

Respiratory Determinants of Diurnal Hypercapnia in Obesity Hypoventilation Syndrome

What Does Weight Have to Do with It?

Shahrokh Javaheri¹ and Loretta A. Simbarti²

¹Cincinnati Veterans Affairs Medical Center, Cincinnati, Ohio; and ²University of Cincinnati College of Medicine, Cincinnati, Ohio

Abstract

Rationale: Among morbidly obese individuals, obstructive sleep apnea (OSA) is highly prevalent, with up to 20% suffering from hypoventilation syndrome. An increased diurnal PaCO_2 , the signature of obesity hypoventilation syndrome (OHS), implies diminished global ventilation, hence the term hypoventilation.

Objectives: We hypothesized that hypercapnic patients with OSA have lower \dot{V}_E than eucapnic patients with OSA.

Methods: In this prospective study we recorded respiratory variables to determine the pathophysiological mechanisms of steady-state diurnal hypercapnia of 12 consecutive hypercapnic and 20 consecutive eucapnic patients with OSA, matched for apnea-hypopnea index. Patients with any known causes of hypercapnia were not included.

Measurements and Main Results: Comparing hypercapnic to eucapnic patients, the mean value (\pm SD) for PaCO_2 (52 ± 5 vs.

40 ± 3 mm Hg) was significantly higher, and the mean PaO_2 (59 ± 8 vs. 75 ± 10 mm Hg) was significantly lower, in the hypercapnic patients. Surprisingly, the mean values for \dot{V}_E (12.2 ± 3.0 vs. 11.6 ± 2.0 L/min), alveolar ventilation, breathing rate, \dot{V}_T , and dead space did not differ significantly. However, hypercapnic patients had a significantly greater CO_2 production (336 ± 79 vs. 278 ± 58 ml/min), which was the main reason for hypercapnia. When adjusted for body surface area, the mean values for CO_2 production were similar between the two groups.

Conclusions: These data emphasize the importance of weight loss, which could potentially reverse hypercapnic OSA to eucapnic OSA, hypothetically even in the absence of improvement in apnea-hypopnea index. In addition, reversal of hypercapnia should also improve oxygenation, both during sleep and while awake, minimizing hypoxia-induced organ dysfunction of OHS.

Keywords: metabolic rate; weight loss; morbid obesity; hypoxia; CO_2 retention

(Received in original form March 6, 2014; accepted in final form May 5, 2014)

Supported by Merit Review Grants from the Department of Veterans Affairs.

Author Contributions: S.J.: design, execution, interpretation, and drafting of the manuscript. L.S.: statistical analysis of the data and drafting of the manuscript.

Correspondence and requests for reprints should be addressed to Shahrokh Javaheri, M.D., Emeritus Professor of Medicine, University of Cincinnati College of Medicine, 231 Albert Sabin Way, Cincinnati, OH 45229. E-mail: shahrokhjavaheri@icloud.com

Ann Am Thorac Soc Vol 11, No 6, pp 945–950, Jul 2014

Published 2014 by the American Thoracic Society

DOI: 10.1513/AnnalsATS.201403-099OC

Internet address: www.atsjournals.org

Obesity is a highly prevalent disorder associated with excess health cost and multiple medical complications. One of these consequences, obesity hypoventilation syndrome (OHS), is a distinct clinical entity characterized by the combination of obesity, daytime steady-state hypercapnia, and the presence of a sleeping breathing disorder, usually, although not always, obstructive sleep apnea (OSA) (1–7). Hypercapnia should not be due to other known diseases.

OHS is associated with a number of comorbidities and in the long run with premature mortality. A major consequence of hypercapnia is the imposed hypoxemia, which contributes to inflammation and organ dysfunction in patients with OHS.

Although the term hypoventilation defines the syndrome by referring to the elevated PaCO_2 , to our knowledge, there are no systematic studies defining the respiratory determinants of diurnal

hypercapnia. In general, when the term hypercapnia is used, it implies diminished global ventilation, hence the term hypoventilation. Although this is the case in some conditions associated with hypercapnia (8), for example opioid-induced respiratory depression when global ventilation and therefore alveolar ventilation is diminished, hypercapnia could be present despite normal global ventilation (8–10).

Equation 1 is the simple formula of alveolar ventilation showing that PaCO_2 is directly proportional to CO_2 production (\dot{V}_{CO_2}) and inversely proportional to alveolar ventilation (\dot{V}_A):

$$\text{PaCO}_2 = K \times \dot{V}_{\text{CO}_2} / \dot{V}_A, \quad (1)$$

where K is a constant.

