Fitting species abundance models with maximum likelihood Quick reference for sads package

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April, 24, 2014

1 Introduction

Species abundance distributions (SADs) are one of the basic patterns of ecological communities (McGill et al., 2007). The empirical distributions are traditionally modelled through probability distributions. Hence, the maximum likelihood method can be used to fit and compare competing models for SADs. The package sads provides functions to fit the most used models to empirical SADs and also to evaluate fits and to compare competing models. The package also allows to simulate SADs expected from samples from communities, with and without aggregation of individuals of the same species.

2 Installation

The package is planned to be published at CRAN soon. Meanwhile you can install the working version from GitHub with the package devtools

```
> library(devtools)
> install_github('sads', 'piklprado')
```

And then load the package:

> library(sads)

3 Exploratory analyses

We'll use two data sets in the sads package:

- > data(moths)# William's moth data
- > data(ARN82.eB.apr77)# Arntz et al. benthos data

3.1 Octaves

Function octav tabulates the number of species in classes of logarithm of abundances at base 2 (Preston's octaves) and returns a data frame ¹:

> (moths.oc <- octav(moths))</pre>

```
Object of class "octav"
   octave upper Freq
                    35
1
         1
                1
2
         2
                    11
         3
3
                4
                    29
4
         4
                8
                    32
5
         5
              16
                    26
6
         6
              32
                    32
7
         7
              64
                    31
         8
              128
8
                    13
9
         9
             256
                    19
             512
10
        10
                     5
11
        11
            1024
                     6
12
        12
            2048
                     0
            4096
13
        13
                      1
```

> (arn.oc <- octav(ARN82.eB.apr77))</pre>

Object of class "octav"

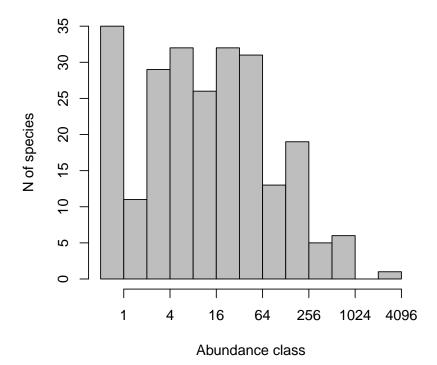
	octave	upper	Freq
1	-5	0.015625	3
2	-4	0.031250	5
3	-3	0.062500	4
4	-2	0.125000	6
5	-1	0.250000	3
6	0	0.500000	5

¹actually an object of class *octav* which inherits from class *dataframe*

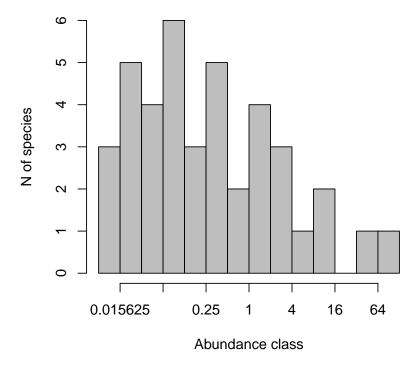
```
7
         1
             1.000000
                           2
8
         2
             2.000000
                           4
9
         3
             4.000000
                           3
10
         4
             8.000000
                           1
            16.000000
11
12
            32.000000
                           0
13
         7
            64.000000
                           1
14
         8 128.000000
                           1
```

A logical argument preston allows to smooth the numbers as proposed by Preston (1948). The octave number is the lower limit of the class in log2 scale. Hence, for abundance values smaller than one (e.g. biomass data) the octave numbers are negative numbers. A Preston plot is a histogram of this table, that you get applying the function plot to the data frame:

> plot(moths.oc)



> plot(arn.oc)



3.2 Rank-abundance plots

Function ${\tt rad}$ returns a data frame of sorted abundances and their ranks 2 :

