R documentation

of '/Users/tarik/Google' etc.

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R topics documented:

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synchrony-package   *Tools for computing spatial, temporal, and spatiotemporal statistics*

Description

Tools for computing spatial, temporal, and spatiotemporal statistics

Details

Package:	synchrony
Type:	Package
Version:	0.1
Date:	2012-11-06
License:	GPL (>=2)
URL:	http://cran.r-project.org/package=synchrony
LazyLoad:	yes

Author(s)

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References

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Bjornstad, O. N., R. A. Ims, and X. Lambin. 1999. Spatial population dynamics: analyzing patterns and processes of population synchrony. *Trends in Ecology & Evolution* 14:427-432.

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

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Gouhier, T. C., F. Guichard, and B. A. Menge. 2010. Ecological processes can synchronize marine population dynamics over continental scales. *Proceedings of the National Academy of Sciences* 107:8281-8286.

Loreau, M., and C. de Mazancourt. 2008. Species synchrony and its drivers: Neutral and nonneutral community dynamics in fluctuating environments. *The American Naturalist* 172:E48-E66.

Vasseur, D. A. 2007. Environmental colour intensifies the Moran effect when population dynamics are spatially heterogeneous. *Oikos* 116:1726-1736.

Zar, J. H. 1999. Biostatistical Analysis, Fourth edition. Prentice-Hall, Inc., Upper Saddle River, NJ.

Examples

```
## Compute phase synchrony
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
sync=phase.sync(t1, t2)
## Distribution of phase difference
hist(sync$deltaphase$mod_phase_diff_2pi)
```

```
## Compute concordant peaks
p=common.peaks.signif(t1, t2)
## Find proportion of time steps where both time series peak together
p$peaks
## Plot (null) distribution of proportion of time steps where both time
## series peak together
hist(p$rand)
## p-value of observed value
p$pval

## Compute Kendall's W
data(bird.traits)
(w=kendall.w(bird.traits))

## Community matrix for 20 species undergoing random fluctuations
comm.rand=matrix(runif(100), nrow=5, ncol=20)
community.sync(comm.rand)
community.sync.signif(comm.rand, nrands=100)$pval
## Community matrix for 20 species undergoing synchronized fluctuations
comm.corr=matrix(rep(comm.rand[,1], 20), nrow=5, ncol=20)
community.sync(comm.corr)
community.sync.signif(comm.corr, nrands=100)$pval
```

---

bird.traits   *bird trait dataset*

Description

Contains the wing length, tail length, and bill length from 12 birds

Usage

```
data(bird.traits)
```

Format

A data frame with 12 observations (birds) on the following 3 variables.

wing.length a numeric vector containing wing length in cm

tail.length a numeric vector containing tail length in cm

bill.length a numeric vector containing bill length in cm

Details

Dataset from Zar (1999; page 444)

Source

Zar, J. H. 1999. Biostatistical Analysis, Fourth edition. Prentice-Hall, Inc., Upper Saddle River, NJ.

Examples

```
data(bird.traits)
(w=kendall.w(bird.traits))
```

---

common.peaks.signif	*Find proportion of common peaks/troughs and compute its significance*

Description

Calculates the proportion of local minima/maxima common to both time series and computes its significance by randomly shuffling the time series

Usage

```
common.peaks.signif (t1, t2, nrands = 999)
```

Arguments

t1	time series 1 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
t2	time series 2 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
nrands	number of randomizations. Default is 999.

Value

Returns a list containing:

pval	p-value computed by randomly shuffling both time series nrands times
rands	proportion of local minima/maxima common to both time series for each randomization
peaks	proportion of local minima/maxima common to both time series in the observed dataset
locations	locations/indices of local minima/maxima common to both time series in the observed dataset

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

See Also

peaks and find.minmax

Examples

```
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
p=common.peaks.signif(t1, t2, nrands=100)
## Find proportion of time steps where both time series peak together
p$peaks
## Plot (null) distribution of proportion of time steps where both time series
## peak together
hist(p$rand)
## p-value of observed value
p$pval
```

---

community.sync	*Compute community-wide synchrony*

Description

Compute community-wide synchrony. If all species fluctuate in perfect unison, the community-wide synchrony will be 1. If species undergo uncorrelated fluctuations, the community-wide synchrony will be 0.

Usage

```
community.sync (comm.matrix)
```

Arguments

comm.matrix   community matrix in wide format where each row contains the abundance at each time step and each column corresponds to a different species.

