Package AquaEnv: an Aquatic modelling Environment in R

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Abstract

AquaEnv is an integrated development toolbox for aquatic chemical model generation focused on (ocean) acidification and CO2 air-water exchange.

- It contains all elements necessary to model the pH, the related CO2 air-water exchange, as well as aquatic acid-base chemistry in general for an arbitrary marine, estuarine or freshwater system. Also chemical batches can be modelled.
- Next to the routines necessary to calculate desired information, AquaEnv also contains a suite of tools to visualize this information.
- Furthermore, AquaEnv can not only be used to build dynamic models of aquatic systems, but it can also serve as a simple desktop tool for the experimental aquatic chemist to generate and visualize all possible derived information from a set of measurements with one single easy to use R function.
- Additionally, the sensitivity of the system to variations in the input variables can be visualized.
- AquaEnv also contains a number of example "applications" that make use of the aquatic modelling toolbox that AquaEnv provides:
 - a theoretical titration simulator
 - and a routine to determine total alkalinity ([TA]), the total dissolved inorganic carbon concentration ([\sum CO2]), as well as additionally the electrode standard potential (E₀) and the first dissociation constant of the carbonate system ($K_{CO_2}^*$) from titration data.

Keywords: aquatic modelling, pH, pH scales, dissolved inorganic carbon, total alkalinity, total alkalinity curve fitting, theoretical titration, revelle factor, omega, solubility products, CO₂, ocean acidification, estuaries, carbonate system, seawater, R.

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1 Introduction

AquaEnv is a toolbox for aquatic modelling that serves several purposes

- It provides functions to calculate the stoichiometric equilibrium constants (K*) for key acid base systems in natural seawater, the Henry's constants (K₀), as well as the solubility products (K_{sp}) for calcite and aragonite. This functionality is provided via the functions K_CO2, K_HCO3, K_BOH3, K_W, K_HSO4, K_HF, K_NH4, K_H2S, K_H3PO4, K_H2PO4, K_HPO4, K_SiOH4, K_SiOOH3, KO_CO2, KO_O2, Ksp_aragonite, and Ksp_calcite.
- It is designed to make its use as easy as possible: all the information that can be calculated from the set of parameters know of a system or sample can be obtained by one single function: aquaenv. This function returns a list of class aquaenv that contains next to the input parameters
 - the clorinity, the ionic strength, $[\sum B(OH)_3]$, $[\sum H_2SO_4]$, $[\sum HF]$, $[Cl^-]$, $[Cl^-]$, $[\sum Br]$, $[Na^+]$, $[Mg^{2+}]$, $[Ca^{2+}]$, $[K^+]$, $[Sr^{2+}]$ calculated from salinity as given in DOE (1994) (Please note that if values for $[\sum B(OH)_3]$, $[\sum H_2SO_4]$, $[\sum HF]$ are given as input parameters, these parameters are used and not the ones calculated from salinity.)
 - the hydrostatic pressure calculated from the given depth and the seawater density calculated from temperature and salinity as given by Millero and Poisson (1981)
 - a set of conversion factors to convert between different pH scales (Dickson 1984;
 Zeebe and Wolf-Gladrow 2001) and between mol/kg-H₂O and mol/kg-solution (inferred from Roy, Roy, Vogel, PorterMoore, Pearson, Good, Millero, and Campbell (1993b) and DOE (1994))
 - the Henry's constants for CO_2 (Weiss 1974) and for O_2 (inferred from Weiss 1970) calculated from temperature and salinity as well as the associated saturation concentrations of CO_2 and O_2 .
 - the ion product of water (Millero 1995), the stoichiometric equilibrium constants of HSO₄⁻ (Dickson 1990a), HF(Dickson and Riley 1979a), CO₂ (Roy et al. 1993b), HCO₃⁻ (Roy et al. 1993b), B(OH)₃ (Dickson 1990a), NH4⁺(Millero, Yao, and Aicher 1995), H2_S (Millero 1995), H₃PO4(Millero 1995), H₂PO4⁻ (Millero 1995), HPO4²⁻ (Millero 1995), SiOH4 (Millero, Plese, and Fernandez 1988), SiOOH3⁻ (Wischmeyer, Del Amo, Brzezinski, and Wolf-Gladrow 2003), HNO2 (Riordan, Minogue, Healy, O'Driscoll, and Sodeau 2005), HNO3, H2SO4 (Atkins 1996), HS (Atkins 1996) mostly calculated as functions of temperature and salinity and pressure corrected according to Millero (1995).
 - the solubility products of calcite and aragonite (Mucci 1983) as well as the associated Ω 's if a full speciation is calculated (see below)
 - the partial pressure of CO₂ if a full speciation is calculated (see below)
 - if $[\sum CO_2]$ and pH are given [TA] is calculated, if $[\sum CO_2]$ and [TA] are given pH is calculated, if $[\sum CO_2]$ and $[CO_2]$ or pCO₂ are given, pH and [TA] are calculated.
 - if either one of the pairs pH and $[CO_2]$ or pCO₂, pH and [TA], or [TA] and $[CO_2]$ or pCO₂ is given, $[\sum CO_2]$ is calculated

- if sufficient information is given and the flag speciation=TRUE is set, a full speciation of $[\sum CO2]$, $[\sum NH4]$, $[\sum H_2S]$, $[\sum HNO3]$, $[\sum HNO2]$, $[\sum H_3PO4]$, $[\sum Si(OH)_4]$, $[\sum B(OH)_3]$, $[\sum H_2SO_4]$, $[\sum HF]$, as well as water itself is calculated
- if the flag revelle=TRUE is set, the revelle factor (Zeebe and Wolf-Gladrow 2001) is calculated. item if the flag revelle=TRUE is set, all necessary quantities for the explicit "direct substitution approach" (DSA) to pH modelling as given in Hofmann, Meysman, Soetaert, and Middelburg (2008b) are calculated. These are the buffer factor (the partial derivative of [TA] with respect to $[H^+]$) and the partial derivatives of [TA] with respect to the other total quantities. Furthermore, the partial derivatives of [TA] with respect to changes in the equilibrium constants (K^*) , multiplied with the partial derivatives of the equilibrium constants with respect to their variables needed for the DSA with time variable equilibrium constants as described in Hofmann, Meysman, Soetaert, and Middelburg (2008a) are calculated. Finally, the ionization fractions as defined by Stumm and Morgan (1996) and used in Hofmann, Middelburg, Soetaert, Wolf-Gladrow, and Meysman (2008c) are calculated for the full speciation.
- Input for aquaenv has to be supplied in standard SI units, the free proton pH scale and in molinity¹ (mol/kg-solution). Conversion of input parameters to this necessary units and pH scale can be done with the generic function convert.
- The information created with aquaenv is also supplied in standard SI units and in molinity. All elements of an object of class aquaenv of a certain unit or pH scale can be converted into other units or pH scales with the function convert as well.
- One can use input vectors of temperature T, salinity S or depth d for aquaenv to obtain vectors of all calculated information as function of the input vector. This can be visualized in a large variety of ways using the plot function specially defined for objects pf class aquaenv.
- Objects of class aquaenv can be used in dynamic models to define the state of the system in each timestep of the numerical integration (done e.g. with **deSolve**). with the function aquaenv and the flag from.data.frame=TRUE it is possible to convert output of those dynamic models into objects pf class aquaenv which allows the user to use the whole suite of visualisation tools that is provided by the function plot in AquaEnv.
- As mentioned above Hofmann et al. (2008b), Hofmann et al. (2008a), and Hofmann et al. (2008c) describe methods for an "explicit" pH modelling which allows for the quantification of the influences of kinetically modelled processes on the pH. Objects pf class aquaenv provide all needed quantities (partial derivatives of [TA], ionization fractions, etc.) to employ both of those methods in dynamic models. Furthermore, AquaEnv provides the functionality to cumulatively plot the obtained influences on the pH.
- As an example of how to use the aquatic chemical toolbox that is provided by AquaEnv, two applications are provided

¹Note that it is not sufficient to give a gravimetric concentration in mol/kg since there is a substancial difference between mol/kg-H₂O (molality) and mol/kg-solution (molinity).

- The function titration: creates theoretical titrations which can be used e.g. to create bjerrum plots with the function plot.aquaenv in AquaEnv.
- The function TAfit: a routine based on a method in DOE (1994) that makes use of that theoretical titration function and allows for determining total alkalinity ([TA]), the total dissolved inorganic carbon concentration ([\sum CO2]), as well as additionally the electrode standard potential (E₀) and the first dissociation constant of the carbonate system ($K_{CO_2}^*$) using the Levenberg-Marquart algorithm (least squares optimization procedure) as provided in the R package **minpack.lm**.

2 The elements of an object of class aquaenv

The function aquaenv, the central function of **AquaEnv**, returns an object of class aquaenv. This object is a list of different elements which can be accesses with the \$\\$ character or with the [[]] operator

```
> test <- aquaenv(10, 35)
> test$Tc

[1] 10
attr(,"unit")
[1] "deg C"

> test[["Tc"]]

[1] 10
attr(,"unit")
[1] "deg C"
```

Maximally, i.e., if the enough input data is supplied to define the pH of the system and the flags speciation, dsa, and revelle are TRUE while the flag skeleton is FALSE, an object of class aquaenv contains the following elements

element	unit	explanation
Tc	°C	temperature
Tk	K	absolute temperature
S	"psu" (no unit)	salinity
Cl	%0	chlorinity
I	mol/kg-H ₂ O	ionic strength
d	m	depth
hydroP	bar	hydrostatic pressure
density	kg/m^3	(seawater) density
SumCO2	mol/kg-soln	$[\sum CO_2]$, total dissolved inorganic carbon concentration
SumNH4	mol/kg-soln	$[\sum NH_4^+]$, total ammonium concentration
SumH2S	mol/kg-soln	$[\sum H_2S]$, total sulfide concentration
SumHNO3	mol/kg-soln	$\sum HNO_3$, total nitrate concentration

SumHNO2	mol/kg-soln	$[\sum HNO_2]$, total nitrite concentration
SumH3PO4	mol/kg-soln	$[\sum H_3PO_4]$, total phosphate concentration
SumSiOH4	mol/kg-soln	$[\sum Si(OH)_4]$, total silicate concentration
SumBOH3	mol/kg-soln	$[\sum B(OH)_3]$, total borates concentration
SumH2SO4	mol/kg-soln	$[\sum H_2SO_4]$, total sulfate concentration
SumHF	mol/kg-soln	$\sum HF$, total fluoride concentration
SumBr	mol/kg-soln	\sum_{i} HBr], total bromide concentration
ClConc	mol/kg-soln	[Cl ⁻], chloride concentration
Na	mol/kg-soln	[Na ⁺], sodium concentration
Mg	mol/kg-soln	[Mg ²⁺], magnesium concentration
Ca	mol/kg-soln	[Ca ²⁺], calcium concentration
K	mol/kg-soln	[K ⁺], potassium concentration
Sr	mol/kg-soln	[Sr ²⁺], strontium concentration
molal2molin	(mol/kg-soln)/(mol/kg-H2O)	concentration conversion factor: from molality to
11101011211101111	(1101/118 50111)/ (11101/118 1120)	molinity
free2tot	-	pH conversion factor: free scale to total scale
free2sws	-	pH conversion factor: free scale to sawater scale
tot2free	-	pH conversion factor: total scale to free scale
tot2sws	-	pH conversion factor: total scale to seawater scale
sws2free	-	pH conversion factor: seawater scale to free scale
sws2tot	-	pH conversion factor: seawater scale to total scale
K0_CO2	mol/(kg-soln*atm)	Henry's constant for CO ₂
K0_O2	mol/(kg-soln*atm)	Henry's constant for O ₂
CO2_sat	mol/kg-soln	${\rm CO_2}$ saturation concentration at an atmospheric partial pressure/fugacity of Fugacity\$CO2
O2_sat	mol/kg-soln	O ₂ saturation concentration at an atmospheric partial pressure/fugacity of Fugacity\$O2
$K_{-}W$	(mol/kg-soln) ² , free pH scale	stoichiometric equilibrium ion product of
		$H_2O: K_W^* = [H^+][OH-]$
K_HSO4	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
		$K_{HSO_4^-}^* = [H^+][SO_4^{2-}]/[HSO_4^-]$
K_HF	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
11111	mor/ ng som, neo pri seme	$K_{HF}^* = [H^+][F^-]/[HF]$
K_CO2	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
		$K_{CO_2}^* = [H^+][HCO_3^-]/[CO_2]$
K_HCO3	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
	mor, ng som, nee pri seeme	$K_{HCO_3^-}^* = [H^+][CO_3^2]/[HCO_3^-]$
K_BOH3	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
1125 0110	moly no com, nee pri seeme	$K_{B(OH)_3}^* = [H^+][B(OH)_4^-]/[B(OH)_3]$
K_NH4	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
	· / G · · / · · · · ·	$K_{NH_4^+}^* = [H^+][NH_3]/[NH_4^+]$
K_H2S	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
		$K_{H_2S}^* = [H^+][HS^-]/[H_2S]$
K_H3PO4	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
		$K_{H_3PO_4}^* = [H^+][H_2PO_4^-]/[H_3PO_4]$
K_H2PO4	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
'	'	

		$K_{H_2PO_4^-}^* = [H^+][HPO_4^{2-}]/[H_2PO_4^-]$
K_HPO4	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
K_IIF 04	mor/kg-som, free pri scare	_
		$K_{HPO_4^{2-}}^* = [H^+][PO_4^{3-}]/[HPO_4^{2-}]$
K_SiOH4	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
		$K_{Si(OH)_4}^* = [H^+][SiO(OH)_3^-]/[Si(OH)_4]$
K_SiOOH3	mol/kg-soln, free pH scale	stoichiometric equilibrium constant
		$K_{SiO(OH)_3^-}^* = [H^+][SiO_2(OH)_2^{2-}]/[SiO(OH)_3^-]$
K_HNO2	mol/kg-soln; mol/kg-H2O; mol/l	approximate value for equilibrium constant
		$K_{HNO_2}^* = [H^+][NO_2^-]/[HNO_2]$
K_HNO3	mol/kg-soln; mol/kg-H2O; mol/l	approximate value for equilibrium constant
		$K_{HNO_3}^* = [H^+][NO_3^-]/[HNO_3]$
K_H2SO4	mol/kg-soln; mol/kg-H2O; mol/l	approximate value for equilibrium constant
		$K_{H_2SO_4}^* = [H^+][HSO_4^-]/[H_2SO_4]$
K_HS	mol/kg-soln; mol/kg-H2O; mol/l	approximate value for equilibrium constant
		$K_{HS^-}^* = [H^+][S^{2-}]/[HS^-]$
Ksp_calcite	(mol/kg-soln) ²	stoichiometric equilibrium solubility product of calcite
		$Ksp_{cal}^* = [Ca^{2+}][CO_3^{2-}]$
Ksp_aragonite	$(\text{mol/kg-soln})^2$	stoichiometric equilibrium solubility product of
		aragonite
		$Ksp_{ara}^* = [Ca^{2+}][CO_3^{2-}]$
TA	mol/kg-soln	[TA], total alkalinity
рН	-, free scale	pH
pCO2	atm,	partial pressure (fugacity) of CO ₂ in the water
CO2	mol/kg-soln	$[CO_2]$
HCO3	mol/kg-soln	$[HCO_3^-]$
CO3	mol/kg-soln	$[CO_3^{2-}]$
ВОН3	mol/kg-soln	[B(OH) ₃]
BOH4	mol/kg-soln	$\left[\mathrm{B}(\mathrm{OH})_{4}^{-} \right]$
ОН	mol/kg-soln	[OH ⁻]
H3PO4	mol/kg-soln	$[\mathrm{H_3PO_4}]$
H2PO4	mol/kg-soln	$[\text{H2PO}_4^-]$
HPO4	mol/kg-soln	$[HPO_4^{2-}]$
PO4	mol/kg-soln	$\left[\mathrm{PO}_4^{3-}\right]$
SiOH4	mol/kg-soln	$[Si(OH)_4]$
SiOOH3	mol/kg-soln	$[SiO(OH)_3^-]$
SiO2OH2	mol/kg-soln	$\left[\mathrm{SiO}_{2}(\mathrm{OH})_{2}^{2-}\right]$
H2S	mol/kg-soln	$[H_2S]$
HS	mol/kg-soln	[HS ⁻]
S2min	mol/kg-soln	$[S^{2-}]$
NH4	mol/kg-soln	[NH ₄ ⁺]
NH3	mol/kg-soln	[NH ₃]
H2SO4	mol/kg-soln	[H ₂ SO ₄]
HSO4	mol/kg-soln	$[HSO_4^-]$
SO4	mol/kg-soln	$\left[\mathrm{SO}_4^{2-}\right]$

HF	mol/kg-soln	[HF]
F	mol/kg-soln	[F-]
HNO3	mol/kg-soln	[HNO ₃]
NO3	mol/kg-soln	$[NO_3^-]$
HNO2	mol/kg-soln	[HNO ₂]
NO2	mol/kg-soln	$[NO_2^-]$
omega_calcite	-	saturation state Ω with respect to calcite
omega_aragonite	_	saturation state Ω with respect to aragonite
revelle	_	Revelle factor
c1	_	ionization fraction $c_1 = [CO_2]/[\sum CO_2]$
c2	_	ionization fraction $c_2 = [HCO_3^-]/[\sum CO_2]$
c3	_	ionization fraction $c_3 = [CO_3^{2-}]/[\sum CO_2]$
dTAdSumCO2	_	$\frac{\partial [TA]}{[\partial \sum CO_2]}$
		with $[TA] = f([H^+], [\sum CO_2],)$
b1	_	ionization fraction $b_1 = [B(OH)_3]/[\sum B(OH)_3]$
b2	_	ionization fraction $b_2 = [B(OH)_4]/[\sum B(OH)_3]$
dTAdSumBOH3	_	$\frac{\partial[TA]}{[\partial \sum B(OH)_3]}$
		with $[TA] = f([H^+], [\sum CO_2],)$
so1	_	ionization fraction $so_1 = [H_2SO_4]/[\sum H_2SO_4]$
so2	_	ionization fraction $so_2 = [HSO_4^-]/[\sum H_2SO_4]$
so3	_	ionization fraction $so_3 = [SO_4^2]/[\sum H_2SO_4]$
dTAdSumH2SO4	_	$\frac{\partial [TA]}{[\partial \sum H_2 SO_4]}$
		with $[TA] = f([H^+], [\sum CO_2],)$
f1		ionization fraction $f_1 = [HF]/[\sum HF]$
f2	_	ionization fraction $f_1 = [F^-]/[\sum HF]$
dTAdSumHF	_	$\frac{\partial[TA]}{[\partial \sum HF]}$
diridgamm		$ \begin{array}{c} [\partial \sum \mathrm{HF}] \\ \mathrm{with} \ [\mathrm{TA}] = \mathrm{f}([\mathrm{H}^+], [\sum \mathrm{CO}_2],) \\ \end{array} $
dTAdH		with $[TA] = I([T], [\underline{\nearrow}, CO_2],)$ $\frac{\partial [TA]}{ \partial [H^+]}$: buffer factor
diAdii		
dTAdKdKdS		with [TA] = $f([H^+], [\sum CO_2],)$ $\sum_{\partial [TA]} \frac{\partial K_i^*}{\partial K_i^*}$
dTAdKdKdS	-	$\sum_{i} \frac{\partial [TA]}{\partial K_{i}^{*}} \frac{\partial K_{i}^{*}}{\partial S}$
1004 177 177 100		with $[TA] = f([H^+], [\sum CO_2],, K_i^*)$
dTAdKdKdT	-	$\sum_{i} \frac{\partial [TA]}{\partial K_{i}^{*}} \frac{\partial K_{i}^{*}}{\partial T}$
		with $[TA] = f([H^+], [\sum CO_2],, K_i^*)$
dTAdKdKdd	-	$\sum_{\mathrm{i}} rac{\partial [\mathrm{TA}]}{\partial \mathrm{K}_{\mathrm{i}}^{*}} rac{\partial \mathrm{K}_{\mathrm{i}}^{*}}{\partial \mathrm{d}}$
		with $[TA] = f([H^+], [\sum CO_2],, K_i^*)$
dTAdKdKdSumH2SO4	-	$\sum_{i} \frac{\partial [TA]}{\partial K_{i}^{*}} \frac{\partial K_{i}^{*}}{\partial [\sum H_{2}SO_{4}]}$
		with $[TA] = f([H^+], [\sum CO_2],, K_i^*)$
${\rm dTAdKdKdSumHF}$	-	$\sum_{i} \frac{\partial [TA]}{\partial K_{i}^{*}} \frac{\partial K_{i}^{*}}{\partial [\sum HF]}$
		with $[TA] = f([H^+], [\sum CO_2],, K_i^*)$
	I	1 . 1 1/122 -1/ / 1/

For elements that are calculated according to certain literature references, those references are given in appendix B.

3 Using AquaEnv

3.1 Basic features

3.1.1 calling the "K" functions directly

The elements K_CO2, K_HCO3, K_BOH3, K_W, K_HSO4, K_HF, K_NH4, K_H2S, K_H3PO4, K_H2PO4, K_HPO4, K_SiOH4, K_SiOOH3, KO_CO2, KO_O2, Ksp_aragonite, and Ksp_calcite can be calculated directly, witout creating an object of class *aquaenv*. This is done via functions that bear the same name as those elements

```
> K_CO2(15, 30)

[1] 9.089117e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"

> KO_CO2(15, 30)

[1] 0.03852158
attr(,"unit")
[1] "mol/(kg-soln*atm)"

> Ksp_calcite(15, 30, 100)

[1] 3.644549e-07
attr(,"unit")
[1] "(mol/kg-soln)^2"
```

3.1.2 Minimal aquaenv definition

Minimally, an object of class aquaenv can be defined with just a temperature and salinity value

```
> ae <- aquaenv(Tc = 15, S = 30)
> ae$K_CO2

[1] 9.089117e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
```

Optionally, a mean depth can be given. As in the above case, the returned object of class aquaenv then contains a standard set of elements as shown by the names command.

```
> ae <- aquaenv(Tc = 15, S = 30, d = 10)
> ae$Ksp_calcite

[1] 3.588429e-07
attr(,"unit")
[1] "(mol/kg-soln)^2"
```

> names(ae)

[1]	"Tc"	"Tk"	"S"	"C1"
[5]	"I"	"d"	"hydroP"	"density"
[9]	"SumCO2"	"SumNH4"	"SumH2S"	"SumHNO3"
[13]	"SumHNO2"	"SumH3PO4"	"SumSiOH4"	"SumBOH3"
[17]	"SumH2SO4"	"SumHF"	"SumBr"	"ClConc"
[21]	"Na"	"Mg"	"Ca"	"K"
[25]	"Sr"	"molal2molin"	"free2tot"	"free2sws"
[29]	"tot2free"	"tot2sws"	"sws2free"	"sws2tot"
[33]	"K0_C02"	"K0_02"	"CO2_sat"	"02_sat"
[37]	"K_W"	"K_HSO4"	"K_HF"	"K_C02"
[41]	"K_HCO3"	"K_BOH3"	"K_NH4"	"K_H2S"
[45]	"K_H3P04"	"K_H2P04"	"K_HPO4"	"K_SiOH4"
[49]	"K_SiOOH3"	"K_HNO2"	"K_HNO3"	"K_H2SO4"
[53]	"K_HS"	"Ksp_calcite"	"Ksp_aragonite"	

A minimal set of elements in an object of class *aquaenv* can be obtained by setting the flag skeleton to TRUE

```
> ae <- aquaenv(Tc = 15, S = 30, d = 10, skeleton = TRUE) > names(ae)
```

```
[1] "Tc"
                                                    "I"
                 "Tk"
                            "S"
                                        "C1"
                                                               "d"
 [7] "hydroP"
                 "density"
                                                               "SumHNO3"
                            "SumCO2"
                                        "SumNH4"
                                                    "SumH2S"
[13] "SumHNO2"
                 "SumH3PO4" "SumSiOH4" "SumBOH3"
                                                   "SumH2SO4" "SumHF"
[19] "K_W"
                 "K_HSO4"
                            "K_HF"
                                        "K_C02"
                                                    "K_HCO3"
                                                               "K_BOH3"
[25] "K_NH4"
                 "K_H2S"
                            "K_H3P04"
                                        "K_H2P04"
                                                   "K_HP04"
                                                               "K_SiOH4"
                            "K_HNO3"
[31] "K_SiOOH3" "K_HNO2"
                                        "K_H2SO4"
                                                    "K_HS"
```