> head(moths.rad <- rad(moths))</pre>

	${\tt rank}$	abund
1	1	2349
2	2	823
3	3	743
4	4	604
5	5	589
6	6	572

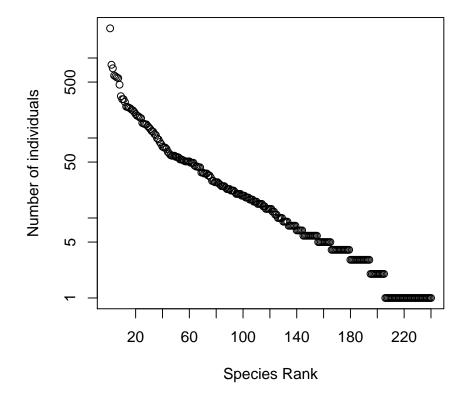
 $^{^2{\}rm actually}$ an object of class rad which inherits from class data frame

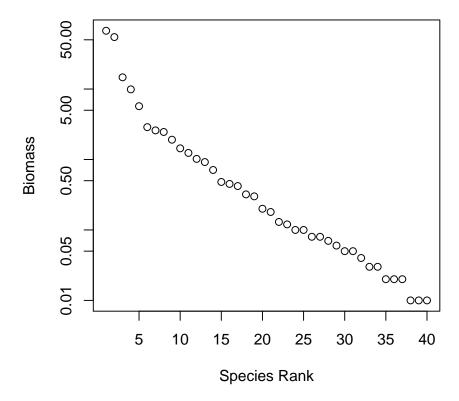
> head(arn.rad <- rad(ARN82.eB.apr77))</pre>

	rank	abund	
sp17	1	67.21	
sp11	2	54.67	
sp33	3	14.67	
sp9	4	9.90	
sp30	5	5.71	
sp10	6	2.88	

To get the rank-abundance or Whitaker's plot apply the function plot on the data frame:

> plot(moths.rad, ylab="Number of individuals")





4 Model fitting

The sads package provides maximum-likelihood fits of many probability distributions to empirical sads. The working horse is the functions fitsad for fitting species abundance distributions and fitrad for fitting rank-abundance distributions. The first argument of these functions is the vector of observed abundances and the second argument is the name of the model to be fitted. Please refer to the help page of the functions for details on the models. For more information on the fitting procedure see also the vignette of the bbmle package, on top of which the package sads is built.

To fit a logseries distribution use the argument sad='ls':

> (moths.ls <- fitsad(moths,'ls'))</pre>

```
Call:
mle2(minuslog1 = LL, start = list(alpha = alfa), method = "Brent",
    data = list(x = x), lower = 0, upper = upper)
Coefficients:
   alpha
40.24728
Log-likelihood: -1087.71
The resulting model object inherits from mle2 (Bolker & R Development Core Team, 2014),
and all usual methods for model objects, such as summaries, log-likelihood, and AIC values:
> summary(moths.ls)
Maximum likelihood estimation
Call:
mle2(minuslog1 = LL, start = list(alpha = alfa), method = "Brent",
    data = list(x = x), lower = 0, upper = upper)
Coefficients:
      Estimate Std. Error z value
                    6.961 5.7818 7.391e-09 ***
alpha
Signif. codes:
0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
-2 log L: 2175.425
> coef(moths.ls)
   alpha
40.24728
> logLik(moths.ls)
'log Lik.' -1087.713 (df=1)
> AIC(moths.ls)
```

[1] 2177.425

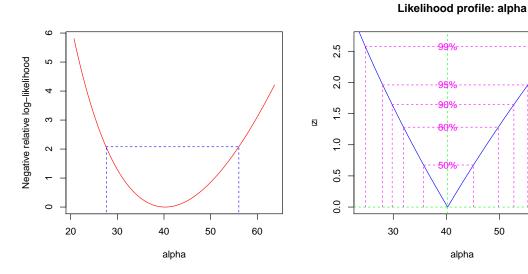
Model diagnostics 4.1

Many other diagnostic and functions are available for sad and rad models. To get likelihood profiles and confidence intervals use:

```
> moths.ls.prf <- profile(moths.ls)
> confint(moths.ls.prf) # conf intervals
  2.5 %
           97.5 %
28.01537 55.36267
```

