# Package 'synchrony'

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Title Tools for computing spatial, temporal, and spatiotemporal statistics

Type Package

Version 0.1

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synchrony-package

Tools for computing spatial, temporal, and spatiotemporal statistics

#### **Description**

Tools for computing spatial, temporal, and spatiotemporal statistics

#### **Details**

Package: synchrony Type: Package Version: 0.1

Date: 2012-11-10 License: GPL (>=2)

URL: http://synchrony.r-forge.r-project.org

LazyLoad: yes

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

Maintainer: Tarik C. Gouhier (tarik.gouhier@gmail.com)

#### References

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Bjornstad, O. N., R. A. Ims, and X. Lambin. 1999. Spatial population dynamics: analyzing patterns and processes of population synchrony. *Trends in Ecology & Evolution* 14:427-432.

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

Fortin, M. J., and M. R. T. Dale. 2005. Spatial Analysis: A Guide for Ecologists. Cambridge University Press.

Gouhier, T. C., and F. Guichard. 2007. Local disturbance cycles and the maintenance of spatial heterogeneity across scales in marine metapopulations. *Ecology* 88:647-657.

Gouhier, T. C., F. Guichard, and A. Gonzalez. 2010. Synchrony and stability of food webs in metacommunities. *The American Naturalist* 175:E16-E34.

Gouhier, T. C., F. Guichard, and B. A. Menge. 2010. Ecological processes can synchronize marine population dynamics over continental scales. *Proceedings of the National Academy of Sciences* 107:8281-8286.

Loreau, M., and C. de Mazancourt. 2008. Species synchrony and its drivers: Neutral and nonneutral community dynamics in fluctuating environments. *The American Naturalist* 172:E48-E66.

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Vasseur, D. A. 2007. Environmental colour intensifies the Moran effect when population dynamics are spatially heterogeneous. *Oikos* 116:1726-1736.

Zar, J. H. 1999. Biostatistical Analysis, Fourth edition. Prentice-Hall, Inc., Upper Saddle River, NJ.

#### **Examples**

```
## Compute phase synchrony
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
sync=phase.sync(t1, t2)
## Distribution of phase difference
hist(sync$deltaphase$mod_phase_diff_2pi)
## Compute concordant peaks
p=peaks(t1, t2, nrands=100)
## Find proportion of time steps where both time series peak together
p$peaks
## Plot (null) distribution of proportion of time steps where both time
## series peak together
hist(p$rand)
## p-value of observed value
p$pval
## Compute Kendall's W
data(bird.traits)
(w=kendall.w(bird.traits))
## Community matrix for 20 species undergoing random fluctuations
comm.rand=matrix(runif(100), nrow=5, ncol=20)
community.sync(comm.rand, nrands=10)
## Community matrix for 20 species undergoing synchronized fluctuations
comm.corr=matrix(rep(comm.rand[,1], 20), nrow=5, ncol=20)
community.sync(comm.corr, nrands=10)
```

bird.traits

bird trait dataset

# Description

Contains the wing length, tail length, and bill length from 12 birds

# Usage

```
data(bird.traits)
```

### **Format**

A data frame with 12 observations (birds) on the following 3 variables.

```
wing.length a numeric vector containing wing length in cm
tail.length a numeric vector containing tail length in cm
bill.length a numeric vector containing bill length in cm
```

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#### **Details**

Dataset from Zar (1999; page 444)

#### **Source**

Zar, J. H. 1999. Biostatistical Analysis, Fourth edition. Prentice-Hall, Inc., Upper Saddle River, NJ.

# **Examples**

```
data(bird.traits)
(w=kendall.w(bird.traits))
```

community.sync

Compute community-wide synchrony and its significance via Monte Carlo randomizations

# **Description**

Compute community-wide synchrony and its the significance via Monte Carlo randomizations. If all species fluctuate in perfect unison, the community-wide synchrony will be 1. If species undergo uncorrelated fluctuations, the community-wide synchrony will be 0. The Monte Carlo randomizations are performed by shuffling the columns of the community matrix independently. This function also returns the mean correlation between the columns of the matrix.

