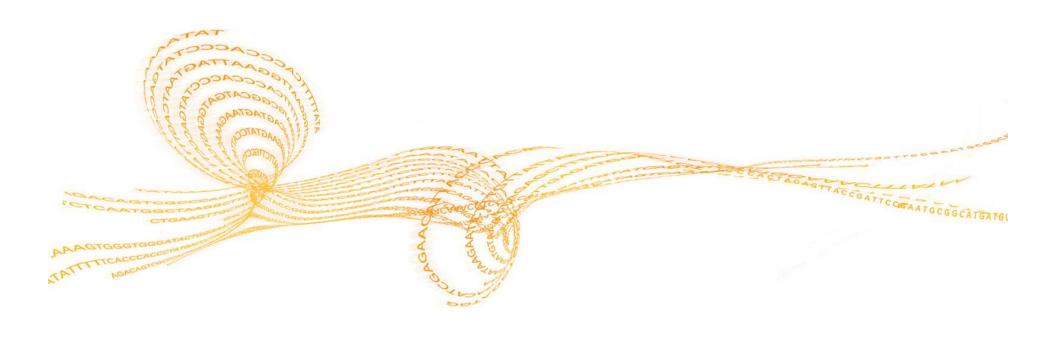


Overview of tailed amplicon sequencing approach with MiSeq

Jonathan Bell





Overview Example Workflow

Three main steps

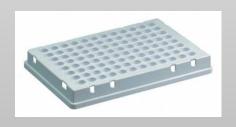
PCR#1

- Amplify genomic regions of interest
- Pool all amplicons from the same sample into a single pool (e.g. 5 single plex PCR reactions pooled into one sample)



PCR #2

- Amplify pooled amplicons from Step 1 using indexed adapter oligos from ILMN
- Produces barcoded amplicons ready for MiSeq
- Pool up to 96 samples into a single library



