

# Day 3, AM Session

PLINK

whole genome association analysis toolkit

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# What is PLINK?

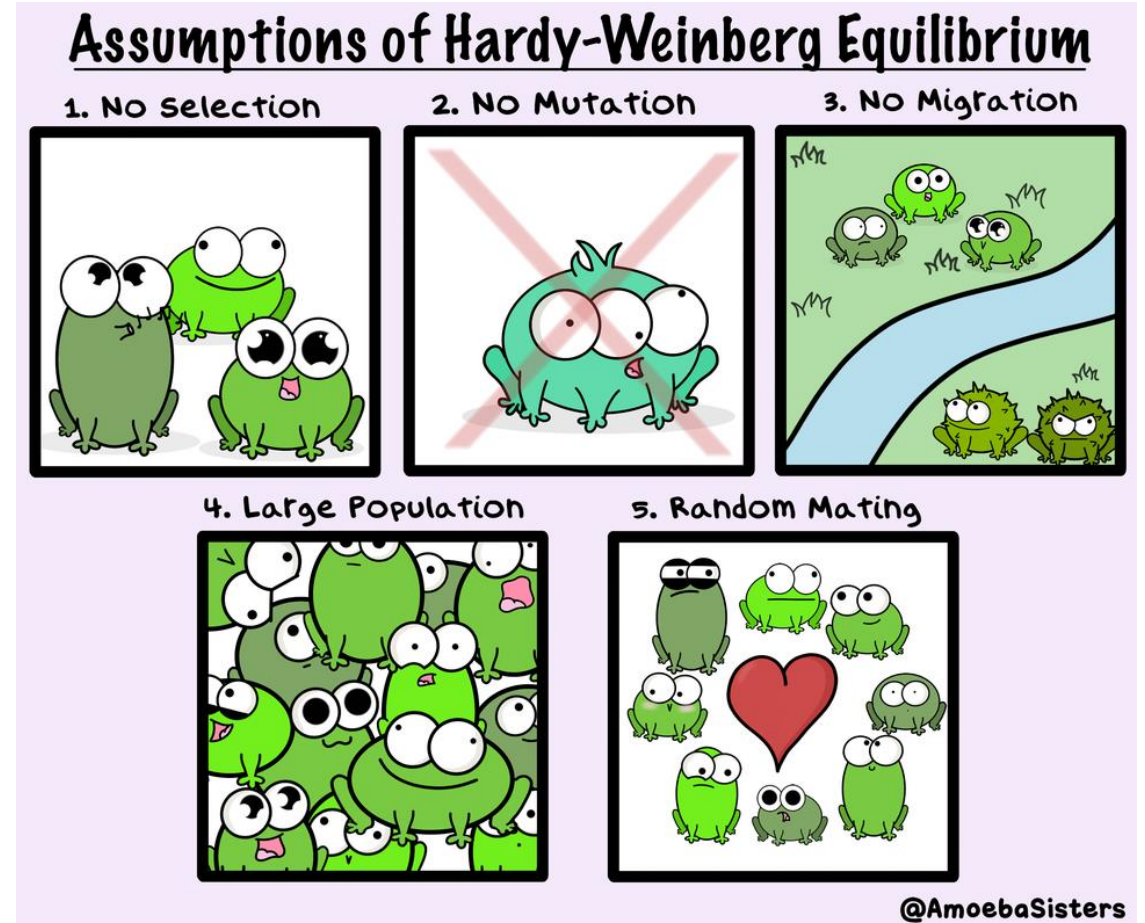
- An open-source tool for analyzing genotype and phenotype data from large omics datasets

<https://zzz.bwh.harvard.edu/plink/index.shtml>

- Variant calling must be done beforehand
- Visualization of results has limited support --> we will visualize results using R and associated packages

# Some PLINK Capabilities

- Data management
  - Read in data in various formats
  - Merge files
  - Extract SNP subset
- Summary statistics
  - Allele frequencies
  - Hardy-Weinberg Equilibrium
  - Missing genotype rates
  - Kinship



# More PLINK Capabilities

- Population stratification
  - Significance tests for whether two individuals belong to same population
- Association testing
  - Case/control
    - Fisher's exact test
    - Logistic regression
  - Quantitative traits
    - Linear regression
- Imputation