And then use plotprofmle to plot likelihood profiles at the original scale (relative negative log-likelihood) and function plot to get plots at chi-square scale (square-root of twice the relative log-likelihood):

```
> par(mfrow=c(1,2))
> plotprofmle(moths.ls.prf)# log-likelihood profile
> plot(moths.ls.prf)# z-transformed profile
> par(mfrow=c(1,1))
```

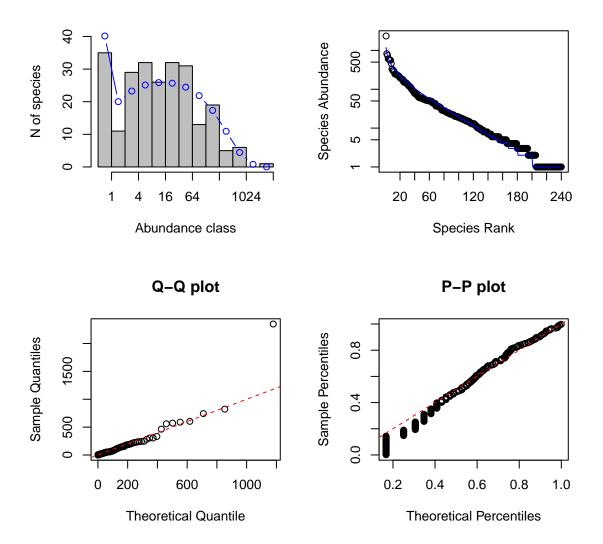


When applied on a sad model object the function plot returns four diagnostic plots:

50

60

```
> par(mfrow=c(2,2))
> plot(moths.ls)
> par(mfrow=c(1,1))
```



The first two plots (top right and left) are the octave and rank-abundance plots with the predicted values of number of species in each octave and abundances of each species. The two last plots (bottom) are quantile-quantile and percentile-percentile graphs of the observed vs. predicted abundances. The straight line indicates the expected relation in case of perfect fit.

4.2 Rank-abundance models

Species-abundance models assigns a probability to each abundance value. Hence these models are probability density functions (PDFs) of abundances of a species taken at random from the community. Rank-abundance models assigns a probability for each **abundance** rank. They are PDFs for rankings. The models are interchangeable (May, 1975), but currently only four rad models are available in package sads trough the argument rad of function fitrad:

- "gs": geometric series (which is NOT geometric PDF, available in fitsads as "geom";
- "rbs": broken-stick model (MacArthur, 1957; May, 1975)
- "zipf": zipf power-law distribution
- "mand": zipf-mandelbrot power-law distribution

Comparison to radfit from *vegan* package:

fits by fitsad, fitrad and radfit of vegan package provide similar estimates of model coefficients but not comparable likelihood values. This is because each function fit models that assigns probability to data in different ways. Function fitsad fit PDFs to observed abundances and fitrad fit PDFs to the ranks of the abundances. Finally, radfit fits a Poisson generalized linear model to the expected abundances deduced from rank-abundance relationships from the correspondending sads and rads models (Wilson, 1991). See also the help page of radfit. Therefore likelihoods obtained from these three functions are not comparable.

