MR example for Supplementary Material

Example R and Stata code to perform a multivariable Mendelian randomization (MVMR) analysis

R code

• Read in the data

```
suppressPackageStartupMessages({
    library(tidyverse)
    library(haven)
})
dat <- read_dta("https://raw.github.com/remlapmot/mrrobust/master/dodata.dta")
dat <- dat %>% filter(ldlcp2 < 1e-8)</pre>
```

Example code using the MendelianRandomization package

• Install the package and load it into memory

library (MendelianRandomization)

• Convert our data frame to the required class

```
datfmt <- mr_mvinput(
  bx = as.matrix(cbind(dat$ldlcbeta, dat$tdbeta, dat$tgbeta)),
  bxse = as.matrix(cbind(dat$ldlcse, dat$tdse, dat$tgse)),
  by = dat$chdbeta,
  byse = dat$chdse,
  exposure = "exposure",
  outcome = "outcome",
  snps = "snp",
  effect_allele = dat$a1,
  other_allele = dat$a2,
  eaf = NA
)</pre>
```

• Fit an MVMR/MVIVW model

```
mvivwfit <- mr_mvivw(datfmt)
mvivwfit</pre>
```

```
##
## Multivariable inverse-variance weighted method
## (variants uncorrelated, random-effect model)
##
## Number of Variants : 73
##
## ------
    Exposure Estimate Std Error 95% CI p-value
##
##
   exposure_1 0.429 0.061 0.309, 0.548 0.000
## exposure_2 -0.194
                     0.131 -0.451, 0.062 0.138
## exposure_3 0.226 0.123 -0.016, 0.468 0.067
## Residual standard error = 1.490
## Heterogeneity test statistic = 155.3766 on 70 degrees of freedom, (p-value = 0.0000)
```

• Fit an MVMR-Egger model

```
mvmreggerfit <- mr_mvegger(datfmt)
mvmreggerfit</pre>
```

```
## Multivariable MR-Egger method
## (variants uncorrelated, random-effect model)
## Orientated to exposure : 1
## Number of Variants : 73
        _____
                                      p-value
##
      Exposure Estimate Std Error 95% CI
##
    exposure_1 0.567 0.100 0.371, 0.764 0.000
##
    exposure_2 -0.136
                        0.133 -0.398, 0.125 0.306
    exposure_3 0.274
                        0.125 0.030, 0.518 0.028
##
## (intercept) -0.009
                        0.005 -0.020, 0.001 0.084
## Residual standard error = 1.469
## Heterogeneity test statistic = 148.9290 on 69 degrees of freedom, (p-value = 0.0000)
```

Example code using the MVMR package

• Install the package and load it into memory

library(MVMR)

• Create a data object of the required structure

```
r_input <- format_mvmr(
   BXGs = dat[,c("ldlcbeta","hdlcbeta","tgbeta")],
   BYG = dat$chdbeta,
   seBXGs = dat[,c("ldlcse","hdlcse","tgse")],
   seBYG = dat$chdse,
   RSID = dat$rsid
)</pre>
```

• Fit an MVMR model

```
mvmrfit <- ivw_mvmr(r_input)

##

## Multivariable MR

##

## Estimate Std. Error t value Pr(>|t|)

## exposure1 0.4286200 0.0609661 7.030464 1.099077e-09

## exposure2 -0.1941989 0.1308289 -1.484372 1.421994e-01

## exposure3 0.2260456 0.1232828 1.833554 7.097168e-02

##

## Residual standard error: 1.49 on 70 degrees of freedom

• Heterogeneity statistic
```

```
strength_mvmr(r_input)
```

```
## Warning in strength_mvmr(r_input): Covariance between effect of genetic variants on each exposure
## not specified. Fixing covariance at 0.
##
## Conditional F-statistics for instrument strength
##
```

```
## exposure1 exposure2 exposure3
## F-statistic 126.7447 35.29937 39.32731

## exposure1 exposure2 exposure3
## F-statistic 126.7447 35.29937 39.32731
```

Stata code

• Load the Statamarkdown package to enable Stata code chunks in an R Markdown file

library(Statamarkdown)

• Read in the data and create an indicator variable to select observations with p-value between the genotype and LDL-C < 10-8

```
use https://raw.github.com/remlapmot/mrrobust/master/dodata, clear
gen byte sel1 = (ldlcp2 < 1e-8)</pre>
```

Example code using the mrrobust package

• Install the mrrobust package using the github package

```
// Note: output suppressed
net install mrrobust, from("https://raw.github.com/remlapmot/mrrobust/master/")
mrdeps
```

• Fit and MVMR model with phenotypes LDL-c and HDL-c (Burgess, Dudbridge, and Thompson 2015).

```
mvmr chdbeta ldlcbeta hdlcbeta [aw=1/(chdse^2)] if sel1==1
```

Number of genotypes = 73 Number of phenotypes = 2 Standard errors: Random effect Residual standard error = 1.514

