# CLINICAL ADVANCEMENT FORECASTING \*

#### **Authors TBD**

Related Sciences

#### **ABSTRACT**

This study examines the extent to which the outcomes of clinical trials can be predicted based on longitudinal properties of drug targets and diseases alone. We find that this is possible by comparing the historical performance of model-based target-disease pair prioritization methods to common baselines. Our primary objective is to demonstrate that statistical learning can effectively optimize such methods with no loss of interpretability. For example, non-negative linear models can produce simple weighting schemes across various types of human, animal and cell model evidence (for targets, diseases and pairings of the two) to identify target-disease pairs that advance beyond phase 2 trials with an average relative risk that is 2x higher than Open Targets composite scores. Other key characteristics of this study include: 1) a comprehensive longitudinal treatment of evidence as well as how it relates to leakage and reverse causality in biomedical research, 2) trial and/or drug details are not used in order to enable ranking for undeveloped targets/diseases, 3) analysis of the space of currently undeveloped, tractable targets with the highest likelihood of clinical success, 3) no data is used outside of Open Targets to ease reproduction and/or deployment, and 4) our method requires no expert knowledge and can easily support the inclusion of more lines of evidence over time, making it easy to operationalize.

#### 1 Introduction

It has been well established that drugs with human genetic evidence linking their respective targets to indications in clinical trials are more likely to succeed [1, 2, 3, 4, 5, 6, 7, 8]. This information has been used to devise many target and target-disease ranking algorithms based primarily on a synthesis of multiple genetic signals alone [9, 10, 11]. It is also possible to expand the breadth of this genetic support to more targets/diseases based on knowledge graphs, protein interactions and/or disease ontologies [12, 13, 14, 15, 16]. To our knowledge, all such expansion methods identify a larger space of opportunities at the expense of expected success rates. This is not a focus of this work as we aim, instead, to establish a framework for identifying targets/diseases with the very highest possible likelihood of success first. This is accomplished by integrating human clinical, genetic/genomic, transcriptomic and proteomic data as well as cell/animal model evidence, pathway information and basic literature metrics from Open Targets [10] in a simple statistical modeling framework. It can be contrasted with far more integrative methods that rely on neural and/or graph models over extensive knowledge graphs [17, 18, 19, 20], which are more complex and difficult to interpret. We believe a desirable middle ground between these approaches and those that aim to combine many orthogonal indicators of success through expert knowledge in heuristic systems [21, 10] would: 1) permit inclusion of many types of evidence from many sources, 2) be highly interpretable, 3) support expert judgement where necessary, and 4) not require manual ranking/weighting schemes.

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A substantial challenge inherent to building such a system is the need to account for the longitudinal nature of knowledge discovery in biomedicine. This is vital because any method that optimizes for likely future clinical success based on historical clinical success may easily be biased by the non-random nature with which evidence is absent otherwise. We use only "temporalized" evidence, i.e. evidence for which timing of its emergence can be determined, and outcomes when training and evaluating our methods before ultimately applying them to present-day evidence with no restrictions on timing. We discuss motivations, prior research and our own analysis on how important this problem is for each source in Section 2.6.

Addressing such problems is common, but not ubiquitous, in studies that attempt to predict clinical trial outcomes using temporalized predictors [22, 23, 24]. The need for this is often clear in that setting where the inclusion of predictors like historical success rates for targets/diseases, trial sponsor track records, eventual patient enrollment, etc. constitute clear information leaks otherwise. This is discussed more in [22] which notes several studies that do not account for this problem, and presents "quasi-prospective" as well as true "prospective" results. The difference between the two is that the former reconstructs timelines for predictors and outcomes based on recorded event dates while the latter relies on frozen predictions that are never evaluated until years later. Nomenclature for these formulations is conflicting though, where this definition of a "quasi-prospective" design is deemed entirely prospective in some cases, e.g. [25]. We will refer to our design in this study as quasi-prospective since this definition is the best fit.

The prior works discussed so far can largely be categorized as either 1) target and target-disease prioritization methods evaluated based on how well they correlate with clinical trial success and 2) clinical trial outcome prediction models. Both are measured against the same outcomes and an important distinction between them lies in how the former methods are **not** directly optimized for those outcomes while the latter methods are. In this study, we attempt to bridge these methodologies by predicting clinical trial advancement for target-disease pairs based solely on information that would be present well in advance of any drug program or individual trial. We then calibrate these predictions to determine what thresholds are necessary to match the observed success rates from benchmarks for genetic support like OMIM [26], ClinVar [27] and GWAS. Finally, we examine how many present-day target-disease pairs are undeveloped (i.e. have never been in clinical trials), have a tractable target and are likely to see success rates matching or exceeding those calibrated benchmarks.

#### 2 Results

In order to model clinical advancement for target-disease (TD) pairs, we first define "advancement" as progression beyond any particular trial phase across all drugs associated with any one TD pair as indicated by the presence of a later-stage trial. All results to follow consider only advancement beyond phase 2 due to limitations described in Section 4. This binary outcome is then predicted based on a list of features shown in Table 1. Information for each of these features is only used when it was published before the year **prior** to the first phase 2 trial observed, with an exception for genetic evidence discussed in Section 2.6. A training dataset is then formed by including only TD pairs where this first phase 2 year is between 1990 and 2015. The evaluation dataset then consists of all TD pairs entering phase 2 between 2016 and 2022, with a 2 year offset from the present year (2024) to allow enough time for some trials to complete. While the average phase 2 trial duration may be as low as 2 years [28], other estimates would suggest half of them take longer than 2.9 years [29]. This means a substantial fraction of outcomes are censored, that this is an important parameter to test sensitivity to and that time itself is likely to be a crucial covariate in this formulation. The distribution of these outcomes, the number of associated targets/diseases and a variety of other statistics on this dataset are presented in Supplementary Figure 7.

#### 2.1 Features

The features used throughout this study consist of 27 target-disease pair predictors, 5 target-specific predictors and 1 disease-specific predictor. These are listed in Table 1. The target and disease specific features are chosen carefully such that they are either capable of being associated with years in which events supporting them occurred or result from large-scale, unbiased methods that do not favor well-studied or drugged targets/diseases. Examples of this include

target-specific tissue expression specificity scores computed from Human Protein Atlas [30] and LOEUF [31] scores from gnomAD. Simply put, our dataset combines scores from Open Targets for target-disease evidence and a select subset of target prioritisation [32] fields with almost no modifications, other than to add target and disease specific indicators of maximum trial phases reached and two extra genetic association features.

#### 2.2 Models

We train a variety of models including constrained and unconstrained linear and tree models. The constrained variants of these models force effects of all features to increase monotonically, i.e. all effects are constrained to be non-negative. This is possible with no underlying feature transformations because all scores in Open Targets are constructed such that higher scores are presumed to be advantageous.

We also apply these models to our evaluation dataset using several feature ablations in order to assess the value of groups of related features. We refer to a "core" feature set consisting of all features listed in Table 1 except for the sole feature capturing the time since a target-disease pair first entered phase 2 trials (i.e. target\_disease\_\_time\_\_transition). Combinations of learning algorithms for the models and the feature sets to which they are applied are referred to using the following convention:

- RDG: Constrained, L2-regularized linear regressor (a.k.a. "Ridge regressor") fit with all core features
- **RDG-T**: **RDG** fit with all features instead of only core features, where the only difference is the inclusion of time since phase 2 transition for a TD pair
- RDG-X: RDG fit without human clinical and genetic evidence features
- GBM: Constrained, gradient-boosted machine with fit with all core features
- GBM-T: GBM fit with all features instead of only core features
- OTS: Open Targets composite score

The omitted human clinical and genetic evidence features for the RDG-X model are all of those in Table 1 with the midfix "clinical" or "genetic\_association". We omit these features specifically because they are known or expected to comprise good predictors of human clinical success, so their exclusion examines the extent to which only literature and animal model evidence along with target-specific properties accomplish this task.

In order to compare these models to an Open Targets composite score (OTS), we use an equally weighted sum of all scores from individual sources except for those assigned lower weights in [33]. Scores from these sources are multiplied by the corresponding weight before being summed and only the TD-specific features of Table 1 are used. Neither the target/disease specific features nor the time since phase 2 transition feature are included in this calculation.

#### 2.3 Performance

#### 2.3.1 Open Targets comparison

Figure 1 demonstrates how well our primary model in this study, RDG, ranks TD pairs by comparison to a composite score from Open Targets, OTS. The relative risk (RR) metric used in this comparison as well as all others where not specified otherwise is defined as:

$$\frac{P(advancement|rank >= N)}{P(advancement|rank < N)}.$$
(1)

The third ranking method presented in Figure 1, "RDG-T", differs from the RDG model only in that it uses time since the phase 2 transition as a predictive factor in addition to all others. We observe that the use of this information almost universally improves standard performance metrics like receiver operating characteristic (ROC) and average precision (AP), however it adds little to no value in rankings beyond a level where substantial relative risk increases

can be observed. In other words, it constitutes an effective but coarse mechanism for ranking TD pairs while lacking the high precision of other factors like genetic support. More implications of this and opportunities it may imply are discussed in Section 4. As a more practical concern, we refrain from focusing on RDG-T, or the similar GBM-T model, because neither is readily applicable to undeveloped TD pairs for which the time since phase 2 transition is not available. They do, however, present a useful performance ceiling towards which future work might build.

