This document is served as the manual to build a new version of FI network

**Set up Eclipse and Load FINetworkBuild Project**

The project used to build the FI network is programmed in Java. Eclipse has been used for all Java related programming. For this project, we have not built a project that can run independent of Eclipse. So you need to download Eclipse from http://www.eclipse.org , set up it and download the FINetworkBuild project from the CVS repository at reactomedev.oicr.on.ca.

After you connect to the CVS repository, find the FINetworkBuild module, and use “Check Out Project As…” popup menu to create a Java project.

You can find an updated document in the doc folder after you check out the CVS module. Use the updated document.

**Notes:**

1. The configuration file in the resources folder is used to configure files and some parameters used by the project. Make sure these two values in the file are correct: RESULT\_DIR, and DATA\_SET\_DIR.
2. RESULT\_DIR is used to hold results generated from the project.
3. DATA\_SET\_DIR is used to hold the download files and other source files for the project.

**Overview of Workflow**

The workflow to build a functional interaction (FI) network as described in Figure 1. Some changes may be made version by version. So the following Figure may be updated periodically.

Macintosh HD:Users:gwu:Documents:EclipseWorkspace:caBigR3:doc:FIConstructionWorkFlow.pdf

*Figure 1. Overall workflow to build the Reactome FI network.*

There are two types of data sources are used: Data sources for predicted FIs and data sources for annotated FIs. The data sources for predicted FIs should be downloaded from protein-protein interaction databases: IntAct, HPRD, and BioGrid, pFam (for domain interactions), and from various journals (see below for details). The data sources for annotated FIs are several human curated pathway databases.

**Data Sources for id mapping**

**UniProt**

UniProt is used as the protein reference database in the FI network construction. We need to download the latest version of UniProt so other data sets can be mapped to it. The download site for UniProt is: [ftp.uniprot.org](ftp://ftp.uniprot.org). You don’t need any registration. We only need to download the human data set.

1. After log into [ftp.uniprot.org](ftp://ftp.uniprot.org), download two files from knowledgebase/taxonomic\_divisions/: uniprot\_sprot\_human.data.gz and uniprot\_trembl\_huma.dat.gz. You may also download reldate.txt or README files for future reference. After download these files, unzip them, and place them into a sub-directory in the dataset folder. Change the configuration value in the configuration file for UNIPROT\_DIR to point to this uniprot directory.

# Used to control the UniProt version

UNIPROT\_DIR=${DATA\_SET\_DIR}/UniProt/release\_2011\_12/

1. Besides these files in step 1, you also need to download the fasta file for Isoform sequences to be used in normalizing features for NBC training and predictions from this web site: <http://www.uniprot.org/downloads>

**IPRO\_CLASS**

A mapping file is needed to map entrez gene id to UniProt accession number. We need to download an original mapping file from the PIR web site, and do some pre-process using a Java method.

1. Download the mapping file from this ftp site: [ftp.pir.georgetown.edu/databases/](ftp://ftp.pir.georgetown.edu/databases/). Go to idmapping/mapping\_by\_sp, and download file h\_sapiens.tb. Remember where you place your file.
2. Modify the following two values in the configuration file:

# This file is used to map Entrez id to UniProt accession number

IPROCLASS\_HUMAN\_FILE = ${DATA\_SET\_DIR}/iproclass/011612/h\_sapiens.tb

ENTREZ\_TO\_UNIPROT\_MAP\_FILE\_NAME=${DATA\_SET\_DIR}/iproclass/011612/EntrezToUniProt.txt

**Data Sources for Annotated FIs**

Annotated FIs are FIs extracted from human curated pathways. Multiple pathway databases have been used. Pathways in other non-Reactome databases are converted to curator tool project first, and then dumped into a modified gk\_central Reactome databases. Different file formats are used for different databases.

**Reactome:**

Reactome is used as the foundation for the FI network build. Public released Reactome database have augmented predicted objects from other non-human species. To minimize the effects of these predicted non-human objects, we use a snapshot of gk\_central, though many non-human pathways have been dumped into gk\_central now (e.g. many objects from plant Cyc).

1. Generate a snapshot for gk\_central @reactomedev.oicr.on.ca: Log into reactomedev, and generate a gk\_central dump by using the following command:

*mysqldump –u{user\_name} –p gk\_central >gk\_central\_{mmddyyyy}.sql*

You will be asked to enter your mysql password. Replace text in {} by correct values.

If you want to do build at your local computer, zip the generate sql dump as following:

*jar –Mcvf gk\_central\_{mmddyyyy}.sql.zip gk\_central\_{mmddyyyy}*

1. Create a snapshot of gk\_central: Copy the above zipped dump file into your build computer (e.g. your desktop machine), unzip it using the following command:

*jar –xvf gk\_central\_{mmddyyyy}.sql.zip*

Log into your mysql database using a user account that can create a database. Create a database as following:

*mysql> create database reactome\_{release\_number}\_plus\_i;*

Here release\_number should be replaced by actual release number (e.g. 39). The release number should be the number of closest public release. For example, we build a FI network in January, 2012. The closest Reactome public release is 39, which was released in Decemeber, 2011. Note: Reactome has a quarterly release schedule.

After the empty database is created, run the following two commands to load the generated gk\_central snapshot mysqldump:

*mysql> use reactome\_{release\_number}\_plus\_i;*

*mysql> source gk\_central\_{mmddyyyy}.sql*

Note: In order to make sure mysql can find the dump file, start mysql from the directory containing the unzipped mysqldump file. Or you have to use absolute path to your dump file (no quotation marks)

1. Modify Reactome database schema: the regular Reactome database schema needs to be expanded for the FI network. A new attribute called “dataSource” should be added to the top-level class, “DatabaseObject”. A new class called “Interaction” should be created. Interaction is a concrete sub-class to Event. (TO\_DO: Modify the final database schema for Perl API: need a new Protégé project)

*mysql> source {absolute\_path\_to /SchemaModification.sql*

Note: You should find a copy of SchemaModifcation.sql in resources folder in project caBigR3 in CVS.

After a successful run of the above sql, check if a new Interaction class, and new attribute dataSource for DatabaseObject have been created by connecting your curator tool to your newly created database. Check the database schema view to make sure these two things have been there.

In order to use this new expanded Reactome schema for future procedures, we need to export the new schema from the database into a local computer. In the curator tool, after connecting to the modified database, do the following:

In the database schema view, choose “Export Schema” in the File menu, and save the schema into the resource folder in your caBigR3 project folder or any working folder that contains the resource folder. This schema file will be needed by the Reactome Java API, which is used extensively in the following procedures.

Note: the file name has to be “schema” without any extension. Don’t change it!

1. Make changes to the configuration file in the source folder for these three properties: REACTOME\_SOURCE\_DB\_NAME, DB\_USER and DB\_PWD, so that the expanded Reactome database can be used in the future as follows:

# local database that is used as the data source

REACTOME\_SOURCE\_DB\_NAME=reactome\_40\_plus\_i

DB\_USER=root

DB\_PWD=macmysql01

**KEGG**

KEGG has many good pathway diagrams and many disease pathways. We import KEGG pathways using its KGML export. The KEGG KGML is downloaded from KEGG ftp folder. You need to have a license first in order to access its ftp site.