5 Model selection

You can fit other models to the same data set, such as the Poisson-lognormal and a truncated lognormal:

```
> (moths.pl <- fitsad(x=moths,sad="poilog"))#default is zero-truncated
Call:
mle2(minuslogl = LL, start = as.list(pl.par), data = list(x = x))
Coefficients:
    mu    sig
1.996469 2.187126
Log-likelihood: -1086.07</pre>
```

```
> (moths.ln <- fitsad(x=moths,sad="lnorm", trunc=0.5)) # lognormal truncated at 0.5
Call:
mle2(minuslog1 = LL, start = list(meanlog = meanlog, sdlog = sdlog),
    data = list(x = x)
Coefficients:
meanlog
            sdlog
2.274346 2.039740
Log-likelihood: -1085.47
and then you can use the function AICtab and friends from the bbmle package to get a
model selection table:
> AICtab(moths.ls, moths.pl, moths.ln, base=TRUE)
         AIC
                 dAIC
                        df
moths.ln 2174.9
                    0.0 2
                    1.2 2
moths.pl 2176.1
moths.ls 2177.4
                    2.5 1
To compare visually fits first get octave tables:
> head(moths.ls.oc <- octavpred(moths.ls))</pre>
  octave upper
1
       1
             1 40.14377
             2 20.02026
2
3
       3
             4 23.27123
4
       4
             8 25.12674
5
       5
            16 25.86285
6
       6
            32 25.67116
> head(moths.pl.oc <- octavpred(moths.pl))</pre>
  octave upper
                    Freq
1
       1
             1 27.58735
       2
             2 19.48216
2
       3
3
             4 26.76472
```

4

5

6

5

6

8 31.88374

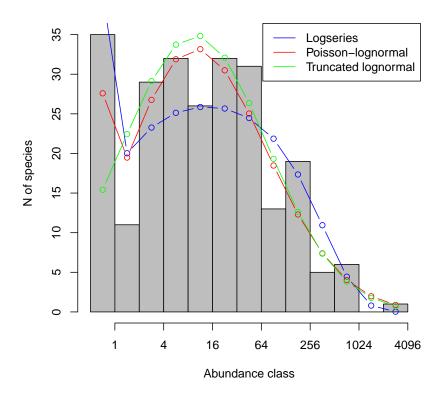
16 33.16140

32 30.49061

```
> head(moths.ln.oc <- octavpred(moths.ln))</pre>
```

```
octave upper
                 Freq
      1
            1 15.41886
1
      2
            2 22.44066
2
3
      3
          4 29.13034
4
      4
          8 33.72746
5
      5 16 34.82976
6
      6
           32 32.08088
```

and then use lines to superimpose the predicted values in the octave plot:



To do the same with rank-abundance plots get the rank-abundance objects:

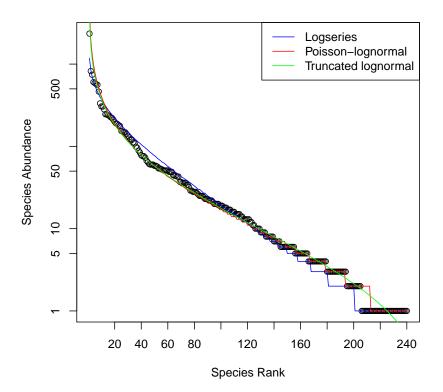
> head(moths.ls.rad <- radpred(moths.ls))</pre>

```
rank abund
1
     1
         1180
2
     2
          854
3
     3
          710
4
     4
          619
     5
5
          554
6
     6
          503
```

> head(moths.pl.rad <- radpred(moths.pl))</pre>

rank abund 1 1 4348

```
2
     2 1973
3
     3 1322
4
     4 1001
5
     5
       807
     6
       676
> head(moths.ln.rad <- radpred(moths.ln))</pre>
  rank
           abund
     1 3524.2394
1
2
     2 1674.8603
3
     3 1148.3539
4
     4 883.6309
5
     5 720.7864
6
     6 609.2707
and then plot observed and predicted values:
> plot(moths.rad)
> lines(moths.ls.rad, col="blue")
> lines(moths.pl.rad, col="red")
> lines(moths.ln.rad, col="green")
> legend("topright",
        c("Logseries", "Poisson-lognormal", "Truncated lognormal"),
        lty=1, col=c("blue","red", "green"))
```



6 Simulations

The function rsad returns random samples of a community which has S species. The mean abundance of the species in the communities are independent identically distributed (iid) variables that follow a given probability distribution. The sample simulates a given number of draws of a fraction a of the total number of individuals of the community. For instance, to simulate two Poisson samples of 10% of a community with 10 species that follows a lognormal distribution with parameters $\mu = 3$ and $\sigma = 1.5$ use:

