

COS-D419 Factor Analysis and Structural Equation Models 2023, Assignment 4

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The texts that reflect my understanding have been highlighted in red color.

1 Task description

The first section is task description, which is copied from the assignment5.rmd. It is for communicating with future “me”. Please skip it.

1.1 Exercise 5.1

Specify and estimate the initial baseline models for the two groups.

Present a brief summary of the model fit and make the first step of the modification by including (**exceptionally, at the same time!**) all the four parameters known to be required for improving the model fit of both models.

Fine-tune the models step by step following the guidelines given in the lecture material, i.e., implement the modifications (**as usually, one change at a time**) testing and studying each step.

Present the final baseline models of each group and draw the graphs

2 Preparation

2.1 Read in the data set:

Start by downloading the **two data files** from Moodle to your Project folder! xie

```
#install the necessary packages
if (!require("pacman")) install.packages("pacman")
pacman::p_load(
  expss,
  tidyverse,
  janitor,
  knitr,
  qualtrics,
  arules,
  arulesViz,
  sjlabelled,
  DT,
  stringr,
  labelled,
  ggstatsplot,
  ggcorplot)

library(tidyverse)
library(readr)

#This week's file name
latest.name1 <- "MBIELM1.CSV"
latest.name2 <- "MBISEC1.CSV"
#read in the data
mbi.elm <- #elementary school
  read_csv(
    file.path(
      here(),
      'data',
      latest.name1
    )
  )

mbi.sec <- #secondary school
  read_csv(
    file.path(
      here(),
      'data',
      latest.name2
    )
  )
```

2.2 Write functions

To control length of reports, codes already shown in the previous homework were not showing in the current report. Yet they are available in .rmd report.

2.2.1 To generate a function for calculating chi square difference was defined.

2.2.2 to generate CFA results with improved readability

2.2.3 Write a function to simplify plotting of merged tables for multi-group fit indices

```
multi.fit.tab <- function(data, title, more.footnote = NULL){
  data <- data |>
  rename(p = 'p value',
         p2 = 'RMSEA p value',
         chi = 'chi square') |>
  mutate(df = as.numeric(df) |> round(0),
         p = case_when(
           as.numeric(p) < 0.001 ~ "<0.001",
           as.numeric(p) >= 0.001 ~ p
         ),
         p2 = case_when(
           as.numeric(p2) < 0.001 ~ "<0.001",
           as.numeric(p2) >= 0.001 ~ p2
         ) |>
  mutate('Chi square (df, p)' =
    paste0(chi, "(", df, ", ", p, ")"),
         'RMSEA(p)' =
    paste0(RMSEA, "(", p2, ")")
  ) |>
  select(
    Model,
    'Chi square (df, p)',
    CFI, TLI,
    'RMSEA(p)',
    SRMR,
    'CSF*' = CSF
  )
  #print the combined table with adjustment of aesthetics
  data |>
  kable(booktabs = T,
        #format = "markdown",
        caption =
          title,
        align = "lrrrrrr"
  ) |>
  kable_styling(full_width = T) |>
  footnote(symbol =
    c("Chi square scaling factor",
      more.footnote)
```

```

    ) |>
column_spec(1, width = "3.5cm") |>
column_spec(2, width = "4cm") |>
column_spec(3, width = "1cm") |>
column_spec(4, width = "1cm") |>
column_spec(5, width = "2.5cm") |>
column_spec(6, width = "1cm") |>
column_spec(7, width = "1cm")
}

```

2.2.4 Write a function to simplify plotting of merged tables for multi-group fit indices with chi square difference statistics

```

delta.fit.tab <- function(data, title, more.footnote = NULL){
  data$'ΔChi-square(df,p)*' <- rep(NA, nrow(data))
  for (i in 2:nrow(data)){
    nested <- paste0("inv", i, ".fit")
    diff<- as.numeric(chisq_mlm(eval(parse(text = nested)), inv1.fit)[1])
    diff.df <- as.numeric(chisq_mlm(eval(parse(text = nested)), inv1.fit)[2])
    diff.p<- as.numeric(chisq_mlm(eval(parse(text = nested)), inv1.fit)[3])
    data$'ΔChi-square(df,p)*'[i] <- paste0(diff, "(", diff.df, ", ", diff.p, ")")
  }

data <- data |>
  rename(p = 'p value',
         p2 = 'RMSEA p value',
         chi = 'chi square') |>
  mutate(df = as.numeric(df) |> round(0),
         p = case_when(
           as.numeric(p) < 0.001 ~ "<0.001",
           as.numeric(p) >= 0.001 ~ p
         ),
         p2 = case_when(
           as.numeric(p2) < 0.001 ~ "<0.001",
           as.numeric(p2) >= 0.001 ~ p2
         )
  ) |>
  mutate('Chi square (df, p)' =
    paste0(chi, "(", df, ", ", p, ")"),
         'RMSEA(p)' =
    paste0(RMSEA, "(", p2, ")")
  )
  ) |>
  select(
    Model,
    'Chi square (df, p)',
    'ΔChi-square(df,p)*',
    CFI, TLI,
    'RMSEA(p)',
    SRMR
  )

```

```

data[1,3] <- "--"
#print the combined table with adjustment of aesthetics
data |>
  kable(booktabs = T,
        #format = "markdown",
        caption =
          title,
        align = "lrrrrrr"
        ) |>
  kable_styling(full_width = T) |>
  footnote(symbol =
    c("Chi square difference statistics of model of the row compared to configural model",
      more.footnote)
    ) |>
  column_spec(1, width = "2.8cm") |>
  column_spec(2, width = "3.5cm") |>
  column_spec(3, width = "3cm") |>
  column_spec(4, width = "1cm") |>
  column_spec(5, width = "1cm") |>
  column_spec(6, width = "2.2cm") |>
  column_spec(7, width = "1cm")
}

```

2.2.5 Write a function to simplify plotting aligned residual variance and co-variance tables

```

align.table <- function(data, num.no.header.col, title){
  data |>
    kable(
      digits = 3,
      booktabs = T,
      #format = "markdown",
      caption = title,
      linesep = ""
    ) |>
    add_header_above(c(" " = num.no.header.col,
                       "Elementary level" = 5,
                       "Secondary level" = 5
                       )
    ) |>
    kable_styling(
      latex_options = "striped"
    ) |>
    footnote(
      symbol = c(
        "Un-standardized estimates",
        "Standardized estimates"
      )
    )
}