Details

Loreau and de Mazancourt (2008) show that community-wide synchrony φ can be quantified by computing the temporal variance $\sigma^2_{x_T}$ of the community time series $x_T(t) = \sum x_i(t)$ and the sum of the temporal standard deviation of the time series across all species $\left(\sum \sigma_{x_i} \right)^2$ such that:
$$\varphi = \frac{\sigma^2_{x_T}}{\left(\sum \sigma_{x_i} \right)^2}$$

Value

Returns community-wide synchrony φ

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Loreau, M., and C. de Mazancourt. 2008. Species synchrony and its drivers: Neutral and nonneutral community dynamics in fluctuating environments. *The American Naturalist* 172:E48-E66.

Examples

```
## Community matrix for 20 species undergoing random fluctuations
comm.rand=matrix(runif(100), nrow=5, ncol=20)
community.sync(comm.rand)
## Community matrix for 20 species undergoing synchronized fluctuations
comm.corr=matrix(rep(comm.rand[,1], 20), nrow=5, ncol=20)
community.sync(comm.corr)
## Compare this metric to Kendall's W
data(bird.traits)
kendall.w(bird.traits)
community.sync(bird.traits)
```

---

community.sync.signif	*Compute significance of community-wide synchrony via Monte Carlo randomizations*

Description

Compute the significance of community-wide synchrony using Monte Carlo randomizations. If all species fluctuate in perfect unison, the community-wide synchrony will be 1. If species undergo uncorrelated fluctuations, the community-wide synchrony will be 0.

Usage

```
community.sync.signif (comm.matrix, nrands = 999)
```

Arguments

comm.matrix	community matrix in wide format where each row contains the abundance at each time step and each column corresponds to a different species.
nrands	number of randomizations to perform (default is 999)

Details

Loreau and de Mazancourt (2008) show that community-wide synchrony φ can be quantified by computing the temporal variance $\sigma^2_{x_T}$ of the community time series $x_T(t) = \sum x_i(t)$ and the sum of the temporal standard deviation of the time series across all species $\left(\sum \sigma_{x_i}\right)^2$ such that:
$$\varphi = \frac{\sigma^2_{x_T}}{\left(\sum \sigma_{x_i}\right)^2}$$

Value

Returns a named list containing:

obs	the observed community synchrony
rands	the community synchrony value of the nrands randomizations
pval	p-value of observed community synchrony

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Loreau, M., and C. de Mazancourt. 2008. Species synchrony and its drivers: Neutral and nonneutral community dynamics in fluctuating environments. *The American Naturalist* 172:E48-E66.

See Also

community.sync

Examples

```
## Community matrix for 20 species undergoing random fluctuations
comm.rand=matrix(runif(100), nrow=5, ncol=20)
community.sync.signif(comm.rand, nrands=20)$pval
## Community matrix for 20 species undergoing synchronized fluctuations
comm.corr=matrix(rep(comm.rand[,1], 20), nrow=5, ncol=20)
community.sync.signif(comm.corr, nrands=20)$pval
## On "real" data
data(bird.traits)
community.sync.signif(bird.traits, nrands=20)$pval
```

---

coord2dist *coord2dist*

Description

Calculate distance between all pairs of sites

Usage

```
coord2dist (coords, is.latlon = TRUE, lower.tri = TRUE)
```

Arguments

coords	n x 4 matrix of coordinates consisting of lat or y, lon or x pairs for each each site
is.latlon	are coordinates latitudes/longitudes? Default is TRUE
lower.tri	Return lower triangular part of the distance matrix? Default is TRUE

Value

Returns the distance between all pairs of sites

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

Examples

```
coords=rbind(c(32, -125), c(43, -130))
## Compute great circle distance
coord2dist(coords)
```

---

find.minmax   *Find min/max of a time series*

Description

Find local minima and maxima of a time series

Usage

```
find.minmax (timeseries)
```

Arguments

timeseries   time series in matrix format (n rows x 2 columns). The first column should
                  contain the time steps and the second column should contain the values.

