

Reactive Oxygen Species Regulate Myocardial Mitochondria through Post-Translational Modification

Nan Hu and Jun Ren

Center for Cardiovascular Research and Alternative Medicine, School of Pharmacy, University of Wyoming College of Health Sciences, Laramie, WY 82071, USA

Correspondence: jren@uwyo.edu (J.R.)

Hu N and Ren J. Reactive Oxygen Species 2(4):264–271, 2016; ©2016 Cell Med Press

<http://dx.doi.org/10.20455/ros.2016.845>

(Received: March 4, 2016; Revised: March 31, 2016; Accepted: April 1, 2016)

ABSTRACT | Post-translational modifications (PTMs) generally refer to covalent and enzymatic modifications of proteins during or after protein biosynthesis, as well as during cleavage or degradation of proteins. These modification processes govern not only protein homeostasis but also new cellular protein functions. As an essential part of complex cellular signal sensing and transduction networks, PTMs respond to various reactive oxygen species (ROS) and contribute to multi-organ injuries although the precise underlying mechanisms of action remain poorly understood. Up to date, a number of PTMs have been postulated to participate in the regulation of mitochondrial function through free radical species and other second messengers. Mitochondrion is believed to serve as a main ROS generator, and at the same time, also an ROS receptor. Modification of mitochondrial proteins may result in detrimental biological consequences through oxidative injury and mitochondrial dysfunction. In this mini-review we will recapitulate some aspects of PTMs in the control of mitochondrial integrity in the heart. The major forms of PTM related to mitochondrial regulation will be discussed here, which include phosphorylation, ubiquitination, acetylation, and oxidative and nitrosative modification.

KEYWORDS | Mitochondria; Oxidative stress; Post-translational modification

ABBREVIATIONS | CoQ10, coenzyme Q10; ETC, electron transport chain; NO, nitric oxide; OXPHOS, oxidative phosphorylation; Ox-PTM, oxidative post-translational modification; PDH, pyruvate dehydrogenase; PTM, Post-translational modification; RNS, reactive nitrogen species; ROS, reactive oxygen species; SNO, S-nitrosylation; STAT3, signal transducer and activator of transcription 3; Ub, ubiquitin

CONTENTS

1. Introduction
2. Phosphorylation
3. Ubiquitination
4. Acetylation
5. Oxidative Modification
6. Nitrosative Modification

7. Conclusion

1. INTRODUCTION

Heart diseases impose a major health threat with a rather complex etiology. A number of mechanisms have been postulated and widely accepted for the onset and progression of heart diseases including genetic and epigenetic alterations in response to various environmental cues [1–3]. Although recent advances in genetics have greatly enriched our understanding of disease mechanisms [4, 5], various factors beyond genetic coding or control, such as post-translational modifications (PTMs) of essential proteins, may also contribute to the cardiac homeostasis and therefore the etiology of heart diseases [6, 7]. PTMs of proteomes are believed to provide the pieces needed to complete the puzzle between unaltered gene levels and changes in metabolism, morphology, and function for the heart [7]. In particular, PTMs have been demonstrated to link regulation of myocardial metabolic or functional changes with energy protein enzymatic activity (e.g., mitochondrial oxidative capacity) [7]. Mitochondria are known as the dynamic organelles for generation of cardiac fuel (ATP) and as the key regulators for main cellular survival function including autophagy, apoptosis, intracellular calcium homeostasis, and lipid metabolism [8]. Many of these cellular machineries help to maintain mitochondrial integrity and function via mitochondrial quality control in various cardiac stress conditions [9]. Mitochondrial variations and uncoupling can function to promote the production of reactive oxygen species (ROS) and thus further exacerbate cardiac pathologies, or trigger pivotal signaling components for development of cardioprotection [10].

Regulation of post-translational protein modifications is paramount to cardiovascular development, homeostasis, and stress response [7]. In general, such protein regulations can be categorized into two major groups: amino acid modification and group attachment [11]. PTMs usually refer to an addition of small chemical moieties to amino acid residues without interfering with DNA coding sequence. These processes can be shown as phosphorylation, ubiquitination, acetylation, oxidative/nitrosative modifications, hydroxylation, and sulfidation [12]. While these PTMs of essential functional proteins may lead to specific cellular and biological events in the heart and other

organs, one main challenge for scientists is how to precisely define the direct functional implications of these modifications [13]. Defining the outcome of modification of a given protein can be challenging as pan protein levels, degree of modification, localization, and activity of the functional component of the protein may contribute to the final pathophysiological outcomes, letting alone the existence of off-target modification for other proteins. More recent evidence has depicted that environmental factors such as nutrients and levels of ROS may help to govern the on/off switch of PTMs for various downstream signal transduction pathways [7].