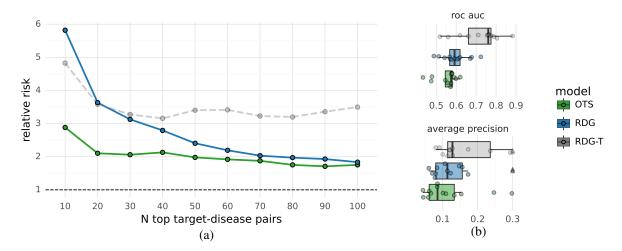


Figure 1: **Performance compared to Open Targets composite scores**. (a) Equally-weighted average relative risk estimates across 13 therapeutic areas, by number of top rankings and 3 methods: RDG (ours), RDG-T (ours) and OTS (Open Targets composite scores). (b) Receiver operating characteristic (ROC) and average precision scores across the same 13 therapeutic areas with no limit on the number of rankings. See Supplementary Figure 10 for raw data underlying (a).

We also note that Figure 1 presents average RR estimates drawn across a subset of therapeutic areas, and the criteria used to select them is described more in Section 5. A full list of therapeutic areas meeting these criteria can be seen in Supplementary Figure 10 along with the RR estimates summarized in Figure 1. Furthermore, a comparison of the distribution of these estimates by model is presented in Supplementary Figure 11 along with the statistical significance of their differences.

#### 2.3.2 Genetic benchmark comparison

In order to establish baseline levels of success and coverage across TD pairs, we examine ranking performance in comparison to well established, univariate indicators of genetic support in Figure 2. This figure presents OMIM and GWAS baselines, in the parlance of [2], [1] and [3], as well as an intermediate baseline from the European Variation Archive (EVA) [34] containing evidence predominantly from ClinVar [27].

One key objective of this study is to determine if any model, e.g. RDG, can sort TD pairs with genetic support such that at least some portion of that sorted list has a likelihood of advancement that consistently exceeding what is expected from any one source of genetic support alone. We find that this goal is met and exceeded by the RDG model, which actually identifies more TD pairs than those that have either EVA or GWAS support alone at an expected rate of advancement exceeding that of the single source (respectively). This does not appear to be the case with the OMIM baseline, however the lack of examples in our evaluation dataset with OMIM support makes any determination difficult. See Section 2.5 for more on how these benchmarks are employed to contextualize opportunities among undeveloped TD pairs and Supplementary Figure 13 for top predictions from the RDG model. This latter, supplementary figure further emphasizes our focus on prioritizing opportunities beyond those with genetic support and provides examples of TD pairs with multiple lines of evidence.

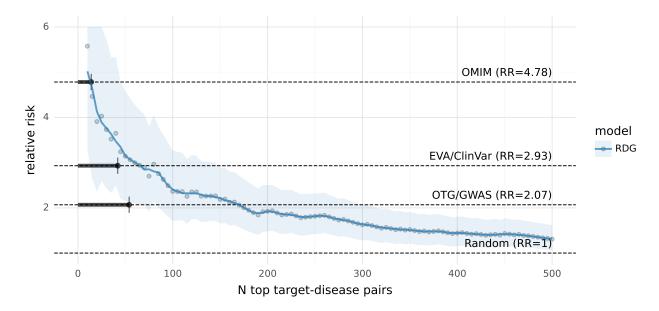


Figure 2: **Performance compared to genetic support benchmarks**. The RR estimates for each benchmark are based on the presence of the corresponding support across all TD pairs, and the number of pairs for which is present is represented by the horizontal bars extending horizontally from the y-axis. The bounds around the RDG RR estimates correspond to a Katz 90% confidence interval.

We also note that Supplementary Figure 8 shows confidence intervals for each of the genetic benchmarks of Figure 2 in isolation, as well as all other target-disease-specific evidence sources, in addition to confidence intervals for the RDG model at various top ranking cutoffs. Similar comparisons for target-specific and disease-specific features can be seen in Supplementary Figure 9. These findings suggest that 1) genetic support for TD pairs is highly predictive but rare, 2) human clinical support for targets and diseases in isolation is also predictive while being more common and 3) constraint and expression specificity of targets exhibit modest but significant effects. While only human clinical evidence appears to have a prognostic value rivaling that of genetic support when considered on a univariate basis, the combined influence of non-genetic, non-clinical evidence is examined in Section 2.3.3 where a model using only this information, RDG-X, still outperforms Open Targets composite scores by all measures.

#### 2.3.3 Model comparison

Figure 3 presents average performance across therapeutic areas for select combinations of learning algorithm, constraint type and feature group described in Section 2.2. Several key findings illustrated in this figure are:

- 1. The RDG-T model achieves far higher ROC and AP scores through the use of the time since transition feature, which indicates the number of years a TD pair has been underdevelopment after having reached phase 2.
- 2. The RDG model, however, matches or exceeds RDG-T in performance among top TD pairs
- 3. The RDG-X model, using no human clinical or genetic evidence linked to a disease, outperforms the Open Targets composite score and nearly matches the performance of the RDG model beyond top rankings
- 4. Linear models outperform gradient-boosting models by nearly all measures
- 5. Constrained linear models outperform unconstrained linear models by nearly all measures

We conclude from these results that constrained linear models are an optimal choice for this problem due both to their greater performance and interpretability. This interpretability is illustrated more in Section 2.4 and owed much to

RDG-T	0.21	0.72	4.83	3.58	3.28	3.16	3.40	3.50	(a)
RDG	0.14	0.60	5.82	3.63	3.13	2.79	2.41	1.84	feature ablations
RDG-X	0.12	0.57	4.27	2.75	2.71	2.75	2.38	1.94	([+] models)
OTS	0.11	0.55	2.88	2.10	2.06	2.13	1.98	1.75	
RDG[+]	0.14	0.60	5.82	3.63	3.13	2.79	2.41	1.84	(b)
RDG[+/-]	0.14	0.59	5.04	3.53	3.13	2.53	2.33	1.86	
GBM[+]	0.13	0.54	4.09	3.34	2.89	2.63	2.54	1.97	model algorithms (core features)
GBM[+/-]	0.13	0.58	4 <mark>.4</mark> 4	3.29	2.64	2.41	2 <mark>.2</mark> 4	1.70	
OTS	0.11	0.55	2.88	2.10	2.06	2.13	1.98	1.75	
	AP	ROC	RR@010	RR@020	RR@030	RR@040	RR@050	RR@100	_

Figure 3: **Performance across model algorithms and feature ablation groups**. Average precision (AP) and receiver operating characteristic (ROC) scores with relative risk (RR) at ranking cutoffs denoted by RR@N. (a) Performance across constrained RDG models using different groups of features as described in Section 2.2 (b) Performance across constrained ([+]) and unconstrained ([+/-]) linear and gradient-boosted models using the core feature set.

the effort Open Targets has already undertaken to construct evidence scores such that they can be assumed to have a monotonically increasing effect on the likelihood that a causal relationship exists between a target and a disease.

## 2.4 Effects

The coefficients learned by the RDG model, and the average effects they have across the evaluation dataset, are shown in Figure 4. This model most highly prioritizes genetic signals that have the greatest coverage, i.e. associations from GWAS studies through the ot\_genetics\_portal feature and associations from any curated clinical genetics source, i.e. EVA, Orphanet, UniProt, Genomics England, ClinGen and gene2phenotype, via the curated feature. Notably, literature and target/disease specific clinical features also have substantial effects, followed by indicators of animal evidence and target genetic constraint / expression specificity. Any features not shown were deflated to have no effect, which is possible in this model due to the non-negativity constraint. One such feature worth emphasizing is transcriptomic evidence from Expression Atlas. We found this somewhat surprising, but it is supported by arguments against transcript over/under expression as an indicator of genes that influence disease rather than the other way around [35].

It is worth noting that the discordance between the coefficients and the average feature effects of Figure 4 arises from both the frequency with which features exist and the distribution of their underlying scores. Scores for many clinical genetics features (e.g. OMIM, Genomics England, UniProt) are very frequently absent or close to 1. By comparison, scores for literature associations are typically far lower, even when limited only to cases where they exist, with a median value of 0.12 (mean=.23) in the evaluation data. This is why the europepmo feature has a relatively large associated coefficient and a much smaller average effect on predictions.

