, J = 1.8 Hz, 1H), 8.74 (dd, J = 4.7, 1.6 Hz, 1H), 8.66 (br s, 1H), 8.63 (d, J = 5.1 Hz, 1H), 8.57 (dt, J = 8.0, 1.8 Hz, 1H), 8.50 (s, 1H), 7.95 (dd, J = 8.0, 1.6 Hz, 1H), 7.57–7.60 (m, 2H), 7.56 (d, J = 5.1 Hz, 1H), 7.43 (t, J = 8.0 Hz, 1H), 7.27–7.34 (m, 4H), 7.19–7.23 (m, 1H), 4.75 (d, J = 5.8 Hz, 2H); ¹³C NMR (150 MHz, DMSO- d_6): δ 166.7, 162.1, 160.6, 160.0, 152.1, 148.7, 141.0, 139.9, 135.0, 133.5, 132.5, 128.7, 128.5, 127.5 (2C), 127.0 (2C), 124.4, 122.7, 121.1, 119.3, 108.9, 47.2; HRMS (ESI, m/z): calcd for $C_{24}H_{22}N_7OS^+$ [M+H]+456.1601, found 456.1599, calcd for $C_{24}H_{21}N_7NaOS^+[M+Na]^+$ 478.1421, found 478.1418.

4.1.6.9. N-phenyl-2-(3-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (7C). Yellow solid (97.88%), 0.043 g, Yield: 59%, mp: 202–203 °C. ¹H NMR (600 MHz, DMSO-d₆): δ 10.51 (s, 1H), 9.93–10.00 (s, 1H), 9.83 (br s, 1H), 9.75 (br s, 1H), 8.75 (m, 1H), 8.68 (d, $J = 5.0 \,\text{Hz}$, 1H), 8.60 (br s, 1H), 8.51 (d, J = 7.8 Hz, 1H), 8.00 (t, J = 7.8 Hz, 1H), 7.94 (d, J = 8.0 Hz, 1H), 7.77 (d, $J = 5.0 \,\text{Hz}$, 1H), 7.62 (d, $J = 6.9 \,\text{Hz}$, 1H), 7.56 (dd, J = 6.9, 5.0 Hz,

1H), 7.42–7.50 (m, 3H), 7.34 (t, J=7.3 Hz, 2H), 7.14–7.19 (t, J=7.3 Hz, 1H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.7, 163.3, 160.5, 160.2, 154.0, 150.1, 141.1, 139.8, 138.1, 133.6, 128.8, 128.4 (2C), 126.2, 125.5 (2C), 122.6, 121.8, 121.2, 119.2, 108.7; HRMS (ESI, m/z): calcd for C₂₃H₂₉N₇OS⁺ [M+H]⁺ 442.1445, found 442.1445, calcd for C₂₃H₁₉N₇NaOS⁺ [M+Na]⁺ 464.1264, found 464.1261.

4.1.6.10. N-benzyl-2-(3-((4-(pyridin-2-yl)pyrimidin-2-yl)amino)benzoyl)hydrazine-1-carbothioamide (7D). Yellow solid (98.95%), 0.052 g, yield: 56%, mp: 203–205 °C . 1 H NMR (600 MHz, DMSO-d₆): δ 10.40 (s, 1H), 9.95 (s, 1H), 9.50 (s, 1H), 8.75 (d, J = 4.5 Hz, 1H), 8.68 (t, J = 5.1 Hz, 1H), 8.67 (d, J = 5.1 Hz, 1H), 8.59 (br s, 1H), 8.52 (d, J = 7.8 Hz, 1H), 8.00 (dt, J = 8.0, 1.6 Hz, 1H), 7.93 (dd, J = 8.0, 1.6 Hz, 1H), 7.97 (d, J = 5.1 Hz, 1H), 7.56–7.60 (m, 2H), 7.44 (t, J = 7.8 Hz, 1H), 7.32 (t, J = 7.3 Hz, 2H), 7.29 (t, J = 7.3 Hz, 2H), 7.22 (t, J = 7.3 Hz, 1H), 4.76 (d, J = 5.8 Hz, 2H); 13 C NMR (150 MHz, DMSO-d₆): δ 166.8, 163.3, 160.4, 160.2, 154.0, 150.1, 141.1, 139.9, 138.1, 133.5, 128.8, 128.5 (2C), 127.5, 127.0 (3 C), 126.3, 122.6, 121.8, 121.1, 119.2, 108.7, 47.2; HRMS (ESI, m/z): calcd for $C_{24}H_{22}N_7OS^+$ [M + H] $^+$ 456.1601, found 456.1599, calcd for $C_{24}H_{21}N_7NaOS^+$ [M + Na] $^+$ 478.1421, found 478.1418.