Value

Returns a list containing:

mins   n x 2 matrix containing the time steps and the local min values

maxs   n x 2 matrix containing the time steps and the local max values

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

See Also

[find.minmax.ind](#)

Examples

```
t1=cbind(1:100, runif(100))
min.max=find.minmax(t1)
min.max$maxs
plot (t1, t="l")
points (min.max$mins, col="blue", bg="blue", pch=19)
points (min.max$maxs, col="red", bg="red", pch=19)
```

---

find.minmax.ind *Find indices of local minima or maxima of a time series*

Description

Find indices of local minima or maxima of a time series

Usage

```
find.minmax.ind (timeseries, mins = FALSE)
```

Arguments

timeseries	time series in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
mins	find local minima (`mins=TRUE`) or maxima (`mins=FALSE`). Default is `FALSE`.

Value

Returns index of local minima or maxima

Note

This is the low-level function used by `find.minmax`. The optimized version of this function is based on code posted by Ben Bolker on StackOverflow.

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

See Also

`find.minmax`

Examples

```
t1=cbind(1:100, runif(100))
## Find local maxima
maxs=find.minmax.ind(t1)
## Find local minima
mins=find.minmax.ind(t1, mins=TRUE)
```

---

kendall.w   *Kendall's W*

Description

Compute Kendall's coefficient of concordance (W)

Usage

```
kendall.w (data)
```

Arguments

data   matrix in wide format where each row represents a different sample and each column represents a different variable.

Details

Kendall's W is a non-parametric statistic that ranges from 0 to 1 and measures the level of agreement between multiple variables. When the number of observations $n > 10$, its significance can be determined by using a χ^2 distribution with $df = n - 1$. Kendall's W is often used to measure the degree of synchrony among different locations.

Value

Returns a named list containing:

w.uncorrected   Kendall's W uncorrected for tied ranks

w.corrected   Kendall's W corrected for tied ranks

pval   p-value of Kendall's W

spearman.corr   Spearman's ranked correlation

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

Gouhier, T. C., and F. Guichard. 2007. Local disturbance cycles and the maintenance of spatial heterogeneity across scales in marine metapopulations. *Ecology* 88:647-657.

Gouhier, T. C., F. Guichard, and A. Gonzalez. 2010. Synchrony and stability of food webs in metacommunities. *The American Naturalist* 175:E16-E34.

Zar, J. H. 1999. Biostatistical Analysis, Fourth edition. Prentice-Hall, Inc., Upper Saddle River, NJ.

Examples

```
data(bird.traits)
(w=kendall.w(bird.traits))
```

---

latlon2dist   *latlon2dist*

Description

Calculate distance between a pair of coordinates

Usage

```
latlon2dist (coords)
```

Arguments

coords   4-element vector of coordinates with format: (lat1, lon1, lat2, lon2)

Value

Returns the great circle distance distance between the pair of coordinates

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

See Also

coord2dist

Examples

```
coords=c(32, -125, 43, -130)
## Compute great circle distance
latlon2dist(coords)
```

---

peaks   *Find common peaks*

Description

Find the proportion of local minima/maxima common to both time series

Usage

```
peaks (t1, t2)
```

Arguments

t1   time series 1 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.

t2   time series 2 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.

Value

Returns a named list containing peaks and locations

peaks	proportion of local minima/maxima common to both time series
locations	locations/indices of local minima/maxima common to both time series

Note

This is the low-level function used by common.peaks.signif

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

See Also

common.peaks.signif

Examples

```
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
p=peaks(t1, t2)
p$peaks
```

---

phase.partnered	*Phase partnered time series*

Description

Create two time series with specific autocorrelation γ and cross-correlation ρ

Usage

```
phase.partnered (n = 2000, rho = 1, gamma = 1, ts.sd = 0.1, ts.mean = 0)
```

Arguments

n	number of time steps in time series. Default is 2000.
rho	cross-correlation between the two time series ($-1 \leq \rho \leq 1$). Default is 1.
gamma	autocorrelation of each time series. Gamma (γ) describes the relationship between frequency f and power P: $P(f) = 1/f^{\gamma}$
	$-2 \leq \gamma \leq 0$: blue noise and $0 \leq \gamma \leq 2$: red noise. Default is 1.
ts.sd	standard deviation of both time series. Default is 0.1.
ts.mean	mean of both time series. Default is 0.

Value

Returns an `n` x 2 matrix containing the time series

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Gouhier, T. C., F. Guichard, and A. Gonzalez. 2010. Synchrony and stability of food webs in metacommunities. *The American Naturalist* 175:E16-E34.

Vasseur, D. A. 2007. Environmental colour intensifies the Moran effect when population dynamics are spatially heterogeneous. *Oikos* 116:1726-1736.