## 2.5 Opportunities

A common method for identifying druggable opportunities within a specific disease context involves first ranking TD pairs according to some prioritization methodology followed by filtering or reprioritizing those ranks based on knowledge of target tractability [36, 37, 11]. We use a similar approach to identify tractable targets associated with TD pairs that have yet to enter clinical trials. To aid in interpreting this approach, we also draw on the results of Figure 2. The data in this figure suggests thresholds for the RDG model that align to expected rates of advancement compared to

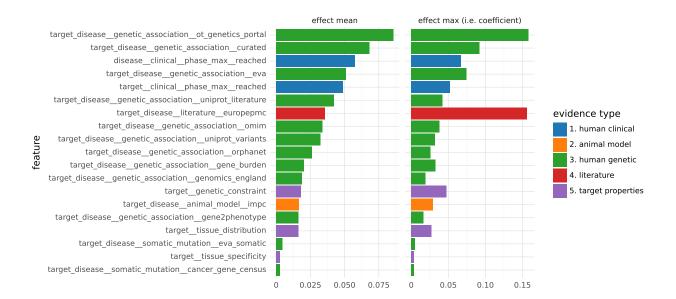


Figure 4: **RDG model feature effects**. The **effect max** values are equivalent to RDG model coefficients for the corresponding feature while the **effect mean** values indicate average values of the product between the coefficient and a particular feature value, when that feature is present.

several genetic support benchmarks. These thresholds are used to bucket undeveloped TD pairs before further bucketing them based on levels of tractability. The tractability buckets in Supplementary Table 2 provide HIGH, MED, and LOW confidence ratings for each type of tractability evidence based on the priorities suggested in [38].

Figure 5 shows the distribution of TD pair counts for select buckets across therapeutic areas as well as across the current maximum phase reached for any one pair. We find that there are  $\sim$ 2,400 small-molecule-enabled,  $\sim$ 1,400 antibody-enabled, and 14 PROTAC-enabled TD pairs with a probability of advancement that is nearly 3x other TD pairs based on the EVA threshold RR=2.93 in Figure 2. Top antibody-enabled pairs are shown in Supplementary Figure 14 along with their corresponding genetic and clinical support.

## 2.6 Inflation

Like most studies of this kind, we assume a "closed-world" [17] over the space of target-disease pairs and any evidence between them. This means that we do not differentiate between evidence that an association for any one pair truly does **not** exist (or is too weak to be relevant under the omnigenic model [39]), and the lack of any attempt to find that evidence in the first place. This also means that our estimate of the prognostic value for any one evidence source is subject to historical trends in biomedical research and the myriad ways that this research can be biased towards particular targets and diseases. We avoid attempting to comprehensively survey these biases in favor of offering an illustrative list of specific examples that are relevant in this study:

- 1. Mendelian randomization research is biased towards cardiovascular diseases as they have a disproportionate number of known, modifiable exposures [40]
- 2. Putative protein interactions that do not result from genome-scale or otherwise unbiased assays result in an overrepresentation of successful drug targets in resources like STRING [41], thereby inflating the success of network expansion methods over these databases to identify such targets [14].
- 3. Transcript expression studies run in late-stage clinical trials for a single indication, e.g. [42] linking SLE to IFN genes, are a degenerate indicator of advancement beyond earlier stage trials when the timing of this evidence is not accounted for.

	stage [threshold=EVA]							threshol		tractability [stage=NONE, confidence=HIGH, threshold=EVA]					
	ALL	NONE	Phase 1	Phase 2	Phase 3	Phase 4	EVA	ОМІМ	отс	АВ	ос	PR	SM		
therapeutic area															
ALL	9724	8560	165	367	274	358	8560	679	20821	1456	655	14	2408		
genetic, familial or congenital disease	4867	4638	32	73	58	66	4638	402	7641	399	271	12	993		
cancer or benign tumor	2063	1531	122	209	125	76	1531	96	6506	430	124	0	639		
nervous system disease	2093	1923	10	54	49	57	1923	166	3710	153	74	3	446		
musculoskeletal or connective tissue disease	1597	1469	15	46	35	32	1469	158	2863	206	129	0	353		
gastrointestinal disease	1149	995	22	48	30	54	995	82	3265	257	67	0	334		
immune system disease	1294	1114	30	69	36	45	1114	107	2762	339	69	4	290		
nutritional or metabolic disease	1491	1409	1	16	16	49	1409	132	2316	94	29	0	310		
endocrine system disease	1071	933	17	42	26	53	933	99	2685	184	80	4	287		
cardiovascular disease	956	834	7	25	32	58	834	81	1908	126	70	0	315		
psychiatric disorder	820	735	2	22	16	45	735	73	1442	59	18	2	168		
disorder of visual system	838	823	1	8	3	3	823	87	1289	40	24	0	131		
integumentary system disease	733	646	14	35	22	16	646	51	1502	115	40	0	164		
hematologic disease	702	589	23	41	31	18	589	45	1394	158	73	1	202		
respiratory or thoracic disease	590	472	8	52	33	25	472	23	1552	177	45	0	134		
urinary system disease	550	490	5	19	19	17	490	42	1164	94	47	0	121		
reproductive system or breast disease	534	452	9	30	18	25	452	33	1240	69	22	6	128		
phenotype	353	307	3	10	10	23	307	13	921	74	54	0	142		
pancreas disease	326	278	7	9	7	25	278	37	770	65	24	0	96		
measurement	7	7	0	0	0	0	7	1	69	1	2	0	3		
disorder of ear	5	5	0	0	0	0	5	0	21	1	0	0	1		

Figure 5: **Present-day target-disease pair counts by stage, likelihood of advancement and tractability**. The **stage** panel contains counts by maximum trial phase reached, the **threshold** panel contains counts of pairs with a RDG model score exceeding that of the associated benchmark for only undeveloped pairs, and the **tractability** panel shows pair frequencies among undeveloped pairs exceeding the EVA threshold that also have a HIGH tractability rating as defined in Supplementary Table 2. This corresponds to targets that have all been in clinical development already, except for the **OC** modality in which case it indicates that a target has been approved.

- 4. Targets tested against more indications in clinical trials enrich for failures because the marginal cost of testing more indications decreases, but the evidence for these indications is often weaker [12].
- 5. Herding effects in pharma R&D pipelines around particular drug targets are becoming increasingly clear over time [43] and generate an excess of clinical evidence for those targets.

We also note that the skew in basic drug target research towards those that already have rich annotations and well characterized molecular function [44] as well as the disproportionate representation of particular target families in pharma R&D pipelines [45, 46] and the fact that literature is well known to be biased away from negative results in general [47] are all problematic.

While it is not possible to address all of these issues, we emphasize that there is a clear pattern across the examples in the list above in that they require **past** clinical successes and/or failures to arise in the first place. This suggests that accounting for when evidence first emerged would limit the extent of these problems. We do so in this study based solely on publication dates associated with any one piece of information linking target-disease pairs. This also offers a novel opportunity to attempt to quantify what kind of evidence suffers most from these biases. Figure 6 presents results for this based on a relative risk statistic defined as:

$$\frac{P(A|B)}{P(A|\neg B)}\tag{2}$$

where:

- A is the event that evidence for a TD pair arises after its first early-stage (phase 1 or 2) trial rather than before
- B is the event that a TD pair advances into late-stage trials (phase 3 or 4)

We refer to this as "inflation risk" so as not to confuse it with the relative risk statistic used in all other contexts, and it can be more simply described as the fraction of TD pairs for which evidence arises **after** the beginning of an ultimately successful early-stage trial divided by that same fraction for TD pairs that do not advance to late-stage trials. The intuition for this statistic is that it will be higher if successful trials lead to the generation of evidence of a particular type, and it should be 1 in cases where the emergence of evidence is independent of clinical success. We also measure this potential lack of independence through the more commonly used Fisher's exact test, e.g. [48], and both are presented in Figure 6.

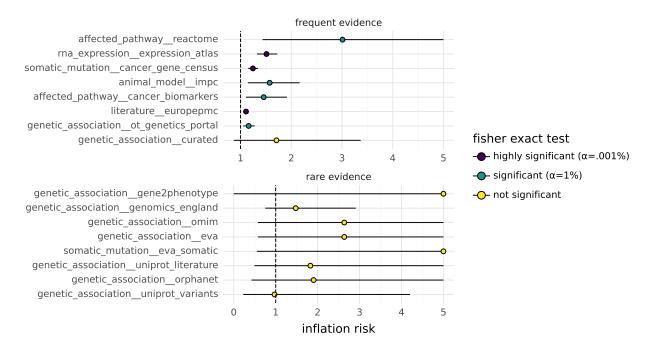


Figure 6: Clinical success drives evidence discovery.

We find that evidence from Reactome is the worst offender by this metric, implying that it often only arises for TD pairs after a certain level of clinical success has been attained. We also find that long-running aggregators/curators of published research often focused on individual diseases/phenotypes, like Expression Atlas, IMPC, CGS and Cancer Biomarkers exhibit this form of inflation as well.

