

## Overview

UniPath provides robust statistical methods to represent every single cell using pathway and gene-set enrichment scores. It can be used with both single cell RNA-seq and single cell ATAC-seq profile with scalability for atlas scale data-sets. UniPath comes with several features like pseudo-temporal ordering using pathway scores and unconventional way of enumerating differences between two cell populations.

## Introduction

This vignette gives a detailed account on the workflow of UniPath tool for analyzing single cell expression data and single cell open chromatin profiles in pathway domain. UniPath is a steadfast statistical method for getting important biological insights from single cells characterized in terms of pathway activity scores and studying temporal dynamics. UniPath is a scalable platform allowing pre-processing and analysis of thousands of single cells by exploiting heterogeneity among cells and uncovering biologically relevant pathways. UniPath can help users with accurate identification of cell types, signaling pathways and doublet cells. Besides these, the user can also perform clustering and pseudo temporal ordering of single cells in pathway space. This may allow the analysis of relevant pathways and genes on single cell lineage transitions or potency.

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binorm	Conversion of non-zero gene FPKM value into p-value using each cell mean and standard deviation
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## Description

Produces a matrix of p-values

## Details

Based on the assumption that non-zero gene expression FPKM follows log normal distribution, non-zero gene expression data is converted into p-values using mean and standard deviation for individual cells.

## Usage

```
binorm(x)
```

## Arguments

**x**     Gene expression matrix

## Value

n\*p matrix of p-values

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combine	Combine p-values using empirical browns method
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### Description

Produces a matrix of combined p-values

### Details

Empirical browns method is used for combining p-values of genes in a gene-set. It combines p-values of genes which are dependent on each other.

### Usage

```
combine(gene_file, expression_matrix, gnames, Pval1)
```

### Arguments

gene_file	Pathway annotation file/gene-set file
expression matrix	Gene expression matrix
gnames	Gene names of expression matrix
Pval1	P-values matrix obtained from binorm

### Value

n\*p matrix of combined p-values

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adjust	Adjusting of combine p-values using null model
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## Description

Adjustment of combined p-values.

## Details

Combined p-values are adjusted using null model to get final pathway scores. Null model helps in highlighting cell-type specific pathways.

## Usage

```
adjust(combp,combp_ref)
```

## Arguments

combp	Combined p-value matrix obtained using gene expression matrix
combp_ref	Combined p-value matrix obtained using null model

## Value

A list contains:

adjpva	n*p matrix of adjusted p-values
adjpvaraw	n*p matrix of adjusted raw p-values
adjpvalog	n*p matrix of adjusted log transformed p-values

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dist_clust	hierarchical clustering of pathway score matrix
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### Description

Performs hierarchical clustering and gives clusters of samples or cells

### Usage

```
dist_clust(pathwayscores,n)
```

### Arguments

pathwayscores	Log transformed adjusted p-value matrix
n	Number of clusters required for pseudo temporal ordering

### Value

A list contains:

distance	Distance matrix
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clusters	Number of clusters
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index	Indices of top k nearest neighbor
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### Description

Produces matrix of indices of k-nearest neighbour

### Usage

```
index(pathwayscores,k)
```

### Arguments

pathwayscores	Log transformed adjusted p-value matrix
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k                      Number of top k nearest neighbour

#### Value

Matrix of indices of nearest neighbour

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KNN      Getting cluster numbers for each of the nearest neighbor of a cell

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#### Description

Produces a matrix with cluster number for top nearest neighbors for each of the cell

#### Usage

KNN(pathwayscores,index,clusters)

#### Arguments

pathwayscores	Log transformed adjusted p-value matrix
clusters	Clusters obtained from hierarchical clustering

#### Value

n\*p matrix having cluster or class number for each of the top nearest neighbor of individual cell

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**class1** Finding how many times nearest neighbor of cells in each class are belonging to different cluster or class.

