#### Overview

UniPath provides robust statistical methods to represent every single cell using pathway and gene-set enrichment scores. It can be used with both single cell RNA-seq and single cell ATAC-seq profile with scalability for atlas scale data-sets. UniPath comes with several features like pseudo-temporal ordering using pathway scores and unconventional way of enumerating differences between two cell populations.

#### Introduction

This vignette gives a detailed account on the workflow of UniPath tool for analyzing single cell expression data and single cell open chromatin profiles in pathway domain. UniPath is a steadfast statistical method for getting important biological insights from single cells characterized in terms of pathway activity scores and studying temporal dynamics. UniPath is a scalable platform allowing pre-processing and analysis of thousands of single cells by exploiting heterogeneity among cells and uncovering biologically relevant pathways. UniPath can help users with accurate identification of cell types, signaling pathways and doublet cells. Besides these, the user can also perform clustering and pseudo temporal ordering of single cells in pathway space. This may allow the analysis of relevant pathways and genes on single cell lineage transitions or potency.

binorm

Conversion of non-zero gene FPKM value into p-value using each cell mean and standard deviation

#### Description

Produces a matrix of p-values

#### **Details**

Based on the assumption that non-zero gene expression FPKM follows log normal distribution, non-zero gene expression data is converted into p-values using mean and standard deviation for individual cells.

#### **Usage**

binorm(x)

#### **Arguments**

**x** Gene expression matrix

#### Value

n\*p matrix of p-values

combine

Combine p-values using empirical browns method

## Description

Produces a matrix of combined p-values

#### **Details**

Empirical browns method is used for combining p-values of genes in a gene-set. It combines p-values of genes which are dependent on each other.

## Usage

combine(gene\_file, expression\_matrix,gnames,Pval1, thr=2)

#### **Arguments**

gene\_file Pathway annotation file/gene-set file

expression matrix Gene expression matrix

gnames Gene names of expression matrix
Pval1 P-values matrix obtained from binorm

thr Based on threshold provided, those gene sets having number of genes

greater than the threshold value will to be considered for covariance matrix

calculation and combining of p values. Default value is 2.

## Value

n\*p matrix of combined p-values

adjust

Adjusting of combine p-values using null model

## Description

Adjustment of combined p-values.

#### **Details**

Combined p-values are adjusted using null model to get final pathway scores. Null model helps in highlighting cell-type specific pathways.

#### Usage

adjust(combp,combp\_ref)

## Arguments

combp Combined p-value matrix obtained using gene expression matrix

combp\_ref Combined p-value matrix obtained using null model

#### Value

A list contains:

adjpva

n\*p matrix of absolute p-values

adjpvaraw

n\*p matrix of raw adjusted p-values

adjpvalog

n\*p matrix of adjusted log transformed p-values

dist\_clust hierarchi

hierarchical clustering of pathway score matrix

## Description

Performs hierarchical clustering and gives clusters of samples or cells

Usage

dist\_clust(pathwayscores,n)

## **Arguments**

pathwayscores Log transformed adjusted p-value matrix

n Number of clusters required for pseudo temporal ordering

Value

A list contains:

distance

Distance matrix

clusters

Number of clusters

index Indices of top k nearest neighbor

#### Description

Produces matrix of indices of k-nearest neighbour

## Usage

index(pathwayscores,k)

### Arguments

pathwayscores Log transformed adjusted p-value matrix k Number of top k nearest neighbour

#### Value

Matrix of indices of nearest neighbour

KNN Getting cluster numbers for each of the nearest neighbor of a cell

### Description

Produces a matrix with cluster number for top nearest neighbors for each of the cell

#### **Usage**

KNN(pathwayscores,index,clusters)

#### **Arguments**

pathwayscores Log transformed adjusted p-value matrix clusters Clusters obtained from hierarchical clustering

#### Value

n\*p matrix having cluster or class number for each of the top nearest neighbor of individual cell

class1 Finding how many times nearest neighbor of cells in each class are belonging to different cluster or class.