Ample clinical and experimental evidence has demonstrated the unique role for ROS-triggered modifications of functional proteins as the main risk factors for oxidative damage in pathological cardiovascular processes [7, 14]. However, the ROS-governed modification theory has been challenged as merely mitigating existing ROS by antioxidants is ineffective to decrease mortality of heart failure [15, 16]. Among various antioxidants with somewhat dismal efficacy, coenzyme Q10 (CoQ10) has shown some clinical promises in the therapeutics of cardiovascular diseases [17, 18]. CoQ10, an endogenously synthesized and dietary lipid-soluble cofactor, acts to transfer electrons from complexes I and II to complex III in the mitochondrial inner membrane, thus enabling CoQ10 to function as a membrane antioxidant and benefit congestive heart failure [17]. One of the hypotheses for the lack of success for antioxidants in heart diseases is possibly related to the inability of “regular” antioxidants to scavenge ROS present within mitochondria. To this end, more effective therapeutic strategies are needed to drastically improve the capacity of mitochondrial quality control in an effort to maintain cardiac function [19]. It has been well established that defective or aged mitochondria can have severe consequences by releasing ROS and cytochrome c into cytosol, inducing apoptosis of cardiomyocytes [20]. Mitochondrial quality control is succinctly controlled by a wide array of factors ranging from mitochondrial dynamics to mitochondrial biogenesis and degradation at various points throughout the life course. These procedures may be grouped under the “mitochondrial biogenesis” umbrella although such dynamic chang-

es are not restricted to newborn mitochondria. In fact, clearance of impaired mitochondrial fragments into the cytosol is well executed through the process of mitophagy [21]. Coordination of mitochondria dynamics in an effort to regulate ROS levels requires not only the mitochondrial outer membrane but also attachment of proteins formed in the cytoplasm [22]. In pathological conditions, intra- or extracellular ROS may regulate mitochondrial ion channels/transporters by way of redox-dependent PTMs in a vicious cycle. These redox-sensitive PTMs may be greatly amplified by mitochondrial ROS production, causing another surge of ROS release from the neighboring mitochondria, and ultimately imposing more PTMs and stress in the mitochondria [23]. Nevertheless, little is known about these mitochondrial protein modifications, for instance, the mitochondrial quality control by ubiquitination or other PTMs. In the following sections, we will briefly review and discuss the critical role of ROS and PTMs in regulating mitochondrial quality control in the pathophysiological process of cardiac injury.

2. PHOSPHORYLATION

Phosphorylation is perhaps one of the most common and well-studied posttranslational modifications in mammals [24]. Phosphorylation involves the addition of a reversible bond of a phosphate ester onto given amino acid residues, including serine (Ser), threonine (Thr), and tyrosine (Tyr). Recent evidence revealed a new link between phosphorylation status and dynamic control in the mitochondrial proteome [13]. Mitochondria are well-known as myocardial “powerhouse” organelles for ATP production. Meanwhile, mitochondria also serve as important regulators for multiple redox signaling pathways as majority of mitochondrial ATP is produced by metabolism of fatty acids and oxidative phosphorylation (OXPHOS) via mitochondrial electron transport chain (ETC) [25]. A small portion of electrons escapes from the ETC to reduce molecular oxygen during OXPHOS, generating superoxide anion ($O_2^{\cdot-}$) continuously, and $O_2^{\cdot-}$ serves as the primary oxygen free radical in mitochondria [23]. OXPHOS defects in phosphorylation have been considered as a main trigger for cardiomyopathy and heart failure due to uncontrollable ROS production, and complications of mitochondrial encephalopathy, neuropathy, and

myopathy [26]. Although lipids may serve as a main dynamic pool for fatty acids and mitochondrial fuels, they may also function as uncouplers and inhibitors of oxidative phosphorylation [27]. A theory of lipid utilization by mitochondria has been put forward, which helps to maintain ROS within levels compatible with signaling while producing robust and reliable energy supply at the same time [27].