```

2.2.6 Write a function for correlation matrix with numbers

2.2.7 to generate a function for histogram overlapping with density plot

2.2.8 to generate a function for violin overlapping with box plot

2.2.9 To generate a function describing continuous data set

2.2.10 Write a function describing continuous data set

2.2.11 Write a function for histogram overlapping with density plot

2.2.12 Write a function to generate dot distribution plot

```
dot.dist <-  
function(data, type, title){  
  data |>  
  t() |>  
  as.data.frame() %>%  
  mutate(Item = rownames(.)) |>  
  rowwise() |>  
  mutate(Median = eval(parse(text = type))(V1:V580)) |>  
  ggstatsplot::ggdotplotstats(  
    point.args = list(color = "red", size = 3, shape = 13),  
    xlab = paste(type, "ratings"),  
    title = title,  
    x = Median,  
    y = Item  
  )  
}
```

2.2.13 Write a function to generate correlation matrix with statistical test

```
mycor <-  
function(data, cols, title){  
  mbi.elm |>  
  select(all_of(cols)) |>  
  ggstatsplot::ggcorrmat(  
    colors = c("#B2182B", "white", "#4D4D4D"),  
    title = "(a) Items on emotional exhaustion,  
    elementary school teacher",  
    matrix.type = "lower"  
  )  
}
```

3 Inspect the data

3.1 Distribution

```
#generate the plots, by subgroup of teachers
p.dist.elm <-
  corr.density(
    mbi.elm,
    fig.num = "1(a)",
    group = "elementary school teacher"
  )
p.dist.sec <-
  corr.density(
    mbi.sec,
    fig.num = "1(b)",
    group = "secondary school teacher"
  )
#print the plot
library(patchwork); p.dist.elm/p.dist.sec
```

Figure 1(a) Distribution of selected items for elementary school teacher

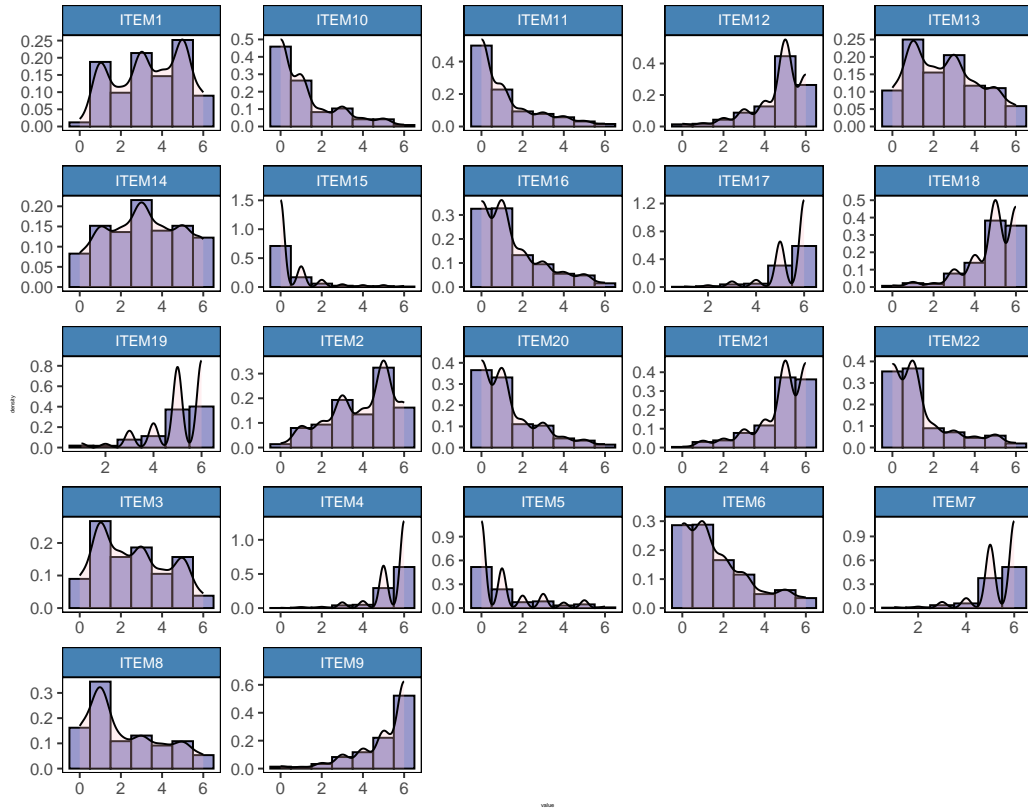
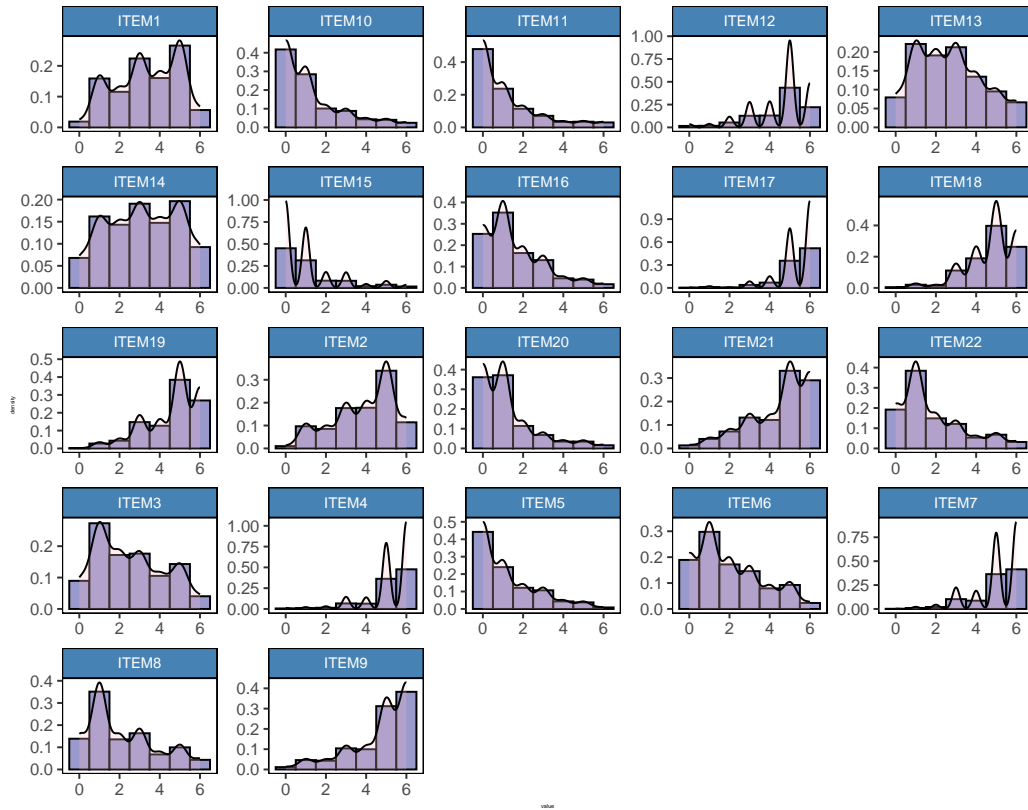