Examples

```
## Positively cross-correlated white noise
pos.corr=phase.partnered(n = 100, rho = 0.7, gamma = 0)
## Negatively cross-correlated white noise
neg.corr=phase.partnered(n = 100, rho = -1, gamma = 0)
par(mfrow=c(2,1))
matplot (pos.corr, t="l", lty=1)
matplot (neg.corr, t="l", lty=1)
```

---

phase.sync *Phase synchrony of quasi-periodic time series*

Description

Compute the phase synchrony between two quasi-periodic time series by quantifying their phase difference at each time step

Usage

```
phase.sync (t1, t2, mins = FALSE)
```

Arguments

t1	time series 1 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
t2	time series 2 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
mins	use local minima instead of local maxima to compute and then interpolate the phase. Default is `FALSE`.

Details

Two time series are phase-locked if the relationship between their phases remains constant over time. This function computes the phase of successive local maxima or minima for each time series and then uses linear interpolation to find the phase at time steps that fall between local maxima/minima. A histogram can be used to determine if the distribution of the phase difference at each time step is uniform (indicating no phase locking) or has a clear peak (indicating phase locking).

Value

Returns a list containing `phases1`, `phases2`, and `deltaphase`

`phases1`	n x 3 matrix containing the timestep, value, and phase of the first time series
`phases2`	n x 3 matrix containing the timestep, value, and phase of the second time series
`deltaphase`	n x 4 matrix containing the timestep, raw phase difference, phase difference modulus 2π between 0 and 2π, phase difference modulus 2π between $-\pi$ and π

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

Examples

```
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
## Compute and interpolate phases using successive local minima
sync.mins=phase.sync(t1, t2, mins=TRUE)
## Compute and interpolate phases using successive local maxima
sync.maxs=phase.sync(t1, t2)
## Plot distribution of phase difference
hist(sync.mins$deltaphase$mod_phase_diff_2pi)
```

`phase.sync.signif`	*Compute significance of phase synchrony*

Description

Compute the significance of phase synchrony between two time series via Monte Carlo randomizations

Usage

```
phase.sync.signif (t1, t2, nrands =  999, type = 1, nbreaks = 10, mins = FALSE)
```

Arguments

`t1`	time series 1 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
`t2`	time series 2 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
`nrands`	number of randomizations to perform (default is 999)
`type`	flag to indicate whether to compute phase difference modulus 2π between 0 and 2π (`type=1`) or phase difference modulus 2π between $-\pi$ and π. Default is 1.
`nbreaks`	number of bins to use to group the values in the time series. Default is 10.
`mins`	use local minima instead of local maxima to compute and the interpolate the phase. Default is `FALSE`.

Details

This function determines whether two time series are phase locked by randomly shuffling the time series. Specifically, the distribution of the phase difference of the original time series is binned using a histogram and an index of phase locking Q based on Shannon's entropy is computed: $Q.obs = (S_{max} - S_{obs})/S_{max}$, where $S_{obs} = -\sum_i^N p_i log(p_i)$, p_i represents the proportion of phase differences in each histogram bin, N the total number of bins, and $S_{max} = log(N)$. $Q.obs = 0$ when there is no phase synchrony and $Q.obs = 1$ when there is perfect phase synchrony. The statistical significance of the phase synchrony $Q.obs$ can then be computed by comparing the observed $Q.obs$ value to the Q values obtained from nrands surrogate time series whose short-term correlations and overall temporal patterns match those of the observed time series. The p-value is then the proportion of surrogate time series whose $Q \geq Q.obs$.

Value

Returns a named list containing:

Q.obs	Observed Q value
pval	p-value of observed Q.obs
rands	Q values from all randomizations
phases1	n x 3 matrix containing the timestep, value, and phase of the first time series
phases2	n x 3 matrix containing the timestep, value, and phase of the second time series
deltaphase	n x 4 matrix containing the timestep, raw phase difference, phase difference modulus 2π between 0 and 2π, phase difference modulus 2π between $-\pi$ and π
icdf	nrands x 2 matrix containing the inverse cumulative distribution of the Q values computed using the surrogate time series

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

See Also

phase.sync and surrogate.ts

Examples

```
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
p=phase.sync.signif(t1, t2, nrands=99)
p$pval
hist(p$rands)
plot (p$icdf, t="l")
abline(v=p$Q.obs, col="red", lty=2)
```

pisco.data	*PISCO multi-year and spatially-explicit mussel and environmental dataset*

Description

Contains the mean annual chl-a concentration, sea surface temperature, upwelling currents, and mussel abundance at 48 intertidal sites along the West Coast of the United States from 2000-2003.