1. Download the following files from the KEGG ftp site: [ftp.bioinformatics.jp](ftp://ftp.bioinformatics.jp)
   1. kegg/xml/kgml/non-metabolic/organisms/hsa.tar.gz: we used human non-metabolic pathways only. Reactome has enough metabolic pathways.
   2. kegg/pathway/pathway.list: the whole list of pathways. From this list, we can choose what pathways we want to use. This file is optional.
   3. kegg/genes/links/genes\_uniprot.list.gz: a mapping file from kegg ids to UniProt ids. After unzipping, this file is around 140 Mb (January 12, 2012). You can use the following command to get a human only mapping file for easy view and quick processing (Mac or Linux only):

*grep hsa: genes\_uniprot.list > hsa\_genes\_uniprot.list*

The output file hsa\_genes\_uniprot.list is around 540 K.

* 1. Also the file map\_title.tab (where is it at the ftp site?), which will be used for generating gene to pathway mapping.

1. Specify values in the configuration file for the KEGG data set:

# These parameters are related to KEGG pathways

KEGG\_DIR=${DATA\_SET\_DIR}/KEGG/011112/

# Unzipped human KGML files should be in this directory

KEGG\_HSA\_KGML\_DIR=${KEGG\_DIR}/KGML/hsa/

# The converted KEGG pathways is saved in this project file

KEGG\_CONVERTED\_FILE=${KEGG\_DIR}/030112.rtpj

# This file is used to map KEGG gene ids to UniProt ids

KEGG\_ID\_TO\_UNIPROT\_MAP\_FILE=${KEGG\_DIR}/hsa\_genes\_uniprot.list

**NCI-PID**

There are two kinds of pathways available in this database: pathways curated by NCI-PID curators and pathways imported from BioCarta and Reactome. Indeed, we don’t need to import Reactome pathways here. BioCarta pathways have not been updated since they were imported in this database. But because the changes of the Reactome schema, these pathways should be re-imported each time if we want to use them.

1. Download pathways from NCI-PID: The data format we are going to use is BioPAX level 2. Download these two BP2 files in its download site, <http://pid.nci.nih.gov/download.shtml>: NCI-Nature Curated Data (BioPAX Level 2), and BioCarta data (BioPAX Level 2). After unzipping, the file names should be:
   1. NCI-Nature\_Curated.bp2.owl
   2. BioCarta.bp2.owl
2. Modified these the following configurations in the configuration file:

# Used for the Nature-PID database files

NATURE\_PID\_DIR=${DATA\_SET\_DIR}/NCI-Pathways/011612/

NATURE\_PID\_CURATED=${NATURE\_PID\_DIR}/NCI-Nature\_Curated.bp2.owl

NATURE\_PID\_CURATED\_CONVERTED=${NATURE\_PID\_DIR}/NCI-Nature\_Curated.bp2.rtpj

NATURE\_PID\_BIOCARTA=${NATURE\_PID\_DIR}/BioCarta.bp2.owl

NATURE\_PID\_BIOCARTA\_CONVERTED=${NATURE\_PID\_DIR}/BioCarta.bp2.rtpj

1. Find the BioPAX to Reactome mapping file: resources/BioPAXToReactomeMappers.xml. Make sure the following XML element is NOT commented out so that we can use NciPIDBToRPostProcessor class in package org.reactome.b2rPostProcessor during mapping:

<!-- This PostProcessor is for NCI Pathways -->

<postProcessor class=*"org.reactome.b2rPostProcessor.NciPIDBToRPostProcessor"* />

**Panther**

Pathways in the Panther database have not been updated since December, 2007, though the version number has been changed from 2.5 to 3.0.1. Basically the annotation of pathways in Panther has stopped for quite a while. However, because our Reactome schema has been evolved continually, we still need to re-convert pathways from Panther into the Curator Tool project in order to merge with other pathways.

1. Download Pathways: download pathways from Panther’s ftp site, [ftp.pantherdb.org/pathway/current\_release](ftp://ftp.pantherdb.org/pathway/current_release). Please download these two files: SBML\_{version}.zip and SequenceAssociationPathway3.0.1.txt. We use the SBML files, which are the file format used by the Panther annotation. The second file is used to map pathway component ids used in pathways into UniProt accession numbers.

Note: One pathway component used in panther pathways can be mapped to multiple UniProt identifiers because these components are actually based on HMM families constructed by Panther. In order to control the quality of imported pathways, we use mappings based on the following reliable confidence only:

"IGI",

"IPI",

"IDA",

"IEP",

"TAS",

"IC",

"IMP",

"RCA"

1. Modify the following values in the configuration file related to Panther:

# Used for the panther database files

# Download on Jan 18, 2011

PANTHER\_DIR=${DATA\_SET\_DIR}/Panther/Version3.0.1/

PANTHER\_FILES\_DIR=${PANTHER\_DIR}/SBML/

PANTHER\_MAPPING\_FILE=${PANTHER\_DIR}/SequenceAssociationPathway3.01.txt

PANTHER\_CONVERTED\_FILE=${PANTHER\_DIR}/Panther\_3\_0\_1.rtpj

*Note: Some part of Panther converted was developed by Andreas Hoelzlwimmer* [*andreash@ebi.ac.uk*](mailto:andreash@ebi.ac.uk) *during his internship at EBI. Needs to give him an acknowledgement in next FI related publication.*

Note: A file called “SpeciesToChEBIId.txt” in the resources folder has been used for converting. This file is old and has not been updated. This may be fine considering all Panther pathways have not been updated for quite a while.

1. For reference, here are some numbers in the converted Curator Tool project:

**CellMap**

Not needed any more!

**TRED**

TRED provides us transcription factors (TF) and their targets interactions. Many of interactions in this database are computationally predicted. We extract only manual curated interactions, which have been supported by literatures if not considering curation errors. The content in the TRED database should not be updated any more. What we need to do is just convert the data in the TRED database into a Curator tool project using the latest Reactome data model. To fetch the data from the TRED database, we use a customized hibernate based API. So have to make sure you have set up your class path correctly in order to use this hibernate API.