#### Usage

```
community.sync (comm.matrix, nrands = 0, method=c("pearson", "kendall", "spearman"), ...)
```

# **Arguments**

comm.matrix community matrix in wide format where each row contains the abundance at

each time step and each column corresponds to a different species.

nrands number of randomizations to perform (default is 0)

method Method to compute mean correlation between columns? Options include pearson,

kendall, and spearman. Default is pearson

Other parameters to cor function.

# **Details**

Loreau and de Mazancourt (2008) show that community-wide synchrony  $\varphi$  can be quantified by computing the temporal variance  $\sigma_{x_T}^2$  of the community time series  $x_T(t) = \sum x_i(t)$  and the sum of the temporal standard deviation of the time series across all species  $(\sum \sigma_{x_i})^2$  such that:  $\varphi = \frac{\sigma_{x_T}^2}{(\sum \sigma_{x_i})^2}$ 

$$\varphi = \frac{\sigma_{x_T}}{\left(\sum \sigma_{x_i}\right)^2}$$

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#### Value

Returns a named list containing:

obs the observed community synchrony

meancorr the mean correlation between the columns of the matrix

rands the community synchrony value the randomizations. This variable is only re-

turned if nrands > 0.

pval p-value of observed community synchrony. This variable is only returned if

nrands > 0.

### Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

#### References

Loreau, M., and C. de Mazancourt. 2008. Species synchrony and its drivers: Neutral and nonneutral community dynamics in fluctuating environments. *The American Naturalist* 172:E48-E66.

# **Examples**

```
## Community matrix for 20 species undergoing random fluctuations
comm.rand=matrix(runif(100), nrow=5, ncol=20)
community.sync(comm.rand, nrands=20)$pval
## Community matrix for 20 species undergoing synchronized fluctuations
comm.corr=matrix(rep(comm.rand[,1], 20), nrow=5, ncol=20)
community.sync(comm.corr, nrands=20)$pval
## On "real" data
data(bird.traits)
community.sync(bird.traits, nrands=20)$pval
```

coord2dist coord2dist

# **Description**

Calculate distance between all pairs of sites

### Usage

```
coord2dist (coords, is.latlon = TRUE, lower.tri = TRUE)
```

# **Arguments**

coords n x 4 matrix of coordinates consisting of lat or y, lon or x pairs for each each site

is.latlon are coordinates latitudes/longitudes? Default is TRUE

lower.tri Return lower triangular part of the distance matrix? Default is TRUE

#### Value

Returns the distance between all pairs of sites

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#### Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

# **Examples**

```
coords=rbind(c(32, -125), c(43, -130))
## Compute great circle distance
coord2dist(coords)
```

correlated.matrix

correlated.matrix

# **Description**

Create an ntimes x nspecies matrix with correlation  $\rho$ , standard deviation  $\sigma$ , and mean  $\mu$ 

# Usage

```
correlated.matrix (rho = 0, sigma = 1, mu = 0, ntimes = 200, nspecies = 10)
```

# **Arguments**

rho Correlation between the columns of the matrix. This can be a single number

describing the correlation between all columns, or the upper triangular portion of a correlation matrix describing the correlation between all pairs of columns.

Default is 0

sigma Standard deviation of the columns. Default is 1

mu Mean of the columns. Default is 0

ntimes Number of rows in the matrix. Default is 200 nspecies Number of columns in the matrix. Default is 10

# **Details**

This function is based on the Cholesky factorization method described by Legendre (2000).

#### Value

Returns a named list containing the following:

rho Correlation(s) between the columns sigma Standard deviation of the columns

mu Mean of the columns

community ntimes x nspecies matrix

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

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#### References

Gouhier, T. C., F. Guichard, and A. Gonzalez. 2010. Synchrony and stability of food webs in metacommunities. *The American Naturalist* 175:E16-E34.

Legendre, P. 2000. Comparison of permutation methods for the partial correlation and partial mantel tests. *Journal of Statistical Computation and Simulation* 67:37-73.

# **Examples**

```
mat=correlated.matrix(rho=0.85, sigma=30, mu=10, nspecies=10)
## Check sd of each column
apply(mat$community, 2, sd)
## Check mean of each column
apply(mat$community, 2, mean)
## Check correlation of matrix
community.sync(mat$community)
```

find.minmax

Find min/max of a time series

# **Description**

Find local minima and maxima of a time series

#### Usage

```
find.minmax (timeseries)
```

# **Arguments**

timeseries

time series in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.