HapMap or 1,000 Genomes	<table><tr><td>0</td><td>0</td><td>1</td><td>1</td><td>1</td><td>0</td><td>0</td><td>1</td><td>1</td><td>0</td><td>0</td><td>0</td><td>1</td><td>1</td><td>1</td></tr><tr><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>1</td><td>1</td><td>1</td><td>0</td><td>1</td><td>1</td><td>1</td><td>0</td><td>0</td><td>1</td></tr><tr><td>1</td><td>1</td><td>1</td><td>1</td><td>1</td><td>0</td><td>0</td><td>0</td><td>1</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td><td>0</td></tr><tr><td>1</td><td>0</td><td>1</td><td>1</td><td>0</td><td>0</td><td>0</td><td>1</td><td>1</td><td>1</td><td>1</td><td>1</td><td>0</td><td>0</td><td>1</td></tr></table>														0	0	1	1	1	0	0	1	1	0	0	0	1	1	1	0	0	0	0	0	1	1	1	0	1	1	1	0	0	1	1	1	1	1	1	0	0	0	1	0	0	0	0	0	0	1	0	1	1	0	0	0	1	1	1	1	1	0	0	1	Reference haplotypes
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# PED (.ped) File

- Pedigree, genotype, and phenotype information
- No header line allowed

```
GNU nano 2.0.6                               File: toy.ped
1 100000000000 0 0 1 1 0 0 A A
1 100000000001 0 0 1 2 C C A G
```

# PED: Columns 1 and 2 (FID and IID)

## Column 1

- Family ID (FID)
- Can be same as individual ID

## Column 2

- Individual ID (IID)
- This must be unique if FIDs are the same!

```
GNU nano 2.0.6                               File: toy.ped
1 10000000000 0 0 1 1 0 0 A A
1 10000000001 0 0 1 2 C C A G
```

# PED: Columns 3 (PATID) and 4 (MATID)

- Required even if pedigree is unknown
- Column 3 = paternal ID (PATID or FATID)
- Column 4 = maternal ID (MATID)
- If pedigree is known, fill in the cell with the appropriate ID
- If the pedigree is not known, fill in the cell with -9 or 0, which code for missing data

```
GNU nano 2.0.6                               File: toy.ped
1 10000000000 0 0 1 1 0 0 A A
1 10000000001 0 0 1 2 C C A G
```

# PED: Column 5 (Sex)

Column 5 = Sex

- If male --> 1
- If female --> 2
- If unknown --> 0 or -9

```
GNU nano 2.0.6 File: toy.ped
1 10000000000 0 0 1 1 0 0 A A
1 10000000001 0 0 1 2 C C A G
```



# PED: Column 6 (Trait of interest)

Column 6 = Phenotype

- Quantitative
  - Plant height: 1.12
  - No commas
- Case/control
  - Unaffected = 1
  - Affected = 2
  - Missing = -9 or 0

```
GNU nano 2.0.6 File: toy.ped
1 100000000000 0 0 1 1 0 0 A A
1 100000000001 0 0 1 2 C C A G
```

# PED: Columns 7 and Beyond (Genotypes)

## Genotype data

- Must be biallelic
- Can code allele as numbers 1,2,3, etc. or letters (A,B,C, etc. or A,T,G,C)
- Missing data
  - Coded as 0 or -9
  - Cannot have one allele present and one allele missing

```
GNU nano 2.0.6 File: toy.ped
1 100000000000 0 0 1 1 0 0 A A
1 100000000001 0 0 1 2 C C A G
```

# MAP (.map) File

- Four columns
- Marker information
- Same order as PED file
- Do not need to be in genomic order

GNU nano 2.0.6			File: toy.map
1	rs0	0	1000
1	rs10	0	1001

# MAP: Columns 1 and 2

## Column 1 = Chromosome

- Number (1, 2, 3, etc.)
- If unplaced --> 0
- X = X chromosome
- Y = Y chromosome
- XY = pseudo-autosomal region
- MT = mitochondrion

## Column 2 = SNP identifier

- rs# for human SNP ID
- chromosome:position for most other organisms

GNU nano 2.0.6		File: toy.map	
1	rs0	0	1000
1	rs10	0	1001

# MAP: Columns 3 and 4

Column 3 = Genetic distance (cM)

- Set to 0 for association testing

Column 4 = Physical base-pair position

- If you preface this with a "-", you exclude the SNP from analysis
  - Example: -1562

GNU nano 2.0.6		File: toy.map	
1	rs0	0	1000
1	rs10	0	1001

# PLINK Exercise

1. Check PLINK is installed on the system.

`plink`

2. Download hapmap1.zip data and unzip. Move this folder to Desktop).

(Do this manually)