3.1.3 Defining the complete aquaenv system in different ways

If enough information is given to define a complete speciation, i.e. either one of the pairs SumCO2 and pH, SumCO2 and TA, SumCO2 and CO2, or SumCO2 and pCO2, a full aquaenv system can be defined.

```
> Tc <- 15
> S <- 30
> d <- 10
```

```
> SumCO2 <- 0.002
> pH <- 8
> TA <- 0.002140323
> pCO2 <- 0.000533576
> CO2 <- 2.055419e-05
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pH = pH)
> ae$TA
[1] 0.002140800
attr(,"unit")
[1] "mol/kg-soln"
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, TA = TA)
> ae$pH
[1] 7.998791
attr(,"pH scale")
[1] "free"
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, CO2 = CO2)
> ae$pH
[1] 7.999293
attr(,"pH scale")
[1] "free"
> names(ae)
                                           "S"
 [1] "Tc"
                        "Tk"
                                                              "C1"
                        "d"
 [5] "I"
                                           "hydroP"
                                                              "density"
 [9] "SumCO2"
                        "SumNH4"
                                           "SumH2S"
                                                              "SumHNO3"
                                                              "SumBOH3"
[13] "SumHNO2"
                        "SumH3P04"
                                           "SumSiOH4"
[17] "SumH2SO4"
                        "SumHF"
                                           "SumBr"
                                                             "ClConc"
                                                              "K"
[21] "Na"
                        "Mg"
                                           "Ca"
[25] "Sr"
                                                              "free2sws"
                        "molal2molin"
                                           "free2tot"
[29] "tot2free"
                        "tot2sws"
                                           "sws2free"
                                                              "sws2tot"
[33] "KO_CO2"
                        "K0_02"
                                           "CO2_sat"
                                                              "02_sat"
[37] "K_W"
                        "K_HS04"
                                           "K_HF"
                                                              "K_C02"
[41] "K_HCO3"
                        "K_BOH3"
                                           "K_NH4"
                                                              "K_H2S"
[45] "K_H3PO4"
                        "K_H2P04"
                                           "K_HP04"
                                                              "K_SiOH4"
[49] "K_SiOOH3"
                        "K_HNO2"
                                           "K_HNO3"
                                                              "K_H2SO4"
[53] "K_HS"
                        "Ksp_calcite"
                                           "Ksp_aragonite"
                                                              "TA"
[57] "pH"
                        "pC02"
                                           "C02"
                                                              "HC03"
[61] "CO3"
                        "BOH3"
                                           "BOH4"
                                                              "OH"
[65] "H3PO4"
                        "H2P04"
                                          "HP04"
                                                             "P04"
[69] "SiOH4"
                        "SiOOH3"
                                          "SiO2OH2"
                                                             "H2S"
```

```
[73] "HS" "S2min" "NH4" "NH3"
[77] "H2S04" "HS04" "S04" "HF"
[81] "F" "HN03" "N03" "HN02"
[85] "N02" "omega_calcite" "omega_aragonite"
```

As seen above, a full speciation is calculated along with the pH or total alkalinity respectively. If only pH or total alkalinity is needed, the calculation of the full speciation can be toggled off. Furthermore, the flag skeleton also works for a full system.

```
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pH = pH, speciation = FALSE)
> names(ae)
```

```
"Tk"
                                        "S"
                                                          "C1"
 [1] "Tc"
 [5] "I"
                       "d"
                                        "hvdroP"
                                                          "density"
 [9] "SumCO2"
                       "SumNH4"
                                        "SumH2S"
                                                          "SumHNO3"
                       "SumH3P04"
                                        "SumSiOH4"
                                                          "SumBOH3"
[13]
     "SumHNO2"
                      "SumHF"
                                        "SumBr"
                                                          "ClConc"
[17] "SumH2SO4"
[21] "Na"
                      "Mg"
                                        "Ca"
                                                          "K"
[25] "Sr"
                       "molal2molin"
                                        "free2tot"
                                                          "free2sws"
                       "tot2sws"
                                        "sws2free"
                                                          "sws2tot"
[29] "tot2free"
[33] "KO_CO2"
                       "K0_02"
                                        "CO2_sat"
                                                          "02_sat"
[37] "K_W"
                      "K_HSO4"
                                        "K_HF"
                                                          "K_C02"
                                        "K_NH4"
                                                          "K_H2S"
[41] "K_HCO3"
                       "K_BOH3"
                                                          "K_SiOH4"
[45] "K_H3PO4"
                      "K_H2P04"
                                        "K_HP04"
[49] "K_SiOOH3"
                      "K_HN02"
                                        "K_HNO3"
                                                          "K_H2S04"
[53] "K_HS"
                       "Ksp_calcite"
                                        "Ksp_aragonite" "TA"
[57] "pH"
                      "pC02"
                                        "C02"
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pH = pH, speciation = FALSE,
      skeleton = TRUE)
> names(ae)
 [1] "Tc"
                             "S"
                                                     "I"
                 "Tk"
                                         "Cl"
                                                                 "d"
                                                                 "SumHNO3"
 [7] "hydroP"
                 "density"
                             "SumCO2"
                                         "SumNH4"
                                                     "SumH2S"
                 "SumH3PO4"
                             "SumSiOH4"
                                                     "SumH2SO4"
                                                                 "SumHF"
[13] "SumHNO2"
                                         "SumBOH3"
```

Furtermore all the quantities needed for the explicit pH modelling approaches as given in Hofmann *et al.* (2008b) and Hofmann *et al.* (2008c) can be calculated by setting the flag dsa to TRUE. The revelle factor can be calculated using the flag revelle.

"K_C02"

"K_H2P04"

"K_H2SO4"

"K_HCO3"

"K_HP04"

"K_HS"

"K_BOH3"

"K_SiOH4" "TA"

```
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pCO2 = pCO2, dsa = TRUE,
+ revelle = TRUE)
> ae$dTAdH
```

"K_HF"

"K_H3P04"

"K_HNO3"

"C02"

[19] "K_W"

[37] "pH"

[25] "K_NH4"

[31] "K_SiOOH3"

"K_HS04"

"K_H2S"

"K_HNO2"

"pC02"

```
[1] -17096.98
attr(,"unit")
[1] "(mol-TA/kg-soln)/(mol-H/kg-soln)"
attr(,"pH scale")
[1] "free"
```

> ae\$revelle

[1] 13.77498

If an ambivalent situation is created because the user enters too much information, an error message is displayed

```
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, CO2 = CO2, pCO2 = pCO2)
[1] "Error! Overdetermined system: entered pCO2: 0.000533576 , calculated pCO2: 0.000533575978437754"
[1] "Please enter only one of: pH, TA, CO2, or pCO2."
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pH = pH, TA = TA)
[1] "Error! Overdetermined system: entered TA: 0.002140323 , calculated TA: 0.00214079972077581"
[1] "Please enter only one of: pH, TA, CO2, or pCO2."
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pH = pH, CO2 = CO2)
[1] "Error! Overdetermined system: entered pH: 8 , calculated pH: 7.99929303203143"
[1] "Please enter only one of: pH, TA, CO2, or pCO2."
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, pH = pH, pCO2 = pCO2)
[1] "Error! Overdetermined system: entered pH: 8 , calculated pH: 7.99929301525035"
[1] "Please enter only one of: pH, TA, CO2, or pCO2."
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, TA = TA, CO2 = CO2)
[1] "Error! Overdetermined system: entered TA: 0.002140323 , calculated TA: 0.002140520813372"
[1] "Please enter only one of: pH, TA, CO2, or pCO2."
> ae <- aquaenv(Tc, S, d, SumCO2 = SumCO2, TA = TA, pCO2 = pCO2)
[1] "Error! Overdetermined system: entered TA: 0.002140323 , calculated TA: 0.00214052080675498"
[1] "Please enter only one of: pH, TA, CO2, or pCO2."
```

3.1.4 Calculating $[\sum CO_2]$

 $[\sum CO_2]$ can be calculated by giving a constant pair of either pH and CO2, pH and TA, TA and CO2, or TA and pCO2

```
> pCO2 <- 0.0006952296
> CO2 <- 2.678134e-05
> pH <- 7.888569
> TA <- 0.0021
> Tc <- 15
> S <- 30
> d <- 10
> ae <- aquaenv(Tc, S, d, SumCO2 = NULL, pH = pH, CO2 = CO2)
> ae$SumCO2
[1] 0.001999999
attr(,"unit")
[1] "mol/kg-soln"
> ae <- aquaenv(Tc, S, d, SumCO2 = NULL, pH = pH, pCO2 = pCO2)
> ae$SumCO2
[1] 0.001999999
attr(,"unit")
[1] "mol/kg-soln"
> ae <- aquaenv(Tc, S, d, SumCO2 = NULL, pH = pH, TA = TA)
> ae$SumCO2
[1] 0.002
attr(,"unit")
[1] "mol/kg-soln"
> ae <- aquaenv(Tc, S, d, SumCO2 = NULL, TA = TA, CO2 = CO2)
> ae$SumCO2
[1] 0.002
attr(,"unit")
[1] "mol/kg-soln"
> ae <- aquaenv(Tc, S, d, SumCO2 = NULL, TA = TA, pCO2 = pCO2)
> ae$SumCO2
[1] 0.002
attr(,"unit")
[1] "mol/kg-soln"
```

3.1.5 Cloning an object of class aquaenv

It is possible to clone an obect of class aquaenv, either 1 to 1 or with different pH, TA, or K_CO2

```
> Tc <- 15
> S <- 30
> SumCO2 <- 0.002
> TA <- 0.00214
> ae <- aquaenv(Tc, S, SumCO2 = SumCO2, TA = TA)
> ae$pH
[1] 7.99838
attr(,"pH scale")
[1] "free"
> ae1 <- aquaenv(ae = ae)</pre>
> ae1$pH
[1] 7.99838
attr(,"pH scale")
[1] "free"
> pH <- 9
> ae2 <- aquaenv(ae = ae, pH = pH)
> ae2$TA
[1] 0.002982758
attr(,"unit")
[1] "mol/kg-soln"
> TA <- 0.002
> ae3 <- aquaenv(ae = ae, TA = TA)
> ae3$pH
[1] 7.548174
attr(,"pH scale")
[1] "free"
> K_CO2 <- 1e-06
> ae4 <- aquaenv(ae = ae, k_co2 = K_CO2)</pre>
> ae4$pH
[1] 7.99838
attr(,"pH scale")
[1] "free"
```

Note that K_CO2 as an input variable is in lower cases!

3.1.6 Preparing input variables

Input variables for the function aquaenv need to be in mol/kg-solution and on the free pH scale. Data in other concentration units or pH scales can be converted using the function convert.

```
> Tc <- 15
> S <- 10
> pH_NBS <- 8.142777
> SumCO2molar <- 0.002016803
> pH_free <- convert(pH_NBS, "pHscale", "nbs2free", Tc = Tc, S = S)
> SumCO2molin <- convert(SumCO2molar, "conc", "molar2molin", Tc = Tc,
+ S = S)
> ae <- aquaenv(Tc, S, SumCO2 = SumCO2molin, pH = pH_free)
> ae$pH

[1] 8
attr(,"pH scale")
[1] "free"
> ae$SumCO2

[1] 0.002000000
attr(,"unit")
[1] "mol/kg-soln"
```

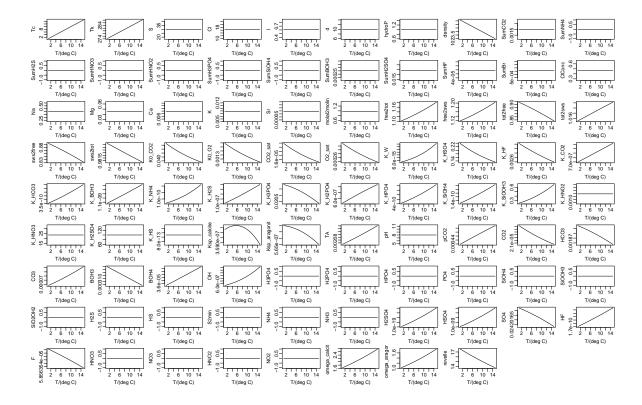
3.1.7 Vectors as input variables

One of the input variables for the function aquaenv may be a vector. All the other input variables are then assumed to be constant. The elements of the resulting two dimensional object of class aquaenv are then vectors containing the elements as a function of the input variable for which a vector is given.

```
> SumC02 <- 0.002
> pH <- 8
> Tc <- 1:15
> S <- 30
> d <- 10
> ae <- aquaenv(Tc, S, d, SumC02 = SumC02, pH = pH, revelle = TRUE)
> ae$revelle

[1] 17.60495 17.30006 16.99846 16.70066 16.40710 16.11818 15.83422 15.55549
[9] 15.28222 15.01459 14.75275 14.49679 14.24679 14.00280 13.76482
```

A two dimensional object of class aquaenv can be visualized using the plot function. For coppletions of the usef,cthexdefault savingsocille pleatheristic for for the class aquaenv results in a new plotting device being opened. Setting the flag newdevice to FALSE prevents that.



The plot function plots all elements of the respective object of class aquaenv. This, however, might not be what the user wants, especially if a larger plotting device cannot properly displayed like in the case above. In this case the parameter what can be used. Note, however, that the default setting for calling plot with the parameter what is that mfrow=c(1,1). So if one wants to plot several elements, mfrow needs to be set to a suitable value.

```
> plot(ae, xval = Tc, xlab = "T/(deg C)", what = c("pH", "C02", + "HC03", "C03"), newdevice = FALSE, mfrow = c(1, 4))

\frac{1}{2} \frac{1}{2} \frac{1}{4} \frac{1}{6} \frac{1}{8} \frac{10}{12} \frac{12}{14} \frac{14}{14} \frac{10}{12} \frac{10}{12} \frac{10}{14} \frac{10}{12} \frac{10}{14} \frac{10}{12} \frac{10}{12} \frac{10}{14} \frac{10}{12} \frac{10}{14} \frac{10}{12} \frac{10
```

The following chunks of example code show other possible definitions of objects of class aquaenv with vectors as input variables.

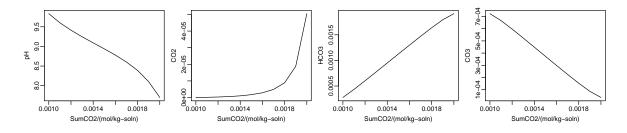
```
> ae <- aquaenv(Tc = 1:15, S = 30, d = 10, SumCO2 = SumCO2, TA = 0.0023) > plot(ae, xval = 1:15, xlab = "T/(deg C)")
```

$$>$$
 ae $<$ - aquaenv($Tc = 15$, $S = 20:30$, $d = 10$, $SumCO2 = SumCO2$, $TA = TA$) $>$ plot(ae, xval = 20:30, xlab = "S")

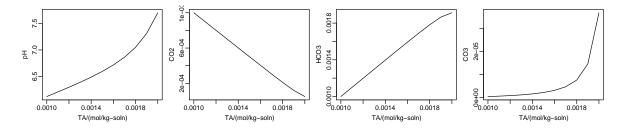
- > ae <- aquaenv(Tc = 15, S = 30, d = seq(1, 1000, 200), SumCO2 = SumCO2,
- + TA = TA, revelle = TRUE, dsa = TRUE)
- > plot(ae, xval = seq(1, 1000, 200), xlab = "depth/m")

Interesting to note is that also, e.g., SumCO2, TA, pH and SumNH4 can be vectors

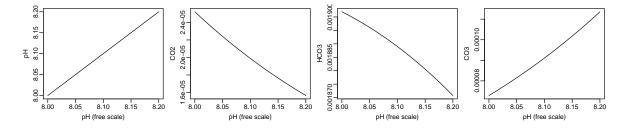
> ae <- aquaenv(10, 20, SumCO2 = seq(0.001, 0.002, 1e-04), TA = 0.002) > plot(ae, xval = ae\$SumCO2, xlab = "SumCO2/(mol/kg-soln)", what = c("pH", "CO2", "HCO3", "CO3"), newdevice = FALSE, mfrow = c(1, 4))



- > ae <- aquaenv(10, 20, SumCO2 = 0.002, TA = seq(0.001, 0.002,
- + 1e-04))
- > plot(ae, xval = aeTA, xlab = "TA/(mol/kg-soln)", what = c("pH",
- + "CO2", "HCO3", "CO3"), newdevice = FALSE, mfrow = c(1, 4))



> ae <- aquaenv(10, 20, SumCO2 = 0.002, pH = seq(8, 8.2, 0.001)) > plot(ae, xval = ae\$pH, xlab = "pH (free scale)", what = c("pH", + "CO2", "HCO3", "CO3"), newdevice = FALSE, mfrow = c(1, 4))



```
> ae <- aquaenv(10, 20, SumCO2 = 0.002, SumNH4 = seq(1e-04, 2e-04,
+ 1e-05), pH = 8)
> ae$NH3

[1] 1.534562e-06 1.688018e-06 1.841474e-06 1.994930e-06 2.148386e-06
[6] 2.301842e-06 2.455299e-06 2.608755e-06 2.762211e-06 2.915667e-06
[11] 3.069123e-06
attr(,"unit")
[1] "mol/kg-soln"
```

3.1.8 Calculating $[\sum CO_2]$ from input vectors

The functionality of calculating SumCO2 can also be used together with vectors as input variables.

```
> ae <- aquaenv(Tc = 11:15, S = 30, SumCO2 = NULL, pH = pH, CO2 = CO2,
+ revelle = TRUE, dsa = TRUE)
> ae$SumCO2

[1] 0.002415982 0.002463453 0.002511189 0.002559171 0.002607383
attr(,"unit")
[1] "mol/kg-soln"

Two further examples
> ae <- aquaenv(Tc = 15, S = 20:30, SumCO2 = NULL, pH = pH, pCO2 = pCO2)
> plot(ae, xval = 20:30, xlab = "S")
> ae <- aquaenv(Tc = 15, S = 30, d = seq(1, 1000, 100), SumCO2 = NULL,
+ pH = pH, TA = TA)
> plot(ae, xval = seq(1, 1000, 100), xlab = "depth/m")
```

3.1.9 Conversion from and to a dataframe

Objects of class aquaenv can be converted to an R data.frame to further post-process them with standard R means. Similarly, R data.frames can be converted to objects of class aquaenv to use the plotting facilities that exist for objects of class aquaenv. This can be helpful for plotting output of a dynamic model run, e.g. from R package **deSolve**, and will be shown later in this document.

```
> aedataframe <- as.data.frame(ae)
> aetest <- aquaenv(ae = aedataframe, from.data.frame = TRUE)</pre>
```

3.1.10 Converting elements in an obect of class aquaenv

Elements of an object of class *aquaenv* are calculated in, e.g., the concentration unit mol/kg-solution (molinity). The function convert can be used to convert all elements in an object of class *aquaenv* that share a common attribute, e.g. the unit.

3.1.11 Quantities needed for explicit pH modelling

As already mentioned above, the quantities needed for the explicit pH modelling approach (direct substitution approach - DSA) as presented by Hofmann *et al.* (2008b) can be calculated with the function aquaenv by setting the flag dsa.

```
> ae <- aquaenv(Tc = 15, S = 30, d = 10, SumCO2 = 0.002, pH = 8, + dsa = TRUE, revelle = TRUE)
```

This command calculated the buffer factor and the partial derivatives of [TA] with respect to other summed quantities referred to in Hofmann *et al.* (2008b)

> ae\$dTAdH

```
[1] -17142.05
attr(,"unit")
[1] "(mol-TA/kg-soln)/(mol-H/kg-soln)"
attr(,"pH scale")
[1] "free"
> ae$dTAdSumCO2
[1] 1.045937
attr(,"unit")
[1] "(mol-TA/kg-soln)/(mol-SumCO2/kg-soln)"
```

as well the sums partial derivatives of [TA] with respect to the equilibrium constants (K^* 's) multiplied with the partial derivatives of the respective equilibrium constant with one of their variables (i.e., S, T, d, SumH2SO4, od SumHF) as introduced in Hofmann *et al.* (2008a).

> ae\$dTAdKdKdS

```
[1] 3.9865e-06
attr(,"unit")
[1] "(mol-TA/kg-soln)/\"psu\""
> ae$dTAdKdKdSumH2SO4
[1] -0.001086853
```

```
[1] -0.001086853
attr(,"unit")
[1] "(mol-TA/kg-soln)/(mol-SumH2SO4/kg-soln)"
```

Furthermore, the ionization fractions used for the pH dependent fractional stoichiometric pH modelling approach described in Hofmann *et al.* (2008c) are calculated as well

> ae\$c1

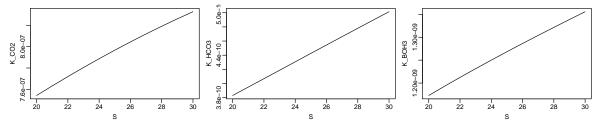
[1] 0.01025961

3.2 The plot.aquaenv function

In the previous sections, the plot function has been introduced. What actually is called if the first element of the arguments list of plot is an object pf class aquaenv is the function plot.aquaenv. This is a multifunctional tool to visualize information contained in an object of class aquaenv. For the convenience of the users, plot.aquaenv combines the call of standard R plotting functions and the previous call of the function par to set parameters like mfrow, mar, etc. as well as the opening of a plotting device with a certain size. As already shown above, setting the flag newdevice to FALSE suppresses the opening of a new plotting device (this feature is needed here to create a plot that will be woven into the LATEX document by Sweave).

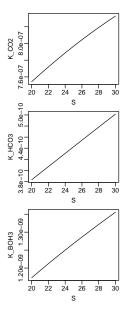
For example

```
> ae <- aquaenv(10, 20:30)
> plot(ae, xval = 20:30, xlab = "S", what = c("K_C02", "K_HC03", "K_B0H3"), size = c(10, 2), mfrow = c(1, 3), newdevice = FALSE)
```



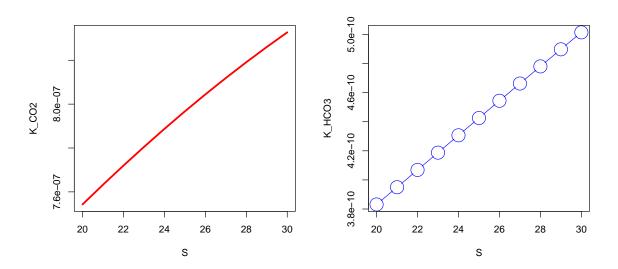
and

```
> plot(ae, xval = 20:30, xlab = "S", what = c("K_C02", "K_HC03", "K_B0H3"), size = c(2, 10), mfrow = c(3, 1), newdevice = FALSE)
```



Furthermore the parameter device can be specified which allows the user to write the plots to .eps and .pdf files. The parameter filename can be used to specify a filename other than the default filename "aquaenv".

These features make the function plot.aquaenv different from standard R plotting functions. However, if the flags newdevice and setpar are set to FALSE, plot.aquaenv behaves like a "normal" R plotting function



Furthermore, the function plot.aquaenv can be used to create "cumulative" plots and "Bjerrum" plots. This will be explained in some of the following sections.

3.3 Using objects of class aquaenv in dynamic models

3.3.1 Ordinary dynamic models

It is convenient to use objects of class *aquaenv* in a dynamic model, e.g. solved using the R package **deSolve**. This can be illustrated with an example. (For information about how to set up a dynamic model with **deSolve**, consult the documentation of **deSolve**).

A list of parameters is specified

```
> parameters <- list(</pre>
                     = 25
         S
                             , # psu
                             , #
         Tc\_min
                    = 5
                                 degrees C
                             , # degrees C
                     = 25
         Tc_max
                     = 10
                             , # m
         d
                                 , # 1/d
                                                         proportionality factor for air-water exchange
         k
                     = 0.4
         r0x
                    = 0.0000003 , \# mol-N/(kg*d)
                                                   maximal rate of oxic mineralisation
         rNitri
                    = 0.0000002 , # mol-N/(kg*d)
                                                   maximal rate of nitrification
                    = 0.000006 , # mol-N/(kg*d)
         rPP
                                                   maximal rate of primary production
         ksDINPP
                     = 0.000001
                                 , # mol-N/kg
         ksNH4PP
                     = 0.000001
                                 , # mol-N/kg
                     = 0.1
                                       1/d
                                                    (dispersive) transport coefficient
                                 , # mol/kg-soln
         02_io
                    = 0.000296
                                 , # mol/kg-soln
         NO3 io
                     = 0.000035
         SumNH4_io
                    = 0.000008
                                   # mol/kg-soln
                                 , # mol/kg-soln
         SumCO2_io
                    = 0.002320
                                 , # mol/kg-soln
                     = 0.002435
         TA\_io
         C_Nratio
                    = 8
                                 , # mol C/mol N
                                                      C:N ratio of organic matter
                     = 30
                                   # timestep from which PP begins
```

```
+ b = 50 , # timestep where PP shuts off again

+ modeltime = 100 # duration of the model

+ )
```

A model function is defined which will be executed every timestep of the numerical integration. An object of class *aquaenv* is created in each timestep, some of its elements are used to calculated kinetic rate expressions and the whole object is returned as output.