#### Description

Produces a matrix with counts of cells belonging same cells have top k nearest neighbor

#### **Usage**

class1(clusters,KNN)

## **Arguments**

KNN Matrix with cluster number for top k nearest neighbors for each of the cell

#### Value

n\*n matrix with number of times cells in same class have top k neighbors in other classes

distance Shrinked distance matrix based on two level of shrinkage

## Description

Two level shrinkage of distance matrix based on nearest neighbour indices and belongingness of cells to same class

#### **Usage**

distance(dist, class, clusters)

## **Arguments**

dist Distance matrix used for hierarchical clustering

class Matrix with number of times cells in same class have top k neighbors in other

classes

clusters Clusters obtained from hierarchical clustering

## Value

Shrinked distance matrix

# Description

Finds minimum spanning tree by creating adjacency graph using shrinked distance matrix

### Usage

minimum\_spanning\_tree(distance)

## Arguments

distance shrinked distance matrix

Value

Minimum spanning tree

mst.plot.mod Plotting minimum spanning tree using netbiov package in R

Usage

UniPath::mst.plot.mod()

temporaldif Differential pathways

### Description

Differential pathways based on Wilcoxon rank sum test

### Usage

temporaldif(data, group)

## **Arguments**

data Adjusted raw p-values matrix

group Group of cell types among which differential pathway analysis needs to be

performed

Value

A list containing p-value based on Wilcoxon rank sum test, fold change based on mean and median

gradient Creates gradient of colors

## Description

Creating gradient of colors for showing continuum of single pathway on minimum spanning tree

#### **Usage**

gradient(pathwayfile,term)

## **Arguments**

pathwayfile Adjusted p-values matrix

term Pathway term for which gradient needs to be plotted

#### Value

Gradient of colors for specific pathway term

makecolor Creates gradient of colors

## Description

Creating gradient of colors for showing continuum of co-occurrence of pathways on minimum spanning tree

## Usage

makecolor (score1, score2)

## **Arguments**

score1 scores of first pathway term score2 scores of second pathway term

#### Value

Matrix of gradient of colors for two pathway terms to be visualized on minimum spanning tree

Counttofpkm Conversion of read count data to fpkm

## Description

Converts raw read count data into fpkm

#### **Usage**

## Counttofpkm (countMatrix,length)

## Arguments

countMatrix Raw read count matrix

length of genes. Order of length of genes and row names of count data should

be same

### Value

n\*p matrix fpkm values

difcoccur

Differential cooccurrence of pathways among two group of cells

#### Description

Differential cooccurrence pathway analysis

## Usage

```
difcoccur(data , group )
```

## **Arguments**

data Adjusted raw p-values matrix

group Group 1 for cells of interest and rest in group 2

#### Value

A list contains:

pval

n\*n matrix of p-values of pathway pairs

dif

n\*n matrix of p-values of pathway pairs

drimpute Imputation

## Description

Imputation of scATAC-seq profiles

## Usage

drimpute(countFile)

# Arguments

countFile scATAC-seq count matrix

#### Value

Imputed count matrix

global\_access

Calculating global accessibility score

## Description

Computes global accessibility scores based on bulk open chromatin profiles

## Usage

global\_access(testfile,referencefile,globalaccess\_scores)

## **Arguments**

testfile Peak list of test data referencefile Peak list of reference data

globalaccess\_scores pre-calculated global accessibility scores

# Value

Matrix of global accessibility scores for test data

nearest\_gene Generation of foreground file

#### Description

Produces a four column file having genes along with their genomic distances.

# Usage

```
nearest_gene (arg1,arg2,arg3,arg4)
```

### Arguments

arg1	nearestGenes.pl script
arg2	genomic coordinate file
arg3	human reference genome file
arg4	output file

#### Value

Matrix having genes along with their genomic distances

runGO Calculating pathway enrichment scores for scATAC-seq profiles

## Description

Produces matrix of hypergeometric and binomial test based pathway enrichment scores respectively.

#### **Details**

UniPath can also help is calculating pathway enrichment scores using scATAC-seq profiles using this function. It gives users to calculate enrichment scores using hypergeometric and binomial test with two options for normalization of data to highlight cell type specific enhancers. User can choose option 1 for normalization using global accessibility scores or option number 2 for local accessibility scores-based normalization of data.

# Usage

runGO(gmtFile,BGfile,countFile,method,globalaccessibility\_scores,FGfile,promoters= FALSE, dist=1000000,threshold=1.25)

#### **Arguments**

gmtFile gene set file or cell marker-based file

BGfile background file

countFile scATAC-seq count matrix

method If method is chosen as 1, data normalization is performed using

global accessibility scores. If selected method is 2 then local

accessibility score-based normalization is performed

globalaccessibility\_scores global accessibility scores

FGfile foreground file

Promoters whether promoters to be used or not for conversion of scATAC-

seq profiles to pathway scores. Default is false

distance to be used for considering nearest gene to a peak.

Default is 1000000

threshold Peaks above the given threshold value are chosen to be in a

foreground set

#### Value

A list contains:

hypergeometric

n`\*p matrix of p-values based on hypergeometric test

binomial

n\*p matrix of p-values based on binomial test