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### Description

Produces a matrix with counts of cells belonging same cells have top k nearest neighbor

### Usage

```
class1(clusters,KNN)
```

### Arguments

clusters	Clusters obtained from hierarchical clustering
KNN	Matrix with cluster number for top k nearest neighbors for each of the cell

### Value

n\*n matrix with number of times cells in same class have top k neighbors in other classes

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**distance** Shrunked distance matrix based on two level of shrinkage

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### Description

Two level shrinkage of distance matrix based on nearest neighbour indices and belongingness of cells to same class

### Usage

distance(dist,class,clusters)

### Arguments

dist	Distance matrix used for hierarchical clustering
class	Matrix with number of times cells in same class have top k neighbors in other classes
clusters	Clusters obtained from hierarchical clustering

### Value

Shrunked distance matrix

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minimum\_spanning\_tree Construction of minimum spanning tree

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### Description

Finds minimum spanning tree by creating adjacency graph using shrunked distance matrix

### Usage

```
minimum_spanning_tree(distance)
```

### Arguments

distance	shrunked distance matrix
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### Value

Minimum spanning tree

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mst.plot.mod   Plotting minimum spanning tree using netbioV package in R

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### Usage

UniPath::mst.plot.mod()

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temporalDif   Differential pathways

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### Description

Differential pathways based on Wilcoxon rank sum test

### Usage

temporalDif(data, group)

### Arguments

data	Adjusted raw p-values matrix
group	Group of cell types among which differential pathway analysis needs to be performed

### Value

A list containing p-value based on Wilcoxon rank sum test, fold change based on mean and median

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gradient      Creates gradient of colors

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### Description

Showing gradient or continuum of pathways on minimum spanning tree

### Usage

```
gradient(pathwayfile,term)
```

### Arguments

pathwayfile	Adjusted p-values matrix
term	Pathway term for which gradient needs to be plotted

### Value

Gradient of colors for specific pathway term

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difcoccur      Differential cooccurrence of pathways among two group of cells

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### Description

Differential cooccurrence pathway analysis

### Usage

```
difcoccur(data , group )
```

### Arguments

data	Adjusted raw p-values matrix
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group      Group 1 for cells of interest and rest in group 2

#### Value

A list contains:

pval

n\*n matrix of p-values of pathway pairs

dif

n\*n matrix of p-values of pathway pairs

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drimpute   Imputation

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#### Description

Imputation of scATAC-seq profiles

#### Usage

drimpute(countFile)

#### Arguments

countFile      scATAC-seq count matrix

#### Value

Imputed count matrix

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global_access	Calculating global accessibility score
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### Description

Computes global accessibility scores based on bulk open chromatin profiles

### Usage

```
global_access(testfile,referencefile,globalaccess_scores)
```

### Arguments

testfile	Peak list of test data
referencefile	Peak list of reference data
globalaccess_scores	pre-calculated global accessibility scores

### Value

Matrix of global accessibility scores for test data

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nearest_gene	Generation of foreground file
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### Description

Produces a four column file having genes along with their genomic distances.

### Usage

nearest\_gene (arg1,arg2,arg3,arg4)

### Arguments

arg1	nearestGenes.pl script
arg2	genomic coordinate file
arg3	human reference genome file
arg4	output file

### Value

Matrix having genes along with their genomic distances

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runGO	Calculating pathway enrichment scores for scATAC-seq profiles
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### Description

Produces matrix of hypergeometric and binomial test based pathway enrichment scores respectively.

### Details

UniPath can also help is calculating pathway enrichment scores using scATAC-seq profiles using this function. It gives users to calculate enrichment scores using hypergeometric and binomial test with two options for normalization of data to highlight cell type specific enhancers. User can choose option 1 for normalization using global accessibility scores or option number 2 for local accessibility scores-based normalization of data.

## Usage

```
runGO(gmtFile,BGfile,countFile,method,globalaccessibility_scores,FGfile,promoters = FALSE)
```

## Arguments

gmtFile	gene set file or cell marker-based file
BGfile	background file
countFile	scATAC-seq count matrix
method	If method is chosen as 1, data normalization is performed using global accessibility scores. If selected method is 2 then local accessibility score-based normalization is performed
globalaccessibility_scores	global accessibility scores
FGfile	foreground file
Promoters	whether promoters to be used or not for conversion of scATAC-seq profiles to pathway scores. Default is false

## Value

A list contains:

hypergeometric

n`p matrix of p-values based on hypergeometric test

binomial

n\*p matrix of p-values based on binomial test