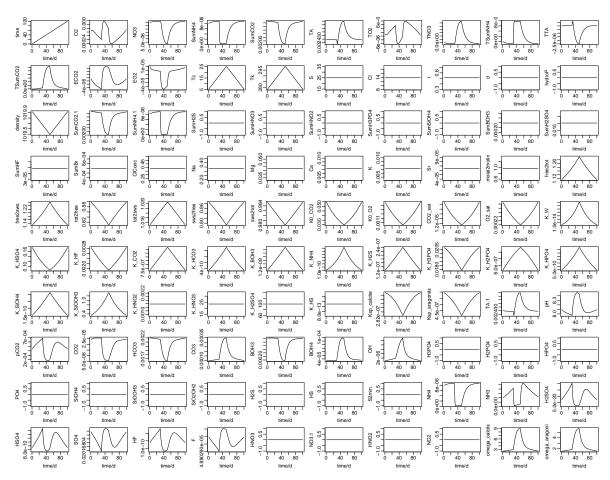
```
> Waddenzeebox <- function(timestep, currentstate, parameters)</pre>
   with (
          as.list(c(currentstate,parameters)),
           Tc \leftarrow c(seq(Tc\_min, Tc\_max, (Tc\_max-Tc\_min)/(modeltime/2)),
                    seq(Tc_max, Tc_min, -(Tc_max-Tc_min)/(modeltime/2)))[[round(timestep)+1]]
            ae <- aquaenv(Tc=Tc, S=S, SumCO2=SumCO2, SumNH4=SumNH4, TA=TA)
            ECO2
                    <- k * (ae$CO2_sat - ae$CO2)
                    <- k * (ae$02_sat - 02)
           E02
                    <- D*(02 io
            TN2
                                    - 02)
                                  - NO3)
            TNO3
                   <- D*(NO3_io
            TSumNH4 <- D*(SumNH4_io - SumNH4)
                                 - TA)
            TTA <- D*(TA_io
            TSumCO2 \leftarrow D*(SumCO2\_io - SumCO2)
           RNit
                      <- rNitri
                     <- r0x
            R\Omega x
           ROxCarbon <- ROx * C_Nratio
           pNH4PP <- 0
           RPP <- 0
            if ((timestep > a) && (timestep < b))</pre>
                      <- rPP * ((SumNH4+NO3)/(ksDINPP + (SumNH4+NO3)))
               pNH4PP <- 1 - (ksNH4PP/(ksNH4PP + SumNH4))
            else
               RPP <- 0
              7
            RPPCarbon <- RPP * C_Nratio
                               + E02 - ROxCarbon - 2*RNit + (2-2*pNH4PP)*RPP + RPPCarbon
            d02
                    <- T02
                    <- TNO3
                              + RNit -(1-pNH4PP)*RPP
            dNO3
            dSumCO2 <- TSumCO2 + ECO2 + ROxCarbon - RPPCarbon
            dSumNH4 <- TSumNH4 + ROx - RNit - pNH4PP*RPP
                              + ROx - 2*RNit -(2*pNH4PP-1)*RPP
            ratesofchanges <- c(d02, dN03, dSumNH4, dSumCO2, dTA)
           transport <- c(T02=T02, TN03=TN03, TSumNH4=TSumNH4, TTA=TTA, TSumC02=TSumC02)
           airseaexchange <- c(ECO2=ECO2, EO2=EO2)</pre>
           return(list(ratesofchanges, list(transport, airseaexchange, ae)))
         }
```

The model is solved

```
> with (as.list(parameters),
+ {
+ initialstate <- c(02=02_io, N03=N03_io, SumNH4=SumNH4_io, SumC02=SumC02_io, TA=TA_io)
+ times <- c(0:modeltime)
+ output <- as.data.frame(vode(initialstate,times,Waddenzeebox,parameters, hmax=1))[-1,]
+ })</pre>
```

and the output can be plotted in the same way as a two dimensional object of class aquaenv by converting it to an object of class aquaenv using the from.data.frame flag of the function aquaenv

```
> plot(aquaenv(ae = output, from.data.frame = TRUE), xval = outputtime,
+ xlab = "time/d", mfrow = c(10, 10), newdevice = FALSE)
```



3.3.2 Models using the explicit pH modelling approach

3.3.2.1 In one single model

Since an object of class *aquaenv* can contain all quantities necessary to employ the explicit pH modelling approaches as introduced by Hofmann *et al.* (2008b,a,c), they can be readily used in an explicit pH model.

As an example, we give a model that calculates the pH in the "classical" way in every timestep using aquaenv, also employs the explicit pH modelling approach (direct substitution approach - DSA) given in Hofmann et al. (2008b) and additionally employs fractional stoichiometry as given in Hofmann et al. (2008c). The pH evolution is thus calculated in three different ways which allows comparing the three values for consistency.

Again, a list of parameters is defined

```
> parameters <- list(</pre>
       \mathcal{S}
                    = 25
                              , # psu
       Tc
                    = 15
                             , # degrees C
                             , # m
                    = 10
       d
                                  , # 1/d
                                                        proportionality factor for air-water exchange
                    = 0.4
       r0x
                    = 0.0000003 , # mol-N/(kg*d) maximal rate of oxic mineralisation
                    = 0.0000002 , # mol-N/(kg*d) maximal rate of nitrification = 0.0000006 , # mol-N/(kg*d) maximal rate of primary production
       rNitri
       rPP
                    = 0.000001 , # mol-N/kg
       ksSumNH4
                                                          (dispersive) transport coefficient
                                 , # mol/kg-soln
       02_io
                    = 0.000296
                                 , # mol/kg-soln
       NO3_io
                    = 0.000035
                                 , # mol/kg-soln
       SumNH4_{io} = 0.000008
                                 , # mol/kg-soln
, # mol/kg-soln
       SumCO2_io
                    = 0.002320
                    = 0.002435
       TA_{-}io
       C Nratio
                                , # mol C/mol\ N
                                                       C:N ratio of organic matter
                    = .30
                                 , # timestep from which PP begins
       b
                    = 50
                                 , # timestep where PP shuts off again
       modeltime
                   = 100
                                   # duration of the model
```

And a model function is defined. Again, an object of class *aquaenv* is created in each timestep and respective elements are used.

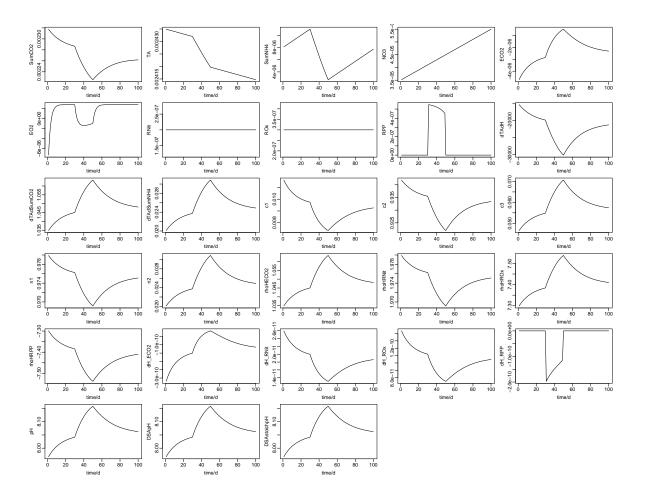
```
> boxmodel <- function(timestep, currentstate, parameters)</pre>
+ {
    with (
          as.list(c(currentstate,parameters)),
            # the "classical" implicit pH calculation method is applied in aquaenv
            ae <- aquaenv(Tc=Tc, S=S, SumCO2=SumCO2, SumNH4=SumNH4, TA=TA, dsa=TRUE)
            EC02
                     <- k * (ae$CO2_sat - ae$CO2)
                     <- k * (ae$02_sat - 02)
            E02
            RNit
                       <- rNitri
            ROx
                      <- r0x
            if ((timestep > a) && (timestep < b))
                RPP <- rPP * (SumNH4/(ksSumNH4 + SumNH4))</pre>
              }
            else
                RPP <- 0
```

```
d02
        <- EO2 - C_Nratio*ROx - 2*RNit + C_Nratio*RPP
dNO3
        <- RNit
dSumCO2 <- ECO2 + C_Nratio*ROx - C_Nratio*RPP
dSumNH4 <- ROx - RNit - RPP
        <- ROx - 2*RNit - RPP
# The DSA pH
dH <- (dTA - (dSumCO2*ae$dTAdSumCO2 + dSumNH4*ae$dTAdSumNH4))/ae$dTAdH
DSApH <- -log10(H)
# The DSA pH using pH dependent fractional stoichiometry (= using partitioning coefficients)
rhoHECO2 <- ae$c2 + 2*ae$c3
rhoHRNit <- 1 + ae$n1
rhoHROx <- C_Nratio * (ae$c2 + 2*ae$c3) - ae$n1
rhoHRPP \leftarrow (C_Nratio * (ae$c2 + 2*ae$c3)) + ae$n1
dH_ECO2 <- rhoHECO2*ECO2/(-ae$dTAdH)</pre>
dH_RNit <- rhoHRNit*RNit/(-ae$dTAdH)</pre>
dH_ROx <- rhoHROx*ROx /(-ae$dTAdH)</pre>
dH_RPP <- rhoHRPP*RPP /(-ae$dTAdH)
dH_stoich <- dH_ECO2 + dH_RNit + dH_ROx + dH_RPP
DSAstoichpH <- -log10(H_stoich)
ratesofchanges \leftarrow c(d02, dN03, dSumNH4, dSumC02, dTA, dH, dH\_stoich)
processrates <- c(ECO2=ECO2, EO2=EO2, RNit=RNit, ROx=ROx, RPP=RPP)</pre>
               <- c(DSApH=DSApH, rhoHECO2=rhoHECO2, rhoHRNit=rhoHRNit, rhoHROx=rhoHROx,</pre>
DSA
                    rhoHRPP=rhoHRPP, dH_ECO2=dH_ECO2, dH_RNit=dH_RNit, dH_ROx=dH_ROx,
                     dH_RPP=dH_RPP, DSAstoichpH=DSAstoichpH)
return(list(ratesofchanges, list(processrates, DSA, ae)))
```

The model is solved

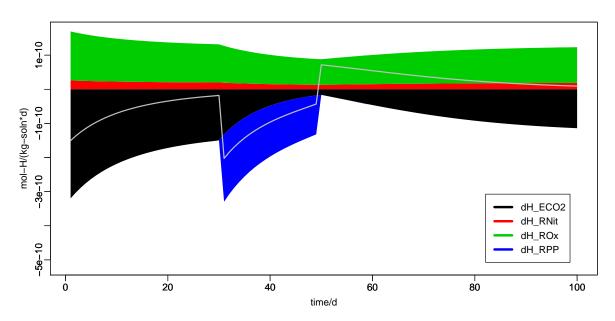
```
> with (as.list(parameters),
+ {
+ H_init <<- 10^(-(aquaenv(Tc=Tc, S=S, SumCO2=SumCO2_io, SumNH4=SumNH4_io, TA=TA_io, speciation=FALSE)$pH))
+ initialstate <<- c(O2=O2_io, NO3=NO3_io, SumNH4=SumNH4_io, SumCO2=SumCO2_io, TA=TA_io, H=H_init, H_stoich=H_init)
+ times <<- c(O:modeltime)
+ output <<- as.data.frame(vode(initialstate, times, boxmodel, parameters, hmax=1))[-1,]
+ })</pre>
```

and output can be plotted. Again using plot.aquaenv. Note that here the parameter what is used.



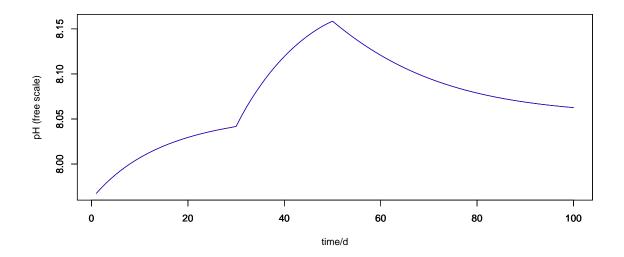
Here, the cumulative plotting functionality of plot.aquaenv can be employed as well to visualize the influences of the different kinetically modelled processes on $[H^+]$.

```
> what <- c("dH_ECO2", "dH_RNit", "dH_ROx", "dH_RPP")
> plot(aquaenv(ae = output, from.data.frame = TRUE), xval = output$time,
+ what = what, xlab = "time/d", size = c(7, 5), ylab = "mol-H/(kg-soln*d)",
+ legendposition = "bottomright", cumulative = TRUE, newdevice = FALSE)
```



Finally, the pH values calculated with the three different methods can be plotted in one single graph to see that they are identical, i.e. the three methods of pH calculation are consistent with each other

```
> ylim <- range(output$DSApH, output$DSAstoichpH, output$pH)
> plot(output$pH, ylim = ylim, type = "l", xlab = "time/d", ylab = "pH (free scale)")
> par(new = TRUE)
> plot(output$DSApH, ylim = ylim, type = "l", col = "red", xlab = "",
+ ylab = "")
> par(new = TRUE)
> plot(output$DSAstoichpH, ylim = ylim, type = "l", col = "blue",
+ xlab = "", ylab = "")
```

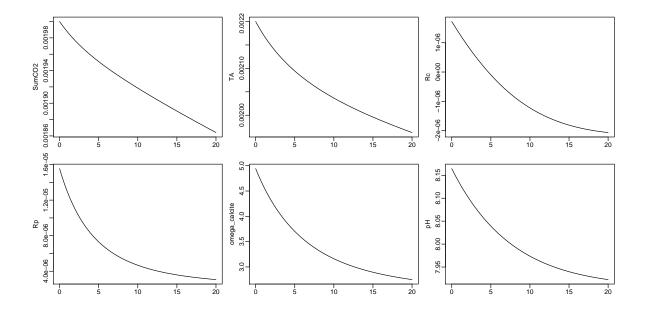


3.3.2.2 In three separate models

3.3.2.2.1 The implicit pH modelling approach

```
A list of parameters
```

```
> parameters <- list(</pre>
                = .35
                            , # psu
                = 15
                            , # degrees C
                            , # mol/kg-soln (comparable to Wang2005)
     SumCO2\_tO = 0.002
                = 0.0022
                            , # mol/kg-soln (comparable to Millero1998)
                            , # 1/d
                = 0.5
                                                  proportionality factor for air-water exchange
                = 0.000001 , \# mol/(kg-soln*d)
                                                    max rate of calcium carbonate precipitation
     kp
                            , # -
                = 2.0
                                                exponent for kinetic rate law of precipitation
     n
     modeltime = 20
                            , # d
                                             duration of the model
     outputsteps = 100
                             #
                                             number of outputsteps
The model function
> boxmodel <- function(timestep, currentstate, parameters)</pre>
   with (
         as.list(c(currentstate,parameters)),
                <- aquaenv(Tc=Tc, S=S, SumCO2=SumCO2, TA=TA, SumSiOH4=0,</pre>
           ae
                           SumBOH3=0, SumH2SO4=0, SumHF=0)
                <- kc * ((ae$CO2_sat) - (ae$CO2))
           Rc
                <- kp * (1-ae$omega_calcite)^n
           dSumCO2 <- Rc - Rp
                  <- -2*Rp
           ratesofchanges <- c(dSumCO2, dTA)
          processrates <- c(Rc=Rc, Rp=Rp)
           return(list(ratesofchanges, list(processrates, ae)))
+ }
Solving the model
> with (as.list(parameters),
         initialstate <<- c(SumCO2=SumCO2_t0, TA=TA_t0)</pre>
                 <<- seq(0,modeltime,(modeltime/outputsteps))</pre>
         times
                     <-- as.data.frame(vode(initialstate,times,boxmodel,parameters, hmax=1))
         output
       })
Visualizing the output
> what <- c("SumCO2", "TA", "Rc", "Rp", "omega_calcite", "pH")
> plot(aquaenv(ae = output, from.data.frame = TRUE), xval = output$time,
       xlab = "", mfrow = c(2, 3), size = c(12, 5), what = what,
       newdevice = FALSE)
```



3.3.2.2.2 The explicit pH modelling approach

A list of parameters

```
> parameters <- list(</pre>
                               , # psu
                  = 35
                  = 15
                               , # degrees C
      Tc
                               , # mol/kg-soln (comparable to Wang2005)
      SumCO2_tO
                  = 0.002
      TA\_t0
                  = 0.0022
                               , # mol/kg-soln
                                                 (comparable to Millero1998)
                               , # 1/d
                  = 0.5
                                                        proportionality factor for air-water exchange
      kc
                                                            \hbox{\it max rate of calcium carbonate precipitation}\\
                  = 0.000001
                               , # mol/(kg-soln*d)
      kр
                  = 2.0
                                                      exponent for kinetic rate law of precipitation
                               , # d
      modeltime
                  = 20
                                                   duration of the model
      outputsteps = 100
                                                   number of outputsteps
```

The model function

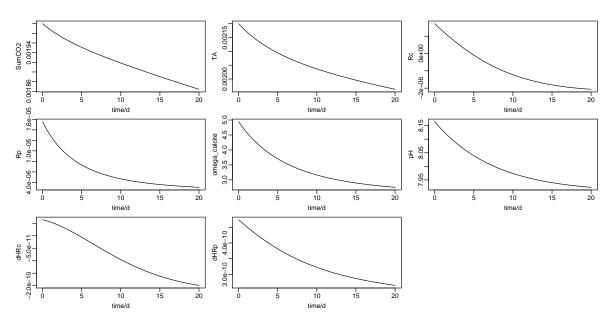
```
> boxmodel <- function(timestep, currentstate, parameters)</pre>
    with (
          as.list(c(currentstate,parameters)),
                  <- aquaenv(Tc=Tc, S=S, SumCO2=SumCO2, pH=-log10(H), SumSiOH4=0,
            ae
                             SumBOH3=0, SumH2SO4=0, SumHF=0, dsa=TRUE)
                  <- kc * ((ae$CO2_sat) - (ae$CO2))
            Rc
                  <- kp * (1-ae$omega_calcite)^n
            Rр
            dSumCO2 <- Rc - Rp
                              -(ae$dTAdSumCO2*Rc ))/ae$dTAdH
            dHRp
                    <- (-2*Rp -(ae$dTAdSumCO2*(-Rp)))/ae$dTAdH
            dН
                    <- dHRc + dHRp
            ratesofchanges <- c(dSumCO2, dH)
```

```
+ processrates <- c(Rc=Rc, Rp=Rp)
+ outputvars <- c(dHRc=dHRc, dHRp=dHRp)
+
+ return(list(ratesofchanges, list(processrates, outputvars, ae)))
+ }
+ )
+ )</pre>
```

Solving the model

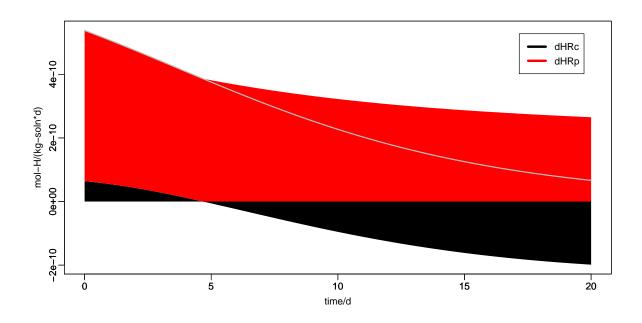
Visualizing the output

```
> what <- c("SumCO2", "TA", "Rc", "Rp", "omega_calcite", "pH",
+    "dHRc", "dHRp")
> plot(aquaenv(ae = output, from.data.frame = TRUE), xval = output$time,
+    xlab = "time/d", mfrow = c(3, 3), size = c(15, 10), what = what,
+    newdevice = FALSE)
```



Cumulatively plotting the influences of the two processes on the pH

```
> what <- c("dHRc", "dHRp")
> plot(aquaenv(ae = output, from.data.frame = TRUE), xval = output$time,
+ xlab = "time/d", what = what, ylab = "mol-H/(kg-soln*d)",
+ legendposition = "topright", cumulative = TRUE, newdevice = FALSE)
```



3.3.2.2.3 The fractional stoichiometric approach

A list of parameters

```
> parameters <- list(</pre>
                                    , # psu
       \mathcal{S}
                     = 35
                      = 15
                                    , # degrees C
       Tc
                                   , # mol/kg-soln (comparable to Wang2005)
, # mol/kg-soln (comparable to Millero1998)
       SumCO2_tO
                     = 0.002
       TA\_tO
                     = 0.0022
                     = 0.5
                                   , # 1/d
, # mol/(kg-soln*d)
                                                                proportionality factor for air-water exchange
       kc
                                                                    max rate of calcium carbonate precipitation
                      = 0.000001
       kр
                      = 2.0
                                                             exponent for kinetic rate law of precipitation
       {\tt modeltime}
                    = 20
                                    , # d
                                                          duration of the model
        outputsteps = 100
                                                          number of outputsteps
```

The model function

```
> boxmodel <- function(timestep, currentstate, parameters)</pre>
    with (
          as.list(c(currentstate,parameters)),
                  <- aquaenv(Tc=Tc, S=S, SumCO2=SumCO2, pH=-log10(H), SumSiOH4=0,
            ae
                             SumBOH3=0, SumH2SO4=0, SumHF=0, dsa=TRUE)
            Rc
                  <- kc * ((ae$CO2_sat) - (ae$CO2))
                  <- kp * (1-ae$omega\_calcite)^n
            Rр
            dSumCO2 <- Rc - Rp
            rhoc
                    <- ae$c2 + 2*ae$c3
            rhop
                    <- 2*ae$c1 + ae$c2
                    <- rhoc*Rc/(-ae$dTAdH)
            dHRc
            dHRp
                    <- rhop*Rp/(-ae$dTAdH)
            dН
                    <- dHRc + dHRp
```

```
ratesofchanges <- c(dSumCO2, dH)
             processrates
                              <- c(Rc=Rc, Rp=Rp)
                              <- c(dHRc=dHRc, dHRp=dHRp, rhoc=rhoc, rhop=rhop)
             outputvars
             return(list(ratesofchanges, list(processrates, outputvars, ae)))
Solving the model
> with (as.list(parameters),
           aetmp <- aquaenv(Tc=Tc, S=S, SumCO2=SumCO2_t0, TA=TA_t0, SumSiOH4=0,</pre>
                              SumBOH3=0, SumH2SO4=0, SumHF=0)
           H_t0 <- 10^(-aetmp$pH)</pre>
           initialstate <<- c(SumCO2=SumCO2_t0, H=H_t0)</pre>
                          <-- seq(0,modeltime,(modeltime/outputsteps))
           times
           output
                          <\!\!<\!\!-\text{ as.data.frame}(\texttt{vode}(\texttt{initialstate},\texttt{times},\texttt{boxmodel},\texttt{parameters},~\texttt{hmax=1}))
Visualizing the output
> what <- c("SumCO2", "TA", "Rc", "Rp", "omega_calcite", "pH",
         "dHRc", "dHRp", "rhoc", "rhop")
> plot(aquaenv(ae = output, from.data.frame = TRUE), xval = output$time,
         xlab = "time/d", mfrow = c(3, 4), size = c(15, 10), what = what,
         newdevice = FALSE)
                                0.00215
  0.00194
                                                                                            1.0e-05
                                                            Rc
0e+00
                              Ϋ́
                                                                                          Вp
                                                              -2e-06
                     15
                                                                                       20
                10
                                             10
  5.0
                                8.15
  4.0 4.5
omega_calcite
                                                            dHRc
-5.0e-11
                                                                                            4.0e-
                                                                                          dHRp
                              PH
8.05
  3.5
                                                                                            3.0e-10
  3.0
                                7.95
                     15
                                                   15
                                                                                 15
                                                                                                               15
                10
                                             10
                                                                           10
                                                                                                         10
                                                                                                        time/d
                                0.94
  1.08
rhoc
                              rhop
                                0.92
  1.06
```

3.4 Titration simulation: the function titration

10

With the function **titration AquaEnv** provides a powerful tool to simulate titrations. A two dimensional object of class *aquaenv* will be created where the second dimension is the

amount of titrant added. For this purpose, three extra elements are added to the *aquaenv* object that will be created:

elemer	\mathbf{nt}	unit	explanation
delta_co	onc_titrant	mol/kg-solution	the offset in concentration of the titrant that is caused by adding the titrant to the sample
delta_m	nass_titrant	kg	the amount of mass of titrant solution added
delta_m	noles_titrant	mol	the amount of moles of titrant added

Each one of this elements is a suitable xval for plotting an aquaenv object generated by titration.