# Value

Returns a named list containing:

mins n x 3 matrix containing the index, time steps, and the local min values maxs n x 3 matrix containing the index, time steps, and the local max values

#### Author(s)

```
Tarik C. Gouhier (tarik.gouhier@gmail.com)
```

```
t1=cbind(1:100, runif(100))
min.max=find.minmax(t1)
min.max$maxs
plot (t1, t="1")
points (min.max$mins, col="blue", bg="blue", pch=19)
points (min.max$maxs, col="red", bg="red", pch=19)
```

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# **Description**

Compute Kendall's coefficient of concordance (W)

#### Usage

```
kendall.w (data, nrands = 0)
```

### Arguments

data matrix in wide format where each row represents a different sample and each

column represents a different variable.

nrands Number of randomizations to perform to determine significance. Default is 0.

#### **Details**

Kendall's W is a non-parametric statistic that ranges from 0 to 1 and measures the level of agreement between multiple variables. When the number of observations n > 10, its significance can be determined by using a  $\chi^2$  distribution with df = n - 1. Legendre (2005) shows that the  $\chi^2$  test is always too conservative (low power) compared to the randomization test. Hence, both tests have been made available in this function. The Monte Carlo randomizations are performed by shuffling the columns of the community matrix independently (Legendre 2005).

#### Value

Returns a named list containing:

w.uncorrected Kendall's W uncorrected for tied ranks
w.corrected Kendall's W corrected for tied ranks

pval p-value of Kendall's W

spearman.corr Spearman's ranked correlation

p-value of Kendall's W based on randomization test. This variable is only re-

turned if nrands > 0

rands randomizations. This variable is only returned if nrands > 0

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

# References

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

Gouhier, T. C., and F. Guichard. 2007. Local disturbance cycles and the maintenance of spatial heterogeneity across scales in marine metapopulations. *Ecology* 88:647-657.

Gouhier, T. C., F. Guichard, and A. Gonzalez. 2010. Synchrony and stability of food webs in metacommunities. *The American Naturalist* 175:E16-E34.

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Legendre, P. 2005. Species associations: the Kendall coefficient of concordance revisited. *Journal of Agricultural, Biological, and Environmental Statistics* 10:226-245.

Zar, J. H. 1999. Biostatistical Analysis, Fourth edition. Prentice-Hall, Inc., Upper Saddle River, NJ.

# **Examples**

```
data(bird.traits)
(w=kendall.w(bird.traits))
```

latlon2dist

latlon2dist

# Description

Calculate distance between a pair of coordinates

# Usage

```
latlon2dist (coords)
```

# **Arguments**

coords

4-element vector of coordinates with format: (lat1, lon1, lat2, lon2)

# Value

Returns the great circle distance distance between the pair of coordinates

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

# See Also

coord2dist

```
coords=c(32, -125, 43, -130)
## Compute great circle distance
latlon2dist(coords)
```

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meancorr	Compute mean column-wise correlation and determine its significance via Monte Carlo randomizations

#### **Description**

Compute mean column-wise correlation and determine its significance via Monte Carlo randomizations. The Monte Carlo randomizations are performed by shuffling the columns of the community matrix independently.

#### Usage

#### **Arguments**

comm.matrix community matrix in wide format where each row contains the abundance at

each time step and each column corresponds to a different species.

nrands number of randomizations to perform (default is 0)

alternative Conduct a one-tailed or a two-tailed test? Default is one. tailed.

method Method to compute correlation? Options include pearson, kendall, and spearman.

Default is pearson

... Other parameters to cor function.

#### Value

Returns a named list containing:

obs the observed mean correlation

rands the mean correlation for each randomization. This variable is only returned if

nrands > 0.

p-value of observed mean correlation. This variable is only returned if nrands > 0.

alternative One-tailed or two-tailed test? This variable is only returned if nrands > 0.

method Method used to compute the mean correlation.