Other than mitochondrial oxidative phosphorylation, other phosphorylation processes also contribute to ROS production and change of mitochondrial function. For example, phosphorylation status and pan-protein levels of signal transducer and activator of transcription 3 (STAT3), a transcriptional molecule, may be dynamic and regulate mitochondrial function. STAT3 is phosphorylated at various residues with functional consequences to protect against oxidative stress [28, 29]. Moreover, evidence from superoxide dismutase 2 (SOD2)-transgenic mice confirmed reduced superoxide production and repressed pyruvate dehydrogenase (PDH) activity and mitochondrial ETC activity in diabetes mellitus. PDH activity, or its phosphorylation, may prevent mitochondrial pyruvate uptake therefore interrupting energy metabolism and promoting ROS production [30]. Elevated PDH phosphorylation was observed in diabetic hearts [31, 32], which may greatly contribute to the overt ROS production and mitochondrial damage along with cardiomyopathy in diabetic hearts [33, 34].

3. UBIQUITINATION

Ubiquitin (Ub), a conserved 76-amino acid globular protein, participates in protein degradation, a process known as ubiquitination essential for mitochondrial quality control. Ubiquitination and Ub receptor proteins (e.g., p62 and ubiquilins) are considered key factors to target misfolded proteins for quality control in a varieties of entities, including the proteasome, lysosomes, and mitophagy to remove defective mitochondria [35]. Ubiquitination process involves a family of Ub-conjugating proteins [36]. An early step for ubiquitination is the ATP-dependent activation of free Ub to form a thiol-ester tie between the protein E1 (a Ub-activating enzyme) and the carboxyl terminals of Ub prior to the transfer to one of the existing E2 (Ub-conjugating enzyme) enzymes. There, E2s ligate with the Ub protein lig-

ase E3 enzymes before attaching onto target proteins. This process normally causes both mono- and multiple-Ub attachments to target proteins, and prepares them for degradation by proteasome [37].

Disrupted mitochondrial dynamics is intimately related to disturbed cellular homeostasis and is linked to accumulation of ROS and onset of various cardiac diseases, including cardiac hypertrophy, heart failure, alcoholic cardiomyopathy, dilated cardiomyopathy, and ischemic heart disease [37, 38]. For example, ROS accumulation in mitochondria is known to lead to increased ubiquitination of SOD2 in type 2 diabetic mice [39]. Ubiquitin ligases and de-ubiquitinating enzymes participate in the regulation of mitochondrial fission and fusion and directly contribute to the pathophysiology of heart diseases through the ubiquitin ligase seven in absentia homolog 2 (Siah2) [37]. Siah2 is a RING finger ligase to regulate prolyl hydroxylases PHD3 and PHD1 under hypoxia, thereby turning on the master regulator of hypoxia—hypoxia-inducible factor (HIF)- α . Recent studies have identified a role for the ubiquitin ligase Siah2 in the control of mitochondrial membrane protein degradation by proteasome, therefore protecting hearts against oxidative stress injury [36]. These authors found that hypoxia-induced mitochondrial fission is mediated by the mitochondrial scaffolding protein A kinase anchor protein 1 (AKAP121). AKAP121 may serve as a substrate of Siah2 for the maintenance of mitochondrial dynamics through modulation of the phosphorylation of dynamin-related protein 1 (Drp1) and the PKA-independent inhibition of the Drp1-Fis1 complex under cardiac stress. Ubiquitination and proteasomal degradation of these proteins were found to play a key role in the regulation of mitochondrial dynamics [37].

4. ACETYLATION

Lysine acetylation is a type of post-translational modifications to regulate mitochondrial homeostasis. Sirtuins, belonging to the family of class III histone deacetylases, require NAD⁺ as a metabolic cofactor pertinent to enzymatic function, cellular stress response and, oxidative stress stimuli [40]. Sirtuins contain seven NAD⁺-dependent protein deacetylases (Sirt1–7). Among them, mitochondrial Sirt3 plays a role in mitochondrial lysine acetylation and contributes to the antioxidant defense leading to preserved