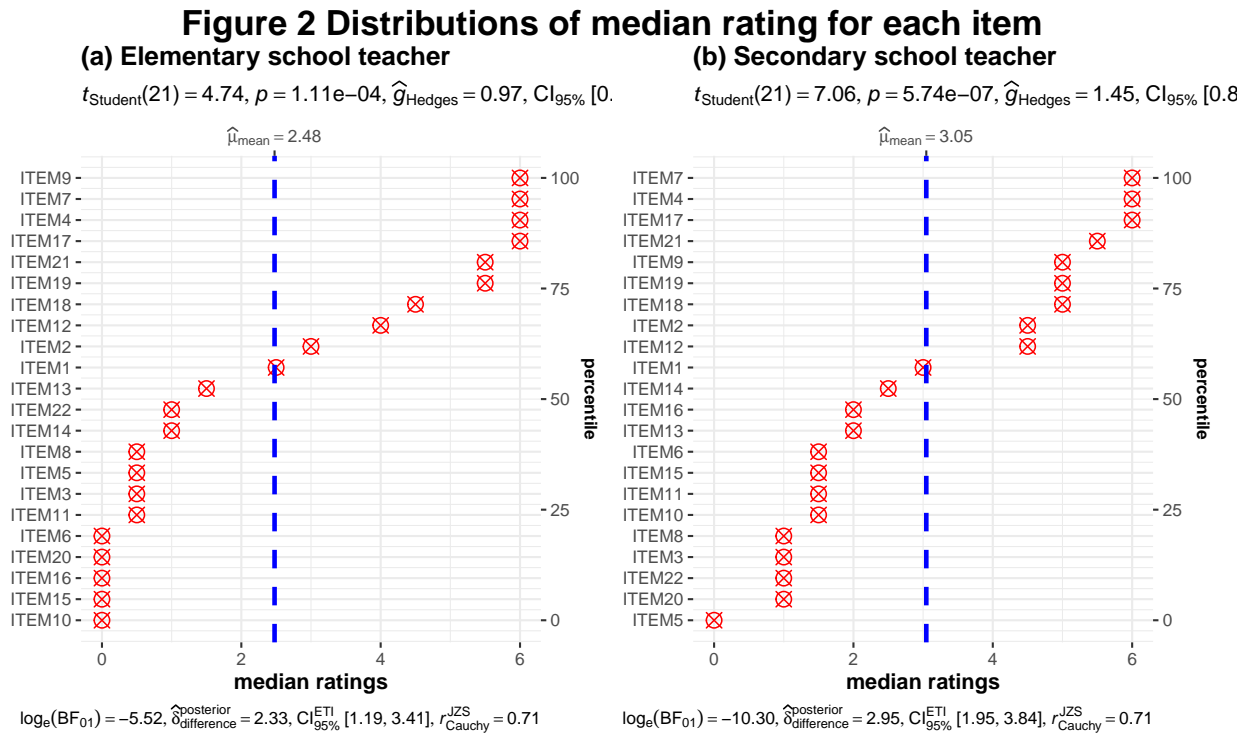
Figure 1(b) Distribution of selected items for secondary school teacher




```

#generate plot by subgroups of teachers
p.dot.elm <-
  dot.dist(
    data = mbi.elm,
    type = "median",
    title = "(a) Elementary school teacher"
  )
p.dot.sec <-
  dot.dist(
    data = mbi.sec,
    type = "median",
    title = "(b) Secondary school teacher"
  )
#plot layout
patchwork <- p.dot.elm|p.dot.sec
#print the plot with a genral title
patchwork+plot_annotation(
  title =
    'Figure 2 Distributions of median rating for each item',
  theme =
    theme(plot.title =
      element_text(
        size = 16,
        face = "bold",
        vjust = -1.5,
        hjust = 0.5)
    )
)