3.4.1 Titration with HCl

The standard titration type is titration with hydrochloric acid (HCl). A simple example will illustrate this best.

An object of class *aquaenv* needs to be created to define the initial conditions of the titration. That is temperature, salinity, depth, the concentrations of all summed quantities and the initial pH (or [TA]).

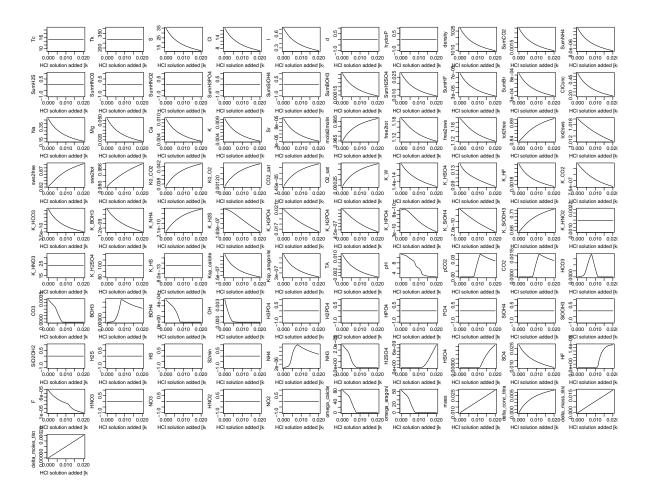
```
> ae_init <- aquaenv(Tc = 15, S = 35, SumCO2 = 0.0035, SumNH4 = 2e-05, pH = 11.3)
```

Then titration can be run to create the object describing the simulated titration. In this example the titrant is HCl of the relatively low concentration of 0.01 mol/kg-solution. The sample solution amounts to 10 g. To sweep a considerable pH range quite a lot of sample needs to be added: 20 g. This means the salinity of the solution in the titration vessel will change due to dilution with the titrant solution. For this reason, the salinity of the titrant solution needs to be given via the parameter S_titrant. However, we assume the titrant does not contain borate, sulfate or fluoride, that is why we do not set the flag seawater_titrant to TRUE.

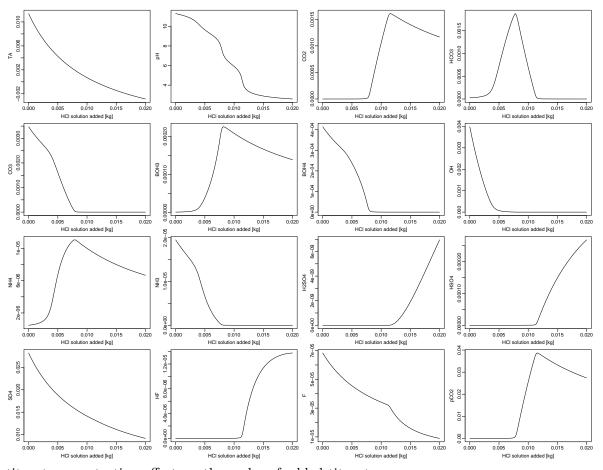
```
> ae <- titration(ae_init, mass_sample = 0.01, mass_titrant = 0.02,
+ conc_titrant = 0.01, S_titrant = 0.5, steps = 100)</pre>
```

To get a quick overview, all elements of the obtained aquaenv object can be plotted

```
> plot(ae, xval = ae$delta_mass_titrant, xlab = "HCl solution added [kg]", + mfrow = c(10, 10), newdevice = FALSE)
```



Then, a selection of elements can be plotted as a function of the added titrant mass,

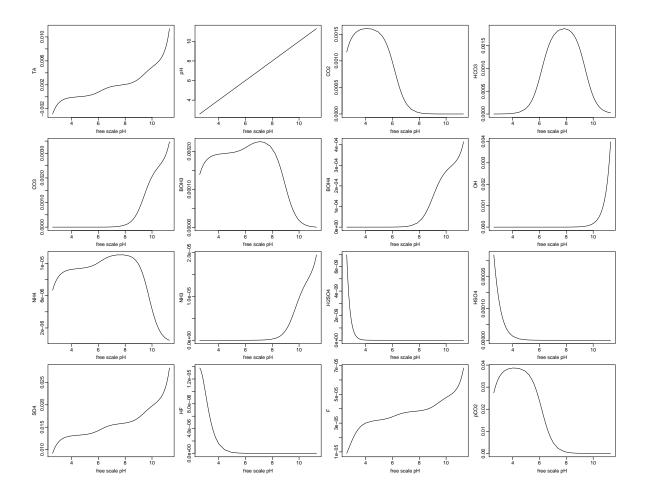


titrant concentration offset, or the moles of added titrant

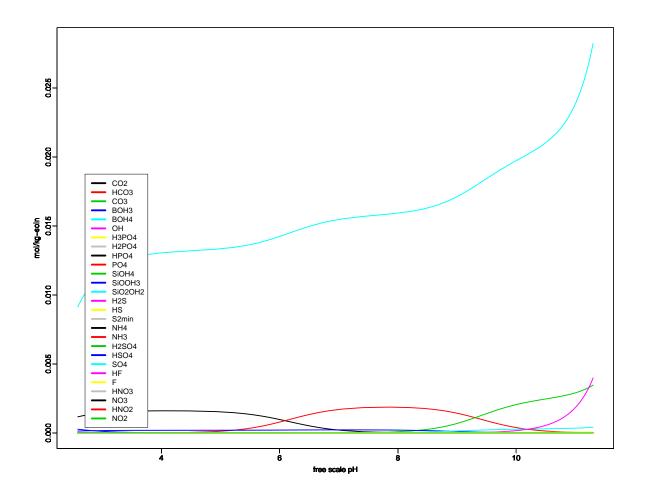
```
> plot(ae, xval = ae\$delta_conc_titrant, xlab = "[HCl] offset added [mol/kg-soln]",
+ what = what, size = c(14, 10), mfrow = c(12, 8))
> plot(ae, xval = ae\$delta_moles_titrant, xlab = "HCl added [mol]",
+ what = what, size = c(14, 10), mfrow = c(12, 8))
```

However, it is also possible to plot this selection of elements against othe variables, e.g., the calculated free scale pH

```
> plot(ae, xval = aepH, xlab = "free scale pH", what = what, size = c(12, + 8), mfrow = c(4, 4), newdevice = FALSE)
```

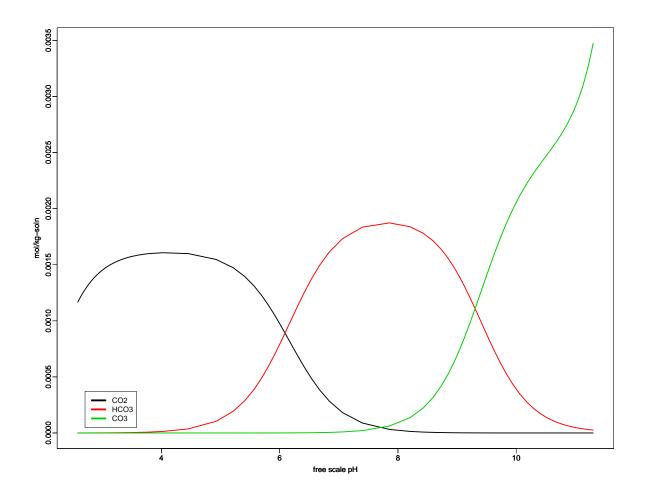


As mentioned earlier, the function plot.aquaenv offers the possibility of creating bjerrum plots from objects obtained with titration. The simplest way to do that is (remember the newdevice=FALSE is just needed to produce plots that are nicely woven into this vignette)

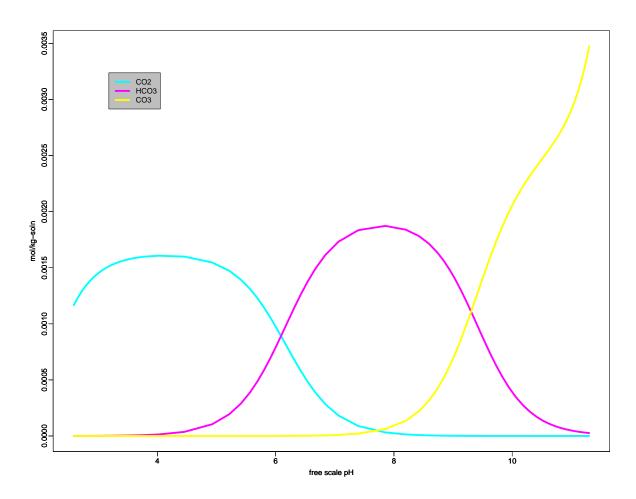


Or just select a few elements

```
> what <- c("CO2", "HCO3", "CO3")
> plot(ae, what = what, bjerrum = TRUE, newdevice = FALSE)
```

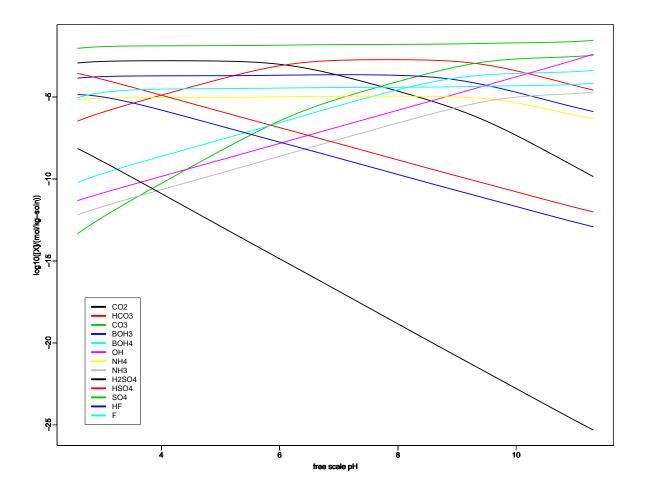


Again, the plotting style can be customized



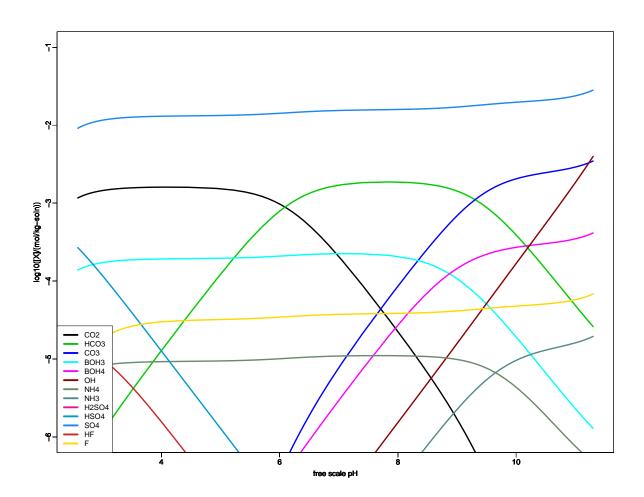
However, generally Bjerrum plots are done on the log scale. This can be accomplished using the flag log

```
> what <- c("CO2", "HCO3", "CO3", "BOH3", "BOH4", "OH", "NH4",
+ "NH3", "H2SO4", "HSO4", "SO4", "HF", "F")
> plot(ae, what = what, bjerrum = TRUE, log = TRUE, newdevice = FALSE)
```



Furthermore, we can zoom in to the region of most interest to marine scientists

```
> plot(ae, what = what, bjerrum = TRUE, log = TRUE, ylim = c(-6, + -1), legendinset = 0, lwd = 3, palette = c(1, 3, 4, 5, 6, colors()[seq(100, 250, 6)]), newdevice = FALSE)
```



3.4.2 Titration with NaOH

Similar to the titration with HCl, also a titration with NaOH can be simulated

```
> ae <- titration(aquaenv(Tc = 15, S = 35, SumCO2 = 0.0035, SumNH4 = 2e-05,
+ pH = 2), mass_sample = 0.01, mass_titrant = 0.02, conc_titrant = 0.01,
+ S_titrant = 0.5, steps = 50, type = "NaOH")</pre>
```

Plottinge everything

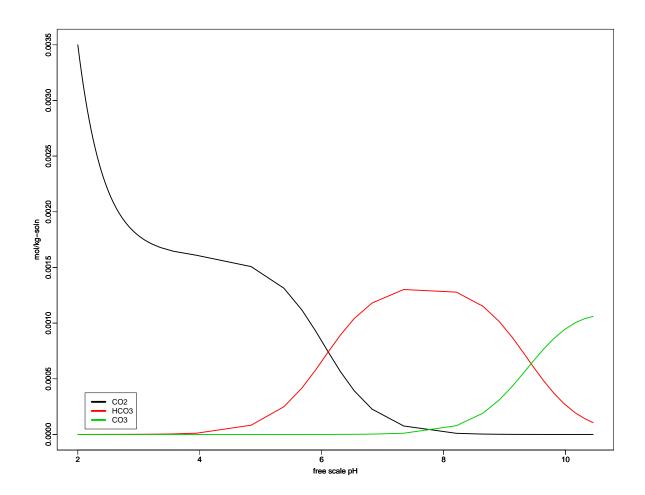
```
> plot(ae, xval = ae$delta_mass_titrant, xlab = "NaOH solution added [kg]", + mfrow = c(10, 10))
```

Plotting selectively

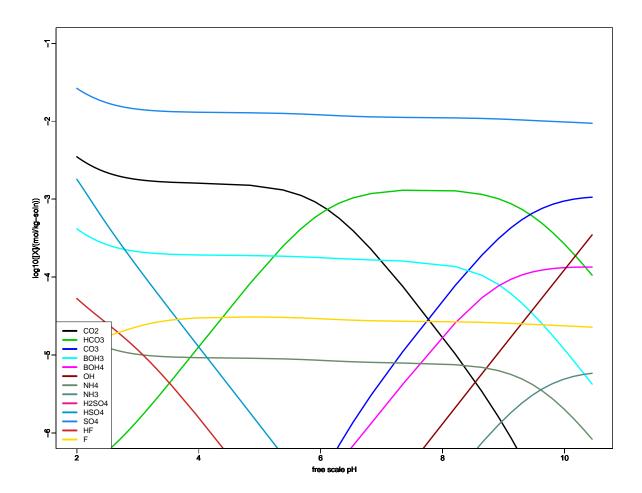
```
> what <- c("TA", "pH", "CO2", "HCO3", "CO3", "BOH3", "BOH4", "OH", + "NH4", "NH3", "H2SO4", "HSO4", "SO4", "HF", "F", "pCO2") > plot(ae, xval = ae$delta_mass_titrant, xlab = "NaOH solution added [kg]", + what = what, size = <math>c(12, 8), mfrow = c(4, 4)) > plot(ae, xval = ae$pH, xlab = "free scale pH", what = what, size = c(12, 8), mfrow = c(4, 4))
```

Bjerrum plots

```
> what <- c("CO2", "HCO3", "CO3")
> plot(ae, what = what, bjerrum = TRUE, newdevice = FALSE)
```



```
> what <- c("CO2", "HCO3", "CO3", "BOH3", "BOH4", "OH", "NH4",
+ "NH3", "H2SO4", "HSO4", "SO4", "HF", "F")
> plot(ae, what = what, bjerrum = TRUE, log = TRUE, ylim = c(-6,
+ -1), legendinset = 0, lwd = 3, palette = c(1, 3, 4, 5, 6,
+ colors()[seq(100, 250, 6)]), newdevice = FALSE)
```



3.4.3 Titration with a titrant with high concentrations and a large sample volume - classical Bjerrum plots

The Bjerrum plots created in the previous two sections do not really look like the classical textbook ones. This is because we simulated a titration with a small sample volume and a titrant with low concentrations. As a result the total concentrations like, e.g., total carbonate decreased due to dilution. In simulating a titration with a rather large volume and a titrant with high concentrations the volume and salinity corrections do not matter any more and graphs known from textbooks (e.g. Zeebe and Wolf-Gladrow 2001) are produced.

```
> ae <- titration(aquaenv(Tc = 15, S = 35, SumCO2 = 0.0035, SumNH4 = 2e-05, + pH = 11.3), mass_sample = 100, mass_titrant = 0.5, conc_titrant = 3, + S_{titrant} = 0.5, steps = 100)
```

Plotting everything

```
> plot(ae, xval = ae$delta_mass_titrant, xlab = "HCl solution added [kg]", + mfrow = c(10, 10))
```

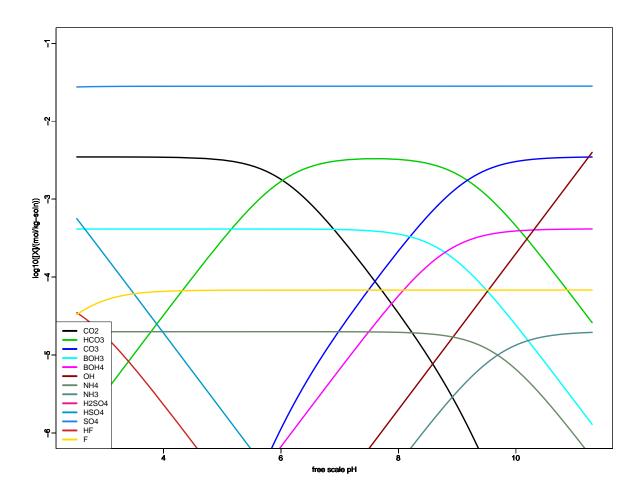
Plotting selectively and with different elements for xval

```
> what <- c("TA", "pH", "CO2", "HCO3", "CO3", "BOH3", "BOH4", "OH", + "NH4", "NH3", "H2SO4", "HSO4", "SO4", "HF", "F", "pCO2")
> plot(ae, xval = ae$delta_mass_titrant, xlab = "HCl solution added [kg]", + what = what, size = c(12, 8), mfrow = c(4, 4))
> plot(ae, xval = ae$pH, xlab = "free scale pH", what = what, size = c(12, + 8), mfrow = c(4, 4))
> plot(ae, xval = ae$delta_conc_titrant, xlab = "[HCl] offset added [mol/kg-soln]", + what = what, size = c(12, 8), mfrow = c(4, 4))
> plot(ae, xval = ae$delta_moles_titrant, xlab = "HCl added [mol]", + what = what, size = c(12, 8), mfrow = c(4, 4))
```

Creating different kinds of Bjerrum plots

and the classical textbook one

```
> plot(ae, what = what, bjerrum = TRUE, log = TRUE, ylim = c(-6, + -1), legendinset = 0, lwd = 3, palette = c(1, 3, 4, 5, 6, + colors()[seq(100, 250, 6)]), newdevice = FALSE)
```



3.5 Calculating information from titration curves: the function TAfit

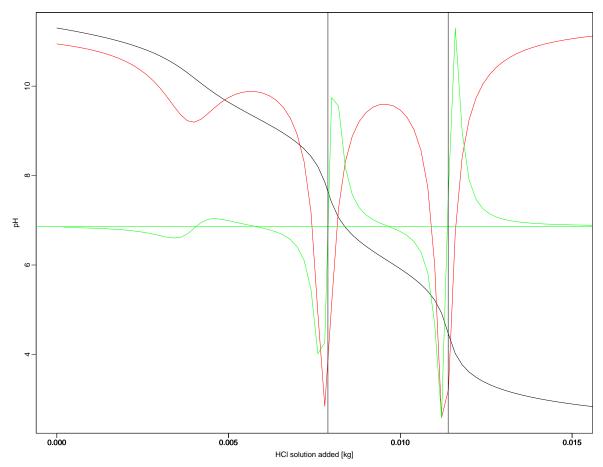
3.5.1 A little theory

While titrating a sample of natural seawater with HCl there one sees two clear equivalence points (Dickson 1981) The second equivalence point is the equivalence point of total alkalinity and the difference between the second and the first equivalence point signifies the total amount of $\sum CO_2$ of the sample Hansson and Jagner (1973).

This can be illustrated with <code>AquaEnv</code>. The respective titration curve can be plotted, together with its first and second derivative. Furthermore, the equivalence points can be marked with vertical lines (Please note that for a titrant concentration of 0.01 mol/kg-solution and 0.01 kg of sample, the value of the concentration (in mol/kg-solution) of total alkalinity and total carbonate equals the value of the total amount (in mol)).

```
> ae_init <- aquaenv(Tc = 15, S = 35, SumCO2 = 0.0035, SumNH4 = 2e-05, 
+ pH = 11.3)
> ae <- titration(ae_init, mass_sample = 0.01, mass_titrant = 0.02, 
+ conc_titrant = 0.01, S_titrant = 0.5, steps = 100)
> plot(ae, xval = ae$delta_mass_titrant, xlab = "HCl solution added [kg]",
```

```
+ what = "pH", xlim = c(0, 0.015), newdevice = FALSE)
> par(new = TRUE)
> plot(ae$delta_mass_titrant[1:100], diff(ae$pH), type = "l", col = "red",
+ xlim = c(0, 0.015), ylab = "", xlab = "", yaxt = "n")
> par(new = TRUE)
> plot(ae$delta_mass_titrant[2:100], diff(diff(ae$pH)), type = "l",
+ col = "green", xlim = c(0, 0.015), ylab = "", xlab = "",
+ yaxt = "n")
> abline(h = 0, col = "green")
> abline(v = ae$TA[[1]])
> abline(v = ae$TA[[1]] - ae$SumCO2[[1]])
```

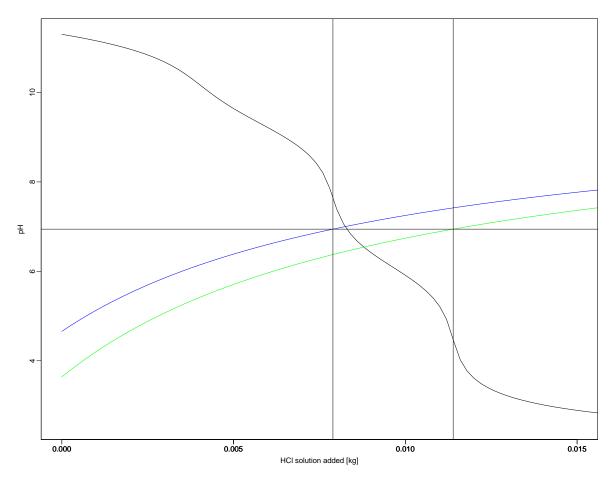


Following classical chemical textbooks (e.g. Skoog and West 1982), one can determine [TA] and $[\sum CO_2]$ of a sample by graphically determining those equivalence points. However, there is no mechanistic understanding of the contents of the solution involved in doing so.

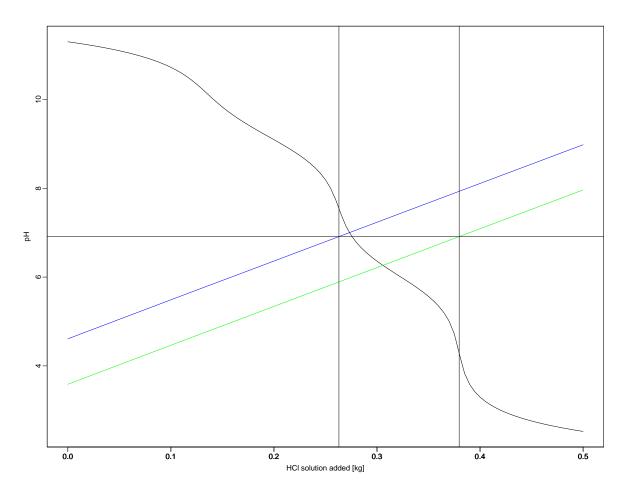
Other methods, called "Gran evaluations" (Gran 1952; Hansson and Jagner 1973; Dickson 1981; Haraldsson, Anderson, Hassellov, Hulth, and Olsson 1997; Anderson, Turner, Wedborg, and Dyrssen 1999), try to linearize the mechanistic model of what is going on in the solution during titration. They define the so called linear "Gran functions" and try to find their roots to determine the equivalence points. We will illustrate that by plotting the Gran functions

F0 (blue) and F2 (-F1, green) and again mark the equivalence points with vertical lines. The y=zero line for the Gran functions is indicated by a horizontal line

```
> plot(ae, xval = ae$delta_mass_titrant, xlab = "HCl solution added [kg]",
      what = "pH", xlim = c(0, 0.015), newdevice = FALSE)
> prot1 <- c()
> for (i in 1:length(ae$pH)) {
     prot1 <- c(prot1, (10^-(ae$pH[[i]]) + ae$HSO4[[i]] + ae$HF[[i]] +</pre>
          ae$CO2[[i]] - ae$CO3[[i]] - ae$BOH4[[i]] - ae$OH[[i]]))
+ }
> par(new = TRUE)
> plot(ae$delta_mass_titrant, prot1, type = "1", col = "blue",
      xlim = c(0, 0.015), ylab = "", xlab = "", yaxt = "n", ylim = c(-0.015,
          0.015))
> prot2 <- c()
> for (i in 1:length(ae$pH)) {
     prot2 <- c(prot2, (10^-(ae$pH[[i]]) + ae$HSO4[[i]] + ae$HF[[i]] -</pre>
          ae$HCO3[[i]] - 2 * ae$CO3[[i]] - ae$BOH4[[i]] - ae$OH[[i]]))
+
+ }
> par(new = TRUE)
> plot(ae$delta_mass_titrant, prot2, type = "1", col = "green",
     xlim = c(0, 0.015), ylab = "", xlab = "", yaxt = "n", ylim = c(-0.015)
         0.015))
> abline(v = ae$TA[[1]])
> abline(v = ae$TA[[1]] - ae$SumCO2[[1]])
> abline(h = 0)
```



One can see that the Gran functions actually are not linear. This is due to volume and salinity change effects during the titration. This can be overcome by either employing "modified Gran functions" (see Haraldsson $et\ al.\ 1997$) that correct for the volume changes or by using a titration with a titrant with high concentrations and a large sample volume (Please not that here the value of the concentration of total alkalinity and total carbonate does not equal their total amount and need to be converted with the factor 100/3)



Another proposed method of determining [TA] and $[\sum CO_2]$ is to not only determine the two equivalence points, but to fit the whole titration curve with a theoretical titration curve based on a mechanistic model of what is going on in the solution during the titration (Dickson 1981; DOE 1994; Anderson *et al.* 1999). The function **titration** of **AquaEnv** provides exactly such a theoretical titration curve and the function **TAfit** makes use of this fact to determine [TA] and $[\sum CO_2]$ of a sample by non linear curve fitting.