1. Install the TRED database into your local computer: If you are building the FI network at your local computer, you may need to install the TRED database for quick performance. If you don’t want to install it, you can access this database at reactomedev.oicr.on.ca. The database name is test\_TRED. To install this database, do a mysqldump from reactomedev, and load the mysqldump into your local mysql database.
2. Configure the hibernate configuration file: Find file TREDHibernate.cfg.xml in the resources folder. Make changes to these three properties so that they point to the correct values:

<property name=*"connection.url"*>jdbc:mysql://localhost:3306/TRED</property>

<property name=*"connection.username"*>root</property>

<property name=*"connection.password"*>macmysql01</property>

1. Make changes to these two values in the configuration file:

# For TRED files

TRED\_DIR=${DATA\_SET\_DIR}/TRED/

TRED\_CONVERTED\_FILE=${TRED\_DIR}/TRED\_030112.rtpj

**Data sources for predicted FIs**

**Ensembl-Compara**

Ensembl-compara is used to map PPIs from non-human species to human. We need to have a local compara database to do this mapping by downloading some files from the Ensembl-Compara database:

1. Download files: In this web page, <http://www.ensembl.org/info/data/ftp/index.html>, choose Comparative/MySQL to its ftp site. From the ftp site, choose ensemble\_compara\_xx (xx for release number, 66 as of February, 2012. However, for some reason, many yeast UniProt ids are not in this current release. So release 62 is used!). Download member.txt.gz, family.txt.gz, and family\_member.txt.gz. Also download the database schema file: ensembl\_compara\_62.sql.gz. Unzip all files.
2. Create ensemble\_compara database: log into a local mysql database, create a ensembl\_compara database by using, “create database ensembl\_compara\_xx” (xx should be the release number for compara). After the database is created, load the schema by using ensembl\_compara\_xx.sql with command: source ensembl\_compara\_62.sql, assuming you start mysql from the directory containing this sql file. Otherwise, provide the absolute path to it.
3. Load data into the database: log out from the mysql database, and run the following command:

mysqlimport –u{mysql\_db\_user} -p --local ensembl\_compara\_62 family.txt family\_member.txt member.txt

Note: make sure you use correct mysql user name and password

1. Make changes to the following values in the configuration file:

# For ensembl related files

ENSEMBL\_DIR=${DATA\_SET\_DIR}/Ensembl/release\_62/

ENSEMBL\_COMPARA\_DATABASE=ensembl\_compara\_62

ENSEMBL\_PROTEIN\_FAMILIES=${ENSEMBL\_DIR}/ProteinFamilies.txt

1. Note: we may also connect to the ensembl public mysql databases. However, it is much faster to load the above tables into a local mysql database.

**Protein-Protein Interactions (PPIs)**

Protein protein interactions are downloaded from iRefIndex, a comprehensive protein-protein interaction databases by grouping many popular protein interaction databases together, including IntAct, BioGrid, HPRD, BIND, and others, and creating a non-redundant interaction database using customized hash keys. One problem using this database is that we need to make sure this data source is updated regularly. As of now (February, 2012, release 9.0), it is updated pretty regularly.

1. Download interaction files: the PPIs in iRefIndex are provided in PSIMI-TAB format. The following files (these files names are for release 9.0. Names may have different date in future releases) should be downloaded from its ftp site via this web page: <http://irefindex.uio.no/wiki/README_MITAB2.6_for_iRefIndex>, and unzip them.

4932.mitab.10182011.txt.zip (baker’s yeast): this file is not used!

6239.mitab.10182011.txt.zip (worm)

7227.mitab.10182011.txt.zip (fly)

9606.mitab.10182011.txt.zip (human)

559292.mitab.10182011.txt.zip (yeast S288c)

10090.mitab.10182011.txt.zip (mouse)

1. Based on the above data files, make changes to the configuration data file:

# Constants for iRefIndex data files

IREFINDEX\_DIR=${DATA\_SET\_DIR}/iRefIndex/9.0/

IREFINDEX\_HUMAN\_FILE=${IREFINDEX\_DIR}/9606.mitab.10182011.txt

IREFINDEX\_HUMAN\_PPI\_FILE=${IREFINDEX\_DIR}/HumanPPIsInUniProt022712.txt

IREFINDEX\_YEAST\_FILE=${IREFINDEX\_DIR}/559292.mitab.10182011.txt

IREFINDEX\_YEAST\_PPI\_FILE=${IREFINDEX\_DIR}/YeastPPIsInUniProt022812.txt

IREFINDEX\_YEAST\_TO\_HUMAN\_PPI\_FILE=${IREFINDEX\_DIR}/HumanPPIsFromYeastInUniProt030112.txt

IREFINDEX\_FLY\_FILE=${IREFINDEX\_DIR}/7227.mitab.10182011.txt

IREFINDEX\_FLY\_PPI\_FILE=${IREFINDEX\_DIR}/FLyPPIsInUniProt022812.txt

IREFINDEX\_FLY\_TO\_HUMAN\_PPI\_FILE=${IREFINDEX\_DIR}/HumanPPIsFromFLyInUniProt030112.txt

IREFINDEX\_WORM\_FILE=${IREFINDEX\_DIR}/6239.mitab.10182011.txt

IREFINDEX\_WORM\_PPI\_FILE=${IREFINDEX\_DIR}/WormPPIsInUniProt022812.txt

IREFINDEX\_WORM\_TO\_HUMAN\_PPI\_FILE=${IREFINDEX\_DIR}/HumanPPIsFromWormInUniProt030112.txt

IREFINDEX\_MOUSE\_FILE=${IREFINDEX\_DIR}/10090.mitab.10182011.txt

IREFINDEX\_MOUSE\_PPI\_FILE=${IREFINDEX\_DIR}/MousePPIsInUniProt031412.txt

IREFINDEX\_MOUSE\_TO\_HUMAN\_PPI\_FILE=${IREFINDEX\_DIR}/HumanPPIsFromMouseInUniProt031412.txt

**Gene-Expression Correlations**

Two gene expression data sets have been used as NBC features: Lee’s Gene Expression and Prieto’s Gene Expression. The original data files were downloaded from their paper’s web sites, and should not be changed any more. However, we need to update protein-pairs to latest version of UniProt.

Note: The original downloaded data files contain gene pairs only. In order to be used as features in NBC, we have to map gene names to UniProt accession numbers. We use downloaded UniProt data file to do this mapping. We use the swissprot part of UniProt data for doing the mapping for the Prieto data file. For Lee’s data file, we use the original downloaded mapping file and normalize with the latest UniProt data.

1. Get the archived source files from CVS: synchronize data\_archive/data\_archive.zip. Unzip this zipped file. You should find two files: GeneExpWith3FromPavlidis.txt under the LeeGeneExp folder, and union60.txt in the PrietoGeneExp folder.
2. Make sure these two constants in the configuration file have been set correctly (usually you don’t need to make a change):

# Two Gene expression data files

LEE\_GENE\_EXP\_FILE\_SOURCE=${DATA\_SET\_DIR}/microarray/Pavlidis/GeneExpWith3FromPavlidis.txt;

LEE\_GENE\_EXP\_FILE=${RESULT\_DIR}/LeeGeneExp.txt

PRIETO\_GENE\_EXP\_FILE=${RESULT\_DIR}/PrietoGeneExp.txt

**Gene Ontology Annotation**

The sharing of GO annotation (Biological Process only) is used as another feature in our NBC training and prediction. In our usage, we check a simple sharing only without considering any parent-child relationships among GO terms.