Usage

```
data(pisco.data)
```

Format

A data frame with 192 observations on the following 7 variables.

latitude latitude (degrees North)

longitude longitude (degrees West)

chl mean annual remote sensed chlorophyll-a concentration

sst mean annual remote sensed sea surface temperature

upwelling mean annual remote sensed upwelling currents

mussel_abund mean annual mussel cover (*Mytilus californianus*)

year sampling year

References

Gouhier, T. C., F. Guichard, and B. A. Menge. 2010. Ecological processes can synchronize marine population dynamics over continental scales. *Proceedings of the National Academy of Sciences* 107:8281-8286.

Examples

```
data(pisco.data)
```

plot.vario	*Plot* vario *objects*

Description

Plot vario objects

Usage

```
## S3 method for class 'vario'
 plot(x, xlab = "Lag distance", ylab = NULL, ylim = NULL,
                    xtype = c("mean.bin.dist", "bins"), rug = FALSE,
                    pch = 21, col.sig="black", col.nonsig="black", bg.sig="black",
                    bg.nonsig="white", alpha = 0.05, ...)
```

Arguments

x	vario object generated by `vario` function.
xlab	xlabel of the figure. Default is "Lag distance"
ylab	ylabel of the figure. Default is `NULL` and will automatically generate the right label
ylim	y-range. Default is `NULL` and will automatically generate the best range based on the metric
xtype	Use either the discrete bin classes (`bins`) or the mean distance of the points within each bin (`mean.bin.dist`) on the x-axis. Default is `mean.bin.dist`
rug	Plot rug indicating the density of data points? Default is `FALSE`
pch	Type of points to use when plotting the variogram. Default is 21
col.sig	Border color of points for significant values. Default is black
col.nonsig	Border color of points for non-significant values. Default is black
bg.sig	Background color of points for significant values. Default is black
bg.nonsig	Background color of points for non-significant values. Default is black
alpha	Significance level. Default is 0.05
...	other graphical parameters.

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

Examples

```
data(pisco.data)
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "sst"))
semiv=vario(data=d)
moran=vario(data=d, type="moran", nrand=100)
geary=vario(data=d, type="geary", nrand=100)

par(mfrow=c(3,1))
plot(semiv)
plot(moran, bg.sig="blue")
plot(geary, bg.sig="red")
```

---

plot.variofit	*Plot* variofit *objects*

Description

Plot `variofit` objects

Usage

```
## S3 method for class 'variofit'
 plot(x, xlab = "Lag distance", ylab = "Variogram",
                        col.pts = "black", col.line = "red", pch = 21, ...)
```

Arguments

x	variofit object generated by `vario.fit` function
xlab	xlabel of the figure. Default is "Lag distance"
ylab	ylabel of the figure. Default is "Variogram"
col.pts	Border color of the points. Default is black
col.line	Color of the fitted variogram. Default is red
pch	Type of points to use when plotting the variogram. Default is 21
...	other graphical parameters.

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

Examples

```
## Environmental variogram
data(pisco.data)
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "upwelling"))
semiv=vario(data=d)
mod.sph=vario.fit(semiv$vario, semiv$mean.bin.dist)
plot(mod.sph)
```

---

surrogate.ts   *Create surrogate time series via Markov process*

Description

Create surrogate time series with the same short-term time correlation and overall temporal pattern as the original time series using the Markov process described by Cazelles and Stones (2003)

Usage

```
surrogate.ts (ts, distr.ts = NULL, trans.ts = NULL, nbreaks = 10)
```

Arguments

ts	time series in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
distr.ts	binning of time series values. This parameter must be specified if `trans.ts` is not set to NULL. Default is NULL.
trans.ts	transition matrix from bin i to bin j. Default is NULL.
nbreaks	number of bins to use to group the time series values. Default is 10.

Details

The values of the time series x_n are grouped into nbreak equally-sized bins. The transition matrix M_{ij} describing the probability of x_{n+1} belonging to bin j based on x_n belonging to bin i is defined using the relative frequencies of the data such that: $M_{ij} = Pr(x_{n+1} \in b_j | x_n \in b_i)$. The surrogate time series is then constructed by randomly selecting a starting value and randomly selecting the next value from the proper bin based on the transition matrix. This process is repeated until the surrogate time series has the same length as the original time series.