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

```
## Community matrix for 20 species undergoing random fluctuations
comm.rand=matrix(runif(100), nrow=5, ncol=20)
meancorr(comm.rand, nrands=20)$pval
## Community matrix for 20 species undergoing synchronized fluctuations
comm.corr=matrix(rep(comm.rand[,1], 20), nrow=5, ncol=20)
meancorr(comm.corr, nrands=20)$pval
## On "real" data
data(bird.traits)
meancorr(bird.traits, nrands=20)$pval
```

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peaks	Find the proportion of local minima/maxima common to both time series and compute its significance via Monte Carlo randomizations

# Description

Find the proportion of local minima/maxima common to both time series and compute its significance via Monte Carlo randomizations

# Usage

```
peaks (t1, t2, nrands = 0)
```

# **Arguments**

t1	time series 1 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
t2	time series 2 in matrix format (n rows $x$ 2 columns). The first column should contain the time steps and the second column should contain the values.
nrands	number of randomizations. Default is 0.

# Value

Returns a named list containing:

pval	p-value computed by randomly shuffling both time series nrands times
rands	proportion of local minima/maxima common to both time series for each randomization
peaks	proportion of local minima/maxima common to both time series in the observed dataset
locations	locations/indices of local minima/maxima common to both time series in the observed dataset

# Author(s)

```
Tarik C. Gouhier (tarik.gouhier@gmail.com)
```

# References

Buonaccorsi, J. P., J. S. Elkinton, S. R. Evans, and A. M. Liebhold. 2001. Measuring and testing for spatial synchrony. *Ecology* 82:1668-1679.

```
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
(p=peaks(t1, t2))
```

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Phase partnered time series

# **Description**

Create two time series with specific autocorrelation  $\gamma$ , cross-correlation  $\rho$ , mean ts.mean, and standard deviation ts.sd using the phase partnered algorithm described by Vasseur (2007)

# Usage

```
phase.partnered (n = 2000, rho = 1, gamma = 1, ts.sd = 0.1, ts.mean = 0)
```

### **Arguments**

n	number of time steps in time series. Default is 2000.
rho	cross-correlation between the two time series ( $-1 \le \rho \le 1$ ). Default is 1.
gamma	autocorrelation of each time series. Gamma $(\gamma)$ describes the relationship between frequency $f$ and power $P$ : $P(f)=1/f^\gamma$
	$-2 \le \gamma \le 0$ : blue noise and $0 \le \gamma \le 2$ : red noise. Default is 1.
ts.sd	standard deviation of both time series. Default is 0.1.
ts.mean	mean of both time series. Default is 0.

#### Value

Returns an n x 2 matrix containing the time series

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

#### References

Gouhier, T. C., F. Guichard, and A. Gonzalez. 2010. Synchrony and stability of food webs in metacommunities. *The American Naturalist* 175:E16-E34.

Vasseur, D. A. 2007. Environmental colour intensifies the Moran effect when population dynamics are spatially heterogeneous. *Oikos* 116:1726-1736.

```
## Positively cross-correlated white noise
pos.corr=phase.partnered(n = 100, rho = 0.7, gamma = 0)
## Negatively cross-correlated white noise
neg.corr=phase.partnered(n = 100, rho = -1, gamma = 0)
par(mfrow=c(2,1))
matplot (pos.corr, t="l", lty=1)
matplot (neg.corr, t="l", lty=1)
```

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phase.sync	Phase synchrony of quasi-periodic time series

# **Description**

Compute the phase synchrony between two quasi-periodic time series by quantifying their phase difference at each time step

#### Usage

```
phase.sync (t1, t2, nrands = 0, type = 1, nbreaks = 10, mins = FALSE)
```

# **Arguments**

t1	time series 1 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
t2	time series 2 in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
nrands	number of randomizations to perform (default is 0)
type	flag to indicate whether to compute phase difference modulus $2\pi$ between 0 and $2\pi$ (type=1) or phase difference modulus $2\pi$ between $-\pi$ and $\pi$ . Default is 1.
nbreaks	number of bins to use to group the values in the time series. Default is 10.
mins	use local minima instead of local maxima to compute and the interpolate the phase. Default is FALSE.

#### **Details**

Two time series are phase-locked if the relationship between their phases remains constant over time. This function computes the phase of successive local maxima or minima for each time series and then uses linear interpolation to find the phase at time steps that fall between local maxima/minima. A histogram can be used to determine if the distribution of the phase difference at each time step is uniform (indicating no phase locking) or has a clear peak (indicating phase locking).