```



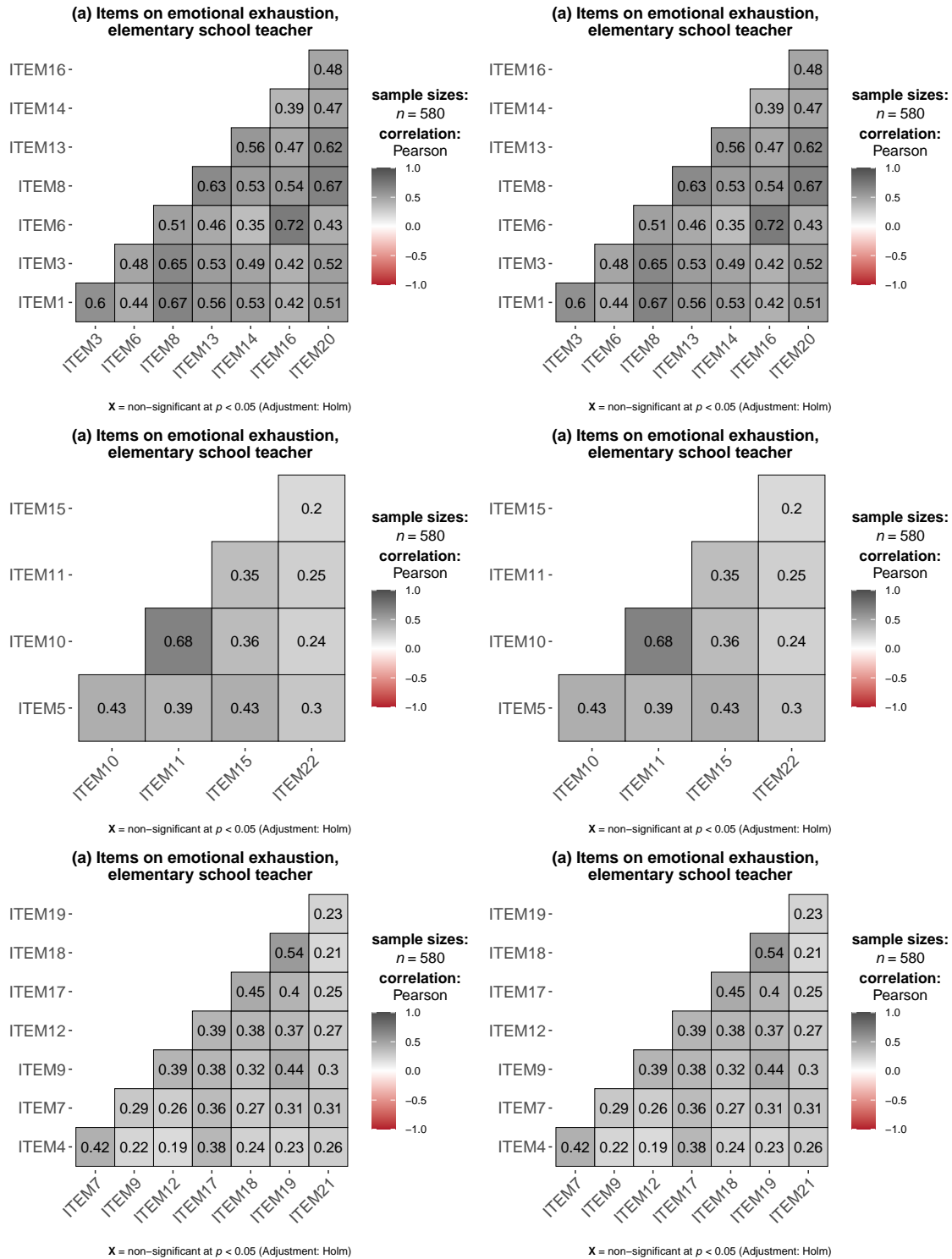
```

fa.ee <- c("ITEM1", "ITEM3", "ITEM6", "ITEM8", "ITEM13", "ITEM14", "ITEM16", "ITEM20")
fa.dp <- c("ITEM5", "ITEM10", "ITEM11", "ITEM15", "ITEM22")
fa.pa <- c("ITEM4", "ITEM7", "ITEM9", "ITEM12", "ITEM17", "ITEM18", "ITEM19", "ITEM21")
#generate 6 plots, 3 factors X 2 subgroups of teachers
p.cor.elm.ee <-
  mycor(
    data= mbi.elm,
    cols = fa.ee,
    "(a) Items on emotional exhaustion,
    elementary school teacher"
  )
p.cor.sec.ee <-
  mycor(
    data = mbi.sec,
    cols = fa.ee,
    "(b) Items on emotional exhaustion,
    secondary school teacher"
  )
p.cor.elm.dp <-
  mycor(
    data = mbi.elm,
    cols = fa.dp,
    "(c) Items on depersonalization,
    elementary school teacher"
  )
p.cor.sec.dp <-
  mycor(
    data = mbi.sec,
    cols = fa.dp,
    "(d) Items on depersonalization,
    secondary school teacher"
  )
p.cor.elm.pa <-
  mycor(
    data = mbi.elm,
    cols = fa.pa,
    "(e) Items on personal accomplishment,
    secondary school teacher"
  )
p.cor.sec.pa <-
  mycor(
    data = mbi.sec ,
    cols = fa.pa,
    "(f) Items on personal accomplishment,
    secondary school teacher"
  )
#plot sub-figure layout
patchwork <-
  p.cor.elm.ee/p.cor.elm.dp/p.cor.elm.pa|p.cor.sec.ee/p.cor.sec.dp/p.cor.sec.pa
#print the plot with a gernal title
patchwork+
  plot_annotation(
    title =

```

```
    'Figure 3 Correlalogram for items on each factor for two groups of teachers',  
  theme =  
    theme(plot.title =  
      element_text(  
        size = 16,  
        face = "bold",  
        vjust = -1.5,  
        hjust =0.5)  
      )  
    )  
  )  
)
```

Figure 3 Correlalogram for items on each factor for two groups of teachers



4 Testing the factorial invariance of MBI inventory between elementary and secondary school teachers

4.1 Define and estimate initial models for both subgroups

The postulated three-factor structure of the MBI that was tested in the previous assignments were re-tested as the initial model for establishing a baseline model.

4.1.1 Define the initial model

```
library(lavaan)
# Define a CFA model using the lavaan (Latent Variable Analysis) syntax:
# see https://lavaan.ugent.be/tutorial/syntax1.html
initial.model <- '
# CFA model for the burnout, the baseline model:
  EE =~ ITEM1 + ITEM2 + ITEM3 + ITEM6 + ITEM8 +
      ITEM13 + ITEM14 + ITEM16 + ITEM20
  DP =~ ITEM5 + ITEM10 + ITEM11 + ITEM15 + ITEM22
  PA =~ ITEM4 + ITEM7 + ITEM9 + ITEM12 +
      ITEM17 + ITEM18 + ITEM19 + ITEM21
'
```

Cited from Byrne: *It is important to note that measuring instruments are often group specific in the way they operate, and, thus, it is possible that baseline models may not be completely identical across groups.*

4.1.2 Estimate indices to examine factorial validity

- (1) Estimate factorial validity for the elementary teacher subgroup

```
cfa.elm <-
cfa(
  initial.model,
  data = mbi.elm,
  estimator = "MLM",
  mimic = "Mplus"
)
```

- (2) Estimate factorial validity for the secondary teacher subgroup

```
cfa.sec <-
cfa(
  initial.model,
  data = mbi.sec,
  estimator = "MLM",
  mimic = "Mplus"
)
```

Table 1: Fit indices for two subgroups, baseline models

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Elementary level	826.573(206, <0.001)	0.857	0.840	0.072(<0.001)	0.068	1.225
Secondary level	999.359(206, <0.001)	0.836	0.816	0.075(<0.001)	0.077	1.284

* Chi square scaling factor

4.1.3 Evaluate model

(1) Fit indices

```
library(knitr);library(kableExtra)
#combine fit indices of both levels
initial.elm.fit <-
  cfa.summary.mlm.a(cfa.elm) |>
  t() |>
  as.data.frame()

initial.sec.fit <-
  cfa.summary.mlm.a(cfa.sec) |>
  t() |>
  as.data.frame()

initial.both <-
  rbind(
    initial.elm.fit[2,],
    initial.sec.fit[2,]
  )

names(initial.both) <-
  initial.elm.fit[1,]

rownames(initial.both) <- NULL

initial.both <-
  initial.both |>
  mutate(Model = c("Elementary level",
    "Secondary level")) |>
  select(Model, everything())

#print the table
multi.fit.tab(initial.both, "Fit indices for two subgroups, baseline models")
```

See table 1. Goodness-of-fit statistics for this baseline model (three factor) reveals that the indices are less than optimal for both elementary (MLM Chi-square[206] = 826.573; CFI = 0.857; RMSEA = 0.072 ; SRMR = 0.068) and secondary (MLM Chi-square[206] = 999.359; CFI = 0.836; RMSEA = 0.075; SRMR = 0.077) levels.