Value

Returns a named list containing:

surr.ts	surrogate time series in matrix format
trans	transition matrix M_{ij}
distr	binning of time series values

Author(s)

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References

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

See Also

[phase.sync.signif](#)

Examples

```
t1=cbind(1:100, runif(100))
surr.t1=surrogate.ts(ts=t1)
plot(t1, t="l")
lines(surr.t1$surr.ts, col="red")
```

---

vario	*vario*

Description

Compute the empirical variogram

Usage

```
vario (nbins = 20, extent = 0.5, data, data2 = NULL, is.latlon = TRUE, centered = FALSE,
                nrands = 0, type = c("semivar", "cov", "pearson",
                "spearman", "kendall", "moran", "geary"),
                mult.test.corr = c(FALSE, "holm", "hochberg", "sidak", "bonferroni"))
```

Arguments

nbins	Number of bins or lags
extent	Proportion of the spatial extent of the data over which to compute the variogram. Default is 0.5 to limit potentially spurious results due to the limited number of data points at large lag distances.
data	n x m matrix containing y-coordinates (or latitude), x-coordinates (or longitude), and values. The values can either be a single column of observations at each site for univariate variograms or a matrix of observations at each site for multivariate variograms (e.g., to compute spatial synchrony).

data2	n x m matrix containing y-coordinates (or latitude), x-coordinates (or longitude), and values for second variable. The values can either be a single column of observations at each site for univariate variograms or a matrix of observations at each site for multivariate variograms (e.g., to compute spatial synchrony).
is.latlon	Are coordinates latitudes/longitudes? Default is TRUE
centered	Should the variogram be centered by subtracting the regional mean from the value in each bin? Default is FALSE
nrands	Number of randomizations to determine statistical significance of variogram. Default is 0.
type	Type of variogram to compute. Default is semivar for semivariance. Other options include cov for covariance, pearson for Pearson correlation, spearman for Spearman correlation, kendall for Kendall correlation, moran for Moran's I, and geary for Geary's C
mult.test.corr	Correct for multiple tests? Default is FALSE. Other options inlcude holm, hochberg, sidak, and bonferroni

Details

This function can be used to compute univariate correlograms using Moran's I, Geary's C, and the covariance function or variograms using the semivariance function. Multivariate (Mantel) correlograms can also be computed using the covariance function, Pearson's, Spearman's or Kendall's correlation coefficients. Cross-correlograms/variograms between data1 and data2 can be computed with the covariance function, Pearson's, Spearman's or Kendall's correlation coefficients for multivariate variograms and Moran's I, Geary's C, the covariance function, or semivariance for univariate variograms.

Value

Returns a named list containing the following variables:

bins	Center of each lag/bin
mean.bin.dist	Mean distance of each lag/bin
vario	Variogram values in each lag/bin
pvals	P-value for each lag/bin
rands	nrands x nbins matrix of randomizations
metric	Type of variogram computed
pvals	P-value for each lag/bin
mult.test.corr	
	Correct for multiple tests?
is.multivar	Was the analysis performed on multivariate data?

Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

References

Bjornstad, O. N., and W. Falck. 2001. Nonparametric spatial covariance functions: Estimation and testing. *Environmental and Ecological Statistics* 8:53-70.

Bjornstad, O. N., R. A. Ims, and X. Lambin. 1999. Spatial population dynamics: analyzing patterns and processes of population synchrony. *Trends in Ecology & Evolution* 14:427-432.

Fortin, M. J., and M. R. T. Dale. 2005. Spatial Analysis: A Guide for Ecologists. Cambridge University Press.