# Value

Returns a list containing phases1, phases2, and deltaphase

phases1 n x 3 matrix containing the timestep, value, and phase of the first time series n x 3 matrix containing the timestep, value, and phase of the second time series deltaphase n x 4 matrix containing the timestep, raw phase difference, phase difference modulus  $2\pi$  between 0 and  $2\pi$ , phase difference modulus  $2\pi$  between  $-\pi$  and  $\pi$ 

#### Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

# References

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

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#### **Examples**

```
t1=cbind(1:100, runif(100))
t2=cbind(1:100, runif(100))
## Compute and interpolate phases using successive local minima
sync.mins=phase.sync(t1, t2, mins=TRUE)
## Compute and interpolate phases using successive local maxima
sync.maxs=phase.sync(t1, t2)
## Plot distribution of phase difference
hist(sync.mins$deltaphase$mod_phase_diff_2pi)
```

pisco.data

PISCO multi-year and spatially-explicit mussel and environmental dataset

# Description

Contains the mean annual chl-a concentration, sea surface temperature, upwelling currents, and mussel abundance at 48 intertidal sites along the West Coast of the United States from 2000-2003.

#### Usage

```
data(pisco.data)
```

### **Format**

A data frame with 192 observations on the following 7 variables.

```
latitude latitude (degrees North)

longitude longitude (degrees West)

chl mean annual remote sensed chlorophyll-a concentration

sst mean annual remote sensed sea surface temperature

upwelling mean annual remote sensed upwelling currents

mussel_abund mean annual mussel cover (Mytilus californianus)

year sampling year
```

# References

Gouhier, T. C., F. Guichard, and B. A. Menge. 2010. Ecological processes can synchronize marine population dynamics over continental scales. *Proceedings of the National Academy of Sciences* 107:8281-8286.

```
data(pisco.data)
```

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# Description

Plot vario objects

# Usage

# **Arguments**

X	vario object generated by vario function.
xlab	xlabel of the figure. Default is "Lag distance"
ylab	ylabel of the figure. Default is NULL and will automatically generate the right label
ylim	y-range. Default is NULL and will automatically generate the best range based on the metric
xtype	Use either the discrete bin classes (bins) or the mean distance of the points within each bin (mean.bin.dist) on the x-axis. Default is mean.bin.dist
rug	Plot rug indicating the density of data points? Default is FALSE
ci	Plot two-tailed $(1-\alpha)\%$ confidence intervals? Default is FALSE
pch	Type of points to use when plotting the variogram. Default is 21
col.sig	Border color of points for significant values. Default is black
col.nonsig	Border color of points for non-significant values. Default is black
bg.sig	Background color of points for significant values. Default is black
bg.nonsig	Background color of points for non-significant values. Default is black
alpha	Significance level. Default is 0.05
	other graphical parameters.

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

```
data(pisco.data)
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "sst"))
semiv=vario(data=d)
moran=vario(data=d, type="moran", nrand=100)
geary=vario(data=d, type="geary", nrand=100)
par(mfrow=c(3,1))
```

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```
plot(semiv)
plot(moran, bg.sig="blue")
plot(geary, bg.sig="red")
```

plot.variofit

Plot variofit objects

# Description

Plot variofit objects

# Usage

# **Arguments**

x	variofit object generated by vario.fit function
xlab	xlabel of the figure. Default is "Lag distance"
ylab	ylabel of the figure. Default is "Variogram"
col.pts	Border color of the points. Default is black
col.line	Color of the fitted variogram. Default is red
pch	Type of points to use when plotting the variogram. Default is 21
	other graphical parameters.

# Author(s)

```
Tarik C. Gouhier (tarik.gouhier@gmail.com)
```

```
## Environmental variogram
data(pisco.data)
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "upwelling"))
semiv=vario(data=d)
mod.sph=vario.fit(semiv$vario, semiv$mean.bin.dist)
plot(mod.sph)
```

surrogate.ts 17

#### **Description**

Create surrogate time series with the same short-term time correlation and overall temporal pattern as the original time series using the Markov process described by Cazelles and Stones (2003)

# Usage