Because $\dot{V}_A = \dot{V}_E$ minus dead space ventilation (\dot{V}_D), the denominator of the equation is changed and:

$$\text{PaCO}_2 = K \times \dot{V}_{\text{CO}_2} / \text{MV} - \dot{V}_D.$$

Meanwhile, $\text{MV} = \text{RR} \times \text{VT}$, and $\dot{V}_D = \text{RR} \times \text{VD}$, where RR is respiratory rate, VT is tidal volume, and VD is dead space volume. Therefore, $\dot{V}_A = \text{RR} \times \text{VT} - \text{RR} \times \text{VD}$, or alternatively, $\dot{V}_A = \text{RR} (\text{VT} - \text{VD})$. Then, $\text{PaCO}_2 = K \times \dot{V}_{\text{CO}_2} / \text{RR} (\text{VT} - \text{VD})$, or

$$\text{PaCO}_2 = K \times \dot{V}_{\text{CO}_2} / \text{RR} (1 - \text{VD} / \text{VT}). \quad (2)$$

Equation 2, a modification of the alveolar ventilation equation, shows the various determinants of steady-state hypercapnia. As an example, the underlying respiratory determinants of hypercapnia in chronic obstructive pulmonary disease (COPD), comparing hypercapnic to eucapnic patients matched for pulmonary function tests, dead space, and CO_2 production, are rapid shallow breathing, which results in increased VD / VT ratio and \dot{V}_D with consequent diminished alveolar ventilation and hypercapnia (9, 10). Importantly, there are no significant differences in global ventilation when the two groups of patients with COPD are compared.

The present study was designed to determine the underlying mechanisms of diurnal hypercapnia in patients with OHS, while exercising extreme care, to the best of our abilities, not to disturb steady state. Frequently, sampling of arterial blood and measuring ventilation are associated with anxiety and other behavioral influences that could disturb steady state. Consequently, the measured PaCO_2 is not accurate. It is for this reason that serum bicarbonate has been suggested as a surrogate (11). The major concern, here, is the potential for "hyperventilation," as the result of which the acid-base disturbance that *in vivo* is chronic respiratory acidosis is interpreted as primary metabolic alkalosis (12), when in reality, the disturbance is post-hypercapnic metabolic alkalosis.

Methods

Design

This was a prospective study of 32 consecutive patients. The entry criteria included a history of loud snoring, witnessed apnea, excessive daytime sleepiness, obesity, and other symptoms of OSA syndrome, and a consistent polysomnography (PSG). Most of the patients who participated in this study were part of a larger study in which we measured ventilatory responses of patients and their family members (13). Data were collected from 12 consecutive hypercapnic and 20 eucapnic patients with obstructive sleep apnea-hypopnea syndrome matched for severity of OSA using the metric apnea-hypopnea index (AHI). As noted above, the purpose of this study was not to shed light on the mechanisms of transition from nocturnal hypercapnia to daytime hypercapnia but rather to specifically investigate the respiratory determinants of steady-state hypercapnia while awake.

All patients were male veterans who were referred to the author (S.J.) at a government-based research hospital. For homogeneity, only male patients were included, because female veterans are seldom referred to this center.

Procedures and Tests

PSG was performed using standard techniques as detailed previously (14, 15). All patients were adapted to the sleep laboratory during the first night by having electrodes placed without recordings being made. PSG was performed during the second night. For staging sleep, we recorded EEG (two channels), chin electromyogram (one channel), and electrooculogram (two channels). Thoracoabdominal excursions were measured qualitatively by respiratory inductance plethysmography (Respirace; Ambulatory Monitoring Inc., Ardsley, NY) placed over the rib cage and abdomen. Airflow was qualitatively monitored using an oral/nasal thermocouple (Model TCT1R; Grass Instrument Co., Quincy, MA). Arterial blood oxyhemoglobin saturation was recorded using an ear oximeter (Biox IIA; BT Inc., Boulder, CO). These variables were recorded on a multichannel polygraph (Model 78D; Grass Instrument Co., Quincy, MA). An apnea was defined as cessation of inspiratory airflow for 10 seconds or more.

An obstructive apnea was defined as the absence of airflow in the presence of rib cage and abdominal excursions. A central apnea was defined as the absence of airflow with absence of rib cage and abdominal excursions. A hypopnea was defined as a discernible reduction of airflow (30%) lasting 10 seconds or more and associated with at least a 4% drop in arterial oxyhemoglobin saturation and/or an arousal (for further details, see References 7, 13–15). The number of apneas and hypopneas per hour of sleep is referred to as the AHI. The number of arousals per hour of sleep is referred to as the arousal index.

Coded polysomnograms were scored blindly page by page by one of the authors for staging of sleep, arousals, and respiratory events.