(2) factor loading

Factor loading of elementary level were extracted.

```
fl.elm <- cfa.summary.b (cfa.elm) #fl is for factor loading)
colnames(fl.elm)[2] <- "Beta*"
```

Factor loading of secondary level were extracted.

```
fl.sec <- cfa.summary.b (cfa.sec) #fl is for factor loading
colnames(fl.sec) <- c("Parameter",
                      "Beta* ",
                      "SE ",
                      "Z ",
                      "p-value ")
```

Factor loading of both levels were merged in one table and printed.

```
fl.both <- left_join(fl.elm,
                    fl.sec,
                    by = "Parameter")

fl.both |>
  kable(
    digits = 3,
    booktabs = T,
    #format = "markdown",
    caption = "Factor loadings for both levels",
    linesep = ""
  ) |>
  add_header_above(c(" " = 1,
                    "Elementary level" = 4,
                    "Secondary level" = 4
                    )
  ) |>
  kable_styling() |>
  row_spec(1:9,
    background = "#E5E4E2"
  ) |>
  row_spec(15:22,
    background = "#E5E4E2"
  ) |>
  row_spec(c(1,10,15), bold = T) |>
  footnote(general =
    "Rows with coeffcient estimates fixed to 1 are highlighted in bold ",
    symbol = c(
      "Standardized estimates"
    )
  )
```

the cross-loading involved the loading of Item 12 on Factor 1 (Emotional Exhaustion) in addition to its targeted Factor 3 (Personal Accomplishment)

(3) Variance

Variance of elementary level were extracted.

Table 2: Factor loadings for both levels

Parameter	Elementary level				Secondary level			
	Beta*	SE	Z	p-value	Beta*	SE	Z	p-value
EE→ITEM1	0.776	0.000	NA	NA	0.756	0.000	NA	NA
EE→ITEM2	0.754	0.032	28.561	<0.001	0.736	0.031	30.236	<0.001
EE→ITEM3	0.740	0.045	21.984	<0.001	0.722	0.043	24.030	<0.001
EE→ITEM6	0.631	0.051	16.064	<0.001	0.626	0.046	18.669	<0.001
EE→ITEM8	0.855	0.042	28.448	<0.001	0.833	0.046	25.968	<0.001
EE→ITEM13	0.754	0.045	22.474	<0.001	0.762	0.045	23.619	<0.001
EE→ITEM14	0.655	0.046	19.939	<0.001	0.634	0.045	20.685	<0.001
EE→ITEM16	0.640	0.047	15.992	<0.001	0.596	0.047	15.261	<0.001
EE→ITEM20	0.734	0.045	18.371	<0.001	0.707	0.048	17.421	<0.001
DP→ITEM5	0.576	0.000	NA	NA	0.453	0.000	NA	NA
DP→ITEM10	0.794	0.115	11.968	<0.001	0.820	0.188	10.259	<0.001
DP→ITEM11	0.793	0.122	11.588	<0.001	0.808	0.197	9.666	<0.001
DP→ITEM15	0.505	0.072	9.287	<0.001	0.472	0.098	10.295	<0.001
DP→ITEM22	0.351	0.091	6.997	<0.001	0.447	0.131	8.226	<0.001
PA→ITEM4	0.447	0.000	NA	NA	0.340	0.000	NA	NA
PA→ITEM7	0.516	0.148	7.308	<0.001	0.545	0.221	7.495	<0.001
PA→ITEM9	0.581	0.280	6.629	<0.001	0.681	0.365	7.432	<0.001
PA→ITEM12	0.611	0.303	6.214	<0.001	0.586	0.283	7.398	<0.001
PA→ITEM17	0.681	0.185	7.796	<0.001	0.546	0.187	7.486	<0.001
PA→ITEM18	0.628	0.276	6.628	<0.001	0.698	0.294	7.431	<0.001
PA→ITEM19	0.643	0.255	6.844	<0.001	0.706	0.324	7.565	<0.001
PA→ITEM21	0.425	0.187	7.018	<0.001	0.410	0.242	6.808	<0.001

Note:

Rows with coefficient estimates fixed to 1 are highlighted in bold

* Standardized estimates


```
var.elm <- cfa.summary.c(cfa.elm, fa.num = 3, item.num = 22)
names(var.elm)[3] <- "Beta*"
names(var.elm)[4] <- "Beta†"
```

Variance of secondary level were extracted.

```
var.sec <- cfa.summary.c(cfa.sec, fa.num = 3, item.num = 22)
var.sec <- var.sec[,-1]
names(var.sec) <-
  c("Indicator",
    "Beta* ",
    "Beta† ",
    "SE ",
    "Z ",
    "p-value ")
)
```

Variance of both levels were merged in one table and printed.

```
var.both <- left_join(var.elm,
                      var.sec,
                      by = "Indicator")

align.table(data = var.both,
            num.no.header.col = 2,
            title = "Residual variance for both levels")
```

(3) Co-variance

Co-variance of elementary level were extracted.

```
cov.elm <- cfa.summary.d(cfa.elm, fa.num = 3, item.num = 22)
colnames(cov.elm)[2:3] <- c("Beta*", "Beta†")
```

Co-variance of secondary level were extracted.

```
cov.sec <- cfa.summary.d(cfa.sec, fa.num = 3, item.num = 22)
colnames(cov.sec) <- c("Parameter", "Beta* ", "Beta† ", "SE ", "Z ", "p-value ")
```

Co-variance of both levels were merged in one table and printed.

```
cov.both <- left_join(cov.elm,
                      cov.sec,
                      by = "Parameter")

align.table(data = cov.both,
            num.no.header.col = 1,
            title = "Residual co-variance for both levels")
```