```
surrogate.ts (ts, distr.ts = NULL, trans.ts = NULL, nbreaks = 10)
```

# **Arguments**

ts	time series in matrix format (n rows x 2 columns). The first column should contain the time steps and the second column should contain the values.
distr.ts	binning of time series values. This parameter must be specified if trans.ts is not set to NULL. Default is NULL.
trans.ts	transition matrix from bin $i$ to bin $j$ . Default is NULL.
nbreaks	number of bins to use to group the time series values. Default is 10.

#### **Details**

The values of the time series  $x_n$  are grouped into nbreak equally-sized bins. The transition matrix  $M_{ij}$  describing the probability of  $x_{n+1}$  belonging to bin j based on  $x_n$  belonging to bin i is defined using the relative frequencies of the data such that:  $M_{ij} = Pr(x_{n+1} \in b_j | x_n \in b_i)$ . The surrogate time series is then constructed by randomly selecting a starting value and randomly selecting the next value from the proper bin based on the transition matrix. This process is repeated until the surrogate time series has the same length as the original time series.

#### Value

Returns a named list containing:

surr.ts surrogate time series in matrix format trans transition matrix  $M_{ij}$ 

distr binning of time series values

#### Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

#### References

Cazelles, B., and L. Stone. 2003. Detection of imperfect population synchrony in an uncertain world. *Journal of Animal Ecology* 72:953-968.

#### See Also

```
phase.sync
```

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# **Examples**

```
t1=cbind(1:100, runif(100))
surr.t1=surrogate.ts(ts=t1)
plot(t1, t="l")
lines(surr.t1$surr.ts, col="red")
```

vario vario

# Description

Compute the empirical variogram and determine its significance via Monte Carlo randomizations

# Usage

# **Arguments**

nbins	Number of bins or lags
extent	Proportion of the spatial extent of the data over which to compute the variogram. Default is 0.5 to limit potentially spurious results due to the limited number of data points at large lag distances.
data	n x m matrix containing y-coordinates (or latitude), x-coordinates (or longitude), and values. The values can either be a single column of observations at each site for univariate variograms or a matrix of observations at each site for multivariate variograms (e.g., to compute spatial synchrony).
data2	n x m matrix containing y-coordinates (or latitude), x-coordinates (or longitude), and values for second variable. The values can either be a single column of observations at each site for univariate variograms or a matrix of observations at each site for multivariate variograms (e.g., to compute spatial synchrony).
is.latlon	Are coordinates latitudes/longitudes? Default is TRUE
centered	Should the variogram be centered by subtracting the regional mean from each value? If so, the zero-line represents the regional mean. Default is FALSE
nrands	Number of randomizations to determine statistical significance of variogram. Default is 0.
type	Type of variogram to compute. Default is semivar for semivariance. Other options include cov for covariance, pearson for Pearson correlation, spearman for Spearman correlation, kendall for Kendall correlation, moran for Moran's I, and geary for Geary's C
alternative	Conduct a one-tailed or a two-tailed test? Note that the statistical test is to determine whether the local value within each lag distance is different from the regional mean. If the variogram is centered, the null hypothesis is that the local values are equal to zero. If the variogram is not centered, the null hypothesis is

that the local values are equal to the regional mean. Default is one.tailed

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mult.test.corr Correct for multiple tests? Default is "none". Other options include holm, hochberg, sidak, and bonferroni

#### **Details**

This function can be used to compute univariate correlograms using Moran's I, Geary's C, and the covariance function or variograms using the semivariance function. Multivariate (Mantel) correlograms can also be computed using the covariance function, Pearson's, Spearman's or Kendall's correlation coefficients. Cross-correlograms/variograms between data1 and data2 can be computed with the covariance function, Pearson's, Spearman's or Kendall's correlation coefficients for multivariate variograms and Moran's I, Geary's C, the covariance function, or semivariance for univariate variograms.

#### Value

Returns a named list containing the following variables:

bins Center of each lag/bin

mean.bin.dist Mean distance of each lag/bin vario Variogram values in each lag/bin

npoints Number of pairs of points in each lag/bin

metric Type of variogram computed centered Is the variogram centered?

regional.mean Regional mean value

p-value for each lag/bin. This variable is only returned if nrands > 0.

rands nrands x nbins matrix of randomizations. This variable is only returned if

nrands > 0.

alternative One-tailed or two-tailed test? This variable is only returned if nrands > 0.

mult.test.corr

Correct for multiple tests? This variable is only returned if nrands > 0.

is.multivar Was the analysis performed on multivariate data?

# Author(s)

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#### References

Bjornstad, O. N., and W. Falck. 2001. Nonparametric spatial covariance functions: Estimation and testing. *Environmental and Ecological Statistics* 8:53-70.

Bjornstad, O. N., R. A. Ims, and X. Lambin. 1999. Spatial population dynamics: analyzing patterns and processes of population synchrony. *Trends in Ecology & Evolution* 14:427-432.

Fortin, M. J., and M. R. T. Dale. 2005. Spatial Analysis: A Guide for Ecologists. Cambridge University Press.

# See Also

vario.func

20 vario.fit

#### **Examples**