mitochondrial integrity. The beneficial effects of caloric restriction on oxidative stress are reported to rely on the antioxidant property of Sirt3 [41]. In response to stress, mitochondrial complex I deficiency leads to ROS accumulation and decreased NAD⁺/NADH ratio in the heart, thus inhibiting Sirt3 activity and favoring protein acetylation [42]. Doxorubicin, a widely used anticancer drug, is known to induce cardiac mitochondrial damage through up-regulated ROS level. However, this anti-neoplastic agent triggered depletion of Sirt3, enormous acetylation of mitochondrial proteins, and mitochondrial DNA damage [43]. Along the same line, deletion of Sirt3 was associated with exacerbated cardiac hypertrophy in doxorubicin-induced dilated cardiomyopathy, and with increased cardiovascular risk factors in low-density lipoprotein (LDL) receptor-knockout mice [43, 44].

Manganese SOD (MnSOD, also known as SOD2) is the essential mitochondrial ROS scavenging enzyme that converts superoxide to hydrogen peroxide (H₂O₂). H₂O₂ is subsequently converted to water by catalase and other peroxidases. It was recently shown that MnSOD activity is tightly regulated by reversible acetylation of evolutionarily conserved lysine(s) [40]. Interestingly, overexpression of Sirt3 decreased cellular levels of acetylation and ROS, activated mitochondrial SOD, and up-regulated major ROS-detoxifying enzymes [1, 45]. Importantly, as a member of the sirtuin family of NAD-dependent deacetylases, Sirt3 is found to preserve cardiac function by way of mitochondrial SOD and transcription factor FoxO3A [41, 46].

5. OXIDATIVE MODIFICATION

Among all post-translational modifications, the oxidative post-translational modification (Ox-PTM) of mitochondrial proteins is perhaps the most likelihood event triggered by ROS [47]. Oxidation refers to a unique covalent PTM of products by ROS, or the secondary products from oxidative stress, which are produced by various metabolic processes [48]. Oxidation can occur in a number of amino acid residues, including histidine, asparagine, lysine, tyrosine, and cysteine. Ox-PTM of the cysteine residues is an essential event to alter protein structure and then cellular function. The exclusive feature of a cysteine side chain permits a range of Ox-PTMs, leading to multi-

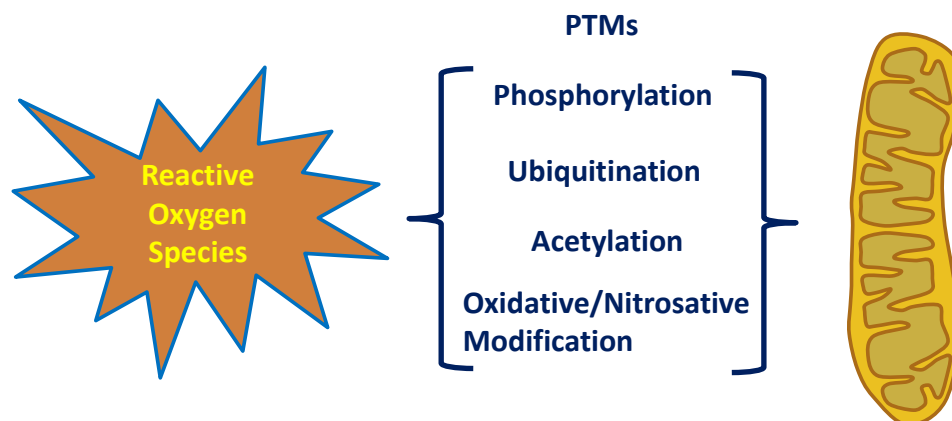


FIGURE 1. Mitochondrial quality control via PTMs. As illustrated, post translational modifications (PTMs) of proteins are critical for regulating mitochondrial biological activity, and they occur mostly via phosphorylation, ubiquitination, acetylation, and oxidative and nitrosative modification.

ple regulatory consequences. Up to date, the majority of cysteine Ox-PTMs involve the reversible reaction between diffusible small molecules and free thiol groups located on a cysteine side chain. These reactions may be reversed by the antioxidant enzyme (back to free thiols) or further converted to other Ox-PTMs dependent on the surrounding redox environment [49].

Oxidative protein modification plays a key role in the onset and development of heart failure [48, 49]. It is suggested that tryptophan residues in mitochondria can be the main targets of oxidative modification [50]. Not surprisingly, one hallmark for heart failure is progressive ROS production. Given that heart failure is associated with progressive ROS production, it is likely that mitochondrial quality control may serve as a safeguard for oxidative damage in cardiomyopathy and ultimately heart failure [49, 51].