3.5.2 Determining [TA] and $[\sum CO_2]$ by non linear curve fitting

3.5.2.1 Proof of concept

First, a proof of concept will show that the function TAfit is implemented consistently. Some "data" can be generated with the titration function.

```
> initial_ae <- aquaenv(Tc = 15, S = 35, SumCO2 = 0.002, TA = 0.0022) > ae <- titration(initial_ae, mass_sample = 0.01, mass_titrant = 0.003, conc_titrant = 0.01, S_titrant = 0.5, steps = 20)
```

Now, the input data for the TAfit routine can be generated: a table with the added mass of the titrant and the resulting free scale pH

```
> titcurve <- cbind(ae$delta_mass_titrant, ae$pH)</pre>
```

Note that For the TAfit all total quantities except SumCO2 (SumNH4, SumH2S, SumH3PO4, SumSiOH4, SumHNO3, SumHNO2, SumBOH3, SumH2SO4, SumHF) need to be known. However, the latter three can be calculated from salinity as it is done in this example.

Thus, we see that TAfit calculates the correct SumCO2 and TA values.

Trying the Lueker, Dickson, and Keeling (2000) (K_CO2 and K_HCO3) and Perez and Fraga (1987b) (K_HF) values

```
$TA

[1] 0.0022

attr(,"unit")

[1] "mol/kg-soln"

$SumCO2

[1] 0.002

attr(,"unit")

[1] "mol/kg-soln"

$sumofsquares

[1] 1.676329e-29
```

TAfit can also take E (V) values as input variables, so we generate E values using E0=0.4 V and the Nernst equation. However, to do so we first need to convert our pH curve to the seawater pH scale. According to (DOE 1994, p.7, ch.4, sop.3), the Nernst equation relates E to the total proton concentration, but, if fluoride is present, its effect (as proton donor/acceptor) is also measured. Hence, we use the seawater scale here.

```
> swstitcurve <- convert(titcurve[, 2], "pHscale", "free2sws", + Tc = 15, S = 35) > Etitcurve <- cbind(titcurve[, 1], (0.4 - ((Constants$R/10) * + initial_ae$Tk/Constants$F) * log(10^-swstitcurve)))
```

Again, TAfit can be executed, this time also calculating E0. Note that the flag verbose=TRUE causes TAfit to show the progress of the fitting procedure in a plot window

```
causes TAfit to show the progress of the fitting procedure in a plot window.
> fit2 <- TAfit(initial_ae, Etitcurve, conc_titrant = 0.01, mass_sample = 0.01,
      S_titrant = 0.5, Evals = TRUE, verbose = TRUE)
> fit2
$TA
[1] 0.002199463
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.001997465
attr(,"unit")
[1] "mol/kg-soln"
$E0
[1] 0.3997121
attr(,"unit")
[1] "V"
```

\$sumofsquares

[1] 1.494212e-07

Furthermore, TAfit can fit K_CO2 as well, however, one single value for the whole titration curve is fitted, i.e. there is no correction for K_CO2 changes due to changing S due to mixing with the titrant

```
> fit3 <- TAfit(initial_ae, titcurve, conc_titrant = 0.01, mass_sample = 0.01,
      S_{titrant} = 0.5, K_{CO2fit} = TRUE
> fit3
$TA
[1] 0.002201253
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002002897
attr(,"unit")
[1] "mol/kg-soln"
$K_C02
[1] 9.169217e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
$sumofsquares
[1] 0.0001173612
> initial_ae$K_CO2
[1] 9.385878e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
```

One can see that the fitted value for K_CO2 is not the same as the value in the initial aquaenv object, which is the "correct" value. That is, because during data creation K_CO2 changed along the course of the titration due to changes in salinity. Assuming that the titrant has the same salinity as the sample (and is made up of natural seawater, i.e. containing SumBOH4, SumH2SO4 and SumHF as functions of S), then the "correct" K_CO2 should be fitted. This can be accomplished in TAfit by not giving the argument S_titrant (i.e. assuming the titrant has the same salinity as the sample) and setting the flag seawater_titrant to TRUE

```
> ae <- titration(initial_ae, mass_sample = 0.01, mass_titrant = 0.003,
+ conc_titrant = 0.01, steps = 20, seawater_titrant = TRUE)
> titcurve <- cbind(ae$delta_mass_titrant, ae$pH)</pre>
```

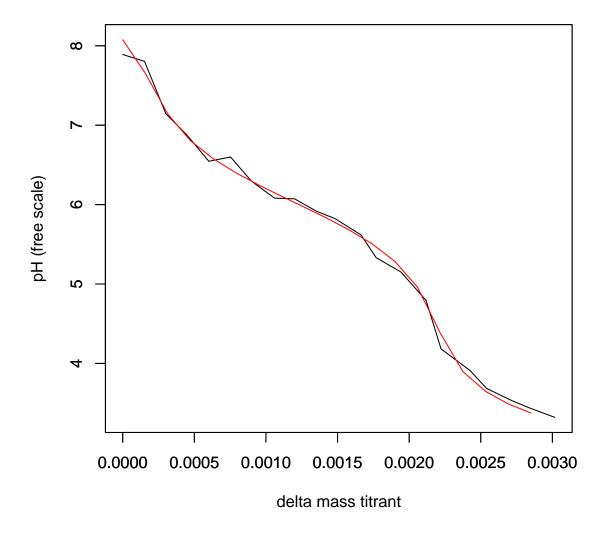
```
> fit4 <- TAfit(initial_ae, titcurve, conc_titrant = 0.01, mass_sample = 0.01,
      K_CO2fit = TRUE, seawater_titrant = TRUE)
> fit4
$TA
[1] 0.0022
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002
attr(,"unit")
[1] "mol/kg-soln"
$K_C02
[1] 9.385878e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
$sumofsquares
[1] 9.72271e-29
Furthermore, TA, SumCO2, K_CO2 and E0 can be fitted at the same time.
> Etitcurve <- cbind(titcurve[, 1], (0.4 - ((Constants$R/10) *
      initial_ae$Tk/Constants$F) * log(10^-titcurve[, 2])))
> fit5 <- TAfit(initial_ae, Etitcurve, conc_titrant = 0.01, mass_sample = 0.01,
      K_CO2fit = TRUE, seawater_titrant = TRUE, Evals = TRUE)
> fit5
$TA
[1] 0.0022
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002
attr(,"unit")
[1] "mol/kg-soln"
$F.0
[1] 0.4048613
attr(,"unit")
[1] "V"
```

```
$K_C02
[1] 9.385878e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
$sumofsquares
[1] 1.590048e-30
Sometimes, the obtained titration curve is not equally spaced on the x axis. TAfit can deal
with such curves if the flag equalspaced is set to FALSE
> neqsptitcurve <- rbind(titcurve[1:9, ], titcurve[11:20, ])</pre>
> fit6 <- TAfit(initial_ae, neqsptitcurve, conc_titrant = 0.01,
      mass_sample = 0.01, seawater_titrant = TRUE, equalspaced = FALSE,
      verbose = TRUE, debug = TRUE)
> fit6
$TA
[1] 0.002199682
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002000279
attr(,"unit")
[1] "mol/kg-soln"
$sumofsquares
[1] 0.002378034
Finally, some "noise" is added to the generated data
> noisetitcurve <- titcurve * rnorm(length(titcurve), mean = 1,</pre>
      sd = 0.01)
> fit7 <- TAfit(initial_ae, noisetitcurve, conc_titrant = 0.01,</pre>
      mass_sample = 0.01, seawater_titrant = TRUE, verbose = TRUE)
> fit7
$TA
[1] 0.002207184
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002007010
```

```
attr(,"unit")
[1] "mol/kg-soln"
$sumofsquares
[1] 0.1288424
```

The flag verbose=TRUE prompts to show the traject of the fitting procedure in a plot window. However, each new fit is plotted over the first one and Sweave includes only the first plot in each code chunk in the resulting LATEX file. Therefore, we use the flag debug=TRUE to visualize the final fit

```
> ylim = range(noisetitcurve[, 2], calc)
> xlim = range(tit$delta_mass_titrant, noisetitcurve[, 1])
> plot(noisetitcurve[, 1], noisetitcurve[, 2], xlim = xlim, ylim = ylim,
+ type = "l", xlab = "delta mass titrant", ylab = "pH (free scale)")
> par(new = TRUE)
> plot(tit$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,
+ type = "l", col = "red", xlab = "", ylab = "")
```



3.5.2.2 Test with generated data from literature

Dickson (1981) provided a synthetic dataset to test total alkalinity fitting programs. This dataset is included in **AquaEnv** as sample_dickson. Following quantities are given

```
> conc_titrant <- 0.3
```

> mass_sample <- 0.2

> S_titrant <- 14.835

> SumBOH3 <- 0.00042

> SumH2SO4 <- 0.02824

> SumHF <- 7e-05

Note that all concentrations are in mol/kg-solution and the mass of the sample is in kg. Note further that the salinity of the titrant has been calculated from its ionic strength of $0.3 \,$ mol/kg-soln.

In the original dataset as represented in sample_dickson, the mass of titrant is given in g which needs to be converted to kg

```
> sam <- cbind(sample_dickson[, 1]/1000, sample_dickson[, 2])
```

Then an attempt to recalculate the [TA] and $[\sum CO_2]$ values given in Dickson (1981) ([TA]=0.00245 mol/kg-soln and $[\sum CO_2]$ 0.00220 mol/kg-soln) can be done

\$sumofsquares

[1] 0.01287255

This shows the fit is not accurate. Why is that so?

3.5.2.2.1 Does the salinity correction (S_titrant) matter?

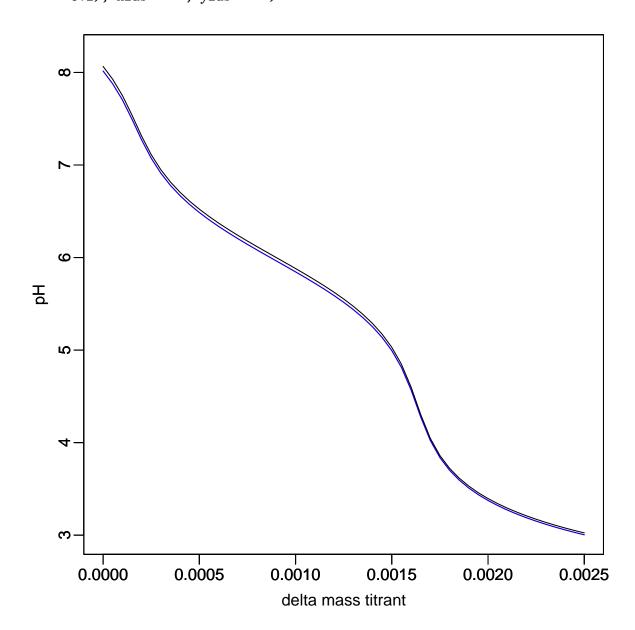
Let us calculate a theoretical titration without salinity correction

```
> dicksontitration1 <- titration(aquaenv(Tc = 25, S = 35, SumCO2 = 0.0022,
+ SumBOH3 = SumBOH3, SumH2SO4 = SumH2SO4, SumHF = SumHF, TA = 0.00245),
+ mass_sample = mass_sample, mass_titrant = 0.0025, conc_titrant = conc_titrant,
+ steps = 50, type = "HC1")

and one with salinity correction
> dicksontitration2 <- titration(aquaenv(Tc = 25, S = 35, SumCO2 = 0.0022,
+ SumBOH3 = SumBOH3, SumH2SO4 = SumH2SO4, SumHF = SumHF, TA = 0.00245),
+ mass_sample = mass_sample, mass_titrant = 0.0025, conc_titrant = conc_titrant,
+ S_titrant = S_titrant, steps = 50, type = "HC1")</pre>
```

Now the difference between both curves (in red and blue) and the "Dickson" curve (in black) can be visualized

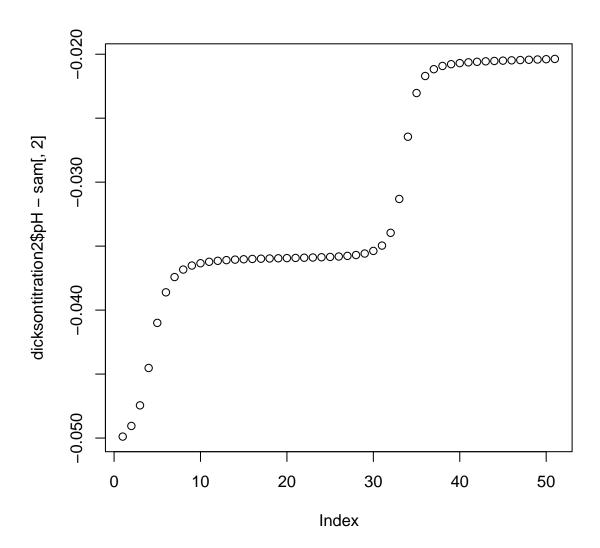
```
> plot(dicksontitration1, xval = dicksontitration1$delta_mass_titrant,
+ what = "pH", xlim = c(0, 0.0025), ylim = c(3, 8.2), newdevice = FALSE,
+ col = "red", xlab = "delta mass titrant")
> par(new = TRUE)
> plot(dicksontitration2, xval = dicksontitration2$delta_mass_titrant,
+ what = "pH", xlim = c(0, 0.0025), ylim = c(3, 8.2), newdevice = FALSE,
+ col = "blue", xlab = "")
> par(new = TRUE)
> plot(sam[, 1], sam[, 2], type = "l", xlim = c(0, 0.0025), ylim = c(3,
+ 8.2), xlab = "", ylab = "")
```



That means, the salinity correction makes no significant difference (the red and the blue curve cannot be discerned), because the relation between the total amount of sample and the added amount of titrant is very large: salinity only drops from 35 to 34.75105.

But there is an offset between the "Dickson" curve and our curve

> plot(dicksontitration2\$pH - sam[, 2])



3.5.2.2.2 Does fitting K_CO2 as well improve the fit?

```
> dicksonfit2 <- TAfit(aquaenv(Tc = 25, S = 35, SumBOH3 = SumBOH3,
+ SumH2SO4 = SumH2SO4, SumHF = SumHF), sam, conc_titrant, mass_sample,</pre>
```

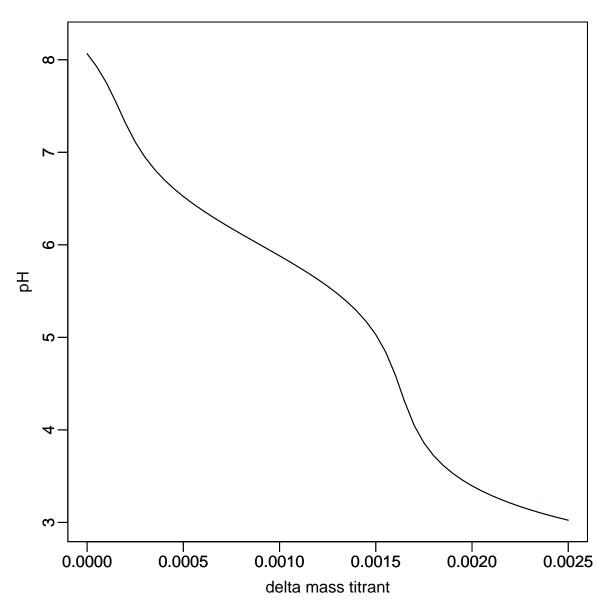
```
S_titrant = S_titrant, debug = TRUE, K_CO2fit = TRUE)
> dicksonfit2
$TA
[1] 0.002458081
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002194006
attr(,"unit")
[1] "mol/kg-soln"
$K_C02
[1] 1.030960e-06
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
$sumofsquares
```

Yes it does, but it is not optimal yet.

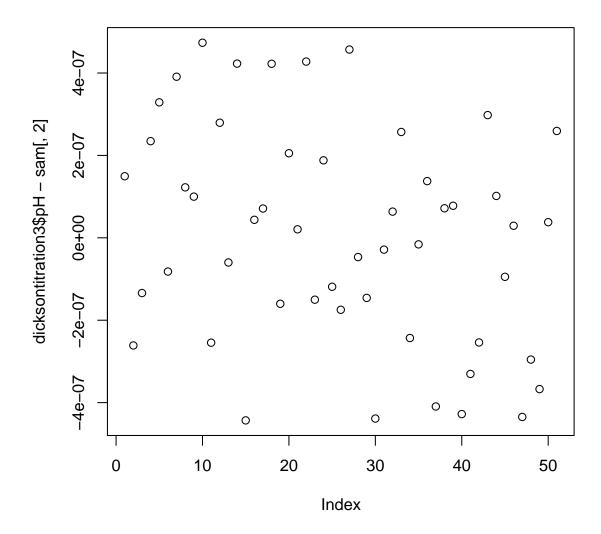
[1] 0.005724457

There still remains one major difference between the calculations carried out in Dickson (1981) and the calculations in **AquaEnv**: Dickson (1981) uses fixed values for the equilibrium constants and does not calculate them as functions of temperature and salinity. Furthermore, the values that are used in Dickson (1981) are not exactly the same as are obtained in **AquaEnv** for the same salinity and temperature.

Let us calculate a theoretical titration curve employing exactly the same equilibrium constant values as used in Dickson (1981) and plot the result together with the "Dickson" curve



Plotting the differences between both curves reveals that they are the same down to 1 umol/kg-soln.



Calculating [TA] and $[\sum CO_2]$ using TAfit and exactly the same equilibrium constant values as used in Dickson (1981)

```
> dicksonfit3 <- TAfit(aquaenv(Tc = 25, S = 35, SumBOH3 = SumBOH3, 
+ SumH2SO4 = SumH2SO4, SumHF = SumHF, k_w = 4.32e-14, k_co2 = 1e-06, 
+ k_co3 = 8.2e-10, k_co3 = 1.78e-09, k_co4 = (1/12.3), k_co5 = (1/408)), 
+ sam, conc_titrant, mass_sample, S_titrant = S_titrant, debug = TRUE, 
+ k_cw = 4.32e-14, k_co2 = 1e-06, k_co3 = 8.2e-10, k_co3 = 1.78e-09, 
+ k_co4 = (1/12.3), k_c6 = (1/408)) 
> dicksonfit3
```

\$TA [1] 0.00245 attr(,"unit")

```
[1] "mol/kg-soln"
```

\$SumCO2 [1] 0.0022 attr(,"unit") [1] "mol/kg-soln"

\$sumofsquares

[1] 3.280551e-12

reveals that now exactly the same values are calculated as are given in Dickson (1981).

3.5.2.3 Real data

3.5.2.3.1 sample1

The dataset sample1 is a titration curve for TA determination, obtained in the lab of the NIOO-CEME, Yerseke, The Netherlands.

Metadata of the sample which is:

x-value	volume of titrant added (in ml) with a Metrohm Omega 665 Dosimat
y-value	pH measured on the NBS scale with a Metrohm Omega 714 pH Meter (Buffer pH 4: citric acid, sodium hydroxide, chlor hydrogen; Buffer pH 7: di-sodium hydrogen phosphate, potassium di-hydrogen phosphate)
Tc	23.5 °C
S	34
volume of the sample	10 ml
titrant type	HCl
titrant concrntration	0.01 N = 0.01 M

We assume that the salinity of the titrant solution is 0.5 (the salinity of a solution with a ionic strength of 0.01 mol/kg-solution (0.01N) according to: I = (19.924 S) / (1000 - 1.005 S)). Furthermore, the the x values have to be converted to kg titrant, assuming the density of the titrant solution is aequivalent to the density of seawater with S=0.5, and last but not least 1 ml = 0.000001 m3.

```
> sample <- cbind((sample1[, 1] * 1e-06 * aquaenv(Tc = 23.5, S = 0.5)$density), + sample1[, 2])
```

We reduce the amount of datapoints: for runtime reasons

```
> sample <- sample[seq(1, length(sample[, 1]), 3), ]</pre>
```

We convert the concentration of the titrant from molarity (mol/l) to mol/kg-soln

```
> conc\_titrant <- convert(0.01, "conc", "molar2molin", Tc = 23.5, + S = 0.5)
```

We calculate the mass of the sample in kg (and 10 ml = 0.00001 m3)

```
> mass_sample <- 1e-05 * aquaenv(Tc = 23.5, S = 34)$density
```

No the fitting procedure can be performed. Note that the datapoints are not equally spaced and the pH scale of the measured pH values is the NBS scale.

Note that AquaEnv uses the Davies equation to calculate the activity coefficient for the proton upon conversion from/to NBS scale. The Davies equation is only valid up to a ionic strength (I) of 0.5 (Zeebe and Wolf-Gladrow 2001), that is around S=24.48. To accurately determine [TA] for samples with higher salinity using the function TAfit, one needs to provide either pH values on the free, total or seawater scale or E (V) values.

The [TA] value obtained with the Peene-Method was 0.0058740 mol/l, i.e.

```
> convert(0.005874, "conc", "molar2molin", Tc = 23.5, S = 34)
[1] 0.005724596
attr(,"unit")
[1] "mol/kg-soln"
```

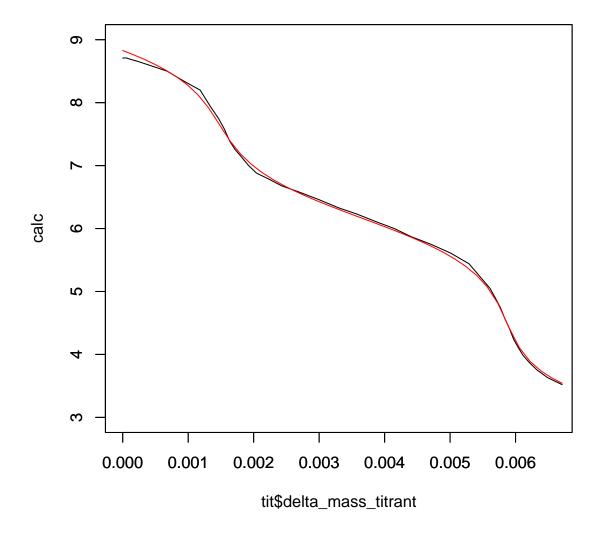
the difference in umol/kg-soln is

attr(,"unit")
[1] "mol/kg-soln"

```
> (0.005724596 - sample1_fit$TA) * 1e+06
[1] 12.68628
attr(,"unit")
[1] "mol/kg-soln"
and the SumCO2 value was 0.0522 mgC/l, i.e.
> convert((0.0522/12), "conc", "molar2molin", Tc = 23.5, S = 34)
[1] 0.004239359
attr(,"unit")
[1] "mol/kg-soln"
the difference in umol/kg-soln is
> (0.004239359 - sample1_fit$SumCO2) * 1e+06
[1] -39.35063
```

Note the deviation of the measured curve from the simulated curve, especially at high pH's (and high salinities) as the titration proceeds, salinity drops due to dilution with non saline titrant (HCl solution) and the agreement between the curves gets better

```
> plot(sample[, 1], sample[, 2], xlim = c(0, 0.0066), ylim = c(3,
+ 9), type = "l", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit$delta_mass_titrant, calc, xlim = c(0, 0.0066), ylim = c(3,
+ 9), type = "l", col = "red")
```



this shows that NBS pH values with initial salinities of more than 25 are not excellent input values for TAfit.