3. Open and observe each file.

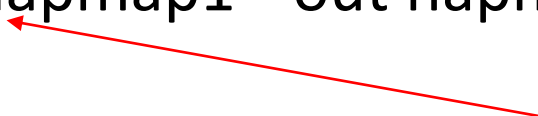
`nano hapmap1.ped`

`nano hapmap1.map` (Exit a nano file with Control X)

4. Identify how many individuals and SNPs are in the initial dataset.

`plink --file hapmap1 --out hapmap1`

Note: this implies there are the files hapmap1.ped and hapmap1.map in the directory



# Binary PED file

- Compact form that saves time and space
- Produced from command `plink --file hapmap1`
  - `.bed` = raw genotype data
    - this is the binary file
    - Cannot be viewed easily
  - `.bim` = raw genotype data + allele names
    - Can be viewed (nano)
  - `.fam` = first six columns of `.ped` file
    - Can be viewed (nano)

Exercise:

`ls`

`nano hapmap1.bim`

(Exit a nano file with Control X)

`nano hapmap1.fam`

(Exit a nano file with Control X)

# Summary Statistics

- `--missing` --> missing data per individual (.imiss) and marker (.lmiss)
- `--freq` --> minor allele frequencies (.frq)
- `--hardy` --> Hardy-Weinberg Equilibrium (.hwe)

Adding the `--chr` modifier (example: `--chr 1`) provides the statistics for only the chromosome of interest

Adding the `--snp` modifier (with a SNP ID) provides the statistics for only the SNP of interest



# Missing Data Exercise

Determine amount of missing data

1. `plink --file hapmap1 --missing --out miss_stat`

2. `nano miss_stat.lmiss`

Gives missing data per marker/locus

`nano miss_stat.imiss`

Gives missing data per individual

Missing data shown as counts and frequencies.

# .lmiss

GNU nano 2.0.6

File: miss\_stat.lmiss

CHR	SNP	N_MISS	N_GENO	F_MISS
1	rs6681049	0	89	0
1	rs4074137	0	89	0
1	rs7540009	0	89	0
1	rs1891905	0	89	0

N\_MISS = number of missing genotypes/locus

F\_MISS = percent of missing data/locus

# .imiss

FID	IID	MISS_PHENO	N_MISS	N_GENO	F_MISS
HCB181	1	N	671	83534	0.008033
HCB182	1	N	1156	83534	0.01384
HCB183	1	N	498	83534	0.005962

N\_MISS = number of missing genotypes/individual

F\_MISS = percent of missing data/individual

# Minor Allele Frequency Exercise

Using the complete dataset, no population subdivision

```
plink --file hapmap1 --freq --out freq_stat
```

```
nano freq_stat.frq
```

# .frq

**A1** = minor allele

**A2** = major allele

**MAF** = minor allele frequency


**NCHROBS** = number of alleles  
without missing data (actually  
genotyped)

GNU nano 2.0.6		File: freq_stat.frq			
CHR	SNP	A1	A2	MAF	NCHROBS
1	rs6681049	1	2	0.2135	178
1	rs4074137	1	2	0.07865	178
1	rs7540009	0	2	0	178
1	rs1891905	1	2	0.4045	178
1	rs9729550	1	2	0.1292	178
1	rs3813196	1	2	0.02809	178
1	rs6704013	0	2	0	174
1	rs307347	0	2	0	154

# MAF (population subdivision)

1. Look at the pop.phe file

`nano pop.phe`



	FID, IID, Population
HCB222	1 1
HCB223	1 1
HCB224	1 1
HCB225	1 1
JPT226	1 2
JPT227	1 2
JPT228	1 2

2. Calculating MAF per subpopulation

`plink --file hapmap1 --freq --within pop.phe --out popfreq_stat`

`nano popfreq_stat.frq.strat`

# .frq.strat

GNU nano 2.0.6

File: popfreq\_stat.frq.strat

CHR	SNP	CLST	A1	A2	MAF	MAC	NCHROBS
1	rs6681049	1	1	2	0.2333	21	90
1	rs6681049	2	1	2	0.1932	17	88
1	rs4074137	1	1	2	0.1	9	90
1	rs4074137	2	1	2	0.05682	5	88
1	rs7540009	1	0	2	0	0	90
1	rs7540009	2	0	2	0	0	88

CLST = cluster

MAC = minor allele count

# Filtering Options

- `--mind` --> allowed missing data/individual to retain individual
  - Example: `--mind 0.05` for retaining individuals with 95% data present
- `--geno` --> allowed missing data/marker to retain marker
  - Example: `--geno 0.05` for retaining markers with genotyping call rate of 95%
- `--maf` --> allow markers with this Minor Allele Frequency and above
  - Example: `--maf 0.05` for retaining markers with MAF 0.05 and above
- `--hwe` --> performs Hardy-Weinberg Exact test at specified threshold
  - Be careful when using this filter
  - Make sure you do this for each population separately
- Syntax example: `plink --file hapmap1 --make-bed --mind 0.05 --out filter1`



# Filtering Options Exercise

Retain markers with a genotype call rate of 85%.