In the morning of PSG, the following tests were obtained: arterial blood gases and pH, ventilation parameters, hypercapnic ventilatory response, and pulmonary function tests. In conducting these tests, extreme caution was exercised to ensure uniformity in technique. Patients were not allowed to drink caffeinated products on the morning of the tests. Tests were performed 2 hours after a meal, and the patients were asked to empty their bladders before the tests. Arterial blood samples were obtained using strict criteria, as detailed previously (13–15). To minimize pain and anxiety, 2% lidocaine was used to anesthetize skin where the radial artery was punctured. Afterward, by touching the patient's skin with a needle, the patient was assured that the procedure was painless. Our hope was to minimize pain and anxiety and, therefore, any attendant change in PaCO_2 . With the patient in a sitting position and breathing comfortably for several minutes, an arterial blood sample was obtained painlessly, as attested by the patient. The care used in drawing the blood sample was to assure steady state in the sense that the *in vitro* measured PaCO_2 accurately reflects the *in vivo* value. Details for measurements can be found elsewhere (15).

Ventilation, CO_2 Production, and O_2 Consumption

After the arterial blood sampling, ventilation parameters were measured, details of which have been described previously (14, 15). The patients were familiarized with the equipment and breathed through the mouthpiece for several minutes before

measurement began. An assistant monitored the patients continuously to ensure that they remained awake. Ventilation was measured in the sitting position with the subject wearing a nose clip and breathing through a mouthpiece connected to a low-resistance two-way valve (Hans-Rudolph, Kansas City, MO). Measurements were started several minutes after a stable baseline had been achieved (i.e., stable end-tidal P_{CO_2} for several minutes).

Airflow and the volume were measured by a pneumoscan (S-301; KL Engineering Co., Sylmar, CA). Calibrations were performed by known flow rates from pneumatic calibrator (model 65-250; Penwalt, Cor, Belleville, NJ) and for volume by a 3-L syringe. End-tidal CO_2 was sampled at the mouthpiece by an infrared CO_2 analyzer (Beckman medical gas analyzer, LB-2; Beckman Cardiopulmonary Instruments, Fullerton, CA), which was calibrated with gases of known CO_2 concentrations.

Resting ventilation was measured as the subject breathed room air for 3 minutes while exhaled gas was collected for measurement of mixed expired CO_2 and O_2 concentrations by gas analyzer. Dead space was calculated using the Bohr equation, the known arterial, and the mixed-expired P_{CO_2} . Alveolar ventilation was calculated by subtracting \dot{V}_E (the product of \dot{V}_T times breathing rate) from dead space ventilation (the product of \dot{V}_D times breathing rate). Respiratory quotient was calculated as the ratio of CO_2 production over oxygen consumption.

Hypercapnic Ventilatory Response

The hypercapnic ventilatory response (HCVR) was measured in the sitting position with the subject wearing a nose clip and breathing through a mouthpiece connected to a low-resistance two-way valve. Details of the test as performed in our laboratory have been previously described (7, 14, 15). Measurements were started several minutes after a stable baseline had been achieved (i.e., ventilation and end-tidal P_{CO_2} had not changed for several minutes). The hyperoxic HCVR was determined by Read's rebreathing method. Linear regression was used to determine the slope according to the equation: $\dot{V} = S \times Pa_{CO_2} - B$, where S is the slope of HCVR and B is the intercept (on the abscissa) of the line that relates ventilation to P_{CO_2} .

Pulmonary Function Tests

For measurements of lung volume, flow rates, and single breath carbon monoxide diffusing capacity, we used an automated system (Collings/DS560; Braintree, MA) as detailed previously (16). Maximum voluntary ventilation was measured by instructing the subject to breathe as quickly and as deeply as possible for 12 seconds. For further reference, these procedures have been described previously (7).

Left and right ventricular ejection fractions were also measured by radioactive technique as reported previously (17), either the day before or after PSG.

The protocol was approved by the Institutional Review Board of the University of Cincinnati College of Medicine. All patients signed informed consent.

Statistical Analysis

Unpaired two-tailed t tests were used to determine differences of statistical significance between the two groups. Data are presented as means \pm SD, and P less than 0.05 was considered significant. Dichotomous categorical values were compared using Chi-square tests. Calculations were performed using SAS 9.0.

Results

Hypercapnic patients were significantly heavier with a greater body mass index than

eucapnic patients (Table 1). Hypercapnic patients, as expected, had a lower FEV_1 and FVC than eucapnic patients; however, the mean values of the percent predicted FEV_1 /FVC ratio were normal and did not differ significantly between the two groups.