3.5.2.3.2 sample1 to sample3

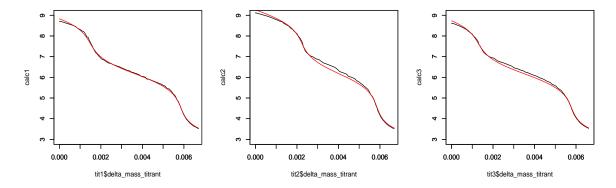
The datasets sample2 and sample3 are similar to sample1 and their metadata can be found in their helpfiles.

Preparing the input for TAfit

```
> sam1 <- cbind((sample1[, 1] * 1e-06 * aquaenv(Tc = 23.5, S = 0.5)$density),  
+ sample1[, 2])
```

```
> sam2 < - cbind((sample2[, 1] * 1e-06 * aquaenv(Tc = 22.9, S = 0.5) density),
      sample2[, 2])
> sam3 < - cbind((sample3[, 1] * 1e-06 * aquaenv(Tc = 23.3, S = 0.5)$density),
      sample3[, 2])
> conc_titrant1 <- convert(0.01, "conc", "molar2molin", Tc = 23.5,</pre>
      S = 0.5)
> conc_titrant2 <- convert(0.01, "conc", "molar2molin", Tc = 22.9,</pre>
      S = 0.5
> conc_titrant3 <- convert(0.01, "conc", "molar2molin", Tc = 23.3,</pre>
      S = 0.5
> mass_sample1 <- 1e-05 * aquaenv(Tc = 23.5, S = 34)$density
> mass_sample2 <- 1e-05 * aquaenv(Tc = 22.9, S = 33.4)$density
> mass_sample3 <- 1e-05 * aquaenv(Tc = 23.3, S = 33.8)$density
Performing the fitting
> sample1_fit <- TAfit(aquaenv(Tc = 23.5, S = 34), sam1, conc_titrant1,
      mass_sample1, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> tit1 <- tit
> calc1 <- calc
> sample2_fit <- TAfit(aquaenv(Tc = 22.9, S = 33.4), sam2, conc_titrant2,
      mass_sample2, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> tit2 <- tit
> calc2 <- calc
> sample3_fit <- TAfit(aquaenv(Tc = 23.3, S = 33.8), sam3, conc_titrant3,
      mass_sample3, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> tit3 <- tit
> calc3 <- calc
Visualizing the final fits
> par(mfrow = c(1, 3))
> plot(sam1[, 1], sam1[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
      type = "1", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit1$delta_mass_titrant, calc1, xlim = c(0, 0.0066), ylim = c(3,
      9), type = "1", col = "red")
> plot(sam2[, 1], sam2[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
      type = "1", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit2$delta_mass_titrant, calc2, xlim = c(0, 0.0066), ylim = c(3, 0.0066))
      9), type = "l", col = "red")
> plot(sam3[, 1], sam3[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
     type = "l", xlab = "", ylab = "")
```

```
> par(new = TRUE)
> plot(tit3$delta_mass_titrant, calc3, xlim = c(0, 0.0066), ylim = c(3,
+ 9), type = "1", col = "red")
```



Comparing the resulting values to the values obtained with the graphical method (J. Peene)

```
> TA1peene <- convert(0.005874, "conc", "molar2molin", Tc = 23.5,
+ S = 34)
> TA2peene <- convert(0.005856, "conc", "molar2molin", Tc = 22.9,
+ S = 33.4)
> TA3peene <- convert(0.005794, "conc", "molar2molin", Tc = 23.3,
+ S = 33.8)
> SumC021peene <- convert(0.0522/12, "conc", "molar2molin", Tc = 23.5,
+ S = 34)
> SumC022peene <- convert(0.043488/12, "conc", "molar2molin", Tc = 22.9,
+ S = 33.4)
> SumC023peene <- convert(0.05376/12, "conc", "molar2molin", Tc = 23.3,
+ S = 33.8)
> c(TA1peene, sample1_fit$TA)
```

[1] 0.005724596 0.005711035

> c(TA2peene, sample2_fit\$TA)

[1] 0.005709335 0.005706368

> c(TA3peene, sample3_fit\$TA)

[1] 0.005647382 0.005641810

> c(SumCO21peene, sample1_fit\$SumCO2)

[1] 0.004239359 0.004274075

```
> c(SumCO22peene, sample2_fit$SumCO2)
[1] 0.003533236 0.003541130
> c(SumCO23peene, sample3_fit$SumCO2)
[1] 0.004366633 0.004396890
```

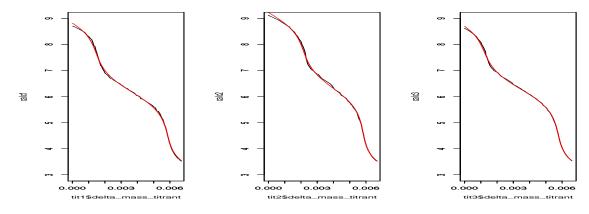
3.5.2.3.3 sample1 to sample3: fitting K_CO2 as well

Performing the fitting. Note that, with fitting K_CO2 as well, it might be necessary to choose a small initial stepwidth-factor and/or bring the parameter TASumCO2guess close to expected values for [TA] and $[\sum CO_2]$, otherwise the optimization routine might create an error, e.g. by "trying" negative K_CO2 values.

```
> sample1_fit <- TAfit(aquaenv(Tc = 23.5, S = 34), sam1, conc_titrant1,
      mass_sample1, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, K_CO2fit = TRUE, nlscontrol = nls.lm.control(factor = 0.1),
      verbose = TRUE)
> tit1 <- tit
> calc1 <- calc
> sample2_fit <- TAfit(aquaenv(Tc = 22.9, S = 33.4), sam2, conc_titrant2,
      mass_sample2, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, K_CO2fit = TRUE, TASumCO2guess = 0.006, nlscontrol = nls.lm.control(fa
      verbose = TRUE)
> tit2 <- tit
> calc2 <- calc
> sample3_fit <- TAfit(aquaenv(Tc = 23.3, S = 33.8), sam3, conc_titrant3,
      mass_sample3, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, K_CO2fit = TRUE, nlscontrol = nls.lm.control(factor = 0.1),
      verbose = TRUE)
> tit3 <- tit
> calc3 <- calc
Visualizing the resulting fits
> par(mfrow = c(1, 3))
> plot(sam1[, 1], sam1[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
      type = "1", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit1$delta_mass_titrant, calc1, xlim = c(0, 0.0066), ylim = c(3, 0.0066))
      9), type = "l", col = "red")
> plot(sam2[, 1], sam2[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
      type = "1", xlab = "", ylab = "")
> par(new = TRUE)
```

> plot(tit2\$delta_mass_titrant, calc2, xlim = c(0, 0.0066), ylim = c(3,

```
+ 9), type = "l", col = "red")
> plot(sam3[, 1], sam3[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
+ type = "l", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit3$delta_mass_titrant, calc3, xlim = c(0, 0.0066), ylim = c(3, 9), type = "l", col = "red")
```



Comparing the resulting values to the values obtained with the graphical method (J. Peene)

```
> TA1peene <- convert(0.005874, "conc", "molar2molin", Tc = 23.5,
+ S = 34)
> TA2peene <- convert(0.005856, "conc", "molar2molin", Tc = 22.9,
+ S = 33.4)
> TA3peene <- convert(0.005794, "conc", "molar2molin", Tc = 23.3,
+ S = 33.8)
> SumC021peene <- convert(0.0522/12, "conc", "molar2molin", Tc = 23.5,
+ S = 34)
> SumC022peene <- convert(0.043488/12, "conc", "molar2molin", Tc = 22.9,
+ S = 33.4)
> SumC023peene <- convert(0.05376/12, "conc", "molar2molin", Tc = 23.3,
+ S = 33.8)
> c(TA1peene, sample1_fit$TA)
```

[1] 0.005724596 0.005708382

> c(TA2peene, sample2_fit\$TA)

[1] 0.005709335 0.005665999

> c(TA3peene, sample3_fit\$TA)

[1] 0.005647382 0.005609916

> c(SumCO21peene, sample1_fit\$SumCO2)

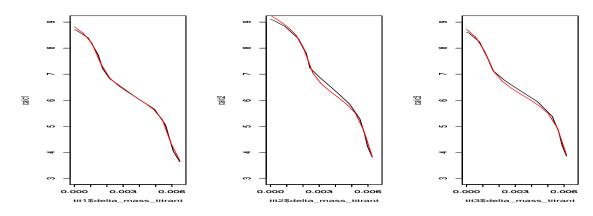
```
[1] 0.004239359 0.004285634
> c(SumCO22peene, sample2_fit$SumCO2)
[1] 0.003533236 0.003566389
> c(SumCO23peene, sample3_fit$SumCO2)
[1] 0.004366633 0.004431569
3.5.2.3.4 sample1 to sample3: fitting \approx 10 datapoints
To test if the fit wil be significantly worse if the amount of datapoints in the titration curve
is reduced, we reduce the amount of datapoints in the curve to around 10.
> sam1 < -cbind((sample1[, 1] * 1e-06 * aquaenv(Tc = 23.5, S = 0.5) * density),
      sample1[, 2])[seq(1, length(sample1[, 1]), 10), ]
> sam2 < - cbind((sample2[, 1] * 1e-06 * aquaenv(Tc = 22.9, S = 0.5)$density),
      sample2[, 2])[seq(1, length(sample2[, 1]), 10), ]
> sam3 < - cbind((sample3[, 1] * 1e-06 * aquaenv(Tc = 23.3, S = 0.5)$density),
      sample3[, 2])[seq(1, length(sample3[, 1]), 10), ]
Performing the fitting
> sample1_fit <- TAfit(aquaenv(Tc = 23.5, S = 34), sam1, conc_titrant1,
      mass_sample1, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> tit1 <- tit
> calc1 <- calc
> sample2_fit <- TAfit(aquaenv(Tc = 22.9, S = 33.4), sam2, conc_titrant2,
      mass_sample2, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> tit2 <- tit
> calc2 <- calc
> sample3_fit <- TAfit(aquaenv(Tc = 23.3, S = 33.8), sam3, conc_titrant3,
      mass_sample3, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> tit3 <- tit
> calc3 <- calc
Visualizing the resulting fit
> par(mfrow = c(1, 3))
```

> plot(sam1[, 1], sam1[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),

type = "l", xlab = "", ylab = "")

> par(new = TRUE)

```
> plot(tit1$delta_mass_titrant, calc1, xlim = c(0, 0.0066), ylim = c(3,
+ 9), type = "l", col = "red")
> plot(sam2[, 1], sam2[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
+ type = "l", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit2$delta_mass_titrant, calc2, xlim = c(0, 0.0066), ylim = c(3,
+ 9), type = "l", col = "red")
> plot(sam3[, 1], sam3[, 2], xlim = c(0, 0.0066), ylim = c(3, 9),
+ type = "l", xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit3$delta_mass_titrant, calc3, xlim = c(0, 0.0066), ylim = c(3,
+ 9), type = "l", col = "red")
```



Comparing the resulting values to the values obtained with the graphical method (J. Peene)

```
> TA1peene <- convert(0.005874, "conc", "molar2molin", Tc = 23.5,
+ S = 34)
> TA2peene <- convert(0.005856, "conc", "molar2molin", Tc = 22.9,
+ S = 33.4)
> TA3peene <- convert(0.005794, "conc", "molar2molin", Tc = 23.3,
+ S = 33.8)
> SumC021peene <- convert(0.0522/12, "conc", "molar2molin", Tc = 23.5,
+ S = 34)
> SumC022peene <- convert(0.043488/12, "conc", "molar2molin", Tc = 22.9,
+ S = 33.4)
> SumC023peene <- convert(0.05376/12, "conc", "molar2molin", Tc = 23.3,
+ S = 33.8)
> c(TA1peene, sample1_fit$TA)
[1] 0.005724596 0.005721581
```

[1] 0.005709335 0.005702284

> c(TA2peene, sample2_fit\$TA)

```
> c(TA3peene, sample3_fit$TA)

[1] 0.005647382 0.005648471

> c(SumCO21peene, sample1_fit$SumCO2)

[1] 0.004239359 0.004296768

> c(SumCO22peene, sample2_fit$SumCO2)

[1] 0.003533236 0.003552380

> c(SumCO23peene, sample3_fit$SumCO2)
```

[1] 0.004366633 0.004429815

That means using just ≈ 10 datapoints is feasible, as the fits do not get worse significantly

3.5.2.3.5 sample4_1 to sample4_2: standard seawater

sample4_1 to sample4_4 are titration curves of reference material (standard seawater) "Batch 82" (A. Dickson) for TA determination, obtained in the lab of the NIOO-CEME, Yerseke, The Netherlands. Metadata can again be obtained from the helpfiles of the datasets.

We assume the salinity of the titrant solution is 0.5 (the salinity of a solution with a ionic strength of 0.01 mol/kg-solution (0.01N) according to: I = (19.924 S) / (1000 - 1.005 S). We convert the x values to kg titrant, and assume the density of the titrant solution is aequivalent to the density of seawater with S=0.5. Finally, 1 ml = 0.000001 m³.

```
> sam4_1 \leftarrow cbind((sample4_1[, 1] * 1e-06 * aquaenv(Tc = 21, S = 0.5)$density), + sample4_1[, 2])
```

We convert the concentration of the titrant from molarity (mol/l) to mol/kg-soln

```
> conc_titrant <- convert(0.01, "conc", "molar2molin", Tc = 21, S = 0.5)
```

We calculate the mass of the sample in kg (10 ml = 0.00001 m3)

```
> mass_sample <- 1e-05 * aquaenv(Tc = 21, S = 35.356)$density
```

We perform the fitting procedure: note that the datapoints are NOT equally spaced and the pH scale of the measured pH values is the NBS scale, (debug=TRUE toggles the debug mode: the last simulated titration tit and the converted pH profile calc are made global variables for investigation and plotting)

```
> sample4_1_fit <- TAfit(aquaenv(Tc = 21, S = 34.356, SumH3P04 = 3e-07, SumSiOH4 = 2e-06, SumHNO2 = 1e-08, SumHNO3 = 8.3e-07), sam4_1, conc_titrant, mass_sample, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs", debug = TRUE) > sample4_1_fit
```

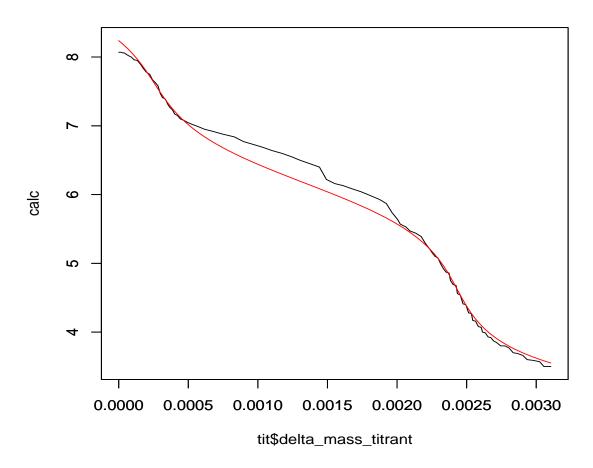
```
$TA
[1] 0.002369027
attr(,"unit")
[1] "mol/kg-soln"

$SumCO2
[1] 0.002122602
attr(,"unit")
[1] "mol/kg-soln"

$sumofsquares
```

[1] 1.484307

The fit can be visualized



(Note that the fit is very good on the steep slopes and not so good inbetween. This is because the sample titration curve is not equally spaced. There are smaller titration steps in the steep parts, resulting in more points there. Since **AquaEnv** minimizes the sum of squares of all points without weighting the points according to their density in a certain region of the curve, the regions with higher point density have implicitly a higher weight during the fitting procedure.)

and the results can be compared to results obtained with the graphical method (J.Peene)

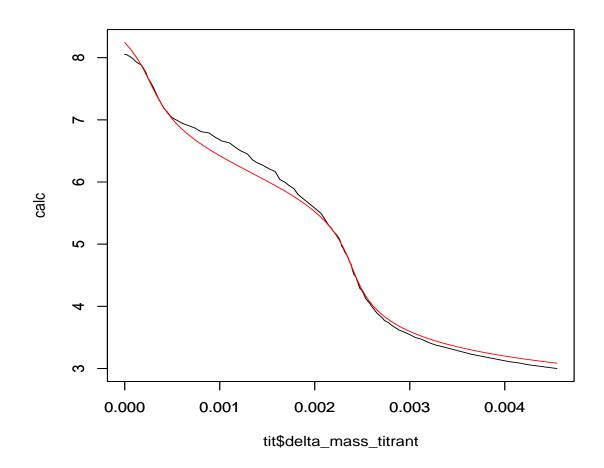
```
> peeneTA <- convert(0.002418, "conc", "molar2molin", Tc = 21,
+ S = 35.356)
> peeneSumC02 <- convert(0.025872/12, "conc", "molar2molin", Tc = 21,
+ S = 35.356)
> c(peeneTA, sample4_1_fit$TA)

[1] 0.002353904 0.002369027
> c(peeneSumC02, sample4_1_fit$SumC02)

[1] 0.002098849 0.002122602
```

```
sample4_2 is a replica titration of sample4_1.
Preparations
> sam4_2 <- cbind((sample4_2[, 1] * 1e-06 * aquaenv(Tc = 22.8,
      S = 0.5)$density), sample4_2[, 2])
> conc_titrant <- convert(0.01, "conc", "molar2molin", Tc = 22.8,</pre>
     S = 0.5
> mass_sample <- 1e-05 * aquaenv(Tc = 22.8, S = 35.356)$density
Fitting
> sample4_2_fit \leftarrow TAfit(aquaenv(Tc = 22.8, S = 34.356, SumH3P04 = 3e-07,
      SumSiOH4 = 2e-06, SumHNO2 = 1e-08, SumHNO3 = 8.3e-07), sam4_2,
      conc_titrant, mass_sample, S_titrant = 0.5, equalspaced = FALSE,
      pHscale = "nbs", debug = TRUE)
> sample4_2_fit
$TA
[1] 0.002331524
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002073307
attr(,"unit")
[1] "mol/kg-soln"
$sumofsquares
[1] 1.316187
Visualisation
> xlim <- range(sam4_2[, 1], tit$delta_mass_titrant)</pre>
> ylim <- range(sam4_2[, 2], calc)</pre>
> plot(sam4_2[, 1], sam4_2[, 2], xlim = xlim, ylim = ylim, type = "l",
      xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,
```

type = "1", col = "red")



Comparison to graphical method

```
> peeneTA <- convert(0.00239, "conc", "molar2molin", Tc = 22.8,
+    S = 35.356)
> peeneSumC02 <- convert(0.02532/12, "conc", "molar2molin", Tc = 22.8,
+    S = 35.356)
> peeneCalcSumC02 <- convert(0.02625722/12, "conc", "molar2molin",
+    Tc = 22.8, S = 35.356)
> c(peeneTA, sample4_2_fit$TA)
```

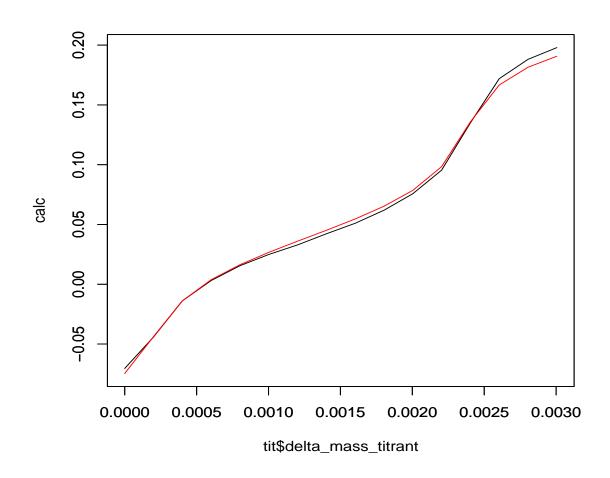
- [1] 0.002326871 0.002331524
- > c(peeneSumCO2, peeneCalcSumCO2, sample4_2_fit\$SumCO2)
- [1] 0.002054267 0.002130305 0.002073307

3.5.2.3.6 sample4_3 to sample4_4: E curves

The datasets sample4_3 and sample4_4 have been obtained by manually titrating samples of the standard seawater mentioned above and recording the "raw" potential (E) values coming from the electrode.

Preparations

```
> sam4_3 < - cbind((sample4_3[, 1] * 1e-06 * aquaenv(Tc = 21, S = 0.5)$density),
      sample4_3[, 2]/1000)
> conc_titrant <- convert(0.01, "conc", "molar2molin", Tc = 21,</pre>
      S = 0.5)
> mass_sample <- 1e-05 * aquaenv(Tc = 21, S = 0)$density
Fitting
> sample4_3_fit \leftarrow TAfit(aquaenv(Tc = 21, S = 34.356, SumH3P04 = 3e-07,
      SumSiOH4 = 2e-06, SumHNO2 = 1e-08, SumHNO3 = 8.3e-07), sam4_3,
      conc_titrant, mass_sample, S_titrant = 0.5, Evals = TRUE,
      verbose = TRUE, electrode_polarity = "neg", debug = TRUE)
> sample4_3_fit
$TA
[1] 0.002375396
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002184827
attr(,"unit")
[1] "mol/kg-soln"
$E0
[1] -0.3842297
attr(,"unit")
[1] "V"
$sumofsquares
[1] 0.0002054284
Visualizing
> xlim <- range(sam4_3[, 1], tit$delta_mass_titrant)</pre>
> ylim <- range(sam4_3[, 2], calc)
> plot(sam4_3[, 1], sam4_3[, 2], xlim = xlim, ylim = ylim, type = "1",
      xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,
      type = "1", col = "red")
```



Fitting $K_{-}CO2$ as well

```
$E0

[1] -0.3917266

attr(,"unit")

[1] "V"

$K_C02

[1] 7.023512e-07

attr(,"unit")

[1] "mol/kg-soln"

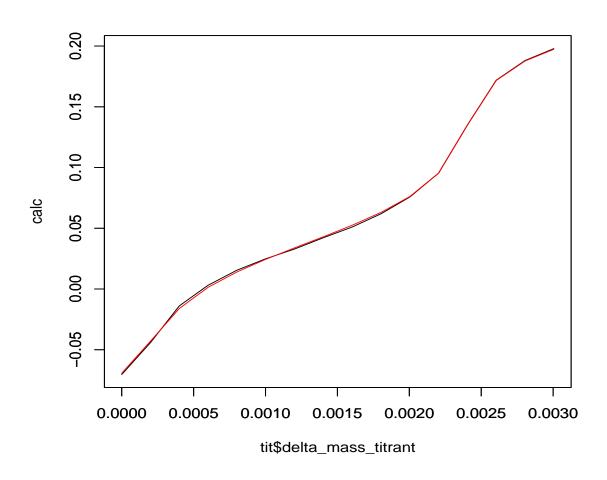
attr(,"pH scale")

[1] "free"

$sumofsquares
```

[1] 1.787927e-05

Visualizing



Fitting with the K values from Dickson (1981)

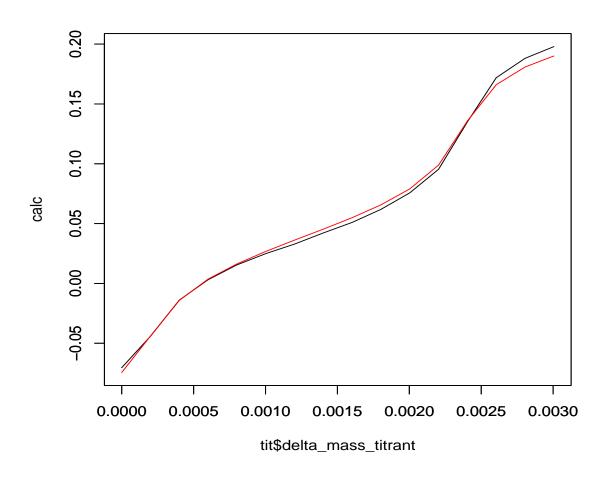
[1] 0.002181591

```
attr(,"unit")
[1] "mol/kg-soln"

$E0
[1] -0.3845453
attr(,"unit")
[1] "V"

$sumofsquares
[1] 0.000243846
```