Table 3: Residual variance for both levels

Parameter	Indicator	Elementary level					Secondary level				
		Beta*	Beta†	SE	Z	p-value	Beta*	Beta†	SE	Z	p-value
Residual	ITEM1	1.095	0.398	0.062	17.641	<0.001	1.078	0.429	0.056	19.329	<0.001
Residual	ITEM2	1.067	0.432	0.063	16.832	<0.001	1.071	0.459	0.053	20.373	<0.001
Residual	ITEM3	1.322	0.452	0.089	14.773	<0.001	1.383	0.479	0.083	16.704	<0.001
Residual	ITEM6	1.655	0.602	0.098	16.924	<0.001	1.656	0.609	0.084	19.730	<0.001
Residual	ITEM8	0.886	0.269	0.068	13.044	<0.001	0.890	0.306	0.061	14.560	<0.001
Residual	ITEM13	1.281	0.431	0.087	14.663	<0.001	1.167	0.419	0.075	15.574	<0.001
Residual	ITEM14	1.897	0.571	0.113	16.728	<0.001	1.883	0.599	0.110	17.084	<0.001
Residual	ITEM16	1.363	0.591	0.066	20.746	<0.001	1.353	0.645	0.071	19.024	<0.001
Residual	ITEM20	0.954	0.461	0.093	10.210	<0.001	0.983	0.500	0.057	17.125	<0.001
Residual	ITEM5	1.459	0.669	0.119	12.289	<0.001	1.711	0.795	0.100	17.052	<0.001
Residual	ITEM10	0.806	0.370	0.094	8.530	<0.001	0.803	0.328	0.090	8.944	<0.001
Residual	ITEM11	0.848	0.372	0.101	8.404	<0.001	0.854	0.347	0.095	9.013	<0.001
Residual	ITEM15	0.934	0.745	0.119	7.870	<0.001	1.562	0.778	0.112	13.964	<0.001
Residual	ITEM22	2.086	0.877	0.143	14.538	<0.001	2.052	0.800	0.124	16.598	<0.001
Residual	ITEM4	0.696	0.800	0.066	10.568	<0.001	1.074	0.884	0.104	10.372	<0.001
Residual	ITEM7	0.562	0.734	0.058	9.605	<0.001	0.907	0.703	0.064	14.108	<0.001
Residual	ITEM9	1.176	0.662	0.115	10.247	<0.001	1.194	0.536	0.097	12.297	<0.001
Residual	ITEM12	1.039	0.627	0.079	13.108	<0.001	1.177	0.657	0.076	15.418	<0.001
Residual	ITEM17	0.418	0.536	0.048	8.653	<0.001	0.649	0.701	0.063	10.319	<0.001
Residual	ITEM18	0.894	0.606	0.109	8.170	<0.001	0.703	0.512	0.068	10.329	<0.001
Residual	ITEM19	0.753	0.587	0.062	12.153	<0.001	0.847	0.501	0.080	10.595	<0.001
Residual	ITEM21	1.360	0.819	0.124	10.949	<0.001	1.889	0.832	0.111	17.056	<0.001
Total	EE	1.657	1.000	0.114	14.585	<0.001	1.436	1.000	0.097	14.854	<0.001
Total	DP	0.723	1.000	0.111	6.515	<0.001	0.442	1.000	0.085	5.188	<0.001
Total	PA	0.174	1.000	0.046	3.814	<0.001	0.141	1.000	0.034	4.108	<0.001

* Un-standardized estimates

† Standardized estimates

Table 4: Residual co-variance for both levels

Parameter	Elementary level					Secondary level				
	Beta*	Beta†	SE	Z	p-value	Beta*	Beta†	SE	Z	p-value
EE $\leftarrow \rightarrow$ DP	0.688	0.628	0.075	9.171	<0.001	0.451	0.566	0.057	7.928	<0.001
EE $\leftarrow \rightarrow$ PA	-0.254	-0.473	0.037	-6.952	<0.001	-0.177	-0.393	0.029	-6.193	<0.001

* Un-standardized estimates

† Standardized estimates

4.1.4 Model re-specification

- (1) Search for mis-specified parameters

To establish baseline models for both panels of teachers that represent good model fit and parsimony, I further investigated the modification indices of the hypothesized models, respectively for two levels.

MIIs of elementary level panel were calculated.

```
#extract needed variables
initial.MI.elm <-
  modindices(cfa.elm,
             standardized = TRUE,
             sort. = TRUE,
             maximum.number = 10)
```

MIIs of secondary level panel were calculated.

```
#extract needed variables
initial.MI.sec <-
  modindices(cfa.sec,
             standardized = TRUE,
             sort. = TRUE,
             maximum.number = 10)
```

MI tables with 10 largest MI parameters was printed in descending order of MI. Potential mis-specification of most concerns were highlighted in red.

```
MI.both <- rbind(initial.MI.elm, initial.MI.sec)

MI.both |>
  mutate(
    op = case_when(op == "~"~"~"~"↔",
                   op == "=~"~"~"~"→"),
    Parameter =
      paste(lhs, op, rhs)
  ) |>
  select(Parameter,
         MI = mi,
         EPC = epc,
         "std EPC" = sepc.all
  ) |>
  kable(digits = 3,
        booktab = T,
        linesep = "",
        caption =
          "Selected modification indices for determining baseline model") |>
  kable_styling(
    latex_options = "striped"
  ) |>
  row_spec(
    c(1:4, 11:14),
    color = "red"
```

Table 5: Selected modification indices for determining baseline model

	Parameter	MI	EPC	std EPC
Elementary level				
183	ITEM6 \leftrightarrow ITEM16	180.298	0.893	0.595
120	ITEM1 \leftrightarrow ITEM2	103.177	0.534	0.494
84	EE \rightarrow ITEM12	81.319	-0.400	-0.400
285	ITEM10 \leftrightarrow ITEM11	67.743	0.688	0.832
348	ITEM18 \leftrightarrow ITEM19	43.669	0.279	0.340
323	ITEM4 \leftrightarrow ITEM7	42.833	0.184	0.294
175	ITEM3 \leftrightarrow ITEM12	28.187	-0.287	-0.245
275	ITEM5 \leftrightarrow ITEM15	25.815	0.273	0.234
96	DP \rightarrow ITEM16	25.652	0.459	0.257
185	ITEM6 \leftrightarrow ITEM5	23.753	0.337	0.217
Secondary level				
1201	ITEM1 \leftrightarrow ITEM2	171.647	0.627	0.583
2851	ITEM10 \leftrightarrow ITEM11	135.841	1.181	1.426
1831	ITEM6 \leftrightarrow ITEM16	127.756	0.686	0.458
841	EE \rightarrow ITEM12	118.156	-0.468	-0.419
2751	ITEM5 \leftrightarrow ITEM15	77.216	0.580	0.355
296	ITEM11 \leftrightarrow ITEM15	60.947	-0.485	-0.420
147	ITEM2 \leftrightarrow ITEM20	53.024	-0.324	-0.316
274	ITEM5 \leftrightarrow ITEM11	48.297	-0.446	-0.369
339	ITEM9 \leftrightarrow ITEM19	46.617	0.360	0.358
77	EE \rightarrow ITEM10	45.623	-0.394	-0.302

Note:

Rows highlighted in red are of special concerns

```

) |>
footnote(general =
  "Rows highlighted in red are of special concerns") |>
pack_rows(index = c(
  "Elementary level" = 10,
  "Secondary level" = 10
))

```

See table 5. Three exceptionally large residual co-variances and one cross-loading contributed to the misfit of the model for both teacher panels. The residual co-variances involved Items 1 and 2, Items 6 and 16, and Items 10 and 11; the cross-loading involved the loading of Item 12 on Factor 1 (Emotional Exhaustion) in addition to its targeted Factor 3 (Personal Accomplishment).