Sources of genetic evidence appear to be much less inflated, or have too little data to reach significance. This is to be expected for GWAS evidence arising from genome-wide, phenome-wide biobank consortia, however much of historical GWAS evidence is not phenome-wide. More context on how much this is likely to matter comes from [49] in which it was estimated that as few as 6% of 500 FDA-approved targets for non-cancer drugs arose from programs highly motivated by pre-existing genetic support and that "the remaining 94% were probably identified using conventional pharmacology, biochemistry or molecular biology approaches". We then speculate that if the initiation of new drug programs was not historically motivated highly by the existence of genetic support, then the incentives for pursing new genetic evidence based on clinical and commercial success are likely to be minimized. This, in conjunction with existing precedent [1, 2, 3, 4, 5, 7] and our inflation results, ultimately led us to the use of genetic evidence without temporalization. In other words, we do not treat genetic evidence as longitudinal features like all others associated with TD pairs. A breakdown of which features are treated in which manner is provided in Table 1.

#### 3 Conclusion

We have demonstrated that simple machine learning methods applied to longitudinal biomedical evidence from many sources can be used to predict clinical outcomes for combinations of drug targets and diseases, without knowledge of molecular properties or trial design details. We have also shown that these methods are more precise in the extremes of their predictions than composite, heuristic scores like those from Open Targets. They also outperform such baselines by more comprehensive, traditional measures of classifier performance; however, we find this less compelling and easier to accomplish than improving performance among the upper tail of the opportunities implied by the very highest predictions. This framework would also support the addition of new lines of evidence over time well as it is designed to automatically determine the relevance of any new information without intervention. Lastly, we find that the space of present-day, undeveloped targets within a disease context that both exceed baseline levels of tractability and have a high predicted likelihood of clinical advancement is substantial. It is likely to grow quickly as well since the breadth of much of the underlying evidence is expanding rapidly [50, 51, 52].

#### 4 Discussion

- Cover limitations with OT concerning temporalization for both evidence and drug approvals, and why only transitions from phase 2 are relevant for this work
- Discuss the possibility to do prospective evaluation with OT snapshots
- Discuss how typical classifier metrics are inadequate for this problem
- Expand on how the inclusion of time greatly improves tail performance and mention that this is consistent with its nature as a necessary but not sufficient condition for success, and it is likely that much of the value it adds in the tail of lower rankings could be captured and enhanced if other early indicators for the many reasons trials fail [4] were also included (see more on this in Section 4)
- We are focusing on prioritizing among targets/indications with genetic support, rather than expanding this space to find opportunities with weaker support
- "There is evidence that a 9.6% vs. 13.8% success rate for drugs from phase 1 trials to approval may mean a \$480 million difference in the median research and development cost required to bring a new drug to the market (Wouters, McKee, and Luyten 2020)." [53]
- From [12]: "It is important to bear in mind therefore that what we are measuring when looking at historical trial outcomes is not an unbiased measure of any given gene's true disease associations, but rather a view on how useful a given evidence source or analytical method has been for choosing drug targets based on current and historical drug discovery practices. Dramatic changes in these practices in the future could render some of our conclusions obsolete, though the fundamental observation that genetic association itself is retained in molecular networks will remain valid."
- Add select tractability and DepMap features?

## 5 Methods

- Discuss why all target prioritisation data fields are not used due to the potential leakage they may impose (e.g. target families, GO annotations, etc.)
- Describe how therapeutic areas were selected based on having at least 100 TD pairs with target-disease-specific
  evidence of any kind and with explicit omissions: ("biological\_process", "pregnancy or perinatal disease",
  "injury, poisoning or other complication", "pregnancy or perinatal disease", "medical procedure", "infectious
  disease", "animal disease")
- RDG models are fit using the Ridge implementation from scikit-learn and GBM models are fit using the LightGBM [54] algorithm

- Training/evaluation results are limited by temporalization while the present-day predictions are not
- OMIM is defined as EVA associations with publications
- The "genetic\_association\_\_curated" field is a union of all genetic association sources other than "gene\_burden" and "ot\_genetics\_portal"
- Mention that the scores for a TD pairs in a year are the maximum score for that source, not the harmonic sum
- Mean imputation is used for target-specific features
- Mention specifics on RDG and GBM implementations, i.e. lightGBM and scikit-learn
- RDG is always trained on all features, but only applied to feature subsets where relevant

## 6 Appendix

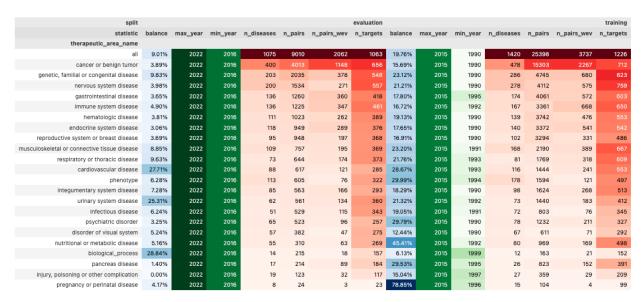


Figure 7: Training and evaluation dataset summary statistics.

Table 1: Features used in modeling and analysis

	feature	entity	kind
1	diseaseclinicalphase_maxreached	disease	temporal
2	targetclinicalphase_maxreached	target	temporal
3	targetgenetic_constraint	target	static
4	targetmouse_ko_score	target	static
5	targettissue_distribution	target	static
6	targettissue_specificity	target	static
7	target_diseaseaffected_pathwaycancer_biomarkers	target_disease	temporal
8	target_diseaseaffected_pathwaycrispr	target_disease	temporal
9	target_diseaseaffected_pathwaycrispr_screen	target_disease	temporal
10	target_diseaseaffected_pathwayprogeny	target_disease	temporal
11	target_diseaseaffected_pathwayreactome	target_disease	temporal
12	target_diseaseaffected_pathwayslapenrich	target_disease	temporal
13	target_diseaseaffected_pathwaysysbio	target_disease	temporal
14	target_diseaseanimal_modelimpc	target_disease	temporal
15	target_diseasegenetic_associationclingen	target_disease	static
16	target_diseasegenetic_associationcurated	target_disease	static
17	target_diseasegenetic_associationeva	target_disease	static
18	target_diseasegenetic_associationgene2phenotype	target_disease	static
19	target_diseasegenetic_associationgene_burden	target_disease	static
20	target_diseasegenetic_associationgenomics_england	target_disease	static
21	target_diseasegenetic_associationomim	target_disease	static
22	target_diseasegenetic_associationorphanet	target_disease	static
23	target_diseasegenetic_associationot_genetics_portal	target_disease	static
24	target_diseasegenetic_associationuniprot_literature	target_disease	static
25	target_diseasegenetic_associationuniprot_variants	target_disease	static
26	target_diseaseknown_drugchembl	target_disease	temporal
27	target_diseaseliteratureeuropepmc	target_disease	temporal
28	target_diseaseoutcomeadvanced	target_disease	temporal
29	target_diseaserna_expressionexpression_atlas	target_disease	temporal
30	target_diseasesomatic_mutationcancer_gene_census	target_disease	temporal
31	target_diseasesomatic_mutationeva_somatic	target_disease	temporal
32	target_diseasesomatic_mutationintogen	target_disease	temporal
33	target_diseasetimetransition	target_disease	temporal

## 7 Supplementary Material

#### 7.1 Performance

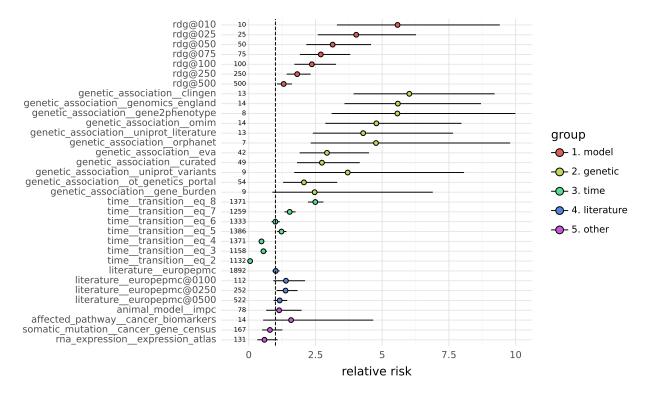


Figure 8: **Performance of individual features and predictive scores as measured by relative risk**. RDG model results denoted by rdg@N indicate performance for the N top TD pairs. The same convention is used for literature evidence and the time\_transition\_eq\_X convention denotes RR estimates when the time since the phase 2 transition is equal to X years. The omim, eva, and ot\_genetics\_portal features correspond to the OMIM, EVA and OTG baselines of Figure 2, respectively. All other features are assessed based on their existence. The counts along the origin indicate how many TD pairs were used to compute the RR numerator.