1. Download GO annotation file for homo sapiens from the gene ontology’s web site: <http://www.geneontology.org/GO.downloads.annotations.shtml>.
2. Modify the following value in the configuration file to point to the correct directory that holds the above downloaded file:

# For GO related files

GO\_DIR=${DATA\_SET\_DIR}/GO/030712/

**Domain-Domain Interactions**

We use domain-domain interactions from the pFam database.

1. Generate UniProt accession to pFam domain ids mapping file: run method genreateUniToPfamMap() in class UniProtAnalyzer.
2. Download domain interaction file from pFam: go to pFam release ftp site, <ftp://ftp.sanger.ac.uk/pub/databases/Pfam>, go to the release folder, the latest release, and database\_files. Dowload two files: pfamA.txt.gz and pfamA\_interactions.txt.gz, unzip them.
3. Specify the following in the configuration file as being used in step 2:

# Directory for pFam

PFAM\_DIR\_NAME=${DATA\_SET\_DIR}/Pfam/26.0/

**Gene Ways Protein-Protein Interactions**

Not used any more because many PPIs are actually extracted from literatures manually!

**Functional Interaction Network Build**

Run the following methods in order in class org.reactome.fi.FINEtworkBuilder

1. **prepareMappingFile()**

Example logging from running this method like this:

2012-04-17 13:38:07,578 [main] INFO org.reactome.fi.FINetworkBuilder - Running UniProtAnalyzer.generateUniProtIDsMap()...

total entries in uniprot: 178240

total entries in uniprot: 79584

2012-04-17 13:38:13,368 [main] INFO org.reactome.fi.FINetworkBuilder - Running UniProtAnalyzer.generateEntrezGeneToUniProt()...

Counter: 70246

2012-04-17 13:38:14,364 [main] INFO org.reactome.fi.FINetworkBuilder - Running UniProtAnalyzer.generateUniToPfamMap()...

After this method running, four files should be generated as following:

Uni2Pfam.txt in the UniProt directory

SwissProtACIDMap.txt in the UniProt directory

ENTREZ\_TO\_UNIPROT\_MAP\_FILE\_NAME as configured

ENSEMBL\_PROTEIN\_FAMILIES as configured

1. **convertPathwayDBs()**

Convert Pathways in KEGG, NCIPID, and Panther, and TF/Target interactions into individual curator tool project files. You may need to open these converted project files in the curator tool to see how they look like and make sure they are correct.

Notes: 1). Make sure to assign enough memory for running this method (e.g. –Xmx1024m).

2). On the KEGG download in January, 2012, there are 26 KEGG ids cannot be mapped to UniProts. Check to ID mapping in UniProt, find three mappings. These unmapped KEGG ids are left without further processing since the number is very small. The unmapped ids are listed in file UnMappedKeggIds.txt (this file was created manually from the output in the Eclipse console).

3). In the version of FI network constructed in 2009, one KEGG id in pathway has been mapped to one UniProt only. This has been changed now. A KEGG id can be mapped to multiple UniProt via DefinedSet. So a DefinedSet converted from KEGG may contain another DefinedSet.

4). ReferenceIsoform instances may have been fetched out from the database during converting. However, our converting cannot take use of isoforms yet. So the Isoform instances should be treated as the top-level ReferenceGeneProduct instance!

5). We use KGML to import KEGG pathways. However, many complexes have not been described in KGML files. So interactions from these complexes cannot be extracted. For example, see complex HAP1-HTT-Dyneim-Dunactin in Huntington’s disease pathway: <http://www.genome.jp/kegg/pathway/hsa/hsa05016.html>. It seems there is no good file that can be used to extract these complexes. Probably a future import can use map files directly (e.g. hsadd05016.conf).

6). Because of the above problem, the pathway to protein/gene mapping file extracted for KEGG uses KGML files directly, instead of imported Reactome pathways (see below).

7). For converting the BioCarta pathways in the NCI-PID pathway, an old GO.terms\_and\_ids.txt in the resource folder has been used. This may be fine since the biocarta file is pretty old itself.

8). For converting pathways in the Panther database, a file called “SpeciesToChEBIId.txt” in the resources folder has been used for converting. This file is old and has not been updated. This may be fine considering all Panther pathways have not been updated for quite a while.

9). For TRED database, As of January, 2012, we use gene names, which are used in the TRED database, in our target Reactome database directly to map to UniProt. Some of gene names in TRED cannot be mapped. See below (partial list):

2012-01-19 14:56:22,763 [main] WARN org.reactome.convert.common.PostProcessTemplate - E2F-4 for EWAS -3 cannot be mapped to UniProt!

2012-01-19 14:56:22,900 [main] WARN org.reactome.convert.common.PostProcessTemplate - DNA Pol alpha for EWAS -42 cannot be mapped to UniProt!

2012-01-19 14:56:22,935 [main] WARN org.reactome.convert.common.PostProcessTemplate - RNAPII large subunit for EWAS -58 cannot be mapped to UniProt!

2012-01-19 14:56:22,947 [main] WARN org.reactome.convert.common.PostProcessTemplate - DNA topoisomerase I for EWAS -68 cannot be mapped to UniProt!

2012-01-19 14:56:22,991 [main] WARN org.reactome.convert.common.PostProcessTemplate - PCNA p120 for EWAS -97 cannot be mapped to UniProt!

2012-01-19 14:56:23,004 [main] WARN org.reactome.convert.common.PostProcessTemplate - p107 for EWAS -110 cannot be mapped to UniProt!

2012-01-19 14:56:23,026 [main] WARN org.reactome.convert.common.PostProcessTemplate - E2F-1 for EWAS -129 cannot be mapped to UniProt!

…………

Total unmapped names: 38

10). We may get a rough idea how many proteins can be merged into the original Reactome by output from the last section like the following (ids for UniProt accession numbers):

Total ids from Reactome: 7447 // This number may be bigger than the release one

Total ids from KEGG: 7897

Total ids after merging: 11525 (0.5163986323920476, 4078)

Total SwissProt ids after merging: 8534 (0.421473725800079)

Total ids from Nature-PID: 2573

Total ids after merging: 11984 (0.17839098328799066, 459)

Total SwissProt ids after merging: 8977 (0.44335242986961676)

Total ids from Biocarta-PID: 1526

Total ids after merging: 12785 (0.5249017038007864, 801)

Total SwissProt ids after merging: 9076 (0.4482418016594232)

Total ids from TRED: 1142

Total ids after merging: 12951 (0.14535901926444833, 166)

Total SwissProt ids after merging: 9242 (0.4564401422362703)

Total ids from Panther: 1424

Total ids after merging: 13120 (0.11867977528089887, 169)

Total SwissProt ids after merging: 9411 (0.4647866455946266)

1. **dumpPathwayDBs()**

Dump the converted curator tool project into the extended Reactome database created before, and generate FI files from these converted projects.

Notes: 1). You are encouraged to compare the numbers from different pathways to ones in the previous reactome\_plus\_i database to make sure there are no big surprise there. You can use the dataSource attribute to get a list of converted Event instances for a specific database (e.g. KEGG, Panther).