There were no significant differences in left (48%, $n = 12$ vs. 54%, $n = 15$) or right (48%, $n = 12$ vs. 49%, $n = 15$) ventricular ejection fractions between and hypercapnic and eucapnic patients (15 of the 20 patients).

As expected, hypercapnic patients also had significantly greater plasma $[HCO_3^-]$ than eucapnic patients, reflecting the chronicity of CO_2 retention (Table 2). The mean value of the hypercapnic ventilatory response was significantly lower in the hypercapnic than in the eucapnic patients. Also in part imposed by hypercapnia, the mean PaO_2 was significantly lower in hypercapnic patients.

There were no significant differences in ventilation parameters, and the mean values of \dot{V}_E and alveolar ventilations were similar between the two groups. Hypercapnic patients had a significantly greater CO_2 production ($P = 0.02$) and trended toward higher oxygen consumption ($P = 0.06$) than eucapnic patients; importantly, when these values were adjusted for body surface area, there were no significant differences between the two groups. Respiratory quotients were within normal range and did not differ significantly between the two groups. The normal values for respiratory

Table 1. Anthropomorphic and pulmonary function tests of hypercapnic and eucapnic patients with sleep apnea-hypopnea syndrome

	Hypercapnic ($n = 12$)	Eucapnic ($n = 20$)	P Value
Age, yr	51 \pm 11	53 \pm 9	0.68
Weight, kg	159 \pm 23	121 \pm 22	0.0001
Height, cm	180 \pm 8	176 \pm 4	0.04
Body mass index, kg/m ²	49 \pm 8	39 \pm 6	0.0004
Body surface area, m ²	2.66 \pm 0.21	2.34 \pm 0.20	0.0002
FEV_1 , L/s	2.34 \pm 0.85 (61 \pm 19)	2.98 \pm 0.82 (84 \pm 21)	0.04 (0.004)
FVC, L	2.93 \pm 0.10 (63 \pm 19)	3.77 \pm 1.04 (87 \pm 21)	0.03 (0.003)
FEV_1 /FVC, %	79 \pm 6 (97 \pm 6)	79 \pm 5 (97 \pm 7)	0.91 (0.96)
FRC, L	2.31 \pm 0.58	2.40 \pm 0.64	0.69
TLC, L	5.06 \pm 0.95 (75 \pm 15)	5.73 \pm 1.06 (90 \pm 16)	0.09 (0.014)
RV, L	2.06 \pm 0.66 (65 \pm 19)	1.86 \pm 0.04 (94 \pm 21)	0.03 (0.0004)
Diffusing capacity for CO, ml/min/mm Hg	27 \pm 8 (93 \pm 28)	28 \pm 8 (97 \pm 28)	0.86 (0.69)
Maximum voluntary ventilation, L/min	93 \pm 37 (58 \pm 20)	118 \pm 36 (87 \pm 32)	0.07 (0.007)
Slope of HCVR, ml/min/mm Hg	1.39 \pm 0.57	2.83 \pm 2.31	0.04

Definition of abbreviations: HCVR = hypercapnic ventilatory response; RV = residual volume. Values are mean \pm SD. Values in parenthesis are percent predicted.

Table 2. Ventilation data of hypercapnic and eucapnic patients with sleep apnea-hypopnea syndrome

	Hypercapnic (n = 12)	Eucapnic (n = 20)	P Value
PaCO ₂ , mm Hg	52 ± 5	40 ± 3	0.0001
PaO ₂ , mm Hg	59 ± 8	75 ± 10	0.0001
[H ⁺], nEq/L	40 ± 4	39 ± 2	0.39
[HCO ₃ ⁻], mEq/L	32 ± 4	25 ± 2	0.0001
Breathing rate, n/min	14.9 ± 3.9	14.0 ± 4.4	0.59
V _T , ml	834 ± 124	853 ± 208	0.78
VE, L/min	12.2 ± 3.0	11.6 ± 2.0	0.52
VE/BSA, L/min/m ²	4.6 ± 1.0	5.0 ± 0.09	0.26
Alveolar ventilation, L/min	5.7 ± 1.5	6.1 ± 1.5	0.53
Alveolar ventilation/BSA, L/min/m ²	2.2 ± 0.5	2.6 ± 0.6	0.04
Dead space, ml	436 ± 59	397 ± 8	0.18
Dead space ventilation, L/min	6.5 ± 2.0	5.5 ± 1.6	0.13
Dead space ventilation/BSA, L/min/m ²	2.4 ± 0.7	2.4 ± 0.7	0.82
Dead space/V _T ratio, %	53 ± 7	48 ± 9	0.10
CO ₂ production, ml/min	336 ± 79	278 ± 58	0.02
CO ₂ production/BSA, ml/min/m ²	126 ± 27	119 ± 22	0.40
Oxygen consumption, ml/min	435 ± 101	360 ± 89	0.06
Oxygen consumption/BSA, ml/min/m ²	164 ± 34	153 ± 36	0.40
Respiratory quotient	0.80 ± 0.10	0.82 ± 0.13	0.60

Definition of abbreviation: BSA = Body surface area.