6. NITROSATIVE MODIFICATION

Some post-translational modifications arise from the attachment of chemical components produced physiologically inside cells, such as nitric oxide (NO). NO acts as a second messenger, originating a wide range of reactive nitrogen species (RNS) [11]. NO-driven reactions can lead to several modification residues, such as the reaction of NO with ROS to form RNS switches from nitration to S-nitrosylation (SNO) of

proteins. In fact, SNO of proteins is a specific PTM where the NO moiety covalently binds to the free cysteine residues of the target protein, which plays an critical part in heart health, especially within myocardial mitochondria [52]. It is understood that unstable levels of ROS/RNS are responsible for cardiac cytotoxicity, which then contribute to contractile dysfunction and heart failure. Indeed, SNO is of principal significance in either promoting or restricting ROS/RNS generation or subsequently reperfusion injury. Critical kinases may be turned on by ROS/RNS to elicit cardioprotection [53]. This process should be able to prevent mitochondrial transition pore opening and ROS-induced cytokine release. Finally, SNO may also represent a shield against oxidative stress, and thus nitrosylated protein may shield cells away from an irreversible oxidation and burst of ROS/RNS challenge [52].

In addition, the S-nitrosylated proteins work as a source of NO as intracellular signaling molecule in cardiomyocyte mitochondria [54]. It seems that most effects of NO are mediated by SNO of essential proteins, a covalent modification process between protein cysteine thiols and NO molecules to form S-nitrosothiol. Mitochondria were found to possess abundance of these tyrosine nitrated proteins as evidenced by enhanced 3-nitrotyrosine staining in the hearts following ischemia-reperfusion challenge [55]. This SNO process may be either an indication of PTM alteration or damaged mitochondrial quality

control in response to myocardial ischemia-reperfusion challenge [55, 56].

7. CONCLUSION

A steadily growing body of evidence points to an essential role of PTMs in mitochondrial homeostasis (Figure 1), due in part to the advance of new technologies. Nonetheless, accurate identification of the modified amino acid residues is still challenging at the present time. Such residue identifications are pertinent to the direct evaluation of the functional outcomes of any given PTMs, such as Ox-PTMs. In addition, elucidation of the precise enzymes responsible for the PTM mechanisms is also pivotal for the understanding of PTMs. This will lead to the possible advancement of therapies to manipulate PTMs along with mitochondrial function to combat against ROS over production and oxidative injury.