2). Some of UniProt identifiers that cannot be mapped to the original ReferenceGeneProducts from different databases may be duplicated in the database. Just leave them as is. But have to remember to merge them when counting how many UniProt accessions in the database. Avoid checking the numbers based on ReferenceGeneProduct instances only.

1. **dumpPathwayFIs()**

Note: Before running method, you have to make sure dataSourcesIds used in method ReactomeAnalyzer.getPathwayDbAnalyzer() are correct by assigning correct values in the following configuration properties. You may need to use the curator tool and connect it to the database containing pathways dumped from step 3 to get the correct DB\_IDs:

# The following values are DB\_IDs for imported non-Reactome pathway databases. These values vary in each build, and should be set by checking

# database for ReferenceDatabase instances after running method dumpPathwaysDBs()

PANTHER\_DB\_ID=191282

NCI\_NATURE\_CURATED\_DB\_ID=2185693

NCI\_NATURE\_BIOCARTA\_DB\_ID=2241489

KEGG\_DB\_ID=1385644

TRED\_DB\_ID=2279928

The dumped pathway FIs should have similar numbers or more as follows (March 2012):

Total interactions from reactions: 128727

Time for looping: 17968

Total interactions from Reactome: 164841

After filtering: 140430

Before sequence consolidating: 140414

After sequence consolidating: 140095

Done data source: Reactome

Total interactions from reactions: 14516

Time for looping: 4055

Total interactions from Reactome: 15400

After filtering: 15400

Before sequence consolidating: 15400

After sequence consolidating: 15400

Done data source: pantherdb

Total interactions from reactions: 15000

Time for looping: 50960

Total interactions from Reactome: 15797

Total interactions from Interaction Event: 0

After filtering: 15716

Before sequence consolidating: 15694

After sequence consolidating: 15694

Done data source: Pathway Interaction Database

Total interactions from reactions: 5701

Time for looping: 21150

Total interactions from Reactome: 6763

Total interactions from Interaction Event: 0

After filtering: 6763

Before sequence consolidating: 6763

After sequence consolidating: 6761

Done data source: BioCarta - Imported by PID

Total interactions from reactions: 0

Time for looping: 207

Total interactions from Reactome: 2159

Total interactions from Interaction Event: 57359

After filtering: 59079

Before sequence consolidating: 59079

After sequence consolidating: 48819

Done data source: Kegg

Total interactions from reactions: 0

Time for looping: 0

Total interactions from Reactome: 0

Total interactions from Interaction Event: 2858

After filtering: 2858

Before sequence consolidating: 2858

After sequence consolidating: 2858

Done data source: TRED

1. **prepareNBCFeatures()**

Output from the above running:

2012-04-17 14:47:23,100 [main] INFO org.reactome.fi.FINetworkBuilder - Running EnsemblAnalyzer.dumpProteinFamilies()...

Size of familyToProteins(): 44475

Total families before filtering: 44475

Total families after filtering: 11202

2012-04-17 14:47:26,823 [main] INFO org.reactome.fi.FINetworkBuilder - Running IRefIndexMITTabAnalyzer.loadHumanPPIs()...

Total PPIs: 112639

Total ids: 20722

in SwissProt: 12154

in UniProt: 14728

Total ids: 13864

in SwissProt: 11538

in UniProt: 13864

Total time: 7245

2012-04-17 14:47:34,068 [main] INFO org.reactome.fi.FINetworkBuilder - Running IRefIndexMITTabAnalyzer.loadFlyPPIs()...

Total PPIs: 38940

Total time: 2634

2012-04-17 14:47:36,703 [main] INFO org.reactome.fi.FINetworkBuilder - Running IRefIndexMITTabAnalyzer.loadWormPPIs()...

Total PPIs: 11626

Total time: 701

2012-04-17 14:47:37,404 [main] INFO org.reactome.fi.FINetworkBuilder - Running IRefIndexMITTabAnalyzer.loadYeastPPIs()...

Total PPIs: 214848

Total time: 9350

2012-04-17 14:47:46,755 [main] INFO org.reactome.fi.FINetworkBuilder - Running IRefIndexMITTabAnalyzer.loadMousePPIs()...

Total PPIs: 12787

Total time: 784

2012-04-17 14:47:47,539 [main] INFO org.reactome.fi.FINetworkBuilder - Running PsiMiOrthologyAnalyzer.generateHumanPPIsFromYeastInUniProt()...

Total yeast proteins: 1272

Mapped human PPIs from other species: 214848 -> 1368910

After filtering: 1368910

Before sequence consolidating: 1368910

After sequence consolidating: 1304755

Before normalization: 1368910, after: 1304755

2012-04-17 14:48:07,954 [main] INFO org.reactome.fi.FINetworkBuilder - Running PsiMiOrthologyAnalyzer.generateHumanPPIsFromWormInUniProt()...

Mapped human PPIs from other species: 11626 -> 129619

After filtering: 129619

Before sequence consolidating: 129619

After sequence consolidating: 122547

Before normalization: 129619, after: 122547

2012-04-17 14:48:21,018 [main] INFO org.reactome.fi.FINetworkBuilder - Running PsiMiOrthologyAnalyzer.generateHumanPPIsFromFlyInUniProt()...

Mapped human PPIs from other species: 38940 -> 394801

After filtering: 394801

Before sequence consolidating: 394801

After sequence consolidating: 376781

Before normalization: 394801, after: 376781

2012-04-17 14:48:35,018 [main] INFO org.reactome.fi.FINetworkBuilder - Running PsiMiOrthologyAnalyzer.generateHumanPPIsFromMouseInUniProt()...

Total mouse proteins: 41655

Mapped human PPIs from other species: 12787 -> 854318

After filtering: 854318

Before sequence consolidating: 854318

After sequence consolidating: 770385

Before normalization: 854318, after: 770385

2012-04-17 14:48:51,936 [main] INFO org.reactome.fi.FINetworkBuilder - Checking odds ratio...

File: datasets/iRefIndex/9.0//HumanPPIsInUniProt022712.txt

Total checked pairs: 95959

Total: 140095

Mapped to ppi: 8542 (0.060973)

Mapped to random: 141 (0.001006)

Mapped to random: 152 (0.001085)

Mapped to random: 154 (0.001099)

Mapped to random: 139 (0.000992)

Mapped to random: 128 (0.000914)

Mapped to random: 145 (0.001035)

Mapped to random: 160 (0.001142)

Mapped to random: 147 (0.001049)

Mapped to random: 135 (0.000964)

Mapped to random: 149 (0.001064)

Average odds ratio: 62.919799976046136 +- 4.227475795333164 (from 10 tests)

File: results/2012/HumanPPIsFromYeast030112.txt

Total checked pairs: 1304755

Total: 140095

Mapped to ppi: 2210 (0.015775)

Mapped to random: 206 (0.001470)

Mapped to random: 216 (0.001542)

Mapped to random: 200 (0.001428)

Mapped to random: 163 (0.001163)

Mapped to random: 178 (0.001271)

Mapped to random: 226 (0.001613)

Mapped to random: 167 (0.001192)