Visualizing

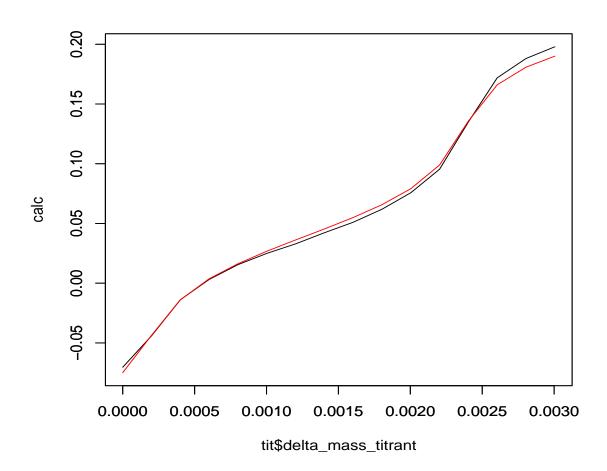


Fitting with titrant salinity assumed equal to sample salinity

> plot(tit\$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,

> par(new = TRUE)

type = "1", col = "red")



Fitting with titrant concentration assumed to be exactly 0.01 mol/kg-soln

```
> sample4_3_fit_4 \leftarrow TAfit(aquaenv(Tc = 21, S = 34.356, SumH3P04 = 3e-07, SumSiOH4 = 2e-06, SumHNO2 = 1e-08, SumHNO3 = 8.3e-07), sam4_3, conc_titrant = 0.01, mass_sample, S_titrant = 0.5, Evals = TRUE, verbose = TRUE, electrode_polarity = "neg", debug = TRUE) > <math>sample4_3_fit_4
```

```
$TA
[1] 0.002379342
attr(,"unit")
[1] "mol/kg-soln"

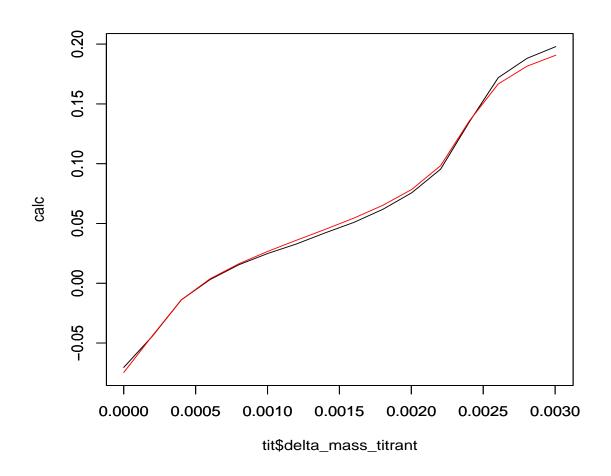
$SumCO2
[1] 0.002188621
attr(,"unit")
[1] "mol/kg-soln"

$EO
[1] -0.3842187
attr(,"unit")
[1] "V"

$sumofsquares
```

[1] 0.0002040564

Visualizing



Preparations for sample4_4

```
> sam4_4 \leftarrow cbind((sample4_4[, 1] * 1e-06 * aquaenv(Tc = 21, S = 0.5)$density), + sample4_4[, 2]/1000) > conc_titrant \leftarrow convert(0.01, "conc", "molar2molin", Tc = 21, + S = 0.5) > mass_sample \leftarrow 1e-05 * aquaenv(Tc = 21, S = 0)$density
```

Fitting

```
> sample4_4_fit <- TAfit(aquaenv(Tc = 22.4, S = 34.356, SumH3P04 = 3e-07,
+ SumSiOH4 = 2e-06, SumHNO2 = 1e-08, SumHNO3 = 8.3e-07), sam4_4,
+ conc_titrant, mass_sample, S_titrant = 0.5, Evals = TRUE,
+ verbose = TRUE, electrode_polarity = "neg", debug = TRUE)
> sample4_4_fit
```

\$TA

[1] 0.002392435

```
attr(,"unit")
[1] "mol/kg-soln"

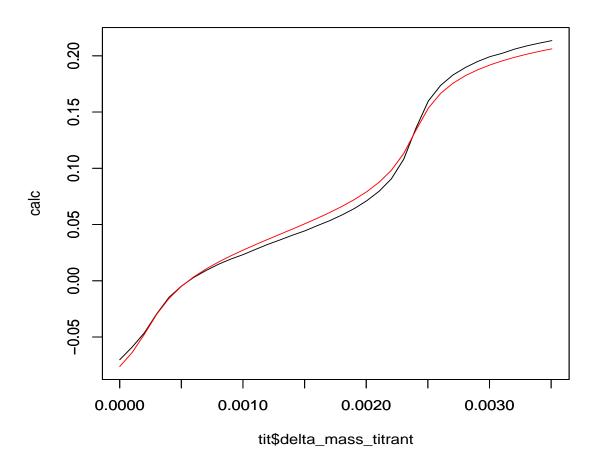
$SumCO2
[1] 0.002171694
attr(,"unit")
[1] "mol/kg-soln"

$EO
[1] -0.3870534
attr(,"unit")
[1] "V"

$sumofsquares
[1] 0.001223981
```

Visualizing

```
> xlim <- range(sam4_4[, 1], tit$delta_mass_titrant)
> ylim <- range(sam4_4[, 2], calc)
> plot(sam4_4[, 1], sam4_4[, 2], xlim = xlim, ylim = ylim, type = "l",
+ xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,
+ type = "l", col = "red")
```



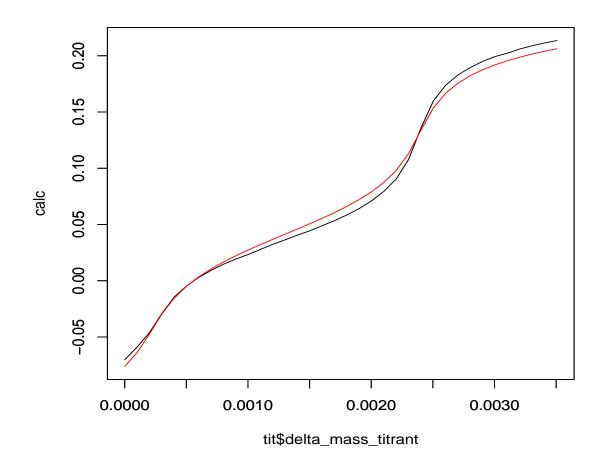
Fitting with concentrations of summed quantities calculated from salinity

```
attr(,"unit")
[1] "V"
```

\$sumofsquares

[1] 0.001224115

Visualizing

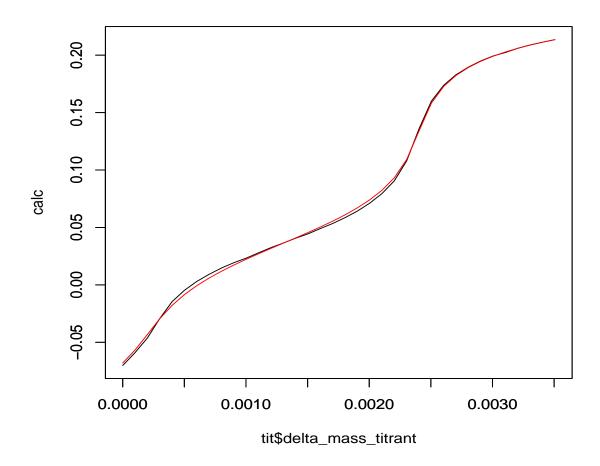


Fitting K_CO2 as well

```
> sample4_4_fit_2 <- TAfit(aquaenv(Tc = 22.4, S = 34.356, SumH3P04 = 3e-07, SumSiOH4 = 2e-06, SumHNO2 = 1e-08, SumHNO3 = 8.3e-07), sam4_4,
```

```
conc_titrant, mass_sample, S_titrant = 0.5, Evals = TRUE,
      verbose = TRUE, electrode_polarity = "neg", K_CO2fit = TRUE,
      debug = TRUE)
> sample4_4_fit_2
$TA
[1] 0.002398211
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.002196573
attr(,"unit")
[1] "mol/kg-soln"
$E0
[1] -0.3945504
attr(,"unit")
[1] "V"
$K_C02
[1] 6.128419e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
$sumofsquares
[1] 0.0001367050
```

Visualizing



3.5.2.3.7 Some rudimentary statistics on sample4_1 to sample4_4

(Beware, these statistics are just "back of the envelope" calculations, since not all the measurements are independent and for different quantities not the same number of measurements have been carried out.)

The graphical method

```
> peeneTA <- c(2353.904, 2326.871)
> peeneDIC <- c(2098.849, 2054.267, 2130.305)
> paste(mean(peeneTA), "+/-", sd(peeneTA))

[1] "2340.3875 +/- 19.1152176158159"

> paste(mean(peeneDIC), "+/-", sd(peeneDIC))

[1] "2094.473666666667 +/- 38.2073555396515"
```

```
pH curve fitting
```

```
> pHTA <- c(sample4_1_fit$TA, sample4_2_fit$TA) * 1e+06
> pHDIC <- c(sample4_1_fit$SumCO2, sample4_2_fit$SumCO2) * 1e+06
> paste(mean(pHTA), "+/-", sd(pHTA))
[1] "2350.27560172219 +/- 26.5186027797206"
> paste(mean(pHDIC), "+/-", sd(pHDIC))
[1] "2097.95465823617 +/- 34.8571361424180"
E curve fitting
> ETA <- c(sample4_3_fit$TA, sample4_3_fit_1$TA, sample4_3_fit_2$TA,
      sample 4\_3\_fit\_3\$TA, \ sample 4\_3\_fit\_4\$TA, \ sample 4\_4\_fit\$TA,
      sample4_4_fit_1$TA, sample4_4_fit_2$TA) * 1e+06
> EDIC <- c(sample4_3_fit$SumCO2, sample4_3_fit_1$SumCO2, sample4_3_fit_2$SumCO2,
      sample4_3_fit_3$SumCO2, sample4_3_fit_4$SumCO2, sample4_4_fit$SumCO2,
      sample 4\_4\_fit\_1\$SumCO2, \ sample 4\_4\_fit\_2\$SumCO2) \ * \ 1e+06
> paste(mean(ETA), "+/-", sd(ETA))
[1] "2385.30719857401 +/- 9.94955717122274"
> paste(mean(EDIC), "+/-", sd(EDIC))
[1] "2184.56596866365 +/- 9.09620796459795"
pH and E curve fitting combined
> totalTA <- c(pHTA, ETA)
> totalDIC <- c(pHDIC, EDIC)</pre>
> paste(mean(totalTA), "+/-", sd(totalTA))
[1] "2378.30087920365 +/- 19.3210666645222"
> paste(mean(totalDIC), "+/-", sd(totalDIC))
[1] "2167.24370657816 +/- 39.1530295205609"
```

3.5.2.3.8 sample5_1 to sample5_2: freshwater samples

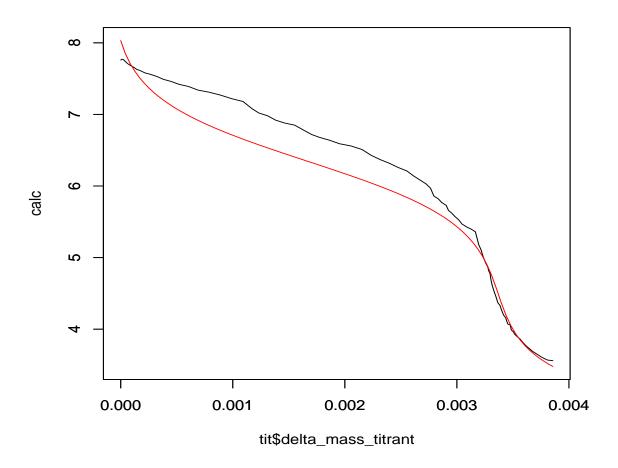
sample5_1 to sample5_3 are titration curves of freshwater samples obtained in the lab of the NIOO-CEME, Yerseke, The Netherlands. sample5_1 and sample5_2 are pH curves.

Preparations

```
> sam5_1 < -cbind((sample5_1[, 1] * 1e-06 * aquaenv(Tc = 21, S = 0.5)$density),
      sample5_1[, 2])
> conc_titrant <- convert(0.01, "conc", "molar2molin", Tc = 21,</pre>
      S = 0.5
> mass_sample <- 1e-05 * aquaenv(Tc = 17.5, S = 0)$density
Fitting
> sample5_1_fit <- TAfit(aquaenv(Tc = 17.5, S = 0), sam5_1, conc_titrant,
      mass_sample, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",
      debug = TRUE, verbose = TRUE)
> sample5_1_fit
$TA
[1] 0.003362829
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.003426125
attr(,"unit")
[1] "mol/kg-soln"
$sumofsquares
[1] 6.008886
Visualizing
> xlim <- range(sam5_1[, 1], tit$delta_mass_titrant)</pre>
> ylim <- range(sam5_1[, 2], calc)</pre>
> plot(sam5_1[, 1], sam5_1[, 2], xlim = xlim, ylim = ylim, type = "l",
      xlab = "", ylab = "")
> par(new = TRUE)
```

> plot(tit\$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,

type = "1", col = "red")



Comparing to graphical method

```
> peeneTA <- convert(0.003314, "conc", "molar2molin", Tc = 17.5,
+ S = 0)
> c(peeneTA, sample5_1_fit$TA)
```

[1] 0.003310610 0.003362829

Preparations for sample5_2

```
> sam5_2 < -cbind((sample5_2[, 1] * 1e-06 * aquaenv(Tc = 21, S = 0.5)$density), + sample5_2[, 2])
> conc_titrant < -convert(0.01, "conc", "molar2molin", Tc = 21, + S = 0.5)
> mass_sample < -1e-05 * aquaenv(Tc = 18.3, S = 0)$density
Fitting
```

```
> sample5_2_fit <- TAfit(aquaenv(Tc = 18.3, S = 0), sam5_2, conc_titrant,
+ mass_sample, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",</pre>
```

```
+ debug = TRUE, verbose = TRUE)
> sample5_2_fit
```

```
$TA
[1] 0.003408187
attr(,"unit")
[1] "mol/kg-soln"

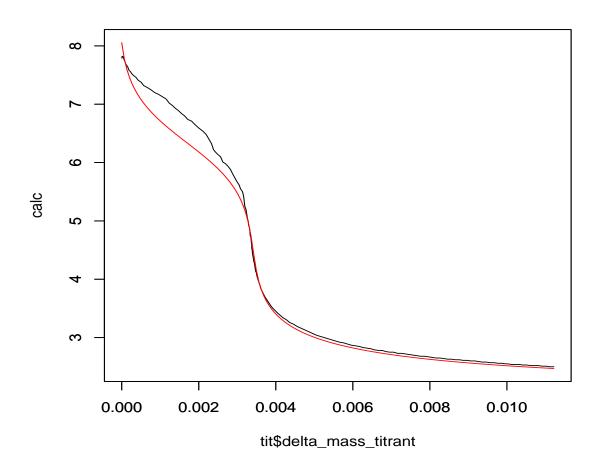
$SumCO2
[1] 0.003465661
attr(,"unit")
[1] "mol/kg-soln"

$sumofsquares
```

Visualizing

[1] 5.05407

```
> xlim <- range(sam5_2[, 1], tit$delta_mass_titrant)
> ylim <- range(sam5_2[, 2], calc)
> plot(sam5_2[, 1], sam5_2[, 2], xlim = xlim, ylim = ylim, type = "l",
+ xlab = "", ylab = "")
> par(new = TRUE)
> plot(tit$delta_mass_titrant, calc, xlim = xlim, ylim = ylim,
+ type = "l", col = "red")
```



Comparing to the graphical method

> sample5_2_fit_1

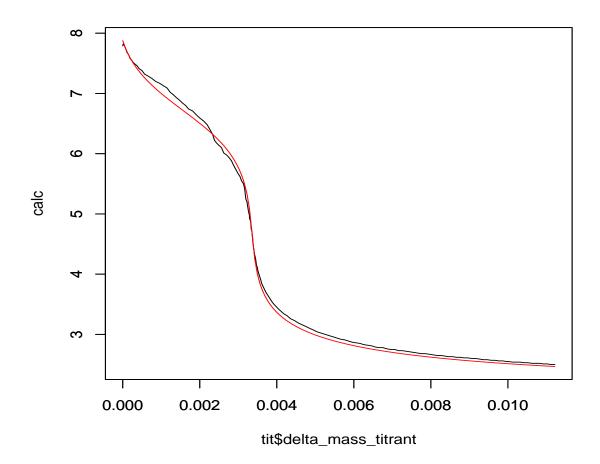
```
> peeneTA <- convert(0.00329, "conc", "molar2molin", Tc = 18.3,
+ S = 0)
> peeneSumCO2 <- convert(0.039647/12, "conc", "molar2molin", Tc = 18.3,
+ S = 0)
> c(peeneTA, sample5_2_fit$TA)
[1] 0.003286456 0.003408187
> c(peeneSumCO2, sample5_2_fit$SumCO2)
[1] 0.003300358 0.003465661
Fitting K_CO2 as well
> sample5_2_fit_1 <- TAfit(aquaenv(Tc = 18.3, S = 0), sam5_2, conc_titrant,</pre>
```

mass_sample, S_titrant = 0.5, equalspaced = FALSE, pHscale = "nbs",

verbose = TRUE, K_CO2fit = TRUE, debug = TRUE)

```
$TA
[1] 0.003351575
attr(,"unit")
[1] "mol/kg-soln"
$SumCO2
[1] 0.003559806
attr(,"unit")
[1] "mol/kg-soln"
$K_C02
[1] 2.022091e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"
$sumofsquares
[1] 0.9145475
```

Visualizing



3.5.2.3.9 sample5_3: E curve

The dataset sample5_3 has been obtained by manually titrating samples of the freshwater sample mentioned above and recording the "raw" potential (E) values coming from the electrode.

Preparations

```
> sam5_3 \leftarrow cbind((sample5_3[, 1] * 1e-06 * aquaenv(Tc = 21, S = 0.5)$density), + sample5_3[, 2]/1000)
> conc_titrant \leftarrow convert(0.01, "conc", "molar2molin", Tc = 21, + S = 0.5)
> mass_sample \leftarrow 1e-05 * aquaenv(Tc = 21, S = 0)$density
Fitting
```

```
> sample5_3_fit <- TAfit(aquaenv(Tc = 21, S = 0.1), sam5_3, conc_titrant,
+ mass_sample, S_titrant = 0.5, Evals = TRUE, verbose = TRUE,</pre>
```

```
+ electrode_polarity = "neg", debug = TRUE)
> sample5_3_fit
```

```
$TA
[1] 0.003360328
attr(,"unit")
[1] "mol/kg-soln"

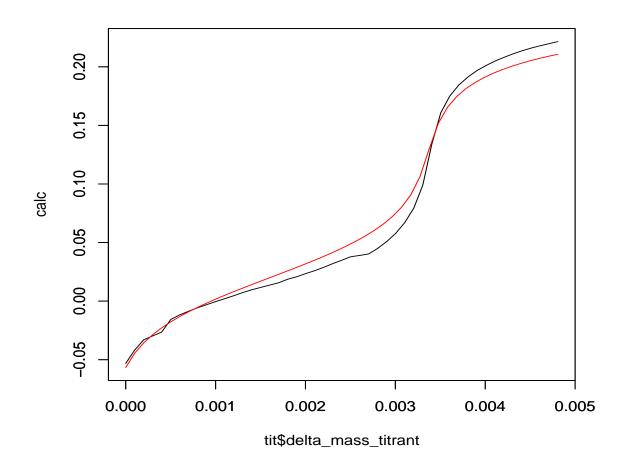
$SumCO2
[1] 0.003518269
attr(,"unit")
[1] "mol/kg-soln"

$EO
[1] -0.3865713
attr(,"unit")
[1] "V"

$sumofsquares
```

Visualizing

[1] 0.003469042



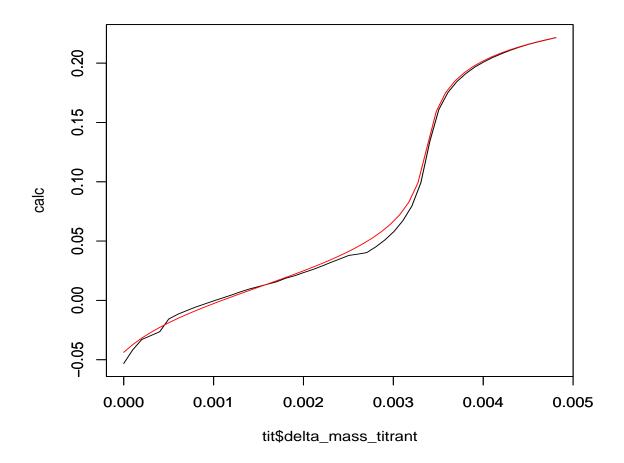
Fitting $K_{-}CO2$ as well

```
attr(,"unit")
[1] "V"

$K_C02
[1] 2.384818e-07
attr(,"unit")
[1] "mol/kg-soln"
attr(,"pH scale")
[1] "free"

$sumofsquares
[1] 0.0006593422
```

Visualizing



3.5.2.3.10 Some rudimentary statistics on sample5_1 to sample5_4 (Beware, these statistics are just "back of the envelope" calculations, since not all the measurements are independent and for different quantities not the same number of measurements have been carried out.)

The graphical method

```
> peeneTA <- c(3310.61, 3286.456)
> peeneDIC <- c(3300.358)
> paste(mean(peeneTA), "+/-", sd(peeneTA))

[1] "3298.533 +/- 17.0794571927799"
> paste(mean(peeneDIC), "+/-", sd(peeneDIC))

[1] "3300.358 +/- NA"
```

```
pH curve fitting
> pHTA <- c(sample5_1_fit$TA, sample5_2_fit$TA, sample5_2_fit_1$TA) *</pre>
> pHDIC <- c(sample5_1_fit$SumCO2, sample5_2_fit$SumCO2, sample5_2_fit_1$SumCO2) *
      1e+06
> paste(mean(pHTA), "+/-", sd(pHTA))
[1] "3374.19705722514 +/- 29.9695150180552"
> paste(mean(pHDIC), "+/-", sd(pHDIC))
[1] "3483.86396474187 +/- 68.6745830284585"
E curve fitting
> ETA <- c(sample5_3_fit$TA, sample5_3_fit_1$TA) * 1e+06
> EDIC <- c(sample5_3_fit$SumCO2, sample5_3_fit_1$SumCO2) * 1e+06
> paste(mean(ETA), "+/-", sd(ETA))
[1] "3363.51887482642 +/- 4.51327230361391"
> paste(mean(EDIC), "+/-", sd(EDIC))
[1] "3629.80333051158 +/- 157.733308339687"
pH and E curve fitting combined
> totalTA <- c(pHTA, ETA)</pre>
> totalDIC <- c(pHDIC, EDIC)</pre>
> paste(mean(totalTA), "+/-", sd(totalTA))
[1] "3369.92578426565 +/- 22.0994433353524"
> paste(mean(totalDIC), "+/-", sd(totalDIC))
[1] "3542.23971104975 +/- 122.341889234195"
```

4 Extending AquaEnv

It is very simple for the user to create own functions that use **AquaEnv** and extend its functionality. We will demonstrate that by creating simple analogons for the **AquaEnv** functions titration and TAfit.