REFERENCES

1. Winnik S, Auwerx J, Sinclair DA, Matter CM. Protective effects of sirtuins in cardiovascular diseases: from bench to bedside. *Eur Heart J* 2015; doi: 10.1093/eurheartj/ehv290.
2. Zhang Y, Ren J. Epigenetics and obesity cardiomyopathy: from pathophysiology to prevention and management. *Pharmacol Ther* 2016; doi: 10.1016/j.pharmthera.2016.03.005.
3. Napoli C, Grimaldi V, De Pascale MR, Sommesse L, Infante T, Soricelli A. Novel epigenetic-based therapies useful in cardiovascular medicine. *World J Cardiol* 2016; 8(2):211–9. doi: 10.4330/wjc.v8.i2.211.
4. Azhar M, Ware SM. Genetic and developmental basis of cardiovascular malformations. *Clin Perinatol* 2016; 43(1):39–53. doi: 10.1016/j.clp.2015.11.002.
5. Edwards JJ, Gelb BD. Genetics of congenital heart disease. *Curr Opin Cardiol* 2016. doi: 10.1097/HCO.0000000000000274.
6. Lee A, Oh JG, Gorski PA, Hajjar RJ, Kho C. Post-translational modifications in heart failure: small changes, big impact. *Heart Lung Circ* 2016; 25(4):319–24. doi: 10.1016/j.hlc.2015.11.008.
7. Wende AR. Post-translational modifications of the cardiac proteome in diabetes and heart failure. *Proteomics Clin Appl* 2016; 10(1):25–38. doi: 10.1002/prca.201500052.
8. Shirihai OS, Song M, Dorn GW, 2nd. How mitochondrial dynamism orchestrates mitophagy. *Circ Res* 2015; 116(11):1835–49. doi: 10.1161/CIRCRESAHA.116.306374.
9. Cao DJ. Epigenetic regulation and heart failure. *Expert Rev Cardiovasc Ther* 2014; 12(9):1087–98. doi: 10.1586/14779072.2014.942285.
10. Kornfeld OS, Hwang S, Disatnik MH, Chen CH, Qvit N, Mochly-Rosen D. Mitochondrial reactive oxygen species at the heart of the matter: new therapeutic approaches for cardiovascular diseases. *Circ Res* 2015; 116(11):1783–99. doi: 10.1161/CIRCRESAHA.116.305432.
11. Victorino VJ, Mencalha AL, Panis C. Post-translational modifications disclose a dual role for redox stress in cardiovascular pathophysiology. *Life Sci* 2015; 129:42–7. doi: 10.1016/j.lfs.2014.11.008.
12. Liddy KA, White MY, Cordwell SJ. Functional decorations: post-translational modifications and heart disease delineated by targeted proteomics. *Genome Med* 2013; 5(2):20. doi: 10.1186/gm424.
13. Kane LA, Van Eyk JE. Post-translational modifications of ATP synthase in the heart: biology and function. *J Bioenerg Biomembr* 2009; 41(2):145–50. doi: 10.1007/s10863-009-9218-6.
14. Kho C, Lee A, Jeong D, Oh JG, Gorski PA, Fish K, et al. Small-molecule activation of SERCA2a SUMOylation for the treatment of heart failure. *Nat Commun* 2015; 6:7229. doi: 10.1038/ncomms8229.
15. Arcaro A, Pirozzi F, Angelini A, Chimenti C, Crotti L, Giordano C, et al. Novel perspectives in redox biology and pathophysiology of failing myocytes: modulation of the intramyocardial redox milieu for therapeutic interventions—a review article from the Working Group of Cardiac Cell Biology, Italian Society of Cardiology. *Oxid Med Cell Longev* 2016; 2016:6353469. doi: 10.1155/2016/6353469.
16. Costa S, Reina-Couto M, Albino-Teixeira A, Sousa T. Statins and oxidative stress in chronic heart failure. *Rev Port Cardiol* 2016; 35(1):41–57. doi: 10.1016/j.repc.2015.09.006.
17. DiNicolantonio JJ, Bhutani J, McCarty MF, O'Keefe JH. Coenzyme Q10 for the treatment of heart failure: a review of the literature. *Open Heart* 2015; 2(1):e000326. doi: 10.1136/openhrt-2015-000326.