See Also

vario.signif, vario.func

Examples

```
data(pisco.data)
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "sst"))
semiv=vario(data=d)
moran=vario(data=d, type="moran", nrands=100)
par(mfrow=c(2,1), mar=c(4.2, 4, 1, 1))
plot(semiv$mean.bin.dist, semiv$vario, xlab="Lag distance (km)", ylab="Semivariance")
plot(moran$mean.bin.dist, moran$vario, xlab="Lag distance (km)", ylab="Moran's I", t="l")
points(moran$mean.bin.dist[moran$pvals >= 0.05], moran$vario[moran$pvals >= 0.05],
       bg="white", pch=21)
points(moran$mean.bin.dist[moran$pvals < 0.05], moran$vario[moran$pvals < 0.05],
       bg="black", pch=21)
abline(h=0, lty=2)


## Compute spatial synchrony
d.upw=subset(pisco.data, select=c("latitude", "longitude", "year", "upwelling"))
d.cov=subset(pisco.data, select=c("latitude", "longitude", "year", "mussel_abund"))
## Reshape the data
d.upw.wide=reshape(data=d.upw, idvar=c("latitude", "longitude"), timevar=c("year"),
                   direction="wide")
d.cov.wide=reshape(data=d.cov, idvar=c("latitude", "longitude"), timevar=c("year"),
                   direction="wide")
## Generate variograms
v.upw=vario(nbins=12, data=d.upw.wide, type="pearson", extent=1, nrands=999)
v.cov=vario(nbins=12, data=d.cov.wide, type="pearson", extent=1, nrands=999)
## Fit variograms
v.cov.per=vario.fit(v.cov$vario, v.cov$mean.bin.dist, type="period",
                    start.vals=list(a=1, b=3, c=0))
v.upw.lin=vario.fit(v.upw$vario, v.upw$mean.bin.dist, type="linear")

par(mfrow=c(2,1))
plot(v.cov, xlab="Lag distance (km)", bg.sig="red", col.nonsig="red",
     main="Mussel cover",
     rug=TRUE, ylim=c(-0.3, 0.3))
lines(v.cov$mean.bin.dist, v.cov.per$fit, col="red")
plot(v.upw, xlab="Lag distance (km)", bg.sig="blue", col.nonsig="blue",
     main="Upwelling", rug=TRUE)
lines(v.upw$mean.bin.dist, v.upw.lin$fit, col="blue")
```

---

vario.fit   *vario.fit*

Description

Fit model to the empirical variogram

Usage

```
vario.fit (vario, bins,
                        type = c("spherical", "gaussian", "nugget", "linear",
                        "exponential", "sill", "periodic", "hole"),
                        start.vals = list(c0 = min(vario), c1 = max(vario),
                                          a = max(bins)/4, b=0.1, c=0.1))
```

Arguments

vario	Empirical variogram from `emp.vario` function
bins	Number of bins or lag distances from `emp.vario` function
type	Type of variogram model to fit to the data. Default is `spherical`. Other options are `gaussian`, `nugget`, `linear`, `exponential`, `sill`, `periodic`, and `hole`
start.vals	Named list containing the start values for the variogram model: `c0`: nugget, `c1`: sill, `a`: spatial range; `b`: slope; `c`: frequency

Value

Return a named list containing the following variables:

vario	Empirical variogram values
bins	Empirical variogram bins/lag distances
AIC	AIC score of the model fit
RMSE	Root Mean Square Error of the model fit
params	Named list containing the best estimates for the model parameters
fit	Predicted variogram values from the model fit
nls.success	did `nls` succeed?

Note

Selecting proper initial values is critical for fitting a reasonable model to the empirical variogram. If these values are off, `nls` will fail and fall-back functions will be used to determine the best parameter values that minimize the Root Mean Square Error (RMSE).

Author(s)

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See Also

[vario.signif](), [vario](), [vario.func]()