					[95% Conf.	Interval]
chdbeta ldlcbeta	 .4670719	.0581901	8.03	0.000	.3530214 5305175	

• Additionally include a third phenotype – trigly cerides.

```
mvmr chdbeta ldlcbeta hdlcbeta tgbeta [aw=1/(chdse^2)] if sel1==1
```

Number of genotypes = 73

Number of phenotypes = 3

Standard errors: Random effect

Residual standard error = 1.490

	Coef.	Std. Err.				Interval]
chdbeta						
ldlcbeta	.42862	.0609661	7.03	0.000	.3091286	.5481113
hdlcbeta	1941989	.1308289	-1.48	0.138	4506189	.0622211
tgbeta	.2260456	.1232828	1.83	0.067	0155842	.4676755

• Report the QA statistic for instrument validity and the conditional F-statistics for instrument strength for each phenotype (Sanderson et al. 2019; Sanderson, Spiller, and Bowden 2020).

mvmr chdbeta ldlcbeta hdlcbeta tgbeta [aw=1/(chdse^2)] if sel1==1, gxse(ldlcse hdlcse tgse)

> e hdlcse tgse)

Coef. Std. Err.

Number of genotypes = 73

Number of phenotypes = 3

Standard errors: Random effect
Residual standard error = 1.490

z P>|z| [95% Conf. Interval]

```
chdbeta | 1dlcbeta | .42862 .0609661 7.03 0.000 .3091286 .5481113 hdlcbeta | -.1941989 .1308289 -1.48 0.138 -.4506189 .0622211 tgbeta | .2260456 .1232828 1.83 0.067 -.0155842 .4676755
```

 Q_A statistic for instrument validity; chi2(70) = 152.88 (p = 0.0000) Conditional F-statistics for instrument strength:

F_x1 = 130.31 (ldlcbeta) F_x2 = 36.29 (hdlcbeta) F_x3 = 40.44 (tgbeta)

• Fit an MVMR-Egger regression (Rees, Wood, and Burgess 2017), orienting the model with respect to the first phenotype in the main *varlist*.

mrmvegger chdbeta ldlcbeta hdlcbeta tgbeta [aw=1/(chdse^2)] if sel1==1

MVMR-Egger model oriented wrt: ldlcbeta

Number of genotypes = 73

Number of phenotypes = 3

Residual standard error = 1.469

	Coef.	Std. Err.	z			Interval]
chdbeta						
ldlcbeta	.5672993	.1002611	5.66	0.000	.370791	.7638075
hdlcbeta	1364113	.1332727	-1.02	0.306	3976209	.1247983
tgbeta	.2739803	.1246927	2.20	0.028	.0295871	.5183735
_cons	0093655	.0054187	-1.73	0.084	019986	.001255

We can also orient the model with respect to HDL-C.

```
mrmvegger chdbeta ldlcbeta hdlcbeta tgbeta [aw=1/(chdse^2)] if sel1==1, orient(2)
```

> t(2)

MVMR-Egger model oriented wrt: hdlcbeta

Number of genotypes = 73

Number of phenotypes = 3

Residual standard error = 1.501

	Coef.	Std. Err.	z	P> z		Interval]
chdbeta						
ldlcbeta	.4286398	.0614056	6.98	0.000	.308287	.5489926
hdlcbeta	1989637	.1541909	-1.29	0.197	5011723	.1032449
tgbeta	.2256794	.1243221	1.82	0.069	0179875	.4693463
_cons	.0002155	.0036218	0.06	0.953	006883	.0073141

Or we can orient the model with respect to triglycerides.

> t(3)

MVMR-Egger model oriented wrt: tgbeta Number of genotypes = 73 Number of phenotypes = 3 Residual standard error = 1.499

	Coef.	Std. Err.	z	P> z	[95% Conf.	Interval]
chdbeta						
ldlcbeta	.4203073	.0660026	6.37	0.000	.2909447	.54967
hdlcbeta	1903089	.1321536	-1.44	0.150	4493252	.0687075
tgbeta	.2065651	.1365427	1.51	0.130	0610537	.474184
_cons	.0013499	.003951	0.34	0.733	0063939	.0090936