```
data(pisco.data)
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "sst"))
semiv=vario(data=d)
moran=vario(data=d, type="moran", nrands=100)
par(mfrow=c(2,1), mar=c(4.2, 4, 1, 1))
plot(semiv$mean.bin.dist, semiv$vario, xlab="Lag distance (km)", ylab="Semivariance")
plot(moran$mean.bin.dist, moran$vario, xlab="Lag distance (km)", ylab="Moran's I", t="l")
points(moran\$mean.bin.dist[moran\$pvals >= 0.05], moran\$vario[moran\$pvals >= 0.05],
       bg="white", pch=21)
points(moran$mean.bin.dist[moran$pvals < 0.05], moran$vario[moran$pvals < 0.05],</pre>
       bg="black", pch=21)
abline(h=0, lty=2)
## Compute spatial synchrony
d.upw=subset(pisco.data, select=c("latitude", "longitude", "year", "upwelling"))
d.cov=subset(pisco.data, select=c("latitude", "longitude", "year", "mussel_abund"))
## Reshape the data
d.upw.wide=reshape(data=d.upw, idvar=c("latitude", "longitude"), timevar=c("year"),
                    direction="wide")
d.cov.wide=reshape(data=d.cov, idvar=c("latitude", "longitude"), timevar=c("year"),
                    direction="wide")
## Generate variograms
v.upw=vario(nbins=12, data=d.upw.wide, type="pearson", extent=1, nrands=999)
v.cov=vario(nbins=12, data=d.cov.wide, type="pearson", extent=1, nrands=999)
## Fit variograms
v.cov.per=vario.fit(v.cov$vario, v.cov$mean.bin.dist, type="period",
                     start.vals=list(a=1, b=3, c=0))
v.upw.lin=vario.fit(v.upw$vario, v.upw$mean.bin.dist, type="linear")
par(mfrow=c(2,1))
plot(v.cov, xlab="Lag distance (km)", bg.sig="red", col.nonsig="red",
     main="Mussel cover",
     rug=TRUE, ylim=c(-0.3, 0.3))
lines(v.cov$mean.bin.dist, v.cov.per$fit, col="red")
plot(v.upw, xlab="Lag distance (km)", bg.sig="blue", col.nonsig="blue",
     main="Upwelling", rug=TRUE)
lines(v.upw$mean.bin.dist, v.upw.lin$fit, col="blue")
```

vario.fit

vario.fit

#### **Description**

Fit model to the empirical variogram

#### Usage

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#### **Arguments**

vario Empirical variogram from emp. vario function

bins Number of bins or lag distances from emp. vario function

weights Vector of weights of the same length as vario. If weights is a vector containing

> the number of points in each distance bin, the model will be fit via weighted least squares with the weights corresponding to the proportion of points within each

bin (i.e., weights sum to 1). Default is a vector of weights equal to 1

type Type of variogram model to fit to the data. Default is spherical. Other options

are gaussian, nugget, linear, exponential, sill, periodic, and hole

start.vals Named list containing the start values for the variogram model: c0: nugget, c1:

sill, a: spatial range; b: slope; c: frequency

#### Value

Return a named list containing the following variables:

Empirical variogram values vario

bins Empirical variogram bins/lag distances

AIC score of the model fit:  $AIC = n \times log\left(\frac{SSE}{n}\right) + 2 \times (p+1)$  where n is the number of points in the variogram,  $SSE = \sum (\hat{x}_i - x_i)^2$ , and p is the number AIC

of parameters

Root Mean Square Error of the model fit:  $\sqrt{\frac{SSE}{n}}$ **RMSE** 

Named list containing the best model parameter estimates params

fit Predicted variogram values from the model fit

nls.success did nls succeed?

# Note

Selecting proper initial values is critical for fitting a reasonable model to the empirical variogram. If these values are off, nls will fail and fall-back functions will be used to determine the best parameter values that minimize the Root Mean Square Error (RMSE).

#### Author(s)

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#### See Also