- a. How many markers were removed?
- b. How many markers were retained?

# Filtering Options

- To keep or remove individuals from an analysis
  - Two column text file (family ID, individual ID), no header
  - `--keep textfile` = keep these individuals
  - `--remove textfile` = remove these individuals
- To keep or remove certain SNP\_IDs from an analysis
  - One column with SNP\_IDs, no header
  - `--extract textfile` = keeps these variants
  - `--exclude textfile` = remove these variants

# What type of data is the phenotype? Exercise

1. Open the hapmap1.ped file.

`nano hapmap1.ped`

2. Is phenotype continuous, binary, or categorical?

3. What type of regression should we perform?

(Options: linear, binary logistic, multinomial logistic, ordered logistic)

# Logistic Regression Exercise

```
plink --file hapmap1 --make-bed --logistic --covar pop.phe --out withpopA  
nano withpopA.assoc.logistic
```

# Output File

GNU nano 2.0.6

File: plink.assoc.logistic

CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P
1	rs6681049	1	1	ADD	89	0.5781	-1.447	0.1479
1	rs6681049	1	1	COV1	89	9.436	4.486	7.247e-06
1	rs4074137	2	1	ADD	89	1.54	0.6577	0.5107
1	rs4074137	2	1	COV1	89	9.764	4.533	5.809e-06
1	rs7540009	3	0	ADD	89	NA	NA	NA
1	rs7540009	3	0	COV1	89	NA	NA	NA
1	rs1891905	4	1	ADD	89	1.079	0.232	0.8166
1	rs1891905	4	1	COV1	89	9.307	4.533	5.822e-06
1	rs9729550	5	1	ADD	89	2.846	2.007	0.04472
1	rs9729550	5	1	COV1	89	11.23	4.604	4.153e-06
1	rs3813196	6	1	ADD	89	1.352	0.2785	0.7806
1	rs3813196	6	1	COV1	89	9.229	4.519	6.205e-06
1	rs6704013	7	0	ADD	87	NA	NA	NA

NA is present when the SNP was monomorphic

Columns:

1. Chromosome
2. SNP ID
3. Base position
4. Minor Allele
5. Test (ADD = additive effects of allele dosage, COV1 = covariate)
6. NMISS = # of nonmissing genotypes
7. Odds ratio
8. T-statistic
9. P-value

# Test Column

- We are interested in the additive effects of allele dosage line.
  - Additive effects of allele dosage = more copies of an allele are more or less strongly correlated with the phenotype.
- This is the significance of the SNP when controlling for the covariate (population).
- We can hide the covariate including the argument `--hide-covar`

## Exercise

```
plink --file hapmap1 --make-bed --logistic --covar pop.phe --hide-covar  
      --out withpopB  
nano withpopB.assoc.logistic
```

# Excluding Covariate Exercise

```
plink --file hapmap1 --make-bed --logistic --out withoutpop  
nano withoutpop.assoc.logistic
```

GNU nano 2.0.6							File: withpopA.assoc.logistic		
CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P	
1	rs6681049	1	1	ADD	89	0.5781	-1.447	0.1479	
1	rs6681049	1	1	COV1	89	9.436	4.486	7.247e-06	

GNU nano 2.0.6							File: withoutpop.assoc.logistic		
CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P	
1	rs6681049	1	1	ADD	89	0.592	-1.534	0.1251	

Adding the covariate, makes the SNP effect less significant.