4.2. Bioassay

All the human cancer cell lines were purchased from the American type culture collection (ATCC, Shanghai, China). Goat antirabbit and anti-mouse secondary antibody conjugated to horseradish peroxidase (HRP) was obtained from Thermo Fisher Scientific (Waltham, MA), anti-rabbit IgG conjugated with Cy3 from Chemicon International (Temecula, CA), anti-RXR α (D-20, sc-553) from Santa Cruz Biotechnology (Santa Cruz, CA), anti-PARP (46D11, 9542) and anti-Clevage Caspase 3 (Asp175, 9661S) from Cell Signalling Technology (Boston, MA), anti- α -tublin antibody from Proteintech (Rosemont, IL), and JC-1 Probe (Cat. T3168) from Thermo Fisher Scientific.

4.2.1. Cell culture and transfection

SW620, LS174T, HepG2, and A549 were maintained in Dulbecco's Modified Eagles Medium supplemented with 10% foetal bovine serum (FBS), HeLa was maintained in Modified Eagles Medium supplemented with 10% FBS and were cultured at $37\,^{\circ}$ C, under 90% humidity with 5% carbon-di-oxide (CO₂).

4.2.2. Antiproliferative activity assay

The logarithmic phase of HepG2 and A549 cells were seeded in 96-well plates at about 1×10^4 cells per well. treated with various concentrations of compounds for 24 h. then 15 μL MTT was added and incubated for 4 h at 37°C and 5% CO2. the light intensity was measured by microplate reader (PerkinElmer, Waltham, MA) at a wavelength of 490 nm. Experiments were repeated three times. The results were analysed by GraphPad Prism 5.01 software, La Jolla, CA.

4.2.3. Luciferase reporter gene assay

HEK293T cells were cotransfected with pG5-luc reporter (50.0 ng) and Pbind-RXR α -LBD (50.0 ng). One day after transfection, cells were treated with DMSO, compounds (1 μ M) or 9-cis-RA (0.1 μ M). After 12 h, cells were lysed and quantitated using the Dual-Luciferase Reporter Assay System (Thermo Fisher Scientific). Transfection and expression efficiency were normalised to Renilla luciferase activity.

4.2.4. RXRE-mediated transcription assay

HEK293T cells were seeded into 48-well plates at 40% confluence and cotransfected with RXRE luciferase reporter gene (100.0 ng), Myc-RXR α (50.0 ng), and Renilla control plasmid (1.0 ng). After 24 h post transfection, the compounds or vehicle were added. Cells were harvested after 12 h of incubation with ligands and then quantitated using the Dual-Luciferase Reporter Assay System (Promega). Luminescence of firefly luciferase values was normalised to the Renilla luciferase activity.

4.2.5. EC₅₀ determination

The EC $_{50}$ values were determined from full dose-response curves for compounds tested ranging from 0.10 to 20.0 μ M. HEK293T cells were plated, transfected, and treated as above using Luciferase Reporter Gene Assay. Then cells were treated with DMSO vehicle (0.1%) or compounds tested (final concentration of 0.1, 0.3125, 0.625, 1.25, 2.5, 5.0, 10.0, and 20.0 μ M) for 24 h. The amount of rexinoid activity at each concentration was measured using the same luciferase assay described above and EC $_{50}$ values were derived from dose-response curves of ligand concentration versus normalised luciferase activity using Graph Pad Prism 5 software.