Values are mean ± SD. All ventilation data are in body temperature pressure saturated except for CO₂ production and O₂ consumption, which are in standard temperature pressure saturated.

quotients are consistent with steady-state conditions critical to our measurements.

Polysomnographic data are presented in Table 3. There were no significant differences in mean values for sleep efficiency, sleep onset, percent REM and non-REM sleep, or AHI (the matching variable) when hypercapnic and eucapnic patients were compared. The major difference was related to desaturation (Table 3).

Discussion

The important findings of the current study are: (1) Patients with OHS have an increased baseline metabolic rate, which is

characterized by increased carbon dioxide production and oxygen consumption. (2) When the parameters of metabolic rate are adjusted for body surface area, the differences between hypercapnic and eucapnic patients become statistically nonsignificant. (3) In this group of patients with OHS, carbon dioxide hyperproduction was the main reason for chronic CO₂ retention, as the mean values for VE and VA were similar when compared with eucapnic patients.

Patients with OHS, by definition, have elevated daytime PaCO₂. The mechanisms that result in spillover from nocturnal hypercapnia to daytime hypercapnia have been speculated (7) and studied in detail by

Berger and colleagues (18–20), Norman and colleagues (21), and Ayappa and colleagues (22). To our knowledge, our study is the first exploring the physiological respiratory mechanisms of diurnal hypercapnia. We had hypothesized that patients with OHS have lower VE and VA than eucapnic patients. To our surprise, this was not the case, and the major mechanism for hypercapnia was CO₂ hyperproduction.

The major function of the respiratory system is to maintain partial pressures of arterial blood oxygen and carbon dioxide and pH normal. Normally, steady-state PaCO₂ is tightly regulated via a negative feedback system. This homeostatic regulation is achieved by adjustment of ventilation to the metabolic carbon dioxide production (Equation 1). In this way, PaCO₂ is tightly controlled and remains constant throughout life (8).

Pathophysiologically, there are four respiratory mechanisms that can result in daytime hypercapnia (8, 23). These include (1) decreased global and alveolar ventilation, as occurs with use of respiratory depressant medications such as opioids, a condition wherein the use of the term hypoventilation is most appropriate; (2) ventilation/perfusion mismatch impairing CO₂ clearance (23) unless ventilation rises enough to increase alveolar ventilation to compensate for ventilation/perfusion mismatch as it commonly occurs in interstitial lung diseases (24); (3) a pattern of breathing characterized by diminished VT and increased breathing rate as it occurs in hypercapnic patients with COPD (9, 10), the so-called blue bloaters; and (4) increased CO₂ production not compensated by increased VE and alveolar ventilation as it occurred in our patients.

As noted above, normally there is coupling of carbon dioxide production with alveolar ventilation, such that PaCO₂ remains constant. A relevant example is exercise, where in spite of several-fold increase in CO₂ production, PaCO₂ remains constant (as long as anaerobic threshold is not reached). Therefore, based on the results of the present study, in patients with OHS, there is uncoupling between CO₂ production, which is increased, and lack of compensation by the respiratory system. Now that this uncoupling has been demonstrated, the question is if this is due to a “cannot breathe” or “do not want to

Table 3. Polysomnographic data of hypercapnic and eucapnic patients with sleep apnea-hypopnea syndrome

	Hypercapnic (n = 12)	Eucapnic (n = 20)	P Value
Total recording time, min	336 ± 88	322 ± 64	0.60
Total sleep time, min	249 ± 95	250 ± 60	1.0
Sleep efficiency, %	70 ± 20	80 ± 12	0.09
Sleep onset, min	10 ± 15	12 ± 18	0.80
Non-REM/total sleep time, %	85 ± 8	88 ± 10	0.40
REM/total sleep time, %	15 ± 8	11 ± 8	0.17
Apnea-hypopnea index, n/h	50 ± 33	47 ± 33	0.80
Baseline saturation, %	92 ± 3	95 ± 2	0.003
Lowest saturation, %	55 ± 17	65 ± 13	0.05