18. Wold LE, Muralikrishnan D, Albano CB, Norby FL, Ebadi M, Ren J. Insulin-like growth factor I (IGF-1) supplementation prevents diabetes-induced alterations in coenzymes Q9 and Q10. *Acta Diabetol* 2003; 40(2):85–90. doi: 10.1007/s005920300010.
19. Liang Q, Kobayashi S. Mitochondrial quality control in the diabetic heart. *J Mol Cell Cardiol* 2015. doi: 10.1016/j.yjmcc.2015.12.025.
20. Carreira RS, Lee P, Gottlieb RA. Mitochondrial therapeutics for cardioprotection. *Curr Pharm Des* 2011; 17(20):2017–35.
21. Zhu J, Wang KZ, Chu CT. After the banquet: mitochondrial biogenesis, mitophagy, and cell survival. *Autophagy* 2013; 9(11):1663–76. doi: 10.4161/auto.24135.
22. Dominy JE, Puigserver P. Mitochondrial biogenesis through activation of nuclear signaling proteins. *Cold Spring Harb Perspect Biol* 2013; 5(7). doi: 10.1101/cshperspect.a015008.
23. J OU, Ryu SY, Jhun BS, Hurst S, Sheu SS. Mitochondrial ion channels/transporters as sensors and regulators of cellular redox signaling. *Antioxid Redox Signal* 2014; 21(6):987–1006. doi: 10.1089/ars.2013.5681.
24. Wang YC, Peterson SE, Loring JF. Protein post-translational modifications and regulation of pluripotency in human stem cells. *Cell Res* 2014; 24(2):143–60. doi: 10.1038/cr.2013.151.
25. Mailloux RJ, Jin X, Willmore WG. Redox regulation of mitochondrial function with emphasis on cysteine oxidation reactions. *Redox Biol* 2014; 2:123–39. doi: 10.1016/j.redox.2013.12.011.
26. Fosslien E. Review: Mitochondrial medicine: cardiomyopathy caused by defective oxidative phosphorylation. *Ann Clin Lab Sci* 2003; 33(4):371–95.
27. Aon MA, Bhatt N, Cortassa SC. Mitochondrial and cellular mechanisms for managing lipid excess. *Front Physiol* 2014; 5:282. doi: 10.3389/fphys.2014.00282.
28. Zouein FA, Kurdi M, Booz GW. Dancing rhinos in stilettos: the amazing saga of the genomic and nongenomic actions of STAT3 in the heart. *JAKSTAT* 2013; 2(3):e24352. doi: 10.4161/jkst.24352.
29. Zouein FA, Altara R, Chen Q, Lesnfsky EJ, Kurdi M, Booz GW. Pivotal importance of STAT3 in protecting the heart from acute and chronic stress: new advancement and unresolved issues. *Front Cardiovasc Med* 2015; 2:36. doi: 10.3389/fcvm.2015.00036.
30. Palomer X, Salvado L, Barroso E, Vazquez-Carrera M. An overview of the crosstalk between inflammatory processes and metabolic dysregulation during diabetic cardiomyopathy. *Int J Cardiol* 2013; 168(4):3160–72. doi: 10.1016/j.ijcard.2013.07.150.
31. Sharma K. Mitochondrial hormesis and diabetic complications. *Diabetes* 2015; 64(3):663–72. doi: 10.2337/db14-0874.
32. Dugan LL, You YH, Ali SS, Diamond-Stanic M, Miyamoto S, DeClevae AE, et al. AMPK dysregulation promotes diabetes-related reduction of superoxide and mitochondrial function. *J Clin Invest* 2013; 123(11):4888–99. doi: 10.1172/JCI66218.
33. He Q, Harris N, Ren J, Han X. Mitochondria-targeted antioxidant prevents cardiac dysfunction induced by tafazzin gene knockdown in cardiac myocytes. *Oxid Med Cell Longev* 2014; 2014:654198. doi: 10.1155/2014/654198.
34. Dong F, Li Q, Sreejayan N, Nunn JM, Ren J. Metallothionein prevents high-fat diet induced cardiac contractile dysfunction: role of peroxisome proliferator activated receptor gamma coactivator 1alpha and mitochondrial biogenesis. *Diabetes* 2007; 56(9):2201–12. doi: 10.2337/db06-1596.
35. Wang X, Terpstra EJ. Ubiquitin receptors and protein quality control. *J Mol Cell Cardiol* 2013; 55:73–84. doi: 10.1016/j.yjmcc.2012.09.012.
36. Pagan J, Seto T, Pagano M, Cittadini A. Role of the ubiquitin proteasome system in the heart. *Circ Res* 2013; 112(7):1046–58. doi: 10.1161/CIRCRESAHA.112.300521.
37. Willis MS, Bevilacqua A, Pulinkunnil T, Kienesberger P, Tannu M, Patterson C. The role of ubiquitin ligases in cardiac disease. *J Mol Cell Cardiol* 2014; 71:43–53. doi: 10.1016/j.yjmcc.2013.11.008.
38. Zungu M, Schisler J, Willis MS. All the little pieces—regulation of mitochondrial fusion and fission by ubiquitin and small ubiquitin-like modifier and their potential relevance in the heart. *Circ J* 2011; 75(11):2513–21.
39. Cho YE, Basu A, Dai A, Heldak M, Makino A. Coronary endothelial dysfunction and mitochondrial reactive oxygen species in type 2 diabetic mice. *Am J Physiol Cell Physiol* 2013; 305(10):C1033–40. doi: 10.1152/ajpcell.00234.2013.
40. Ozden O, Park SH, Kim HS, Jiang H, Coleman MC, Spitz DR, et al. Acetylation of MnSOD directs enzymatic activity responding to cellular