In reviewing both the MIs and expected parameter change (EPC) statistics for elementary teachers (table 5, upper part), it is clear that all four parameters are contributing substantially to model misfit, with the residual covariance between Item 6 and Item 16 exhibiting the most profound effect.

We see precisely the same pattern on secondary teachers, albeit the effect would appear to be even more pronounced than it was for elementary teachers. One slight difference between the two groups of teachers regards the impact of these four parameters on model misfit. Whereas the residual covariance between Items 6 and 16 was found to be the most seriously misfitting parameter for elementary teachers; for secondary teachers, the residual covariance between Items 1 and 2 was most pronounced.

(2) Re-specify initial model to model 2

The good practice is relaxing one parameter each time. Nonetheless, according to the knowledge derived from our previous work, I included all four mis-specified parameters in a post-hoc model (common to the groups).

First, the 4 parameters were relaxed in model statement.

```
respecified4 <- 'EE =~ ITEM12
                ITEM6 ~~ ITEM16
                ITEM10 ~~ ITEM11
                ITEM1 ~~ ITEM2
                '
model2 <- paste(initial.model, respecified4)
```

Then, the model fit were re-estimated for both group, respectively

```
#for elementary
cfa2.elm <-
  cfa(
    model2,
    data = mbi.elm,
    estimator = "MLM",
    mimic = "Mplus"
  )
#for secondary
cfa2.sec <-
  cfa(
    model2,
    data = mbi.sec,
    estimator = "MLM",
    mimic = "Mplus"
  )
```

4.1.5 Examine Model 2

(1) Inspect fit indices of model2 (comparing to initial model)

```
#combine fit indices of both levels
model2.elm.fit <-
  cfa.summary.mlm.a(
    cfa2.elm
  ) |>
  t() |>
  as.data.frame()

model2.sec.fit <-
  cfa.summary.mlm.a(
    cfa2.sec
  ) |>
  t() |>
  as.data.frame()
```

Table 6: Fit indices for two subgroups, model 2, comparing to initial model

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Initial model						
Elementary level	826.573(206, <0.001)	0.857	0.840	0.072(<0.001)	0.068	1.225
Secondary level	999.359(206, <0.001)	0.836	0.816	0.075(<0.001)	0.077	1.284
Model 2						
Elementary level	477.667(202, <0.001)	0.936	0.927	0.049(0.679)	0.050	1.224
Secondary level	587.538(202, <0.001)	0.920	0.909	0.053(0.168)	0.056	1.278

* Chi square scaling factor

```

model2.both <-
  rbind(
    model2.elm.fit[2,],
    model2.sec.fit[2,]
  )

names(model2.both) <- model2.elm.fit[1,]

rownames(model2.both) <- NULL

model2.both <-
  model2.both |>
  mutate(Model = c("Elementary level",
    "Secondary level")) |>
  select(Model, everything())

#combine model 1 and 2 tables
compare12 <- rbind(initial.both, model2.both)

#print the table
multi.fit.tab(compare12,
  "Fit indices for two subgroups, model 2, comparing to initial model") |>
  pack_rows(index = c(
    "Initial model" = 2,
    "Model 2" = 2
  )
)

```

Estimation of this re-specified model, for each teacher group, yielded greatly improved model fit statistics than initial model. See table 6. However, we should note that several statistics, albeit improved comparing to initial model, still fall below the preferable value. For example, CFI from both groups were <0.95.

(2) Modification indices of model 2

To establish baseline models for both panels of teachers that represent good model fit and parsimony, I further investigated the modification indices of model 2, respectively for two groups, to decide if there was any more model mis-fit and mis-specification

MIIs of elementary level panel were calculated.

```
#extract needed variables
model2.MI.elm <-
  modindices(cfa2.elm,
             standardized = TRUE,
             sort. = TRUE,
             maximum.number = 10)
```

MIIs of secondary level panel were calculated.

```
#extract needed variables
model2.MI.sec <-
  modindices(cfa2.sec,
             standardized = TRUE,
             sort. = TRUE,
             maximum.number = 10)
```

MI tables with 10 largest MI parameters was printed in descending order of MI. Potential mis-specification of most concerns were highlighted in red.

```
MI2.both <- rbind(model2.MI.elm, model2.MI.sec)

MI2.both |>
  mutate(
    op = case_when(op == "~::~~"↔",
                   op == "=::~~"→"),
    Parameter =
      paste(lhs, op, rhs)
  ) |>
  select(Parameter,
         MI = mi,
         EPC = epc,
         "std EPC" = sepc.all
  ) |>
  kable(digits = 3,
        booktab = T,
        linesep = "",
        caption =
          "Selected modification indices for determining baseline model") |>
  kable_styling(
    latex_options = "striped"
  ) |>
  row_spec(
    c(1:2, 11:12),
    color = "red"
  ) |>
  footnote(general =
    "Rows highlighted in red are of special concerns") |>
  pack_rows(index = c(
    "Elementary level" = 10,
    "Secondary level" = 10
  ))
  )
```