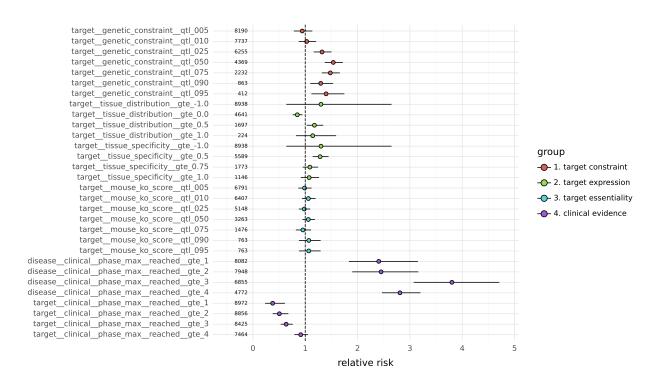


Figure 9: **Relative risk scores for target/disease features**. The features ending with  $qtl_Q$  denote binary indicators constructed from cases where the feature meets or exceeds quantile Q of its distribution. The features ending with  $gte_X$  denote indicators for when the feature meets or exceeds a specific value X.

		ОМІМ	EVA/ClinVar	OTG/GWAS	0TS@010	OTS@020	OTS@030	01S@050	OTS@100	RDG@010	RDG@020	RDG@030	RDG@050	RDG@100
therapeutic_area	n_pairs													
average		9.65	5.78	1.95	2.88	2.10	2.06	1.98	1.75	5.82	3.63	3.13	2.41	1.84
all	9010	4.78	2.93	2.07	6.70	3.91	3.35	2.69	2.13	5.58	3.91	3.73	3.14	2.37
cancer or benign tumor	4013		3.69	0.00	2.58	3.91	5.31	3.65	2.68	5.20	5.00	4.40	4.28	3.26
genetic, familial or congenital disease	2035	3.43	2.37	0.68	3.08	1.53	1.02	0.81	0.91	3.08	2.58	1.71	1.23	0.80
nervous system disease	1534	3.64	1.95	0.00	2.54	1.15	1.70	1.54	2.14	2.54	1.26	2.59	2.08	1.56
gastrointestinal disease	1260		0.00	3.48	5.68	4.33	2.68	3.17	2.82	8.72	5.90	3.90	4.34	2.44
immune system disease	1225	21.09	21.44	2.88	6.39	4.30	4.43	2.61	1.99	11.05	5.48	4.43	3.10	2.53
hematologic disease	1023	26.89	27.59	0.00	5.48	2.71	1.79	2.05	1.68	8.44	4.18	3.78	2.22	1.64
endocrine system disease	949	0.00	0.00	11.26	0.00	0.00	2.12	2.88	2.21	10.83	5.36	4.90	2.88	1.77
reproductive system or breast disease	948		0.00	0.00	0.00	0.00	0.00	1.68	1.72	8.79	4.35	3.95	2.99	2.51
musculoskeletal or connective tissue disease	757	8.82	6.67	2.68	4.74	3.62	2.73	2.48	2.18	6.02	5.00	4.76	3.09	2.80
respiratory or thoracic disease	644		0.00	0.00	4.37	2.74	1.80	1.97	1.52	3.22	2.15	2.19	1.51	1.83
cardiovascular disease	617	3.65	1.66	2.09	1.83	1.66	1.48	0.86	0.87	2.21	1.85	1.35	0.93	0.68
integumentary system disease	563			0.00	0.00	0.00	0.82	1.43	1.49	4.37	2.94	1.92	1.76	1.12
urinary system disease	561		3.99	2.29	0.79	1.40	0.92	0.61	0.59	1.19	1.19	0.78	0.86	0.93

Figure 10: **Relative risk scores by method, benchmark and therapeutic area**. The **average** therapeutic area indicates mean values across all others except for **all**, which is an ungrouped estimate across all diseases regardless of therapeutic area.

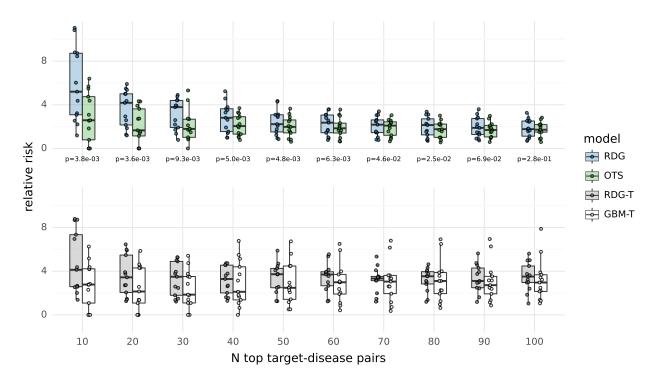


Figure 11: **Relative risk distributions across select therapeutic areas**. P-values are computed from a one-sided Wilcoxon signed-rank test with the alternative that the RDG model RR averages across therapeutic areas exceed OTS averages.

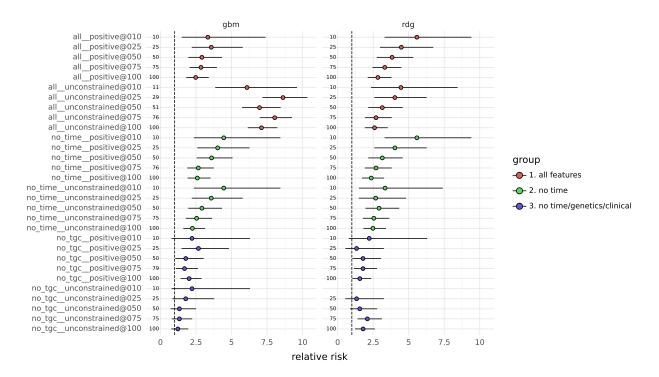


Figure 12: Performance by algorithm, constraint type and feature grouping.

#### 7.2 Predictions

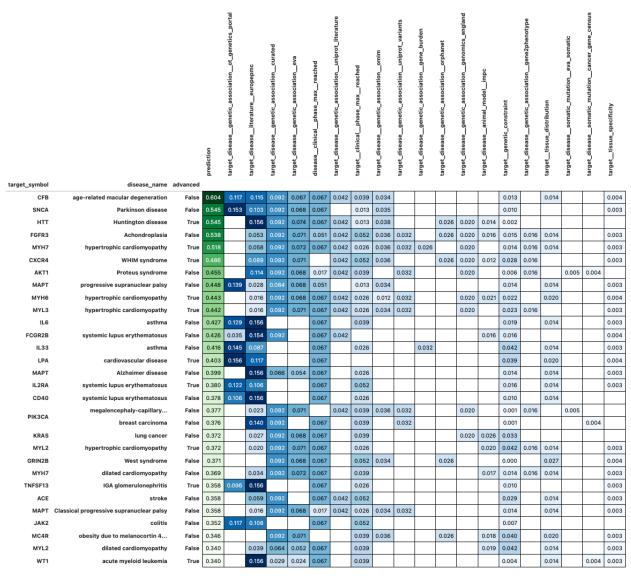


Figure 13: **Top RDG model evaluation dataset predictions**. Feature contributions are shown as the product of their underlying values and the RDG coefficients. The **advanced** field indicates whether the associated TD pair advanced beyond phase 2 as of 2024.

# 7.3 Opportunities

Table 2: Tractability bucket assignments

	evidence	modality	
1	Phase 1 Clinical	OC	LOW
2	Advanced Clinical	OC	MED
3	Approved Drug	OC	HIGH
4	GO CC med conf	AB	LOW
5	Human Protein Atlas loc	AB	LOW
6	UniProt SigP or TMHMM	AB	LOW
7	UniProt loc med conf	AB	LOW
8	GO CC high conf	AB	MED
9	UniProt loc high conf	AB	MED
10	Advanced Clinical	AB	HIGH
11	Approved Drug	AB	HIGH
12	Phase 1 Clinical	AB	HIGH
13	Database Ubiquitination	PR	LOW
14	Half-life Data	PR	LOW
15	Small Molecule Binder	PR	LOW
16	Literature	PR	MED
17	UniProt Ubiquitination	PR	MED
18	Advanced Clinical	PR	HIGH
19	Phase 1 Clinical	PR	HIGH
20	Druggable Family	SM	LOW
21	High-Quality Pocket	SM	LOW
22	Med-Quality Pocket	SM	LOW
23	High-Quality Ligand	SM	MED
24	Structure with Ligand	SM	MED
25	Advanced Clinical	SM	HIGH
26	Approved Drug	SM	HIGH
27	Phase 1 Clinical	SM	HIGH