breathe” mechanism, or the combination of the two? This study was not designed to answer this question, and the discussion of evolution of diurnal hypercapnia is beyond the scope of this manuscript. However, we suggest that decreased ventilatory response could be the consequence rather than the cause of CO₂ retention (7, 25). The massive obesity imposes both mechanical impairment (poor chest wall compliance, curtailing maximum ventilation when needed, for example, in response to an apnea) and increased CO₂ load (respiratory and nonrespiratory production). These are critical factors that impair CO₂ clearance during interapneic periods when ventilation needs to increase to unload the huge amount of metabolically produced CO₂ and retained during obstructive apneic periods. It is hypothesized that with prolonged nocturnal CO₂ retention, plasma HCO₃⁻ concentration gradually rises (7, 19, 25), decreasing ventilatory response to CO₂ (12). However, it is certainly plausible that a diminished ventilatory response could predispose to chronic daytime hypercapnia, although we emphasize that diminished hypercapnic ventilatory response in patients with OHS does not appear to have a genetic predisposition (13). On the other hand, a diminished ventilatory response to CO₂ could contribute to maintenance of diurnal hypercapnia.

Studies in the literature suggest respiratory muscle fatigue and impaired endurance (for reviews, *see* References 1–3, 5, 6). In the present study, maximum voluntary ventilation was trending lower in patients with OHS compared with eucapnic patients. Considering that the test was only 12 seconds in duration and with small number of patients, our data are potentially consistent with the fatigue paradigm.

We should note that a number of studies are available showing that weight loss (26, 27), as well as treatment with positive airway pressure devices (28–33), improves PaCO₂, although residual hypercapnia may persist in spite of excellent adherence to positive airway pressure therapy (31). So far, the results of such studies have been interpreted as if the

improvement in sleep apnea was the only mechanism by which PaCO₂ improved. The results of the current study shed light on an additional and important mechanism contributing to the normalization of PaCO₂. Furthermore, residual hypercapnia after treatment with positive airway pressure devices may be due to persistent CO₂ hyperproduction of obesity.

In fact, comparing the collective information of both groups, the polysomnographic (matched AHI values, Table 2) and ventilation data (Table 3) suggest that weight loss by itself, even in the absence of any improvement in AHI, could potentially convert the phenotype of OSA from hypercapnic to eucapnic. We emphasize, however, the small number of patients enrolled, which could have affected our findings.

In the present study, there were no significant differences in \dot{V}_E between the two groups, as noted earlier. These results appear to be somewhat similar to the study of Pankow and colleagues (34). These authors measured supine \dot{V}_E in six patients with OHS, seven eucapnic patients with OSA, and five obese participants.

Comparing the first two groups, respective values for \dot{V}_E were 10 and 8.4 L/min (*P* value not reported). Authors did not measure CO₂ production; however, respective mean values for body mass indices were 44 and 46 kg/m². This difference in body weight, resulting in excess CO₂ production in patients with OHS, perhaps along with lower \dot{V}_E , could have accounted for hypercapnia. We emphasize the small number of patients and differences in design, which include use of face mask, insertion of an esophageal balloon, and supine measurements. Regarding increased CO₂ production and increased O₂ consumption associated with obesity, our results confirm those of previous studies (35, 36).

Clinical Implications

When compared with patients with OSA and normal PaCO₂, patients with OSA and hypoventilation have decreased quality of life, excess healthcare cost, increased

morbidity, and premature mortality (37, 38). These adverse consequences are not the result of high PaCO₂ adversely affecting enzymatic function by acidemia, because with renal compensation, pH is normalized (Table 2). However, based on Dalton’s law, a high PaCO₂ imposes a low PaO₂ (Table 2), and hypoxemia results in a number of adverse consequences, including activation of redox-sensitive genes with consequent production of reactive oxygen species and activation of an inflammatory cascade (39). Yet, we should emphasize that administration of supplemental oxygen could be associated with worsening hypercapnia (40).

An important implication of our data is that excess CO₂ production is directly related to body surface area. Thus, weight loss should have a major impact on metabolic production of CO₂ (and consumption of oxygen) with a decrease in PaCO₂ and, based on Dalton’s law, an incremental increase in PaO₂. Normalization of oxygen balance is critical in prevention of organ dysfunction due to hypoxia.

Summary

Patients with OSA and OHS, when matched with eucapnic patients with OSA, are more obese, have much higher CO₂ production, but have similar \dot{V}_E and alveolar ventilation. Therefore, CO₂ hyperproduction accounts for their hypercapnia. And when CO₂ production is adjusted for body surface area, there is no significant difference between hypercapnic and eucapnic patients. In part due to hypercapnia, patients with OHS have hypoxemia, which should contribute to activation of inflammatory pathways and organ dysfunction. Our data suggest that weight loss *per se* could change the hypercapnic OSA to eucapnic phenotype and simultaneously improve oxygenation. Improved oxygen balance should minimize long-term organ damage associated with OHS. ■

Author disclosures are available with the text of this article at www.atsjournals.org.