Mapped to random: 177 (0.001263)

Mapped to random: 172 (0.001228)

Mapped to random: 187 (0.001335)

Average odds ratio: 11.988223137514396 +- 1.326957494425449 (from 10 tests)

File: results/2012/HumanPPIsFromWorm030112.txt

Total checked pairs: 122547

Total: 140095

Mapped to ppi: 593 (0.004233)

Mapped to random: 33 (0.000236)

Mapped to random: 26 (0.000186)

Mapped to random: 30 (0.000214)

Mapped to random: 38 (0.000271)

Mapped to random: 26 (0.000186)

Mapped to random: 41 (0.000293)

Mapped to random: 27 (0.000193)

Mapped to random: 31 (0.000221)

Mapped to random: 31 (0.000221)

Mapped to random: 37 (0.000264)

Average odds ratio: 19.04322216358146 +- 3.001409423056792 (from 10 tests)

File: results/2012/HumanPPIsFromFly030112.txt

Total checked pairs: 376781

Total: 140095

Mapped to ppi: 1944 (0.013876)

Mapped to random: 65 (0.000464)

Mapped to random: 75 (0.000535)

Mapped to random: 75 (0.000535)

Mapped to random: 71 (0.000507)

Mapped to random: 75 (0.000535)

Mapped to random: 74 (0.000528)

Mapped to random: 70 (0.000500)

Mapped to random: 80 (0.000571)

Mapped to random: 71 (0.000507)

Mapped to random: 79 (0.000564)

Average odds ratio: 26.897110981818983 +- 1.6644923160996432 (from 10 tests)

File: results/2012/HumanPPIsFromMouse031412.txt

Total checked pairs: 770385

Total: 140095

Mapped to ppi: 4645 (0.033156)

Mapped to random: 125 (0.000892)

Mapped to random: 117 (0.000835)

Mapped to random: 166 (0.001185)

Mapped to random: 131 (0.000935)

Mapped to random: 141 (0.001006)

Mapped to random: 139 (0.000992)

Mapped to random: 129 (0.000921)

Mapped to random: 147 (0.001049)

Mapped to random: 142 (0.001014)

Mapped to random: 127 (0.000907)

Average odds ratio: 35.49923808754484 +- 3.4322625312095267 (from 10 tests)

2012-04-17 14:48:58,373 [main] INFO org.reactome.fi.FINetworkBuilder - Running MicroarrayDataAnalyzer.normalizeLeeGeneExp()...

Pairs before normalizing: 209883

After filtering: 205936

Before sequence consolidating: 205903

After sequence consolidating: 205903

Pairs after normalizing: 205903

2012-04-17 14:49:11,354 [main] INFO org.reactome.fi.FINetworkBuilder - Checking its odds ratio...

Total Co-expression: 205903

Total checked pairs: 205903

Total: 140095

Mapped to ppi: 5853 (0.041779)

Mapped to random: 339 (0.002420)

Mapped to random: 348 (0.002484)

Mapped to random: 354 (0.002527)

Mapped to random: 349 (0.002491)

Mapped to random: 359 (0.002563)

Mapped to random: 343 (0.002448)

Mapped to random: 321 (0.002291)

Mapped to random: 351 (0.002505)

Mapped to random: 319 (0.002277)

Mapped to random: 344 (0.002455)

Average odds ratio: 17.804913094911186 +- 0.7129963491801969 (from 10 tests)

2012-04-17 14:49:12,476 [main] INFO org.reactome.fi.FINetworkBuilder - Running MicroarrayDataAnalyzer.generatePrietoCarlosGeneExpFile()...

Total gene pairs: 15841

Total time for loading: 1225

Size of geneNameToUnAcces: 37951

Total unmapped: 2442

Total UniProt pairs before filtering: 19205

After filtering: 19205

Before sequence consolidating: 19205

After sequence consolidating: 19204

After filtering: 19204

2012-04-17 14:49:26,458 [main] INFO org.reactome.fi.FINetworkBuilder - Check its odds ratio...

Co-expressed genes: 19204

Total checked pairs: 19204

Total: 140095

Mapped to ppi: 1477 (0.010543)

Mapped to random: 23 (0.000164)

Mapped to random: 28 (0.000200)

Mapped to random: 26 (0.000186)

Mapped to random: 25 (0.000178)

Mapped to random: 26 (0.000186)

Mapped to random: 28 (0.000200)

Mapped to random: 30 (0.000214)

Mapped to random: 22 (0.000157)

Mapped to random: 37 (0.000264)

Mapped to random: 31 (0.000221)

Average odds ratio: 55.20614231655218 +- 8.106345475156546 (from 10 tests)

2012-04-17 14:49:27,598 [main] INFO org.reactome.fi.FINetworkBuilder - Running PfamAnalyzer.convertIntToPfamIDs()...

Cannot be mapped: 83 10359

Cannot be mapped: 83 10367

Cannot be mapped: 1670 10194

Cannot be mapped: 8683 8683

Cannot be mapped: 10163 10163

Cannot be mapped: 10194 1670

Cannot be mapped: 10359 83

Cannot be mapped: 10359 10359

Cannot be mapped: 10359 10367

Cannot be mapped: 10367 83

Cannot be mapped: 10367 10359

Cannot be mapped: 10367 10367

2012-04-17 14:49:27,946 [main] INFO org.reactome.fi.FINetworkBuilder - Checking its odds ratio...

Total: 140095

Mapped to ppi: 26214 (0.187116)

Mapped to random: 2385 (0.017024)

Mapped to random: 2438 (0.017402)

Mapped to random: 2518 (0.017974)

Mapped to random: 2506 (0.017888)

Mapped to random: 2426 (0.017317)

Mapped to random: 2390 (0.017060)

Mapped to random: 2387 (0.017038)

Mapped to random: 2492 (0.017788)

Mapped to random: 2517 (0.017966)

Mapped to random: 2585 (0.018452)

Average odds ratio: 12.86455345326133 +- 0.3638005629354243 (from 10 tests)

2012-04-17 14:49:30,551 [main] INFO org.reactome.fi.FINetworkBuilder - Checking GO features...