R session information for reproducibility

library(sessioninfo) session_info()

```
- Session info -----
setting value
version R version 4.1.1 (2021-08-10)
     Windows 10 x64
```

system x86_64, mingw32 RTerm ui language (EN)

collate English United Kingdom. 1252

collate	English_United	Kingdom.	1252						
ctype	English_United	Kingdom.	1252						
tz	Europe/London								
date	2021-09-20								
- Packages	3							 	
package		* version	date	lib	sour	се			
arrangeme	ents	1.1.9	2020-09-13	[1]	CRAN	(R	4.1.0)		
asserttha	at	0.2.1	2019-03-21	[1]	CRAN	(R	4.1.0)		
backports	3	1.2.1	2020-12-09	[1]	CRAN	(R	4.1.0)		
broom		0.7.9	2021-07-27	[1]	CRAN	(R	4.1.0)		
cellrange	er	1.1.0	2016-07-27	[1]	CRAN	(R	4.1.0)		
cli		3.0.1	2021-07-17	[1]	CRAN	(R	4.1.0)		
codetools	3	0.2-18	2020-11-04	[2]	CRAN	(R	4.1.1)		
colorspac	ce	2.0-2	2021-06-24	[1]	CRAN	(R	4.1.0)		
conquer		1.0.2	2020-08-27	[1]	CRAN	(R	4.1.0)		
crayon		1.4.1	2021-02-08	[1]	CRAN	(R	4.1.0)		
curl		4.3.2	2021-06-23	[1]	CRAN	(R	4.1.0)		
data.tab]	Le	1.14.0	2021-02-21	[1]	CRAN	(R	4.1.0)		
DBI		1.1.1	2021-01-15	[1]	CRAN	(R	4.1.0)		
dbplyr		2.1.1	2021-04-06	[1]	CRAN	(R	4.1.0)		

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                          0.6.27
                                  2020-10-24 [1] CRAN (R 4.1.0)
digest
dplyr
                        * 1.0.7
                                  2021-06-18 [1] CRAN (R 4.1.0)
                          0.3.2
                                  2021-04-29 [1] CRAN (R 4.1.0)
ellipsis
evaluate
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                                  2019-05-28 [1] CRAN (R 4.1.0)
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                                  2021-05-25 [1] CRAN (R 4.1.0)
fansi
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fastmap
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forcats
foreach
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fs
generics
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                        * 3.3.5
ggplot2
glmnet
                          4.1 - 2
                                  2021-06-24 [1] CRAN (R 4.1.0)
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                                  2020-05-19 [1] CRAN (R 4.1.0)
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                                  2018-06-12 [1] CRAN (R 4.1.0)
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                          4.9.4.1 2021-06-18 [1] CRAN (R 4.1.0)
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```

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                      1.4.6 2021-05-19 [1] CRAN (R 4.1.0)
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SparseM
                       1.81
                               2021-02-18 [1] CRAN (R 4.1.0)
Statamarkdown
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                      1.7.4 2021-08-25 [1] CRAN (R 4.1.1)
                     * 1.4.0 2019-02-10 [1] CRAN (R 4.1.0)
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survival
                       3.2-13 2021-08-24 [2] CRAN (R 4.1.1)
                     * 3.1.4 2021-08-25 [1] CRAN (R 4.1.1)
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withr
                       2.4.2 2021-04-18 [1] CRAN (R 4.1.0)
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                       0.26 2021-09-14 [1] CRAN (R 4.1.1)
                       1.3.2 2020-04-23 [1] CRAN (R 4.1.0)
xml2
yaml
                       2.2.1 2020-02-01 [1] CRAN (R 4.1.0)
```

- [1] C:/Users/eptmp/Documents/R/win-library/4.1
- [2] C:/Program Files/R/R-4.1.1/library

Stata session information for reproducibility

```
di c(version)
ado describe mrrobust
16.1
[89] package mrrobust from https://raw.github.com/remlapmot/mrrobust/master
______
TITLE
     'mrrobust': Stata package for two-sample Mendelian randomization analyses
DESCRIPTION/AUTHOR(S)
     Author: Tom Palmer
     Distribution-Date: 20210917
INSTALLATION FILES
     m\mrmedian.ado
     m\mrmedian.sthlp
     m\mbox{\em mrmedianobs.ado}
     m\mrmedianobs work.ado
     m\mrmedianobs.sthlp
     m\mregger.ado
     m\mregger.sthlp
```

```
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m\mrrobust-author.ihlp
```

INSTALLED ON

20 Sep 2021

References

Burgess, S, F Dudbridge, and SG Thompson. 2015. "Multivariable Mendelian randomization: the use of pleiotropic genetic variants to estimate causal effects." *American Journal of Epidemiology* 181: 251--260. https://doi.org/10.1093/aje/kwu283.

Rees, J, A Wood, and S Burgess. 2017. "Extending the MR-Egger method for multivariable Mendelian randomization to correct for both measured and unmeasured pleiotropy." Statistics in Medicine 36: 4705–18. https://doi.org/10.1002/sim.7492.

Sanderson, E, G Davey Smith, F Windmeijer, and J Bowden. 2019. "An examination of multivariable Mendelian randomization in the single-sample and two-sample summary data settings." *International Journal of Epidemiology* 48: 713–27. https://doi.org/10.1093/ije/dyy262.

Sanderson, E, W Spiller, and J Bowden. 2020. "Testing and Correcting for Weak and Pleiotropic Instruments in Two-Sample Multivariable Mendelian Randomisation." bioRxiv. https://doi.org/10.1101/2020.04.02.021980.