4.2.6. Western blot analysis

Equal amounts of cell lysates were boiled in sodium dodecyl sulphate (SDS) sample loading buffer, then separated through 10% SDS-polyacrylamide gel electrophoresis and electro transferred to nitrocellulose membranes. The membranes were blocked with 5% non-fat milk in 10 mM TriseHCl (pH 8.0), 150 mM NaCl, 0.05% Tween 20 (TBST) buffer for 1 h at room temperature. The membranes were incubated overnight at 4 °C, with specific primary antibodies, after washing, followed by incubated with HRP-conjugated anti-mouse or anti-rabbit secondary antibodies, immunoreactive products were visualised with a chemiluminescence solution.

4.2.7. Generation of RXR[∞] knock-out cell by CRISPR/Cas9 system

For RXR α genes, sgRNAs were designed based on the original Cas9 approach. Then RXR α sgRNA were colonised into gRNA cloning vector PX330 (add gene 71707) and confirmed by sequencing. The cells were infected with control vector and gRNA expression vectors for 24h and selected single colonies using G418 (0.5 mg/mL). The knockout cell was determined using western blot analysis

4.2.8. SPR

BIAcore-T200 machine (GE Healthcare) was used to measure the binding kinetics of compound **6A** with RXR α -LBD, ER α -LBD, PPAR γ -LBD, and TR3-LBD. Purified RXR α -LBD, ER α -LBD, PPAR γ -LBD, and TR3-LBD were respectively immobilised on the CM5 chips according to the manufacturer's instructions. Then various concentrations of **6A** was injected into the flow cells in running buffer (phosphate buffer saline (PBS), 0.4% DMSO) at a flow rate of 30.0 μ L/min, when all cycle were collected, The results were analysed using Biacore T200 Evaluation Software.

4.2.9. Analysis of mitochondrial membrane potential

For detection of apoptosis by FACS, A549 cells were incubated with **6A** at $3.0\,\mu\text{M}$ for 12 h. After treatment, the cells were harvested and washed with PBS followed by stained with JC-1 for 30 min at room temperature in the dark, Flow-cytometric analysis was performed using FACS.



4.2.10. Immunofluorescence microscopy

A549 cells were seeded on glass cover slips, incubated for 12 h in the presence of compound **6A** at a final concentration of $3.0 \,\mu\text{M}$. Cells mounted on glass slides were fixed in 4% paraformaldehyde and permeabilised with PBS containing 0.1% Triton X-100 and 0.10 mol/l glycine for 10 min, and then blocked with 1% bovine serum albumin in PBS for 30 min at room temperature, followed with incubation with cleaved-caspase 3 antibodies (1:100) at room temperature for 2h, and detected by anti-rabbit IgG conjugated with Cv3 (1:200) at room temperature for 2 h. Cells were contained with 4'6'-diamidino-2-phenylindole (DAPI) (1:10,000) to visualise nuclei. The images were analysed by using MRC-1024 MP laserscanning confocal microscope (Leica).

4.3. Molecular docking studies

The molecular docking studies were performed with the Schrodinger 2015.1 Glide 26 . The crystal structure of RXRlpha (PDB code: 3A9E) were downloaded from the RCSB protein data bank and prepared with Protein Preparation Wizard panel as implemented in Maestro 10.5: water molecules were removed, missing hydrogens and residues were added. The glide grid centre was setting according the geometrical centre of original inhibitor (LG100754) and the grid size was $20 \times 20 \times 20 \text{ Å}^3$. Compound **6A** were prepared with LigPrep panel with OPLS2005 force field, generating the protonation and tautomeric states at $7 \pm 2 \,\mathrm{pH}$ units²⁷. Compound **6A** was docked into the RXR α binding site using Glide in XP mode. All of docking results were processed maestro 10.1 and PyMOL software.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by the National Natural Science Foundation of China [No. 81773600], the Natural Science Foundation of Fujian Province of China [No. 2018J01132], and the Fundamental Research Funds for the Central Universities [No. 20720180051].

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