		prediction	target_clinical_phase_max_reached	disease_clinical_phase_max_reached	target_diseasegenetic_associationcurated	target_diseasegenetic_associationeva	target_diseasegenetic_associationot_genetics_portal	target_diseasegenetic_associationomim	target_diseasegenetic_associationgenomics_england	target_diseasegenetic_associationuniprot_literature	target_diseasegenetic_associationuniprot_variants	target_diseasegenetic_associationorphanet	target_diseasegenetic_associationclingen	target_diseasegenetic_associationgene2phenotype	target_disease_genetic_association_gene_burden
target_symbol	disease_name	0.000				0.0	0.7	0.0	1.0	4.0	4.0				
SNCA	Lewy body dementia	0.669	2	4	1.0	0.9	0.7	0.9	1.0	1.0	1.0				
CTLA4	systemic lupus erythematosus	0.614	4	4	1.0	0.9	0.7			1.0					-
FGFR2	Hashimoto's thyroiditis  Apert syndrome	0.596	4	4	1.0	0.9	8.0	0.9	1.0	1.0	1.0	1.0	1.0	1.0	$\dashv$
TNFSF4	systemic lupus erythematosus	0.574	2	4	1.0	0.9	0.8	0.5	1.0	1.0	1.0	1.0	1.0	1.0	
MAPT	Pick disease	0.568	2	4	1.0	0.9	0.0	0.9	1.0	1.0	1.0				
VWF	von Willebrand disease 1	0.534	3		1.0	0.9		0.9	1.0	1.0	1.0				
CTLA4	celiac disease	0.533	4	3	1.0	0.9	0.7	0.5	1.0	1.0					
CSF1R	Hereditary diffuse leukoencephalopath	0.529	4		1.0	0.9		0.9	1.0	1.0	1.0	1.0			
CTLA4	type 1 diabetes mellitus	0.513	4	4	0.9	0.9	0.8								
VWF	von Willebrand disease 2	0.513	3		1.0	0.9		0.9	1.0	1.0	1.0				$\Box$
F2	Congenital factor II deficiency	0.504	4	1	1.0	0.9		0.9	1.0	1.0	1.0	1.0			
HLA-DRB1	rheumatoid arthritis	0.500	2	4	1.0		0.4			1.0					
FGFR3	hypochondroplasia	0.490	4	2	1.0	0.9		0.9	1.0	1.0		1.0	1.0	1.0	
F11	deep vein thrombosis	0.489	4	4			0.9								
CTLA4	autoimmune disease	0.484	4	4	1.0		0.8		1.0						
FGFR2	Crouzon syndrome	0.482	4		1.0	0.9		0.9	1.0	1.0	1.0	1.0	1.0	1.0	
FGFR3	camptodactyly-tall stature-scoliosis	0.473	4		1.0	0.9		0.9	1.0		1.0	1.0	0.5		
FGF23	${\it autosomal\ dominant\ hypophosphatemic\ r}$	0.472	4		1.0	0.9		0.9	1.0		1.0	1.0			
IL33	allergic rhinitis	0.471	3	4			0.7								1.0
IGF2	type 2 diabetes mellitus	0.466	2	4	0.9	0.9	0.6								
ITGB2	Leukocyte adhesion deficiency type I	0.464	4	1	1.0	0.9		0.9	1.0	1.0	1.0	1.0			
FGFR3	thanatophoric dysplasia type 1	0.463	4		1.0	0.9		0.9	1.0	1.0	1.0	1.0	1.0	1.0	
IL12B	inflammatory bowel disease	0.458	4	4			8.0								
RPS19	Diamond-Blackfan anemia	0.458	4	2	1.0	0.9			1.0			1.0	1.0		
IL33	infection	0.457	3	4			8.0								
TACSTD2	gelatinous drop-like corneal dystrophy	0.457	4		1.0	0.9		0.9	1.0	1.0		1.0		1.0	
TNFSF11	primary biliary cirrhosis	0.456	4	4	1.0	0.0	0.8	0.0	1.0	1.0	1.0	1.0	1.0	1.0	$\dashv$
FGFR3	Muenke syndrome	0.456	4		1.0	0.9	0.7	0.9	1.0	1.0	1.0	1.0	1.0	1.0	$\dashv$
TSLP	Nasal Cavity Polyp	0.455	4	4			0.7								

Figure 14: **Top ranked undeveloped, tractable target-disease pairs**. The highest scoring TD pairs per the RDG model that have not entered clinical trials despite having a target that has been in trials of an antibody-based drug. All values shown other than **prediction** are raw feature values, unweighted by RDG coefficients.

## 7.4 Sensitivity

In order to validate the stability of our findings in Section 2.3, we repeat this analysis across 18 different configurations listed in Supplementary Table 3. This includes 3 separate versions of Open Targets, 3 choices for the year defining the split between training and evaluation data and 2 choices for the length of the minimum advancement window (in years).

We find that the mean RR values from the RDG model consistently exceed the OTS model in all configurations among the very highest rankings (N=10) and also exceed the OTS model in all configurations except for 1 for N between 20 and 60. This data is shown in Supplementary Figure 15. The significance of these differences drops notably after N=40, which can be seen in the distribution of p-values from a Wilcoxon signed-rank test shown in Supplementary Figure 16.

Table 3: Configurations for sensitivity analysis

	open_targets_version	max_training_year	min_time_to_advancement_years
1	23.09	2017	4
2	23.12	2017	2
3	23.09	2015	2
4	23.12	2015	2
5	23.09	2017	2
6	23.06	2015	4
7	23.06	2017	2
8	23.06	2013	2
9	23.06	2015	2
10	23.06	2017	4
11	23.12	2013	4
12	23.09	2015	4
13	23.09	2013	2
14	23.12	2013	2
15	23.09	2013	4
16	23.12	2017	4
17	23.12	2015	4
18	23.06	2013	4

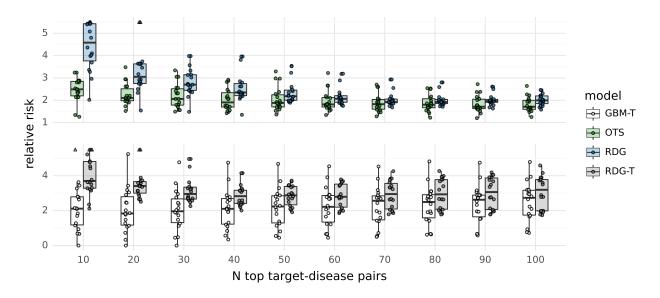


Figure 15: **Relative risk distributions across configurations in sensitivity analysis**. The distribution of the mean RR values displayed for a single configuration in Figure 1 is shown here across 18 configurations.

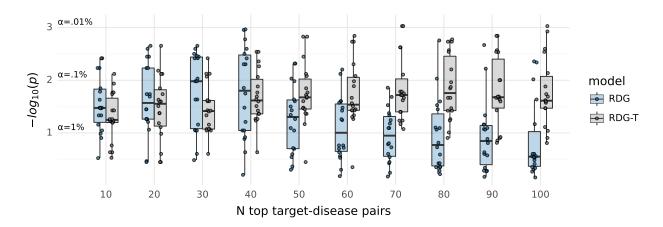


Figure 16: **P-value distributions across configurations in sensitivity analysis**. The distribution of the p-values displayed for a single configuration in Supplementary Figure 11 is shown here across 18 configurations.

## References

- [1] Matthew R Nelson, Hannah Tipney, Jeffery L Painter, Judong Shen, Paola Nicoletti, Yufeng Shen, Aris Floratos, Pak Chung Sham, Mulin Jun Li, Junwen Wang, Lon R Cardon, John C Whittaker, and Philippe Sanseau. The support of human genetic evidence for approved drug indications. *Nat. Genet.*, 47(8):856–860, August 2015.
- [2] Emily A King, J Wade Davis, and Jacob F Degner. Are drug targets with genetic support twice as likely to be approved? revised estimates of the impact of genetic support for drug mechanisms on the probability of drug approval. *PLoS Genet.*, 15(12):e1008489, December 2019.