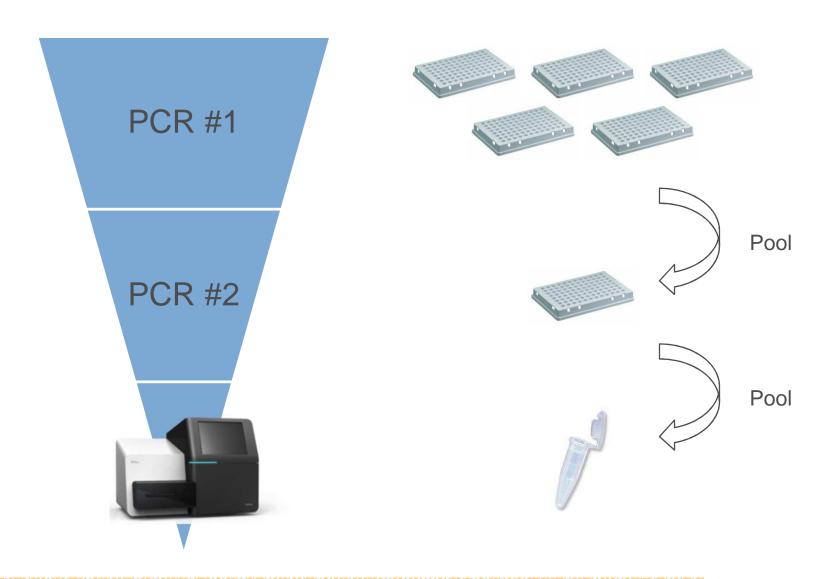
MiSeq

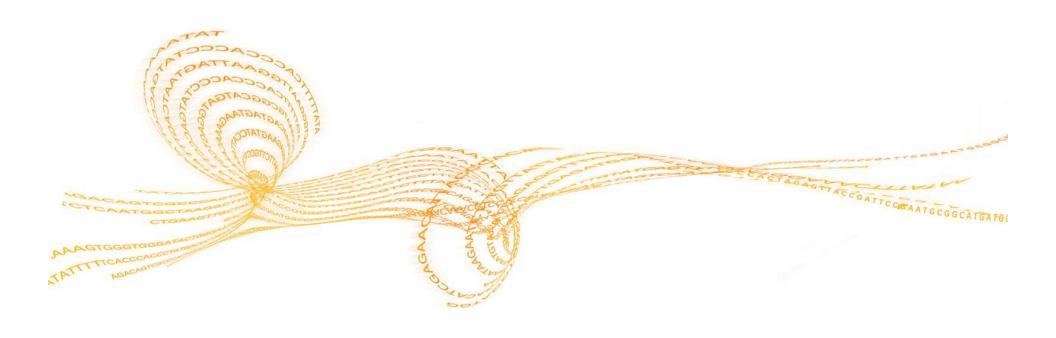
- Sequence pooled library of up to 96 samples in a single MiSeq run
- MiSeq analysis software automatically demultiplexes data to uniquely assign reads to samples



Throughput advantage of NGS

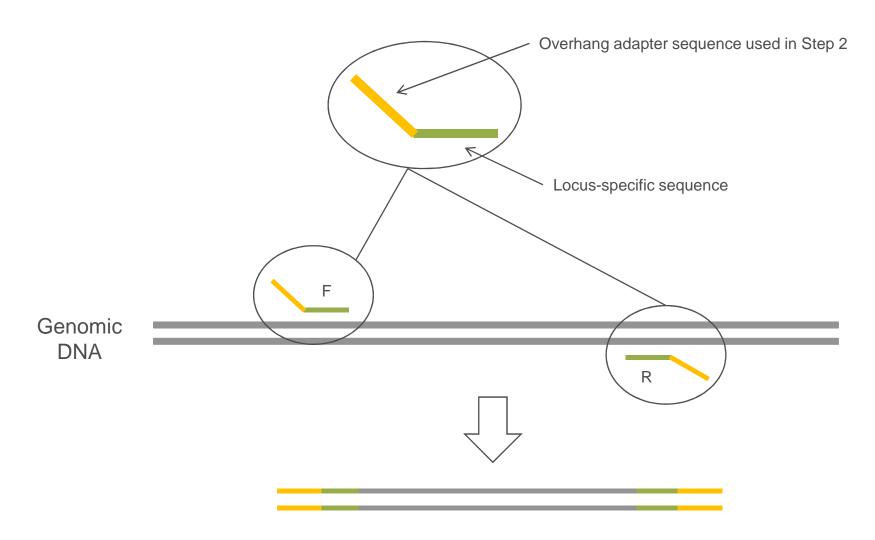
Example project of 5 amplicons per sample, 96 sample project