# Correcting for Multiple Testing Exercise

```
plink --file hapmap1 --make-bed --logistic --covar pop.phe --hide-covar -  
-adjust --out withpopC
```

```
nano withpopC.assoc.logistic.adjusted
```



# Correcting for Multiple Testing

GNU nano 2.0.6

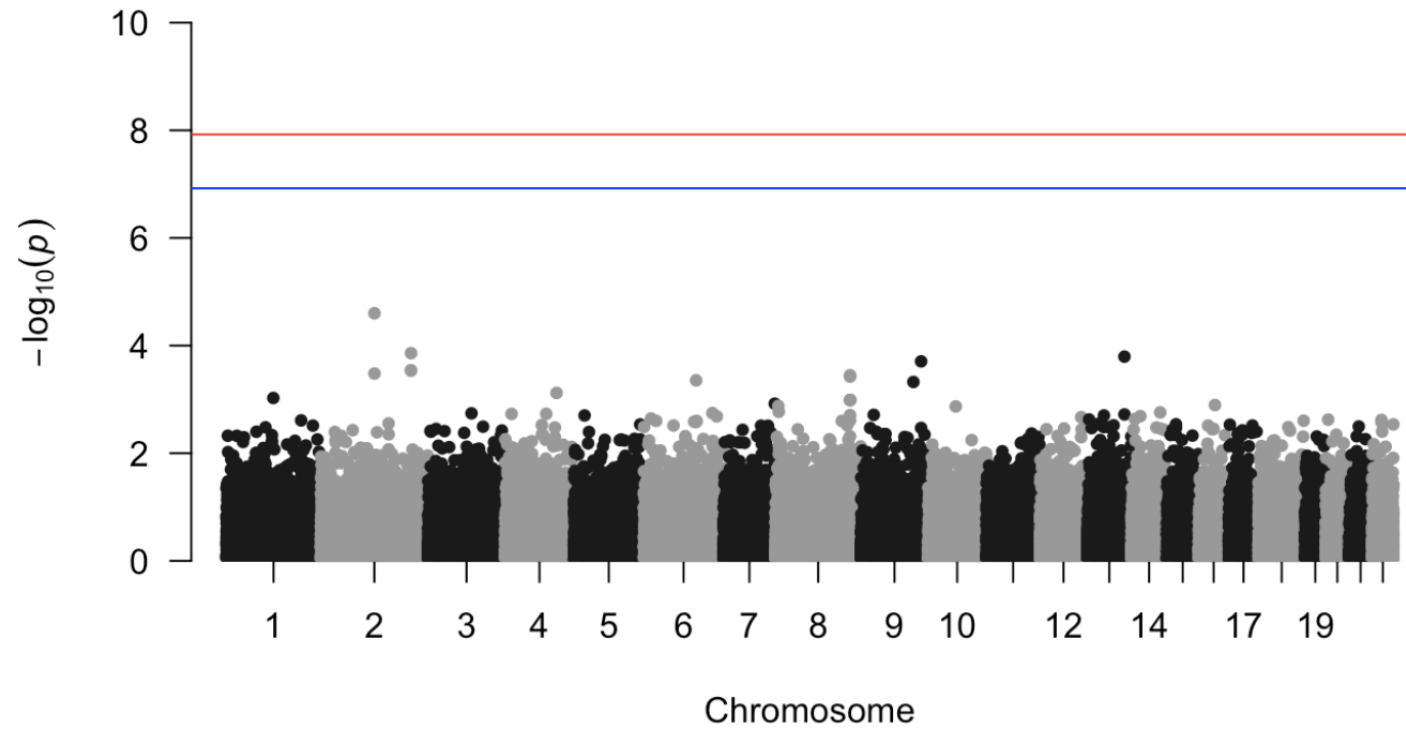
File: withpopC.assoc.logistic.adjusted

CHR	SNP	UNADJ	GC	BONF	HOLM	SIDAK_SS	SIDAK_SD	FDR_BH	FDR_BY
2	rs2222162	2.507e-05	2.507e-05	1	1	0.8056	0.8056	0.9945	1
2	rs4675607	0.0001382	0.0001382	1	1	0.9999	0.9999	0.9945	1
13	rs9585021	0.0001603	0.0001603	1	1	1	1	0.9945	1
9	rs7046471	0.0001963	0.0001963	1	1	1	1	0.9945	1
2	rs4673349	0.0002892	0.0002892	1	1	1	1	0.9945	1

# Manhattan Plot Exercise

- If you remember, we produced a file “withpopB.assoc.logistic” today.
- We are going to use this file, to produce a Manhattan plot.
- Please run the R Markdown named ‘ManhattanPlot.Rmd’ in RStudio.

# Manhattan Plot Answer



# References

- [grunwaldlab.github](https://github.com/grunwaldlab)
- [https://bookdown.org/kdonovan125/ibis\\_data\\_analysis\\_r4/documenting-your-results-with-r-markdown.html](https://bookdown.org/kdonovan125/ibis_data_analysis_r4/documenting-your-results-with-r-markdown.html)