- [3] Eric Vallabh Minikel, Jeffery L Painter, Coco Chengliang Dong, and Matthew R. Nelson. Refining the impact of genetic evidence on clinical success. *medRxiv*, 2023.
- [4] Olesya Razuvayevskaya, Irene Lopez, Ian Dunham, and David Ochoa. Why clinical trials stop: The role of genetics. *medRxiv*, 2023.
- [5] Ben Kinnersley, Amit Sud, Elizabeth A Coker, Joseph E Tym, Patrizio Di Micco, Bissan Al-Lazikani, and Richard S Houlston. Leveraging human genetics to guide cancer drug development. *JCO clinical cancer informatics*, 2:1—11, December 2018.
- [6] David Cook, Dearg Brown, Robert Alexander, Ruth March, Paul Morgan, Gemma Satterthwaite, and Menelas N Pangalos. Lessons learned from the fate of astrazeneca's drug pipeline: a five-dimensional framework. *Nature reviews. Drug discovery*, 13(6):419—431, June 2014.
- [7] David Ochoa, Mohd Karim, Maya Ghoussaini, David G Hulcoop, Ellen M McDonagh, and Ian Dunham. Human genetics evidence supports two-thirds of the 2021 fda-approved drugs. *Nature reviews. Drug discovery*, 21(8):551, August 2022.
- [8] Maya Ghoussaini, Matthew R Nelson, and Ian Dunham. Future prospects for human genetics and genomics in drug discovery. *Current opinion in structural biology*, 80:102568, June 2023.
- [9] Áine Duffy, Ben Omega Petrazzini, David Stein, Joshua K Park, Iain S Forrest, Kyle Gibson, Ha My Vy, Robert Chen, Carla Márquez-Luna, Matthew Mort, Marie Verbanck, Avner Schlessinger, Yuval Itan, David N Cooper, Ghislain Rocheleau, Daniel M Jordan, and Ron Do. Development of a human genetics-guided priority score for 19,365 genes and 399 drug indications. *Nature genetics*, 56(1):51—59, January 2024.
- [10] Gautier Koscielny, Peter An, Denise Carvalho-Silva, Jennifer A Cham, Luca Fumis, Rippa Gasparyan, Samiul Hasan, Nikiforos Karamanis, Michael Maguire, Eliseo Papa, Andrea Pierleoni, Miguel Pignatelli, Theo Platt, Francis Rowland, Priyanka Wankar, A Patrícia Bento, Tony Burdett, Antonio Fabregat, Simon Forbes, Anna Gaulton, Cristina Yenyxe Gonzalez, Henning Hermjakob, Anne Hersey, Steven Jupe, Şenay Kafkas, Maria Keays, Catherine Leroy, Francisco-Javier Lopez, Maria Paula Magarinos, James Malone, Johanna McEntyre, Alfonso Munoz-Pomer Fuentes, Claire O'Donovan, Irene Papatheodorou, Helen Parkinson, Barbara Palka, Justin Paschall, Robert Petryszak, Naruemon Pratanwanich, Sirarat Sarntivijal, Gary Saunders, Konstantinos Sidiropoulos, Thomas Smith, Zbyslaw Sondka, Oliver Stegle, Y Amy Tang, Edward Turner, Brendan Vaughan, Olga Vrousgou, Xavier Watkins, Maria-Jesus Martin, Philippe Sanseau, Jessica Vamathevan, Ewan Birney, Jeffrey Barrett, and Ian Dunham. Open targets: a platform for therapeutic target identification and validation. *Nucleic Acids Res.*, 45(D1):D985–D994, January 2017.
- [11] Hai Fang, ULTRA-DD Consortium, Hans De Wolf, Bogdan Knezevic, Katie L Burnham, Julie Osgood, Anna Sanniti, Alicia Lledó Lara, Silva Kasela, Stephane De Cesco, Jörg K Wegner, Lahiru Handunnetthi, Fiona E McCann, Liye Chen, Takuya Sekine, Paul E Brennan, Brian D Marsden, David Damerell, Chris A O'Callaghan, Chas Bountra, Paul Bowness, Yvonne Sundström, Lili Milani, Louise Berg, Hinrich W Göhlmann, Pieter J Peeters, Benjamin P Fairfax, Michael Sundström, and Julian C Knight. A genetics-led approach defines the drug target landscape of 30 immune-related traits. *Nature genetics*, 51(7):1082—1091, July 2019.
- [12] Aidan MacNamara, Nikolina Nakic, Ali Amin Al Olama, Cong Guo, Karsten B Sieber, Mark R Hurle, and Alex Gutteridge. Network and pathway expansion of genetic disease associations identifies successful drug targets. *Scientific reports*, 10(1):20970, December 2020.
- [13] Chaohui Bao, Hengru Wang, and Hai Fang. Genomic evidence supports the recognition of endometriosis as an inflammatory systemic disease and reveals disease-specific therapeutic potentials of targeting neutrophil degranulation. *Front. Immunol.*, 13:758440, March 2022.
- [14] Marie C Sadler, Chiara Auwerx, Patrick Deelen, and Zoltán Kutalik. Multi-layered genetic approaches to identify approved drug targets. *Cell Genom.*, 3(7):100341, July 2023.
- [15] Inigo Barrio-Hernandez and Pedro Beltrao. Network analysis of genome-wide association studies for drug target prioritisation. *Current opinion in chemical biology*, 71:102206, December 2022.

- [16] Inigo Barrio-Hernandez, Jeremy Schwartzentruber, Anjali Shrivastava, Noemi Del-Toro, Asier Gonzalez, Qian Zhang, Edward Mountjoy, Daniel Suveges, David Ochoa, Maya Ghoussaini, Glyn Bradley, Henning Hermjakob, Sandra Orchard, Ian Dunham, Carl A Anderson, Pablo Porras, and Pedro Beltrao. Network expansion of genetic associations defines a pleiotropy map of human cell biology. *Nature genetics*, 55(3):389—398, March 2023.
- [17] Saee Paliwal, Alex de Giorgio, Daniel Neil, Jean-Baptiste Michel, and Alix Mb Lacoste. Preclinical validation of therapeutic targets predicted by tensor factorization on heterogeneous graphs. *Sci. Rep.*, 10(1):18250, October 2020.
- [18] Camilo Ruiz, Marinka Zitnik, and Jure Leskovec. Identification of disease treatment mechanisms through the multiscale interactome. *Nature communications*, 12(1):1796, March 2021.
- [19] Srivamshi Pittala, William Koehler, Jonathan Deans, Daniel Salinas, Martin Bringmann, Katharina Sophia Volz, and Berk Kapicioglu. Relation-weighted link prediction for disease gene identification, 2020.
- [20] Ozlem Muslu, Charles Tapley Hoyt, Mauricio Lacerda, Martin Hofmann-Apitius, and Holger Frohlich. Guiltytargets: Prioritization of novel therapeutic targets with network representation learning. *IEEE/ACM transactions on computational biology and bioinformatics*, 19(1):491—500, 2022.
- [21] Petrina Kamya, Ivan V Ozerov, Frank W Pun, Kyle Tretina, Tatyana Fokina, Shan Chen, Vladimir Naumov, Xi Long, Sha Lin, Mikhail Korzinkin, Daniil Polykovskiy, Alex Aliper, Feng Ren, and Alex Zhavoronkov. Pandaomics: An ai-driven platform for therapeutic target and biomarker discovery. *Journal of chemical information and modeling*, February 2024.
- [22] Alex Aliper, Roman Kudrin, Daniil Polykovskiy, Petrina Kamya, Elena Tutubalina, Shan Chen, Feng Ren, and Alex Zhavoronkov. Prediction of clinical trials outcomes based on target choice and clinical trial design with multi-modal artificial intelligence. *Clinical pharmacology and therapeutics*, 114(5):972—980, November 2023.
- [23] Kien Wei Siah, Nicholas W Kelley, Steffen Ballerstedt, Björn Holzhauer, Tianmeng Lyu, David Mettler, Sophie Sun, Simon Wandel, Yang Zhong, Bin Zhou, Shifeng Pan, Yingyao Zhou, and Andrew W Lo. Predicting drug approvals: The novartis data science and artificial intelligence challenge. *Patterns (New York, N.Y.)*, 2(8):100312, August 2021.
- [24] Andrew W. Lo, Kien Wei Siah, and Chi Heem Wong. Machine Learning With Statistical Imputation for Predicting Drug Approvals. *Harvard Data Science Review*, 1(1), jul 1 2019. https://hdsr.mitpress.mit.edu/pub/ct67j043.
- [25] David Narganes-Carlón, Daniel J Crowther, and Ewan R Pearson. A publication-wide association study (pwas), historical language models to prioritise novel therapeutic drug targets. *Scientific reports*, 13(1):8366, May 2023.
- [26] Ada Hamosh, Alan F Scott, Joanna S Amberger, Carol A Bocchini, and Victor A McKusick. Online mendelian inheritance in man (omim), a knowledgebase of human genes and genetic disorders. *Nucleic acids research*, 33(Database issue):D514—7, January 2005.
- [27] Melissa J Landrum, Jennifer M Lee, George R Riley, Wonhee Jang, Wendy S Rubinstein, Deanna M Church, and Donna R Maglott. Clinvar: public archive of relationships among sequence variation and human phenotype. *Nucleic acids research*, 42(Database issue):D980—5, January 2014.
- [28] FDA. Step 3: Clinical research fda.gov. https://www.fda.gov/patients/drug-development-process/step-3-clinical-research, Apr 2018.
- [29] Chi Heem Wong, Kien Wei Siah, and Andrew W Lo. Estimation of clinical trial success rates and related parameters. *Biostatistics (Oxford, England)*, 20(2):273—286, April 2019.
- [30] Mathias Uhlén, Linn Fagerberg, Björn M Hallström, Cecilia Lindskog, Per Oksvold, Adil Mardinoglu, Åsa Sivertsson, Caroline Kampf, Evelina Sjöstedt, Anna Asplund, IngMarie Olsson, Karolina Edlund, Emma Lundberg, Sanjay Navani, Cristina Al-Khalili Szigyarto, Jacob Odeberg, Dijana Djureinovic, Jenny Ottosson Takanen, Sophia Hober, Tove Alm, Per-Henrik Edqvist, Holger Berling, Hanna Tegel, Jan Mulder, Johan Rockberg, Peter Nilsson, Jochen M Schwenk, Marica Hamsten, Kalle von Feilitzen, Mattias Forsberg, Lukas Persson, Fredric Johansson, Martin Zwahlen, Gunnar von Heijne, Jens Nielsen, and Fredrik Pontén. Proteomics. tissue-based map of the human proteome. Science (New York, N.Y.), 347(6220):1260419, January 2015.