GO BP:

Total: 140095

Mapped to ppi: 83248 (0.594225)

Mapped to random: 12389 (0.088433)

Mapped to random: 12002 (0.085670)

Mapped to random: 12287 (0.087705)

Mapped to random: 12243 (0.087391)

Mapped to random: 12353 (0.088176)

Mapped to random: 12202 (0.087098)

Mapped to random: 12196 (0.087055)

Mapped to random: 12298 (0.087783)

Mapped to random: 12131 (0.086591)

Mapped to random: 12258 (0.087498)

Average odds ratio: 15.303754902615841 +- 0.1542690878940338 (from 10 tests)

GO MF:

Total: 140095

Mapped to ppi: 70072 (0.500175)

Mapped to random: 29013 (0.207095)

Mapped to random: 29045 (0.207324)

Mapped to random: 29404 (0.209886)

Mapped to random: 28955 (0.206681)

Mapped to random: 28887 (0.206196)

Mapped to random: 29213 (0.208523)

Mapped to random: 29037 (0.207266)

Mapped to random: 29047 (0.207338)

Mapped to random: 28900 (0.206289)

Mapped to random: 28959 (0.206710)

Average odds ratio: 3.8260116856207893 +- 0.02579312511474479 (from 10 tests)

GO CP:

Total: 140095

Mapped to ppi: 96108 (0.686020)

Mapped to random: 48300 (0.344766)

Mapped to random: 48418 (0.345608)

Mapped to random: 48146 (0.343667)

Mapped to random: 48362 (0.345209)

Mapped to random: 48440 (0.345765)

Mapped to random: 48444 (0.345794)

Mapped to random: 48144 (0.343653)

Mapped to random: 48293 (0.344716)

Mapped to random: 48430 (0.345694)

Mapped to random: 48385 (0.345373)

Average odds ratio: 4.147761248626852 +- 0.014979309329591423 (from 10 tests)

Notes: 1). For gene expression, we use the SwissProt part of UniProt data only in order to map gene names to UniProt accessions. This is used to control the size of the final UniProt pairs. There are just two many mappings if we use STREMBL too: many of HLA proteins. Since our FI network focuses on the SwissProt proteins, this should be fine.

2). The odds ratio for GO BP is much higher than others. This is the reason why we have not chosen other two GO aspects.

3). The odds ratio for domain-domain interaction should be around 10.

1. **trainNBC()**
   1. Before running trainNBC(), make sure the following methods are correct in class org.reactome.weka.FeatureHandlerForV3:
      1. In method getFeatureList(), make sure all features to be used by NBC have been listed there.
      2. In method loadFatureToPairs(), make sure all PPIs features have point to correct file names. Usually this should be correct if you have set up correct constants in org.reactome.r3.util.R3Constants class.
   2. Set the following two properties in the configuration file:

# The following propertied are used during NBC training

ROC\_CURVE\_FILE=${RESULT\_DIR}/ROC\_100\_041712.txt

BP\_DOMAIN\_SHARED\_PAIRS=${RESULT\_DIR}/BP\_Domain\_Shared\_Pairs.txt

Output from this method running should be similar as follows (pretty slow) (edited not all):

2012-04-17 16:15:57,479 [main] INFO org.reactome.fi.FINetworkBuilder - Running NBCAnalyzer.calculateNBCBasedOnReactome()...

Total pairs: 140095

After filtering: 92342

Prior probability: 0.0054197030050645596

Total negative: 14009500

test fis before filtering: 80521

Removing fis in the training data set: 51196

Total test fis: 19316

Cutoff False\_Positive\_Rate True\_Positive\_Rate

0.0 1.0 1.0

0.1 0.007751132592107361 0.23389935804514392

0.2 0.007594250208700323 0.2263926278732657

0.30000000000000004 0.007010356834695319 0.20066266307724168

0.4 0.001490382642366867 0.11539656243528681

0.5 0.001331941824687573 0.10669910954648995

0.6000000000000001 9.76618810613353E-4 0.08764754607579209

0.7000000000000001 4.77400365400889E-4 0.05161524125077656

0.8 4.722055844933712E-4 0.051149306274591015

0.9 3.942838708806037E-4 0.0479913025471112

1.0 0.0 0.0

One feature contribution:

pavlidisGeneExp: 0.006976872361487769

mousePPI: 0.014120356643917708

celPPI: 0.0073210054704194935

humanInteraction: 0.02392075252844076

pfamDomainInt: 0.0071525124710044765

carlosGeneExp: 0.01915376111547854

dmePPI: 0.010067552716758948

scePPI: 0.004875978648523508

goBPSharing: 0.024191346728450212

Learning probabilties:

carlosGeneExp\_false|false: 0.999721617473857

carlosGeneExp\_false|true: 0.9840051114335838

carlosGeneExp\_true|false: 2.7838252614297444E-4

carlosGeneExp\_true|true: 0.01599488856641615

celPPI\_false|false: 0.9997069131660659

celPPI\_false|true: 0.9935782200948647

celPPI\_true|false: 2.930868339341161E-4

celPPI\_true|true: 0.006421779905135258

dmePPI\_false|false: 0.9992931225240016

dmePPI\_false|true: 0.9789478243919343

dmePPI\_true|false: 7.068774759984297E-4

dmePPI\_true|true: 0.02105217560806567

goBPSharing\_false|false: 0.8900556765052285

goBPSharing\_false|true: 0.09848173095666113

goBPSharing\_true|false: 0.1099443234947714

goBPSharing\_true|true: 0.9015182690433389

humanInteraction\_false|false: 0.998610514293872

humanInteraction\_false|true: 0.9074960473024193

humanInteraction\_true|false: 0.0013894857061279846

humanInteraction\_true|true: 0.09250395269758073

mousePPI\_false|false: 0.9987644098647347

mousePPI\_false|true: 0.9496978622945139

mousePPI\_true|false: 0.0012355901352653557

mousePPI\_true|true: 0.050302137705486125

pavlidisGeneExp\_false|false: 0.9967881794496591

pavlidisGeneExp\_false|true: 0.9366160576985554

pavlidisGeneExp\_true|false: 0.00321182055034084

pavlidisGeneExp\_true|true: 0.06338394230144463

pfamDomainInt\_false|false: 0.9819248367179414

pfamDomainInt\_false|true: 0.7161205085443243

pfamDomainInt\_true|false: 0.018075163282058604

pfamDomainInt\_true|true: 0.28387949145567565

scePPI\_false|false: 0.9983287769013883

scePPI\_false|true: 0.9760672283467978

scePPI\_true|false: 0.0016712230986116563

scePPI\_true|true: 0.023932771653202227

2012-04-17 16:18:44,757 [main] INFO org.reactome.fi.FINetworkBuilder - Running NBCAnalyzer.checkSharedBPPairAndDomainPairs()...

Total GO ids: 29701

Time for looping: 237994

Total shared: 942953

2012-04-17 16:22:44,603 [main] INFO org.reactome.fi.FINetworkBuilder - Running checkCutoffValueForPredictedFIs()...