References

- 1 Mokhlesi B. Obesity hypoventilation syndrome: a state-of-the-art review. *Respir Care* 2010;55:1347–1362, discussion 1363–1365.
- 2 Chau EH, Lam D, Wong J, Mokhlesi B, Chung F. Obesity hypoventilation syndrome: a review of epidemiology, pathophysiology, and perioperative considerations. *Anesthesiology* 2012;117:188–205.
- 3 Piper AJ, Grunstein RR. Obesity hypoventilation syndrome: mechanisms and management. *Am J Respir Crit Care Med* 2011;183:292–298.
- 4 Olson AL, Zwillich C. The obesity hypoventilation syndrome. *Am J Med* 2005;118:948–956.
- 5 Mokhlesi B, Kryger MH, Grunstein RR. Assessment and management of patients with obesity hypoventilation syndrome. *Proc Am Thorac Soc* 2008;5:218–225.

- 6 Piper AJ, Grunstein RR. Big breathing: the complex interaction of obesity, hypoventilation, weight loss, and respiratory function. *J Appl Physiol* (1985) 2010;108:199–205.
- 7 Javaheri S, Colangelo G, Lacey W, Gartside PS. Chronic hypercapnia in obstructive sleep apnea-hypopnea syndrome. *Sleep* 1994;17:416–423.
- 8 Javaheri S. Determinants of carbon dioxide tension. In: Gennari FJ, Adroque HJ, Galla JH, Madias NE, editors. *Acid-base disorders and their treatment*. Boca Raton, FL: Taylor and Francisco Group; 2005. pp 47–77.
- 9 Sorli J, Grassino A, Lorange G, Milic-Emili J. Control of breathing in patients with chronic obstructive lung disease. *Clin Sci Mol Med* 1978;54:295–304.
- 10 Javaheri S, Blum J, Kazemi H. Pattern of breathing and carbon dioxide retention in chronic obstructive lung disease. *Am J Med* 1981;71:228–234.
- 11 Hart N, Mandal S, Manuel A, Mokhlesi B, Pépin JL, Piper A, Stradling JR. Obesity hypoventilation syndrome: does the current definition need revisiting? *Thorax* 2014;69:83–84.
- 12 Javaheri S, Kazemi H. Metabolic alkalosis and hypoventilation in humans. *Am Rev Respir Dis* 1987;136:1011–1016.
- 13 Javaheri S, Colangelo G, Corser B, Zahedpour MR. Familial respiratory chemosensitivity does not predict hypercapnia of patients with sleep apnea-hypopnea syndrome. *Am Rev Respir Dis* 1992;145:837–840.
- 14 Javaheri S. A mechanism of central sleep apnea in patients with heart failure. *N Engl J Med* 1999;341:949–954.
- 15 Javaheri S, Sands SA, Edwards BA. Acetazolamide attenuates Hunter-Cheyne-Stokes breathing but augments the hypercapnic ventilatory response in patients with heart failure. *Ann Am Thorac Soc* 2014;11:80–86.
- 16 Javaheri S, Bosken CH, Lim SP, Dohn MN, Greene NB, Baughman RP. Effects of hypoventilation on lung functions in humans. *Am Rev Respir Dis* 1987;135:597–599.
- 17 Javaheri S, Parker TJ, Wexler L, Liming JD, Lindower P, Roselle GA. Effects of theophylline on sleep disordered breathing in stable heart failure. *N Engl J Med* 1996;335:562–567.
- 18 Berger KI, Goldring RM, Rapoport DM. Obesity hypoventilation syndrome. *Semin Respir Crit Care Med* 2009;30:253–261.
- 19 Berger KI, Ayappa I, Sorkin IB, Norman RG, Rapoport DM, Goldring RM. CO₂ homeostasis during periodic breathing in obstructive sleep apnea. *J Appl Physiol* (1985) 2000;88:257–264.
- 20 Berger KI, Ayappa I, Chatr-Amontri B, Marfatia A, Sorkin IB, Rapoport DM, Goldring RM. Obesity hypoventilation syndrome as a spectrum of respiratory disturbances during sleep. *Chest* 2001;120:1231–1238.
- 21 Norman RG, Goldring RM, Clain JM, Oppenheimer BW, Charney AN, Rapoport DM, Berger KI. Transition from acute to chronic hypercapnia in patients with periodic breathing: predictions from a computer model. *J Appl Physiol* (1985) 2006;100:1733–1741.
- 22 Ayappa I, Berger KI, Norman RG, Oppenheimer BW, Rapoport DM, Goldring RM. Hypercapnia and ventilatory periodicity in obstructive sleep apnea syndrome. *Am J Respir Crit Care Med* 2002;166:1112–1115.