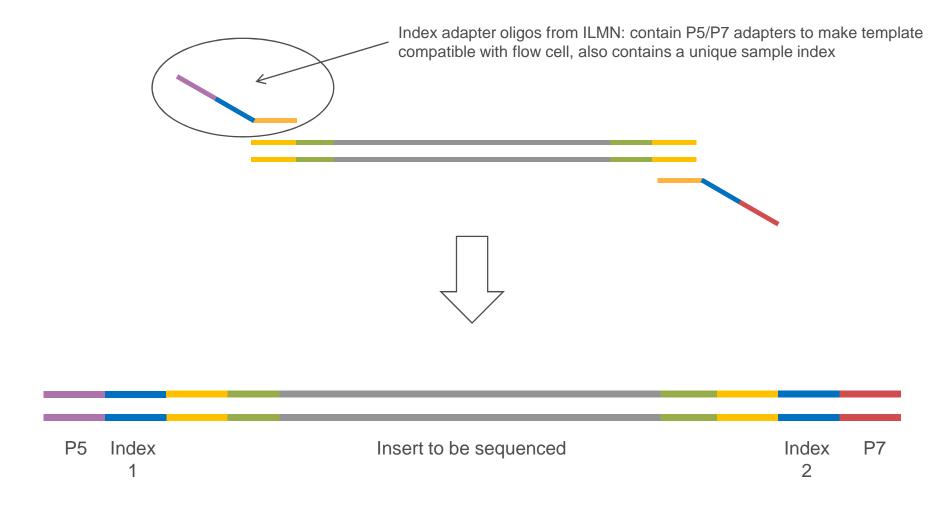
Detailed breakdown

Step 1: PCR to amplify regions of interest



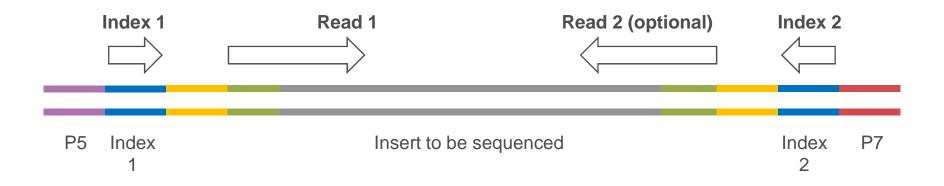


Step 2: 2nd round of PCR to add ILMN indices and sequencing adapters





Step 3: Sequence on MiSeq



Sequencing order on MiSeq system

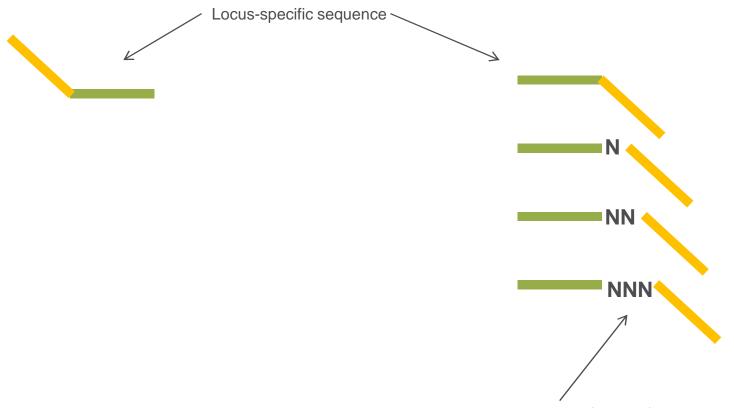
- Read 1 sequence amplicons in Forward direction up to 150 bp
- Index 1 read first barcode
- Index 2 read second barcode (software can now uniquely identify the sample)
- Read 2 sequence amplicons in Reverse direction up to 150 bp

What materials are necessary?

	Description	Step used	Customer Provided	ILMN Provided
Locus-specific PCR primers	Used to amplify genomic regions of interest to be sequenced on MiSeq (e.g. 5 exons from a sample). Oligo primers include overhang adapter sequence	Step 1	✓	
Index Adapter Primers for second PCR	Used to add sequencing adapters and sample- specific indices to samples Nextera Index Kit (96 Indices, 384 Samples) FC-121-1012	Step 2		\checkmark
PCR Reagents	Mastermix (includes nucleotides and polymerase) for PCR reactions	Steps 1 & 2	✓	
Sequencing Reagents	TruSeq SBS kits for sequencing with MiSeq System MiSeq Reagent Kit (300-cycles – PE) MS-102-1001	Step 3		✓



Primer considerations for PCR #1



Amplicons will be variable length (+0-3 bp) resulting in even distribution of all four bases for any one sequencing cycle

1 Forward PCR primer

4 Reverse PCR primers



Sequences of Round 1 PCR Primers: Assuming complex, or mixed, types of amplicons

- Append to 5' end of forward PCR primer:
- ▶ 5' TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-[locus specific sequence]
- Append to 5' end of reverse PCR primers:
- ▶ 5' GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-[locus specific sequence]

Sequences of Round 1 PCR Primers: For simple amplicons with very low complexity

- Append to 5' end of forward PCR primer:
- 5' TCGTCGGCAGCGTCAGATGTGTATAAGAGACAG-[locus specific sequence]
- ▶ 5' TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGN-[locus specific sequence]
- ▶ 5' TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGNN-[locus specific sequence]
- ▶ 5' TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGNNN-[locus specific sequence]
- Append to 5' end of reverse PCR primers:
- 5' GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAG-[locus specific sequence]
- 5' GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGN-[locus specific sequence]
- ▶ 5' **GTCTCGTGGGCTCGG**AGATGTGTATAAGAGACAGNN-[locus specific sequence]
- ▶ 5' GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGNNN-[locus specific sequence]
- ▶ 14 or 15 nt PCR Overlap Sequences
- N, NN, and NNN are mixed sequence bases added to introduce sequence complexity