- nutrient status or oxidative stress. *Aging (Albany NY)* 2011; 3(2):102–7.
41. Klotz LO, Sanchez-Ramos C, Prieto-Arroyo I, Urbanek P, Steinbrenner H, Monsalve M. Redox regulation of FoxO transcription factors. *Redox Biol* 2015; 6:51–72. doi: 10.1016/j.redox.2015.06.019.
 42. Karamanlidis G, Lee CF, Garcia-Menendez L, Kolwicz SC, Jr., Suthammarak W, Gong G, et al. Mitochondrial complex I deficiency increases protein acetylation and accelerates heart failure. *Cell Metab* 2013; 18(2):239–50. doi: 10.1016/j.cmet.2013.07.002.
 43. Pillai VB, Bindu S, Sharp WW, Fang YH, Kim GH, Gupta M, et al. Sirt3 protects mitochondrial DNA damage and blocks the development of doxorubicin-induced cardiomyopathy in mice. *Am J Physiol Heart Circ Physiol* 2016;ajpheart.00832.2015. doi: 10.1152/ajpheart.00832.2015.
 44. Winnik S, Gaul DS, Preitner F, Lohmann C, Weber J, Miranda MX, et al. Deletion of Sirt3 does not affect atherosclerosis but accelerates weight gain and impairs rapid metabolic adaptation in LDL receptor knockout mice: implications for cardiovascular risk factor development. *Basic Res Cardiol* 2014; 109(1):399. doi: 10.1007/s00395-013-0399-0.
 45. Cencioni C, Spallotta F, Mai A, Martelli F, Farsetti A, Zeiher AM, et al. Sirtuin function in aging heart and vessels. *J Mol Cell Cardiol* 2015; 83:55–61. doi: 10.1016/j.yjmcc.2014.12.023.
 46. Sundaresan NR, Gupta M, Kim G, Rajamohan SB, Isbatan A, Gupta MP. Sirt3 blocks the cardiac hypertrophic response by augmenting Foxo3a-dependent antioxidant defense mechanisms in mice. *J Clin Invest* 2009; 119(9):2758–71. doi: 10.1172/JCI39162.
 47. Bartz RR, Piantadosi CA. Clinical review: oxygen as a signaling molecule. *Crit Care* 2010; 14(5):234. doi: 10.1186/cc9185.
 48. Bak DW, Weerapana E. Cysteine-mediated redox signalling in the mitochondria. *Mol Biosyst* 2015; 11(3):678–97. doi: 10.1039/c4mb00571f.
 49. Chung HS, Wang SB, Venkatraman V, Murray CI, Van Eyk JE. Cysteine oxidative posttranslational modifications: emerging regulation in the cardiovascular system. *Circ Res* 2013; 112(2):382–92. doi: 10.1161/CIRCRESAHA.112.268680.
 50. Marchi S, Giorgi C, Suski JM, Agnoletto C, Bononi A, Bonora M, et al. Mitochondria-ROS crosstalk in the control of cell death and aging. *J Signal Transduct* 2012; 2012:329635. doi: 10.1155/2012/329635.
 51. Grimsrud PA, Xie H, Griffin TJ, Bernlohr DA. Oxidative stress and covalent modification of protein with bioactive aldehydes. *J Biol Chem* 2008; 283(32):21837–41. doi: 10.1074/jbc.R700019200.
 52. Tullio F, Angotti C, Perrelli MG, Penna C, Pagliaro P. Redox balance and cardioprotection. *Basic Res Cardiol* 2013; 108(6):392. doi: 10.1007/s00395-013-0392-7.
 53. Chen YR, Zweier JL. Cardiac mitochondria and reactive oxygen species generation. *Circ Res* 2014; 114(3):524–37. doi: 10.1161/CIRCRESAHA.114.300559.
 54. Mieyal JJ, Gallogly MM, Qanungo S, Sabens EA, Shelton MD. Molecular mechanisms and clinical implications of reversible protein S-glutathionylation. *Antioxid Redox Signal* 2008; 10(11):1941–88. doi: 10.1089/ars.2008.2089.
 55. Liu B, Tewari AK, Zhang L, Green-Church KB, Zweier JL, Chen YR, et al. Proteomic analysis of protein tyrosine nitration after ischemia reperfusion injury: mitochondria as the major target. *Biochim Biophys Acta* 2009; 1794(3):476–85. doi: 10.1016/j.bbapap.2008.12.008.
 56. Hill BG, Benavides GA, Lancaster JR, Jr., Ballinger S, Dell'Italia L, Jianhua Z, et al. Integration of cellular bioenergetics with mitochondrial quality control and autophagy. *Biol Chem* 2012; 393(12):1485–512. doi: 10.1515/hsz-2012-0198.