Examples

```
## Load data
data(pisco.data)
## Environmental variogram
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "upwelling"))
semiv=vario(data=d)
plot(semiv, xlab="Lag distance (km)")
mod.sph=vario.fit(semiv$vario, semiv$mean.bin.dist)
mod.exp=vario.fit(semiv$vario, semiv$mean.bin.dist, type="expo")
mod.gau=vario.fit(semiv$vario, semiv$mean.bin.dist, type="gauss")
mod.lin=vario.fit(semiv$vario, semiv$mean.bin.dist, type="lin")
lines(semiv$mean.bin.dist, mod.sph$fit, col="red")
lines(semiv$mean.bin.dist, mod.exp$fit, col="black")
lines(semiv$mean.bin.dist, mod.gau$fit, col="blue")
lines(semiv$mean.bin.dist, mod.lin$fit, col="green")
legend(x="topleft", legend=paste(c("Spherical AIC:", "Exponential AIC:",
                                    "Gaussian AIC:", "Linear AIC:"),
                                  c(format(mod.sph$AIC, dig=2),
                                    format(mod.exp$AIC, dig=2),
                                    format(mod.gau$AIC, dig=2),
        format(mod.lin$AIC, dig=2))), lty=1, col=c("red", "black", "blue", "green"),
        box.lty=0)

## Correlogram
cover=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "mussel_abund"))
moran=vario(data=cover, type="moran")
mod.hol=vario.fit(moran$vario, moran$mean.bin.dist, type="hole", start.vals=list(c0=0.6, a=25, c1=0.01))
mod.per=vario.fit(moran$vario, moran$mean.bin.dist, type="period", start.vals=list(a=1, b=3, c=0))
mod.lin=vario.fit(moran$vario, moran$mean.bin.dist, type="linear")
plot(moran, xlab="Lag distance (km)", ylim=c(-0.6, 0.8))
lines(moran$mean.bin.dist, mod.per$fit, col="red")
lines(moran$mean.bin.dist, mod.hol$fit, col="black")
lines(moran$mean.bin.dist, mod.lin$fit, col="blue")
legend(x="topleft", legend=paste(c("Periodic AIC:", "Hole AIC:",
                                    "Linear AIC:"),
                                  c(format(mod.per$AIC, dig=2),
                                    format(mod.hol$AIC, dig=2),
                                    format(mod.lin$AIC, dig=2))),
                                  lty=1, col=c("red", "black", "blue"), bty="n")
```

---

vario.func	*vario.func*

Description

Compute the empirical variogram values for each bin

Usage

```
vario.func (x, y, glob.mean, glob.sd, glob.N, is.multivar = FALSE,
                     type=c("semivar", "cov", "pearson",
                             "spearman", "kendall", "moran", "geary"))
```

Arguments

x	First set of sites within bin/lag distance
y	Second set of sites within bin/lag distance
glob.mean	Global mean
glob.sd	Global standard deviation
glob.N	Global number of points
is.multivar	Is the data multivariate? Default is FALSE
type	Type of variogram to compute. Default is semivar for semivariance. Other options include cov for covariance, pearson for Pearson correlation, spearman for Spearman correlation, kendall for Kendall correlation, moran for Moran's I, and geary for Geary's C

Value

Return the value.

Author(s)

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See Also

vario, vario.signif

Examples

```
## Internal function used by vario
```

---

vario.signif   *vario.signif*

Description

Determine the significance of the empirical variogram via Monte Carlo randomizations

Usage

```
vario.signif (nrands = 999, bins, all.combs, grpdata, data, data2, type,
                          vario, glob.mean = NULL, glob.sd = NULL,
                          glob.N = NULL, is.multivar = FALSE,
                          mult.test.corr=c(FALSE, "holm", "hochberg",
                                                "sidak", "bonferroni"))
```

Arguments

`nrands`	Number of randomizations
`bins`	Number of bins or lag distances
`all.combs`	All combinations of pairs of sites
`grpdata`	Grouped sites based on distance bins
`data`	n x m matrix containing y-coordinates (or latitude), x-coordinates (or longitude), and values. The values can either be a single column of observations at each site for univariate variograms or a matrix of observations at each site for multivariate variograms (e.g., to compute spatial synchrony).
`data2`	n x m matrix containing y-coordinates (or latitude), x-coordinates (or longitude), and values. The values can either be a single column of observations at each site for univariate variograms or a matrix of observations at each site for multivariate variograms (e.g., to compute spatial synchrony).
`type`	Type of variogram to compute. Default is `semivar` for semivariance. Other options include `cov` for covariance, `pearson` for Pearson correlation, `spearman` for Spearman correlation, `kendall` for Kendall correlation, `moran` for Moran's I, and `geary` for Geary's C
`vario`	Empirical variogram
`glob.mean`	Global mean
`glob.sd`	Global standard deviation
`glob.N`	Global number of points
`is.multivar`	Is the data multivariate? Default is `FALSE`
`mult.test.corr`	Correct for multiple tests? Default is `FALSE`. Other options inlcude `holm`, `hochberg`, `sidak`, and `bonferroni`

Value

Returns a named list containing the following variables:

`rands`	`nrands` x `nbins` matrix of randomizations
`pvals`	p-values based on one-tailed test
`vario`	Variogram values in each lag/bin
`mult.test.corr`	Corrected for multiple tests?

Author(s)

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See Also

[vario,](#) [vario.func](#)

Examples

```
## Internal function used by vario
```

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