Total pairs: 3693723

Cutoff: 0.1

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 1307694 (27683)

SwissProt IDs: 12690

SwissProt Coverage: 0.6267285657842749

FIs merged: 1501595 (29794)

SwissProt IDs: 13440

SwissProt Coverage: 0.6637692611615962

Cutoff: 0.2

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 1260342 (26405)

SwissProt IDs: 12610

SwissProt Coverage: 0.6227775582773607

FIs merged: 1454243 (28588)

SwissProt IDs: 13376

SwissProt Coverage: 0.6606084551560648

Cutoff: 0.30000000000000004

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 1021057 (22947)

SwissProt IDs: 12480

SwissProt Coverage: 0.6163571710786251

FIs merged: 1214958 (25397)

SwissProt IDs: 13269

SwissProt Coverage: 0.655323982615567

Cutoff: 0.4

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 72977 (10893)

SwissProt IDs: 8456

SwissProt Coverage: 0.4176214934808376

FIs merged: 266878 (15276)

SwissProt IDs: 10897

SwissProt Coverage: 0.538176610035559

Cutoff: 0.5

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 67892 (10366)

SwissProt IDs: 8024

SwissProt Coverage: 0.3962860529435006

FIs merged: 261793 (14884)

SwissProt IDs: 10591

SwissProt Coverage: 0.523064006321612

Cutoff: 0.6000000000000001

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 51293 (9169)

SwissProt IDs: 7141

SwissProt Coverage: 0.3526768075859344

FIs merged: 245194 (14155)

SwissProt IDs: 10124

SwissProt Coverage: 0.5

Cutoff: 0.7000000000000001

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 37767 (6663)

SwissProt IDs: 4761

SwissProt Coverage: 0.23513433425523508

FIs merged: 231668 (12933)

SwissProt IDs: 9013

SwissProt Coverage: 0.4451303832477282

Cutoff: 0.8

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 34390 (6442)

SwissProt IDs: 4751

SwissProt Coverage: 0.2346404583168708

FIs merged: 228291 (12740)

SwissProt IDs: 9010

SwissProt Coverage: 0.4449822204662189

Cutoff: 0.9

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 25230 (5735)

SwissProt IDs: 4532

SwissProt Coverage: 0.223824575266693

FIs merged: 219131 (12157)

SwissProt IDs: 8885

SwissProt Coverage: 0.43880877123666534

Notes: 1). You should also run NBCGUITest.main() to study the results using different combinations of feature selections. Here is a screenshot from this application:



2). ROC curve and AUC should be plotted and calculated by using the generated file in the results directory, ROC\_100\_031512.txt. In order to do these, use a R script, ROCCurveDrawing.R, in the RSource folder. Change the value fileName to the above generated ROC point file. Run this R script in R. You should get the ROC curve similar to the following:



Based on the above results, calculate AUC using R as following:

> calculate.AUC(rocData)

[1] "AUC: 0.858892192875038"

3). Based on results from above, choose a reasonable cutoff value for high specificity. Usually cutoff value 0.50 should be chosen though you may get a pretty low true positive rate (recall rate).

1. **predictFIs()**
   1. Before running the method, set the cutoff value in the configuration file (usually it should be 0.50):

CUT\_OFF\_VALUE=0.50d

PREDICTED\_FI\_FILE=${RESULT\_DIR}/PredictedFIs\_031512.txt

Output may be like the following:

2012-04-17 16:37:28,524 [main] INFO org.reactome.fi.FINetworkBuilder - Running NBCAnalyzer.generatePredictedFIs()...

FIs from pathways: 193901 (9609)

SwissProt IDs: 7378

SwissProt Coverage: 0.3643816673251679

FIs from prediction: 67892 (10366)

SwissProt IDs: 8024

SwissProt Coverage: 0.3962860529435006

FIs merged: 261793 (14884)

SwissProt IDs: 10591

SwissProt Coverage: 0.523064006321612

1. **buildFIDb()**
   1. Before running the method, create an empty mysql database named as “FI\_yyyy” (yyyy should the year the FI network is building).
   2. Before running the method, make sure the following values in the hibernate configuration file, resources/ funcIntHibernate.cfg.xml , are correct:

<property name=*"connection.url"*>jdbc:mysql://localhost:3306/FI\_2012</property>

<property name=*"connection.username"*>root</property>

<property name=*"connection.password"*>macmysql01</property>

Output from the method running (logging information from hibernate has been removed):

2012-04-17 16:47:20,257 [main] INFO org.reactome.fi.FINetworkBuilder - Running FIDBBuilder.generateSchema()...

……

2012-04-17 16:47:22,447 [main] INFO org.hibernate.tool.hbm2ddl.SchemaExport - schema export complete

Interaction should be empty: 0

2012-04-17 16:47:22,548 [main] INFO org.reactome.fi.FINetworkBuilder - Running FIDBBuilder.dump()...

Total empty interactions: 0

Total empty interactions: 0

……

Total empty interactions: 505

……

Total empty interactions: 199

total interactions: 193901

Total time to extract: 130507

……

Total time for saving: 144991

2012-04-17 16:51:58,046 [main] INFO org.reactome.fi.FINetworkBuilder - Running FIDBBuilder.dumpPredicted()...

Total predicted FIs: 67892

Total protein from db: 9609

Total FIs will be added to the FI database: 67892

Save proteins: 5275

Save evidences: 67892

1. **generateCytoscapePlugInFiles()**
   1. Before running the method, the following properties should be set up in the configuration file:

GENE\_FI\_BIG\_COMP\_FILE\_NAME=${RESULT\_DIR}/FIsInGene\_032112\_BigComp.txt

GENE\_FI\_FILE\_NAME=${RESULT\_DIR}/FIsInGene\_031612.txt

GENE\_FI\_PATHWAY\_FILE\_NAME=${RESULT\_DIR}/FIsInGene\_Pathway\_031612.txt

GENE\_FI\_PREDICTED\_FILE\_NAME=${RESULT\_DIR}/FIsInGene\_Predicted\_031612.txt

# File for mapping accession to names dumped from the FI network

PROTEIN\_ACCESSION\_TO\_NAME\_FILE = ${RESULT\_DIR}/ProteinAccessionToName\_032112.txt

# We need a flattened list of pathways from Reactome for enrichment analysis

REACTOME\_PATHWAYS = ${RESULT\_DIR}/ReactomePathways.txt

# Gene sets based on pathways

PROTEIN\_ID\_TO\_TOPIC = ${RESULT\_DIR}/ProteinIdToTopic032112.txt

GENE\_TO\_TOPIC = ${RESULT\_DIR}/ProteinNameToTopics032112.txt

Output from the method running should be similar to the following:

2012-04-17 17:33:41,044 [main] INFO org.reactome.fi.FINetworkBuilder - Running HibernateFIReader.generateFIFileInGeneInHiberante()...

……

Total interactions from prediction: 67892

Total interactions from pathways: 193901

Time for getting interactions: 5318

Total predicted FIs: 38450

Total pathway FIs:135432

Total FIs: 172235

Total predicted proteins: 8102

Total pathway proteins: 7402

Total proteins: 10696

2012-04-17 17:33:47,972 [main] INFO org.reactome.fi.FINetworkBuilder - Running FIGraphAnalyzer.analyzeComponents()...

Total interactions: 172235

Total components: 81

0: 10501

……

2012-04-17 17:33:49,860 [main] INFO org.reactome.fi.FINetworkBuilder - Running HiberanteFIReader.generateAccessionToProteinNameMap()...

2012-04-17 17:33:50,353 [main] INFO org.reactome.fi.FINetworkBuilder - Running ReactomeAnalyzer.generateListOfPathways()...

Total Pathways: 141

2012-04-17 17:34:12,681 [main] INFO org.reactome.fi.FINetworkBuilder - Running PathwayGeneSetGenerator.generateProteinNameToPathwayMap()...

……

*======The END======*