The function simpletitration will take the following arguments

```
aquaenv an object of class aquaenv: minimal definition, contains all information about the system: T, S, d, total concentrations of nutrients etc

volume the volume of the (theoretical) titration vessel in l

amount the amount of titrant added in mol

steps the amount of titrant is added in

type the type of titrant: either "HCl" or "NaOH"
```

The function is defined as

```
> simpletitration <- function(aquaenv,
                                                          # an object of class aquaenv: minimal definition,
                                                          # contains all information about the system:
                                                          # T, S, d, total concentrations of nutrients etc
                                volume.
                                                          \mbox{\tt\#} the volume of the (theoretical) titration vessel in 1
                                                          \mbox{\it\#} the amount of titrant added in mol
                                amount,
                                                         # the amount of steps the amount of titrant is added in
                                steps,
                                                          # the type of titrant: either "HC1" or "NaOH"
                                type)
    {
      directionTAchange <- switch(type, HCl = -1, NaOH = +1)</pre>
      TAconcchangeperstep <- convert(((amount/steps)/volume), "conc", "molar2molin", aquaenv$Tc, aquaenv$S)
      aquaenvtemp <- aquaenv
      for (i in 1:steps)
                       <- aquaenvtemp$TA + (directionTAchange * TAconcchangeperstep)</pre>
          aquaenvtemp <- aquaenv(ae=aquaenvtemp, TA=TA)</pre>
                       <- merge(aquaenv, aquaenvtemp)</pre>
      aquaenv[["DeltaCTitrant"]] <- convert((amount/volume)/steps*(1:(steps+1)),
                                               "conc", "molar2molin", aquaenv$Tc, aquaenv$S)
      {\tt return(aquaenv)} \quad {\tt\#\ object\ of\ class\ aquaenv\ which\ contains\ a\ titration\ simulation}
```

and can be used to create a bjerrum plot

```
> ae <- simpletitration(aquaenv(Tc = 15, S = 35, SumCO2 = 0.0035,

+ SumNH4 = 2e-05, pH = 11.3), volume = 100, amount = 1.5, steps = 100,

+ type = "HCl")

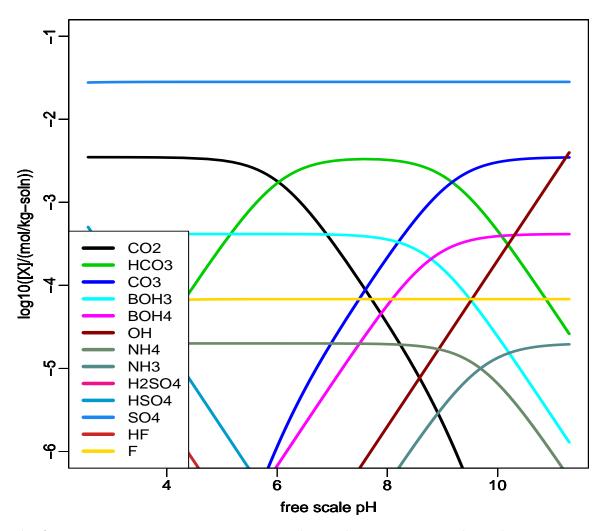
> what <- c("CO2", "HCO3", "CO3", "BOH3", "BOH4", "OH", "NH4",

+ "NH3", "H2SO4", "HSO4", "SO4", "HF", "F")

> plot(ae, what = what, bjerrum = TRUE, log = TRUE, ylim = c(-6,

+ -1), legendinset = 0, lwd = 3, palette = c(1, 3, 4, 5, 6,

+ colors()[seq(100, 250, 6)]), newdevice = FALSE)
```



The function simpletitration in turn can be used to create a simple analogon to TAfit with the arguments

ae an object of class aquaenv: minimal definition, contains all infor-

mation about the system: T, S, d, total concentrations of nutri-

ents etc

pHmeasurements a table containing the titration curve: basically a series of pH

values (pH on free proton scale)

volume the volume of the titration vessel
amount the total amount of the titrant added

TAguess=0.0025 a first guess for [TA] and [SumCO2] to be used as initial values

for the optimization procedure

type="HCl" the type of titrant: either "HCl" or "NaOH"

defined as

```
# basically a series of pH values (pH on free proton scale)
                    volume,
                                           # the volume of the titration vessel
                                           # the total amount of the titrant added
                     amount.
                     TAguess=0.0025,
                                           # a first guess for [TA] and [SumCO2] to be used as
                                           \mbox{\tt\#} initial values for the optimization procedure
                     type="HC1")
                                           # the type of titrant: either "HCl" or "NaOH"
ae$Na <- NULL  # make sure ae gets cloned as "skeleton": cloneaquaenv determines "skeleton"
                # TRUE or FALSE from the presence of a value for Na
residuals <- function(state)</pre>
    ae$SumCO2 <- state[[1]]</pre>
              <- simpletitration(aquaenv(ae=ae, TA=state[[2]]), volume=volume,</pre>
                                   {\tt amount=amount, steps=(length(pHmeasurements)-1), type=type)\$pH}
    residuals <- pHmeasurements-pHcalc
    return(residuals)
require(minpack.lm)
out <- nls.lm(fn=residuals, par=c(TAguess, TAguess)) #guess for TA is also used as guess for SumCO2
                           <- list(out$par[[2]], out$par[[1]], out$deviance)</pre>
attr(result[[1]], "unit") <- "mol/kg-soln"
attr(result[[2]], "unit") <- "mol/kg-soln"</pre>
names(result)
                          <- c("TA", "SumCO2", "sumofsquares")
return(result) # a list of three values
                # ([TA] in mol/kg-solution, [SumCO2] in mol/kg-solution, sum of the squared residuals)
```

The function simpleTAfit can be used to calculate TA and SumCO2

A Abbreviations for references used throughout the code and in the helpfiles

Atkins1996 Atkins (1996) Boudreau1996 Boudreau (1996) DOE1994 DOE (1994) Dickson1979a Dickson and Riley (1979a) Dickson1981 Dickson (1981) Dickson1984 Dickson (1984) Dickson1987 Dickson and Millero (1987) Dickson1990 Dickson (1990a) Emerson and Hedges (2008) Emerson2008 Follows2006 Follows, Ito, and Dutkiewicz (2006) Hofmann et al. (2008b) Hofmann2008 Lewis1998 Lewis and Wallace (1998) Lueker2000 Lueker $et \ al. \ (2000)$ Millero1981 Millero and Poisson (1981) Millero1988 Millero *et al.* (1988) Millero1995 Millero (1995) Millero et al. (1995) Millero1995a Mucci1983 Mucci (1983) Perez and Fraga (1987a) Perez1987a ${\rm Riordan} 2005$ Riordan et al. (2005) Roy1993b Roy et al. (1993b) Sundquist1979 Sundquist, Plummer, and Wigley (1979) Weiss1970 Weiss (1970) Weiss1974 Weiss (1974) Wischmeyer et al. (2003) Wischmeyer 2003 Zeebe2001 Zeebe and Wolf-Gladrow (2001)

B References for the elements of an object of class aquaenv

element	references
Cl	(DOE 1994, chapter 5, p. 11), and (Zeebe and Wolf-Gladrow 2001, p. 100, footnote 3)
I	(DOE 1994, chapter 5, p. 13, 15), (Zeebe and Wolf-Gladrow 2001, p.12), and (Roy et al. 1993b, p.257). Note that the approximation $I/(mol/kg\text{-solution}) \approx 0.0199201$ S is given in (Millero 1982, p. 428.). This relationship converted into mol/kg-H ₂ O and the last digits adjusted (from 0.0199201 to 0.019924) results in the formula used here.
density	Millero and Poisson (1981) and (DOE 1994, chapter 5, p. 6f).

SumBr, ClConc, (DOE 1994, chapter 5, p.11) Na, Mg, Ca, K, Srmolal2molin (Roy et al. 1993b, p.257), and (DOE 1994, chapter 5, p. 15) free2tot. (Dickson 1984, p.2302), (DOE 1994, chapter 4, p.16), (Zeebe and Wolf-Gladrow 2001, p.57, 261) tot2free free2sws, (Dickson 1984, p.2303), (Zeebe and Wolf-Gladrow 2001, p.57) tot2sws, sws2free, sws2tot $K0_{-}CO2$ Weiss (1974), (DOE 1994, chapter 5, p. 13) (here it is stated that the unit is mol/(kg-solution*atm)), (Millero 1995, p.663), (Zeebe and Wolf-Gladrow 2001, p.257) derived from a formula for the oxygen saturation concentration in ml-O₂/kg-solution by Weiss $K0_O2$ (1970) using the first virial coefficient of oxygen (Atkins 1996, p. 41, 1029) and the atmospheric oxygen fugacity (Williams 2004) $K_{-}W$ (Millero 1995, p.670) (original reference, but slightly different formula for seawater pH), (DOE 1994, chapter 5, p. 18) (NOT the original reference! DOE (1994) cites in an update from 1997 Millero (1995)! However the version of the formula used here is the one converted to total pH scale given in DOE (1994)), and (Zeebe and Wolf-Gladrow 2001, p. 258). Constant type (stoichiometric), pH scale (total, converted to free here), and concentration unit (mol/kg-solution squared): (DOE 1994, chapter 5, p. 12,18), pH scale also in (Zeebe and Wolf-Gladrow 2001, p. 258). $K_{-}HSO4$ (DOE 1994, chapter 5 page 13), (Zeebe and Wolf-Gladrow 2001, p. 260), Dickson (1990b) (original reference). Constant type (stoichiometric), pH scale (free), and concentration unit (mol/kg-H₂O converted to mol/kg-solution here): (DOE 1994, chapter 5, p. 13). K_HF

(Dickson and Riley 1979b, p. 91) (original reference), (DOE 1994, c. 5, p. 15), (Roy et al. 1993b, p. 257), (Dickson and Millero 1987, p. 1783), (Millero 1995, p. 664), (Zeebe and Wolf-Gladrow 2001, p. 260) (converted to molinty and total scale). Constant type (stoichiometric), pH scale (free), and concentration unit (mol/kg-H₂O converted to mol/kg-solution here): (DOE 1994, chapter 5, p. 15, 16)

| 1

K_CO2, K_HCO3 (Roy et al. 1993b, p. 254) (original reference), (DOE 1994, chapter 5, p.14) (in a version converted to mol/kg-H₂O), (Millero 1995, p. 664), (Zeebe and Wolf-Gladrow 2001, p. 255). Constant type (stoichiometric) and concentration unit (mol/kg-H₂O converted to mol/kg-solution here): (DOE 1994, chapter 5, p. 14, 15), pH scale (total, converted to free here): In (DOE 1994, chapter 5, p. 12) the total scale is stated for the formula for high salinities and thus can be inferred for the formula for low salinities. The scale is also indirectly stated for both formulations in the original reference Roy et al. (1993b). Note that in Roy et al. (1993b) a function for pure water (Millero 1979) is cited and a function for seawater is derived. In Millero (1995) it is stated that for S<5 the fresh water formula of (Millero 1979) should be used and for S>=5 the seawater formula derived in Roy et al. (1993b). However, both formulations do not always intersect at S=5. The true intersection with respect to salinity S is a function of temperature Tk. Here, we first calculate this intersection by numerical root finding and then decide which formulation to use. This practise results in a continuous function with respect to S. (Note that Millero (1979) is restated wrongly in Roy et al. (1993b): one of the numerical values for the function for $K_{CO_2}^*$ is given as 310.48919, but correct is 2310.48919. However, in Millero (1995) this value is stated correctly.)

 K_BOH3

(Dickson 1990a, p. 763) (original, but mol/kg-H₂O version), (DOE 1994, ch. 5, p. 14), (Zeebe and Wolf-Gladrow 2001, p. 262), (Millero 1995, p.669) (mol/kg-H₂O version) , agrees with data in Roy, Roy, Lawson, Vogel, Moore, Davis, and Millero (1993a). Constant type (stoichiometric) and concentration unit (mol/kg-solution): (DOE 1994, chapter 5, p. 14), pH scale (total): (DOE 1994, chapter 5, p. 12) and (Zeebe and Wolf-Gladrow 2001, p.263).

K_NH4

Millero et al. (1995) (original reference), (Millero 1995, p.671). Constant type (stoichiometric) and concentration unit (mol/kg-solution): (Millero 1995, p.671), pH scale (seawater, converted to free here): Lewis and Wallace (1998) (in corrections of Millero (1995)).

Millero et al. (1988) (original reference), (Millero 1995, p.671). Constant type (stoichiometric) and

 $K_{-}H2S$

K_1125	concentration unit (mol/kg-solution): (Millero 1995, p.671), PH scale (seawater, converted to free here): Lewis and Wallace (1998) (in corrections of Millero (1995)).
K_H3PO4, K_H2PO4, K_HPO4	(Millero 1995, p.670) (original reference, but formula for seawater scale pH), (DOE 1994, ch. 5, p 16,17), agrees with data in Dickson and Riley (1979a). Constant type (stoichiometric), concentration unit (mol/kg-solution), and pH scale (total, converted to free here): (DOE 1994, chapter 5, p. 12, 16, 17).
K_SiOH4	Millero et al. (1988) (original reference), (DOE 1994, chapter 5, p 17), (Millero 1995, p.671) (formula for seawater scale pH) Constant type (stoichiometric), concentration unit (mol/kg-H ₂ O converted to mol/kg-solution here by omitting the conversion summand ln(1-0.001005 S)), and pH scale (total, converted to free here): (DOE 1994, chapter 5, p. 12, 17).
K_SiOOH3	Wischmeyer et al. (2003) (original reference), corrected due to personal communication with Dieter Wolf-Gladrow (one of the authors). The corrected version can be obtained from either Dieter Wolf-Gladrow or Andreas F Hofmann (a.hofmann@nioo.knaw.nl). Constant type (stoichiometric), concentration unit (mol/kg-solution), and pH scale (total, converted to free here): Wischmeyer et al. (2003).
K_HNO2	Constant value, not a function of temperature and salinity! Obtained as a hybrid pk value (featuring the activity of the proton but the concentration of other species (see Zeebe and Wolf-Gladrow (2001) for a treatment of different types of equilibrium constants) in molar concentration (mol/l) on the NBS pH scale (Durst 1975) from Riordan <i>et al.</i> (2005). Used as an approximation for the stoichiometric $K_{\rm HNO_2}^*$ in mol/kg-solution on the free proton pH scale here.
K_H2SO4	Constant value, not a function of temperature and salinity! Obtained as a standard pK value from (Atkins 1996, p. 1045). Used as an approximation for the stoichiometric $K_{H_2SO_4}^*$ in mol/kg-solution on the free proton pH scale here.
K_HS	Constant value, not a function of temperature and salinity! Obtained as a standard pK value from (Atkins 1996, p. 1045). Used as an approximation for the stoichiometric $K_{Hs^-}^*$ in mol/kg-solution on the free proton pH scale here.
Ksp_calcite, Ksp_aragonite	Mucci (1983) (original reference), Boudreau (1996). Note that in there are errors in Boudreau (1996): b_0 for calcite is not 0.77712 but 0.77712 and b_1 for aragonite is not 0.001727 but 0.0017276.
рН	As given in Dickson (1984), p. 2303 (use of "m") and Dickson and Riley (1979a), p. 91f all concentrations appearing in the definition of the total and the seawater pH scale are molal (mol/kg-H ₂ O) concentrations. But in Roy <i>et al.</i> (1993b), p. 257 and in DOE (1994), chapter 4, SOP 6, p. 1 it is stated, that concentrations for the seawater and total pH scale are in mol/kg-solution. To be consistent with DOE (1994) molin concentrations (mol/kg-solution) are chosen for calculating the pH.
revelle	(Zeebe and Wolf-Gladrow 2001, p.73)
dTAdKdKdS, dTAdKdKdT, dTAdKdKddd, dTAdKdKd- SumH2SO4, dTAdKdKd-	Hofmann et al. (2008a)
SumHF	

The values for K_W, K_HSO4, K_HF, K_CO2, K_HCO3, K_BOH3, K_NH4, K_H2S, K_H3PO4, K_H2PO4, K_HPO4, K_SiOH4, K_SiOOH3, Ksp_calcite, Ksp_aragonite obtained as functions of salinity S and temperature Tc from the above references are pressure corrected using the given depth d and the calculated hydrostatic pressure hydroP according to Millero (1995) with corrections by Lewis and Wallace (1998).

In general it is to be said that all corrections from Lewis and Wallace (1998) have been applied.

References

- Anderson LG, Turner DR, Wedborg M, Dyrssen D (1999). "Determination of total alkalinity and total dissolved inorganic carbon." In K Grasshoff, K Gremling, M Ehrhardt (eds.), "Methods of Seawater Analysis," Wiley-VCH.
- Atkins PW (1996). Physikalische Chemie. VCH Weinheim, 2nd edition.
- Boudreau BP (1996). "A method-of-lines code for carbon and nutrient diagenesis in aquatic sediments." Computers & Geosciences, 22(5), 479–496. URL <GotoISI>://A1996UQ20500003.
- Dickson AG (1981). "An Exact Definition of Total Alkalinity and a Procedure for the Estimation of Alkalinity and Total Inorganic Carbon from Titration Data." *Deep-Sea Research Part a-Oceanographic Research Papers*, **28**(6), 609–623. URL <GotoISI>://A1981LY51900007.
- Dickson AG (1984). "Ph Scales and Proton-Transfer Reactions in Saline Media Such as Sea-Water." *Geochimica Et Cosmochimica Acta*, **48**(11), 2299–2308. URL <GotoISI>: //A1984TT08000013.
- Dickson AG (1990a). "Standard Potential of the Reaction AgCl(S)+1/2h-2(G)=Ag(S)+HCl(Aq) and the Standard Acidity Constant of the Ion HSO4⁻ in Synthetic Sea-Water from 273.15-K to 318.15-K." *Journal of Chemical Thermodynamics*, **22**(2), 113–127. URL <GotoISI>://A1990CY50200001.
- Dickson AG (1990b). "Thermodynamics of the Dissociation of Boric-Acid in Synthetic Seawater from 273.15-K to 318.15-K." Deep-Sea Research Part a-Oceanographic Research Papers, 37(5), 755-766. URL <GotoISI>://A1990DK56300004.
- Dickson AG, Millero FJ (1987). "A Comparison of the Equilibrium-Constants for the Dissociation of Carbonic-Acid in Seawater Media." *Deep-Sea Research Part a-Oceanographic Research Papers*, **34**(10), 1733–1743. URL <GotoISI>://A1987L675400008.
- Dickson AG, Riley JP (1979a). "Estimation of Acid Dissociation-Constants in Seawater Media from Potentiometric Titrations with Strong Base .1. Ionic Product of Water Kw." *Marine Chemistry*, **7**(2), 89–99. URL <GotoISI>://A1979GJ26800001.
- Dickson AG, Riley JP (1979b). "Estimation of Acid Dissociation-Constants in Seawater Media from Potentiometric Titrations with Strong Base .2. Dissociation of Phosphoric-Acid." *Marine Chemistry*, **7**(2), 101–109. URL <GotoISI>://A1979GJ26800002.
- DOE (1994). Handbook of Methods for the Analysis of the Various Parameters of the Carbon Dioxide System in Sea Water. ORNL/CDIAC-74.
- Durst A (1975). Standard Reference Materials: Standardization of pH Measurements, volume 260-53 of NBS Spec. Publ. National Bur. Standards, Washington, D.C.
- Emerson S, Hedges JI (2008). "Carbonate Chemistry." In "Chemical Oceanography and the Marine Carbon Cycle," pp. 101–133. Cambridge University Press, Cambridge.

- Follows MJ, Ito T, Dutkiewicz S (2006). "On the solution of the carbonate chemistry system in ocean biogeochemistry models." *Ocean Modelling*, **12**(3-4), 290–301. URL <GotoISI>: //000235695500003.
- Gran G (1952). "Determination of the Equivalence Point in Potentiometric Titrations .2." Analyst, 77(920), 661–671. URL <GotoISI>://A1952UG08300014.
- Hansson I, Jagner D (1973). "Evaluation of Accuracy of Gran Plots by Means of Computer Calculations Application to Potentiometric Titration of Total Alkalinity and Carbonate Content in Sea-Water." Analytica Chimica Acta, 65(2), 363–373. URL <GotoISI>://A1973Q206600014.
- Haraldsson C, Anderson LG, Hassellov M, Hulth S, Olsson K (1997). "Rapid, high-precision potentiometric titration of alkalinity in ocean and sediment pore waters." *Deep-Sea Research Part I-Oceanographic Research Papers*, **44**(12), 2031–2044. URL <GotoISI>: //000072760700007.
- Hofmann A, Meysman F, Soetaert K, Middelburg J (2008a). "Factors governing the pH in a heterotrophic, turbid, tidal estuary." *Biogeosciences Discussion*.
- Hofmann AF, Meysman FJR, Soetaert K, Middelburg JJ (2008b). "A step-by-step procedure for pH model construction in aquatic systems." *Biogeosciences J1 BG*, **5**(1), 227–251. URL http://www.biogeosciences.net/5/227/2008/L1-http://www.biogeosciences.net/5/227/2008/bg-5-227-2008.pdf.
- Hofmann AF, Middelburg J, Soetaert K, Wolf-Gladrow DA, Meysman F (2008c). "pH modelling in aquatic environments: from an alkalinity-based to a proton-based view." in preparation.
- Lewis EL, Wallace DWR (1998). "Program Developed for CO₂ System Calculations."
- Lueker TJ, Dickson AG, Keeling CD (2000). "Ocean pCO(2) calculated from dissolved inorganic carbon, alkalinity, and equations for K-1 and K-2: validation based on laboratory measurements of CO2 in gas and seawater at equilibrium." *Marine Chemistry*, **70**(1-3), 105–119. URL <GotoISI>://000087534100009.
- Millero FJ (1979). "Thermodynamics of the Carbonate System in Seawater." Geochimica Et Cosmochimica Acta, 43(10), 1651–1661. URL <GotoISI>://A1979HP85900006.
- Millero FJ (1982). "The Thermodynamics of Seawater, Part 1. The PVT Properties." Ocean Science and Engineering, 7(4), 403–460.
- Millero FJ (1995). "Thermodynamics of the Carbon-Dioxide System in the Oceans." Geochimica Et Cosmochimica Acta, 59(4), 661–677. URL <GotoISI>://A1995QH99100003.
- Millero FJ, Plese T, Fernandez M (1988). "The Dissociation of Hydrogen-Sulfide in Seawater." Limnology and Oceanography, 33(2), 269–274. URL <GotoISI>://A1988N101700008.
- Millero FJ, Poisson A (1981). "International One-Atmosphere Equation of State of Seawater." Deep-Sea Research Part a-Oceanographic Research Papers, 28(6), 625–629. URL <GotoISI>://A1981LY51900008.

- Millero FJ, Yao WS, Aicher J (1995). "The Speciation of Fe(II) and Fe(III) in Natural-Waters." Marine Chemistry, 50(1-4), 21–39. URL <GotoISI>://A1995RV93400004.
- Mucci A (1983). "The Solubility of Calcite and Aragonite in Seawater at Various Salinities, Temperatures, and One Atmosphere Total Pressure." American Journal of Science, 283(7), 780–799. URL <GotoISI>://A1983RF42100006.
- Perez FF, Fraga F (1987a). "Association Constant of Fluoride and Hydrogen-Ions in Seawater." *Marine Chemistry*, **21**(2), 161–168. URL <GotoISI>://A1987J311800005.
- Perez FF, Fraga F (1987b). "The Ph Measurements in Seawater on the Nbs Scale." Marine Chemistry, 21(4), 315–327. URL <GotoISI>://A1987K470200002.
- Riordan E, Minogue N, Healy D, O'Driscoll P, Sodeau JR (2005). "Spectroscopic and optimization modeling study of nitrous acid in aqueous solution." *Journal of Physical Chemistry* A, 109(5), 779–786. URL <GotoISI>://000226779800009.
- Roy RN, Roy LN, Lawson M, Vogel KM, Moore CP, Davis W, Millero FJ (1993a). "Thermodynamics of the Dissociation of Boric-Acid in Seawater at S=35 from 0-Degrees-C to 55-Degrees-C." Marine Chemistry, 44(2-4), 243-248. URL <GotoISI>://A1993MT06600012.
- Roy RN, Roy LN, Vogel KM, PorterMoore C, Pearson T, Good CE, Millero FJ, Campbell DM (1993b). "The dissociation constants of carbonic acid in seawater at salinities 5 to 45 and temperatures O to 45 degrees C (vol 44, pg 249, 1996)." Marine Chemistry, 52(2), 183–183. URL <GotoISI>://A1996UH75200007.
- Skoog DA, West DM (1982). Fundamentals of Analytical Chemistry. Holt-Saunders International Editions. Holt-Saunders.
- Stumm W, Morgan JJ (1996). Aquatic Chemistry: Chemical Equilibria and Rates in natural Waters. Wiley Interscience, New York.
- Sundquist ET, Plummer LN, Wigley TML (1979). "Carbon-Dioxide in the Ocean Surface Homogeneous Buffer Factor." Science, **204**(4398), 1203–1205. URL <GotoISI>: //A1979GY02300024.
- Weiss RF (1970). "Solubility of Nitrogen, Oxygen and Argon in Water and Seawater." Deep-Sea Research, 17(4), 721–735. URL <GotoISI>://A1970H126100004.
- Weiss RF (1974). "Carbon dioxide in water and seawater: the solubility of a non-ideal gas." *Marine Chemistry*, **2**, 203–215.
- Williams DR (2004). "NASA Earth Fact Sheet." URL http://nssdc.gsfc.nasa.gov/planetary/factsheet/earthfact.html.
- Wischmeyer AG, Del Amo Y, Brzezinski M, Wolf-Gladrow DA (2003). "Theoretical constraints on the uptake of silicic acid species by marine diatoms." *Marine Chemistry*, 82(1-2), 13–29. URL <GotoISI>://000183605000002.
- Zeebe RE, Wolf-Gladrow D (2001). CO₂ in Seawater: Equilibrium, Kinetics, Isotopes. Number 65 in Elsevier Oceanography Series. Elsevier, first edition.

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