```
vario, vario.func
```

```
## Load data
data(pisco.data)
## Environmental variogram
d=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "upwelling"))
semiv=vario(data=d)
plot(semiv, xlab="Lag distance (km)")
mod.sph=vario.fit(semiv$vario, semiv$mean.bin.dist)
## Weighted least squares fit based on the number of points
```

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```
mod.exp=vario.fit(semiv$vario, semiv$mean.bin.dist,
                                         weights=semiv$npoints/sum(semiv$npoints),
                                         type="expo")
mod.gau=vario.fit(semiv$vario, semiv$mean.bin.dist, type="gauss")
mod.lin=vario.fit(semiv$vario, semiv$mean.bin.dist, type="lin")
lines(semiv$mean.bin.dist, mod.sph$fit, col="red")
lines(semiv$mean.bin.dist, mod.exp$fit, col="black")
lines(semiv$mean.bin.dist, mod.gau$fit, col="blue")
lines(semiv$mean.bin.dist, mod.lin$fit, col="green")
\label{legend} legend(x="topleft", legend=paste(c("Spherical AIC:", "Exponential AIC
                                                                                "Gaussian AIC:", "Linear AIC:"),
                                                                                c(format(mod.sph$AIC, dig=2),
                                                                                format(mod.exp$AIC, dig=2),
                                                                                format(mod.gau$AIC, dig=2),
                format(mod.lin$AIC, dig=2))), lty=1, col=c("red", "black", "blue", "green"),
                bty="n")
## Correlogram
cover=subset(pisco.data, subset=year==2000, select=c("latitude", "longitude", "mussel_abund"))
moran=vario(data=cover, type="moran")
mod.hol=vario.fit(moran$vario, moran$mean.bin.dist, type="hole", start.vals=list(c0=0.6, a=25, c1=0.01))
mod.per=vario.fit(moran$vario, moran$mean.bin.dist, type="period", start.vals=list(a=1, b=3, c=0))
mod.lin=vario.fit(moran$vario, moran$mean.bin.dist, type="linear")
plot(moran, xlab="Lag distance (km)", ylim=c(-0.6, 0.8))
lines(moran$mean.bin.dist, mod.per$fit, col="red")
lines(moran$mean.bin.dist, mod.hol$fit, col="black")
lines(moran$mean.bin.dist, mod.lin$fit, col="blue")
legend(x="topleft", legend=paste(c("Periodic AIC:", "Hole AIC:",
                                                                                "Linear AIC:"),
                                                                               c(format(mod.per$AIC, dig=2),
                                                                                format(mod.hol$AIC, dig=2),
                                                                                format(mod.lin$AIC, dig=2))),
                                                                                lty=1, col=c("red", "black", "blue"), bty="n")
```

vario.func

vario.func

# Description

Compute the empirical variogram values for each bin

### Usage

# Arguments

x First set of sites within bin/lag distancey Second set of sites within bin/lag distanceglob.mean Global mean

glob.sd Global standard deviation

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glob.N Global number of points

is.multivar Is the data multivariate? Default is FALSE

type Type of variogram to compute. Default is semivar for semivariance. Other

options include cov for covariance, pearson for Pearson correlation, spearman for Spearman correlation, kendall for Kendall correlation, moran for Moran's

I, and geary for Geary's C

# Value

Return the value.

# Author(s)

Tarik C. Gouhier (tarik.gouhier@gmail.com)

# See Also

vario

# **Examples**

## Internal function used by vario

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