- 23 West JB. Causes of carbon dioxide retention in lung disease. *N Engl J Med* 1971;284:1232–1236.
- 24 Javaheri S, Sicilian L. Lung function, breathing pattern, and gas exchange in interstitial lung disease. *Thorax* 1992;47:93–97.
- 25 Almoosa KF, Almoosa KF, Javaheri S. Obesity and the control of breathing. In: Ward DS, Dahan A, Teppema LJ, editors. *Pharmacology and pathophysiology of the control of breathing; lung biology in health and Disease*. Vol 202. Boca Raton, FL: Taylor and Francis; 2005. pp. 383–412.
- 26 Boone KA, Cullen JJ, Mason EE, Scott DH, Doherty C, Maher JW. Impact of vertical banded gastroplasty on respiratory insufficiency of severe obesity. *Obes Surg* 1996;6:454–458.
- 27 Marti-Valeri C, Sabaté A, Masdevall C, Dalmau A. Improvement of associated respiratory problems in morbidly obese patients after open Roux-en-Y gastric bypass. *Obes Surg* 2007;17:1102–1110.
- 28 Sullivan CE, Berthon-Jones M, Issa FG. Remission of severe obesity-hypoventilation syndrome after short-term treatment during sleep with nasal continuous positive airway pressure. *Am Rev Respir Dis* 1983;128:177–181.
- 29 Piper AJ, Wang D, Yee BJ, Barnes DJ, Grunstein RR. Randomised trial of CPAP vs bilevel support in the treatment of obesity hypoventilation syndrome without severe nocturnal desaturation. *Thorax* 2008;63:395–401.
- 30 Han F, Chen E, Wei H, He Q, Ding D, Strohl KP. Treatment effects on carbon dioxide retention in patients with obstructive sleep apnea-hypopnea syndrome. *Chest* 2001;119:1814–1819.
- 31 Mokhlesi B, Tulaimat A, Evans AT, Wang Y, Itani AA, Hassaballa HA, Herdegen JJ, Stepanski EJ. Impact of adherence with positive airway pressure therapy on hypercapnia in obstructive sleep apnea. *J Clin Sleep Med* 2006;2:57–62.
- 32 Masa JF, Celli BR, Riesco JA, Hernández M, Sánchez De Cos J, Disdier C. The obesity hypoventilation syndrome can be treated with noninvasive mechanical ventilation. *Chest* 2001;119:1102–1107.
- 33 Chouri-Pontarollo N, Borel JC, Tamisier R, Wuyam B, Levy P, Pépin JL. Impaired objective daytime vigilance in obesity-hypoventilation syndrome: impact of noninvasive ventilation. *Chest* 2007;131:148–155.
- 34 Pankow W, Hijeh N, Schüttler F, Penzel T, Becker HF, Peter JH, von Wichert P. Influence of noninvasive positive pressure ventilation on inspiratory muscle activity in obese subjects. *Eur Respir J* 1997;10:2847–2852.
- 35 Kress JP, Pohlman AS, Alverdy J, Hall JB. The impact of morbid obesity on oxygen cost of breathing (VO₂(RESP)) at rest. *Am J Respir Crit Care Med* 1999;160:883–886.
- 36 Gilbert R, Sipple JH, Auchincloss JH Jr. Respiratory control and work of breathing in obese subjects. *J Appl Physiol* 1961;16:21–26.
- 37 Berg G, Delaive K, Manfreda J, Walld R, Kryger MH. The use of health-care resources in obesity-hypoventilation syndrome. *Chest* 2001;120:377–383.
- 38 Nowbar S, Burkart KM, Gonzales R, Fedorowicz A, Gozansky WS, Gaudio JC, Taylor MR, Zwillich CW. Obesity-associated hypoventilation in hospitalized patients: prevalence, effects, and outcome. *Am J Med* 2004;116:1–7.
- 39 Borel JC, Roux-Lombard P, Tamisier R, Arnaud C, Monneret D, Arnol N, Baguet JP, Levy P, Pepin JL. Endothelial dysfunction and specific inflammation in obesity hypoventilation syndrome. *PLoS ONE* 2009;4:e6733.
- 40 Hollier CA, Harmer AR, Maxwell LJ, Menadue C, Willson GN, Unger G, Flunt D, Black DA, Piper AJ. Moderate concentrations of supplemental oxygen worsen hypercapnia in obesity hypoventilation syndrome: a randomised crossover study. *Thorax* 2014;69:346–353.