- [31] Konrad J Karczewski, Laurent C Francioli, Grace Tiao, Beryl B Cummings, Jessica Alföldi, Qingbo Wang, Ryan L Collins, Kristen M Laricchia, Andrea Ganna, Daniel P Birnbaum, Laura D Gauthier, Harrison Brand, Matthew Solomonson, Nicholas A Watts, Daniel Rhodes, Moriel Singer-Berk, Eleina M England, Eleanor G Seaby, Jack A Kosmicki, Raymond K Walters, Katherine Tashman, Yossi Farjoun, Eric Banks, Timothy Poterba, Arcturus Wang, Cotton Seed, Nicola Whiffin, Jessica X Chong, Kaitlin E Samocha, Emma Pierce-Hoffman, Zachary Zappala, Anne H O'Donnell-Luria, Eric Vallabh Minikel, Ben Weisburd, Monkol Lek, James S Ware, Christopher Vittal, Irina M Armean, Louis Bergelson, Kristian Cibulskis, Kristen M Connolly, Miguel Covarrubias, Stacey Donnelly, Steven Ferriera, Stacey Gabriel, Jeff Gentry, Namrata Gupta, Thibault Jeandet, Diane Kaplan, Christopher Llanwarne, Ruchi Munshi, Sam Novod, Nikelle Petrillo, David Roazen, Valentin Ruano-Rubio, Andrea Saltzman, Molly Schleicher, Jose Soto, Kathleen Tibbetts, Charlotte Tolonen, Gordon Wade, Michael E Talkowski, Genome Aggregation Database Consortium, Benjamin M Neale, Mark J Daly, and Daniel G MacArthur. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature*, 581(7809):434—443, May 2020.
- [32] Open targets platform 23.12 has been released! https://blog.opentargets.org/open-targets-platform-23-12-release/#target-prioritisation, Nov 2023.
- [33] Target-disease associations | Open Targets Platform Documentation. https://platform-docs.opentargets.org/associations#data-source-weights, 2023.
- [34] Timothe Cezard, Fiona Cunningham, Sarah E Hunt, Baron Koylass, Nitin Kumar, Gary Saunders, April Shen, Andres F Silva, Kirill Tsukanov, Sundararaman Venkataraman, Paul Flicek, Helen Parkinson, and Thomas M Keane. The european variation archive: a fair resource of genomic variation for all species. *Nucleic acids research*, 50(D1):D1216—D1220, January 2022.
- [35] Eleonora Porcu, Marie C Sadler, Kaido Lepik, Chiara Auwerx, Andrew R Wood, Antoine Weihs, Maroun S Bou Sleiman, Diogo M Ribeiro, Stefania Bandinelli, Toshiko Tanaka, Matthias Nauck, Uwe Völker, Olivier Delaneau, Andres Metspalu, Alexander Teumer, Timothy Frayling, Federico A Santoni, Alexandre Reymond, and Zoltán Kutalik. Differentially expressed genes reflect disease-induced rather than disease-causing changes in the transcriptome. *Nature communications*, 12(1):5647, September 2021.
- [36] Chris Finan, Anna Gaulton, Felix A Kruger, R Thomas Lumbers, Tina Shah, Jorgen Engmann, Luana Galver, Ryan Kelley, Anneli Karlsson, Rita Santos, John P Overington, Aroon D Hingorani, and Juan P Casas. The druggable genome and support for target identification and validation in drug development. *Science translational medicine*, 9(383):eaag1166, March 2017.
- [37] Chaohui Bao, Hengru Wang, and Hai Fang. Genomic evidence supports the recognition of endometriosis as an inflammatory systemic disease and reveals disease-specific therapeutic potentials of targeting neutrophil degranulation. *Frontiers in immunology*, 13:758440, 2022.
- [38] Open Targets Tractability Pipeline (version 2). https://github.com/chembl/tractability\_pipeline\_v2, Jul 2023.
- [39] Evan A Boyle, Yang I Li, and Jonathan K Pritchard. An expanded view of complex traits: From polygenic to omnigenic. *Cell*, 169(7):1177—1186, June 2017.
- [40] Stephen Burgess, Amy M Mason, Andrew J Grant, Eric A W Slob, Apostolos Gkatzionis, Verena Zuber, Ashish Patel, Haodong Tian, Cunhao Liu, William G Haynes, G Kees Hovingh, Lotte Bjerre Knudsen, John C Whittaker, and Dipender Gill. Using genetic association data to guide drug discovery and development: Review of methods and applications. *American journal of human genetics*, 110(2):195—214, February 2023.
- [41] Damian Szklarczyk, Rebecca Kirsch, Mikaela Koutrouli, Katerina Nastou, Farrokh Mehryary, Radja Hachilif, Annika L Gable, Tao Fang, Nadezhda T Doncheva, Sampo Pyysalo, Peer Bork, Lars J Jensen, and Christian von Mering. The string database in 2023: protein-protein association networks and functional enrichment analyses for any sequenced genome of interest. *Nucleic acids research*, 51(D1):D638—D646, January 2023.
- [42] Robert W Hoffman, Joan T Merrill, Marta M E Alarcón-Riquelme, Michelle Petri, Ernst R Dow, Eric Nantz, Laura K Nisenbaum, Krista M Schroeder, Wendy J Komocsar, Narayanan B Perumal, Matthew D Linnik, David C

- Airey, Yushi Liu, Guilherme V Rocha, and Richard E Higgs. Gene expression and pharmacodynamic changes in 1,760 systemic lupus erythematosus patients from two phase iii trials of baff blockade with tabalumab. *Arthritis and rheumatology (Hoboken, N.J.)*, 69(3):643—654, March 2017.
- [43] Christian Fougner, Julie Cannon, Lydia The, Jeffrey F Smith, and Olivier Leclerc. Herding in the drug development pipeline. *Nature reviews. Drug discovery*, 22(8):617—618, August 2023.
- [44] Winston A Haynes, Aurelie Tomczak, and Purvesh Khatri. Gene annotation bias impedes biomedical research. *Scientific reports*, 8(1):1362, January 2018.
- [45] Rita Santos, Oleg Ursu, Anna Gaulton, A Patrícia Bento, Ramesh S Donadi, Cristian G Bologa, Anneli Karlsson, Bissan Al-Lazikani, Anne Hersey, Tudor I Oprea, and John P Overington. A comprehensive map of molecular drug targets. *Nature reviews. Drug discovery*, 16(1):19—34, January 2017.
- [46] Chris Finan, Anna Gaulton, Felix Kruger, Tom Lumbers, Tina Shah, Jorgen Engmann, Luana Galver, Ryan Kelly, Anneli Karlsson, Rita Santos, John Overington, Aroon Hingorani, and Juan Pablo Casas. The druggable genome and support for target identification and validation in drug development, 2016.
- [47] Arielle Marks-Anglin and Yong Chen. A historical review of publication bias. *Research synthesis methods*, 11(6):725—742, November 2020.
- [48] Luca Abatangelo, Rosalia Maglietta, Angela Distaso, Annarita D'Addabbo, Teresa Maria Creanza, Sayan Mukherjee, and Nicola Ancona. Comparative study of gene set enrichment methods. *BMC bioinformatics*, 10:275, September 2009.
- [49] Katerina Trajanoska, Claude Bhérer, Daniel Taliun, Sirui Zhou, J Brent Richards, and Vincent Mooser. From target discovery to clinical drug development with human genetics. *Nature*, 620(7975):737—745, August 2023.
- [50] Ruth J F Loos. 15 years of genome-wide association studies and no signs of slowing down. *Nature communications*, 11(1):5900, November 2020.
- [51] Abdel Abdellaoui, Loic Yengo, Karin J H Verweij, and Peter M Visscher. 15 years of gwas discovery: Realizing the promise. *American journal of human genetics*, 110(2):179—194, February 2023.
- [52] Michael J Bamshad, Deborah A Nickerson, and Jessica X Chong. Mendelian gene discovery: Fast and furious with no end in sight. *American journal of human genetics*, 105(3):448—455, September 2019.
- [53] Catherine S Storm, Demis A Kia, Mona M Almramhi, Sara Bandres-Ciga, Chris Finan, International Parkinson's Disease Genomics Consortium (IPDGC), Aroon D Hingorani, and Nicholas W Wood. Finding genetically-supported drug targets for parkinson's disease using mendelian randomization of the druggable genome. *Nature communications*, 12(1):7342, December 2021.
- [54] Guolin Ke, Qi Meng, Thomas Finley, Taifeng Wang, Wei Chen, Weidong Ma, Qiwei Ye, and Tie-Yan Liu. Lightgbm: A highly efficient gradient boosting decision tree. In I. Guyon, U. Von Luxburg, S. Bengio, H. Wallach, R. Fergus, S. Vishwanathan, and R. Garnett, editors, *Advances in Neural Information Processing Systems*, volume 30. Curran Associates, Inc., 2017.