2	1	2	4
3	1	3	7
4	1	4	2
5	1	5	4
6	1	6	1
7	1	7	25
8	1	8	3
9	1	9	45
10	1	10	1
11	2	1	17
12	2	2	2
13	2	3	0
14	2	4	3
15	2	5	6
16	2	6	2
17	2	7	18
18	2	8	0
19	2	9	53
20	2	10	4

The function returns a data frame with a sample numeric label, species numeric label and species abundance in each sample. By default, rsad returns a vector of abundances of single Poisson sample with zeroes ommitted:

[1]	155	697	4	7	48	5	40	56	105	8	48
[12]	1	3	1	14	21	6	66	2	3	32	259
[23]	8	51	21	1	312	42	23	20	48	12	28
[34]	14	20	40	267	5	209	36	107	93	58	1
[45]	7	39	2	7	56	70	31	3	4	305	25
[56]	15	12	3	48	8	12	101	69	255	5	51
[67]	253	4	1	2	17	49	187	121	599	3	23
[78]	12	9	16	21	10	17	3	5	2	9	5
[89]	3214	1	19	1	31						

Once this is a Poisson sample of a lognormal community, the abundances in the sample should follow a Poisson-lognormal distribution with parameters $\mu + \log a$ and σ (Grøtan & Engen, 2008). We can check this by fitting a Poisson-lognormal model to the sample:

```
> (samp2.pl <- fitsad(samp2, 'poilog'))</pre>
Call:
mle2(minuslog1 = LL, start = as.list(pl.par), data = list(x = x))
Coefficients:
      mu
               sig
2.709138 1.884220
Log-likelihood: -453.22
> ## checking correspondence of parameter mu
> coef(samp2.pl)[1] - log(0.1)
      mu
5.011723
Not bad. By repeating the sampling and the fit many times you can evaluate the bias and
variance of the maximum likelihood estimates:
> results <- matrix(nrow=500,ncol=2)</pre>
> for(i in 1:500){
     x \leftarrow rsad(S = 100, frac=0.1, sad="lnorm",
                meanlog=5, sdlog=2)
     y <- fitsad(x, "poilog")
     results[i,] <- coef(y)
> results[,1] <- results[,1]-log(0.1)
Bias is estimated as the difference between the mean of estimates and the value of parameters
> ##Mean of estimates
> apply(results,2,mean)
[1] 4.983124 1.993854
> ## relative bias
> (c(5,2)-apply(results,2,mean))/c(5,2)
```

And the precision of the estimates are their standard deviations

[1] 0.003375221 0.003072756

```
> ##Mean of estimates
```

```
> apply(results,2,sd)
```

[1] 0.2869798 0.2180105

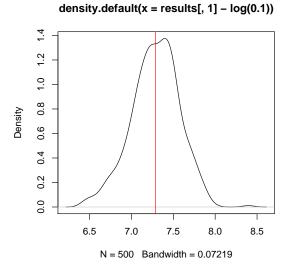
```
> ## relative precision
```

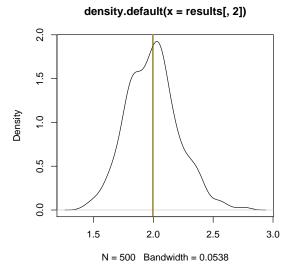
> apply(results,2,sd)/apply(results,2,mean)

[1] 0.05759033 0.10934122

Finally, a density plot with lines indicating the mean of estimates and the values of parameters:

```
> par(mfrow=c(1,2))
> plot(density(results[,1]-log(0.1)))
> abline(v=c(mean(results[,1]-log(0.1)),5), col=2:3)
> plot(density(results[,2]))
> abline(v=c(mean(results[,2]), 2), col=2:3)
> par(mfrow=c(1,1))
```





References

Bolker, B. & R Development Core Team, 2014. bbmle: Tools for general maximum likelihood estimation. R package version 1.0.16.

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