Table 7: Selected modification indices for determining baseline model

	Parameter	MI	EPC	std EPC
Elementary level				
323	ITEM4 \leftrightarrow ITEM7	38.931	0.174	0.284
348	ITEM18 \leftrightarrow ITEM19	38.744	0.266	0.333
115	PA \rightarrow ITEM14	24.435	0.864	0.205
177	ITEM3 \leftrightarrow ITEM12	23.978	-0.250	-0.227
227	ITEM13 \leftrightarrow ITEM12	20.493	0.231	0.211
147	ITEM2 \leftrightarrow ITEM14	16.441	0.245	0.163
99	DP \rightarrow ITEM16	15.733	0.310	0.197
216	ITEM13 \leftrightarrow ITEM14	14.838	0.281	0.180
82	EE \rightarrow ITEM11	14.750	0.250	0.206
105	DP \rightarrow ITEM17	12.788	-0.173	-0.188
Secondary level				
821	EE \rightarrow ITEM11	67.177	0.472	0.339
339	ITEM9 \leftrightarrow ITEM19	43.690	0.355	0.357
276	ITEM5 \leftrightarrow ITEM15	35.576	0.416	0.310
296	ITEM11 \leftrightarrow ITEM15	29.016	-0.297	-0.206
247	ITEM16 \leftrightarrow ITEM20	28.900	0.227	0.201
98	DP \rightarrow ITEM14	22.145	-0.490	-0.239
345	ITEM17 \leftrightarrow ITEM18	21.583	0.147	0.219
335	ITEM7 \leftrightarrow ITEM21	21.370	0.247	0.191
346	ITEM17 \leftrightarrow ITEM19	20.742	-0.159	-0.217
149	ITEM2 \leftrightarrow ITEM20	20.020	-0.171	-0.162

Note:

Rows highlighted in red are of special concerns

See table 7. In reviewing this information for elementary teachers, we observe two MIs larger than all other MIs (ITEM7 with ITEM4; ITEM19 with ITEM18); both represent residual co-variances. I followed Byrne's step in addressing these parameters. According to Byrne, of the two, only the residual covariance between Items 7 and 4 is substantively viable in that there is a clear overlapping of item content. In contrast, the content of Items 19 and 18 exhibits no such redundancy, and, thus, there is no reasonable justification for including this parameter in a succeeding Model 3.

However, in checking the MI for secondary teachers, the decision was made: more work is needed in establishing an appropriate baseline model. Two parameters were of special concern due to their large MI and substantive meaningfulness. They are Item 11 cross-loads onto factor EE, and item 19 co-varies with item 9. This time I operated by the good practice of specifying one parameter each time. Given the substantially large MI representing the cross-loading of Item 11 on factor EE, this parameter alone was included in our next post-hoc model (Model 3 for secondary teachers).

Byrne noted the reasons for making this decision (to further re-specifying model secondary teachers), which I quoted here for future reflection: (a) The model does not yet reflect a satisfactorily good fit to the data (CFI = 0.920); and (b) in reviewing the MIs in Table 7.2, we observe one very large mis-specified parameter representing the loading of Item 11 on Factor 1 (F1 by ITEM11), as well as another substantially large MI representing a residual covariance between Items 19 and 9, both of which can be substantiated as substantively meaningful parameters.

(3) Model re-specification of model 2 to model 3

```
respecified3.elm <- 'ITEM4 ~~ ITEM7
                    '
respecified3.sec <- 'EE =~ ITEM11
                    '
model3.elm <- paste(model2, respecified3.elm)
model3.sec <- paste(model2, respecified3.sec)
```

Then, the model fit were re-estimated for both group, separately.

```
#for elementary
cfa3.elm <-
  cfa(
    model3.elm,
    data = mbi.elm,
    estimator = "MLM",
    mimic = "Mplus"
  )
#for secondary
cfa3.sec <-
  cfa(
    model3.sec,
    data = mbi.sec,
    estimator = "MLM",
    mimic = "Mplus"
  )
```

4.1.6 Examine Model 3

(1) Inspect fit indices of model3 (comparing to model 2)

```

#combine fit indices of both levels
model3.elm.fit <-
  cfa.summary.mlm.a(
    cfa3.elm
  ) |>
  t() |>
  as.data.frame()

model3.sec.fit <-
  cfa.summary.mlm.a(
    cfa3.sec
  ) |>
  t() |>
  as.data.frame()

model3.both <-
  rbind(
    model3.elm.fit[2,],
    model3.sec.fit[2,]
  )

names(model3.both) <- model3.elm.fit[1,]

rownames(model3.both) <- NULL

model3.both <-
  model3.both |>
  mutate(Model = c("Elementary level",
    "Secondary level")) |>
  select(Model, everything())

#combine model 1 and 2 tables
compare123 <- rbind(initial.both, model2.both, model3.both)

#print the table
multi.fit.tab(compare123,
  "Fit indices for two subgroups, model 3, comparing to preceding models") |>
  pack_rows(index = c(
    "Initial model" = 2,
    "Model 2" = 2,
    "Model 3" = 2
  )
)

```

See table 8. Results from the estimation of Model 3 for elementary teachers yielded goodness-of-fit statistics that represented a satisfactorily good fit to the data (MLM chi square[201] = 451.061; CFI = 0.942; RMSEA = 0.046; SRMR = 0.049). Although a review of Table 9 (find below) reveals several additional moderately large MIs, for balancing goodness-of-fit and parsimony, [the decision was model 3 can serve as the baseline model for elementary teachers.](#)

Results from the estimation of Model 3 for secondary teachers, on the other hand, further substantiated the residual covariance between Items 19 and 9 as representing an acutely mis-specified parameter in the model. Thus, for secondary teachers only, model 4 was put to the test with this residual covariance specified as a freely estimated parameter.

Table 8: Fit indices for two subgroups, model 3, comparing to preceding models

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Initial model						
Elementary level	826.573(206, <0.001)	0.857	0.840	0.072(<0.001)	0.068	1.225
Secondary level	999.359(206, <0.001)	0.836	0.816	0.075(<0.001)	0.077	1.284
Model 2						
Elementary level	477.667(202, <0.001)	0.936	0.927	0.049(0.679)	0.050	1.224
Secondary level	587.538(202, <0.001)	0.920	0.909	0.053(0.168)	0.056	1.278
Model 3						
Elementary level	451.061(201, <0.001)	0.942	0.934	0.046(0.876)	0.049	1.210
Secondary level	535.759(201, <0.001)	0.931	0.920	0.049(0.629)	0.053	1.275

* Chi square scaling factor

(2) Modification indices of model 3

MI of model 3 for each groups were calculated.

```
#elementary
model3.MI.elm <-
  modindices(cfa3.elm,
             standardized = TRUE,
             sort. = TRUE,
             maximum.number = 10)
#secondary
model3.MI.sec <-
  modindices(cfa3.sec,
             standardized = TRUE,
             sort. = TRUE,
             maximum.number = 10)
```

MI tables with 10 largest MI parameters was printed in descending order of MI. Potential mis-specification of most concerns were highlighted in red.

```
MI3.both <- rbind(model3.MI.elm, model3.MI.sec)

MI3.both |>
  mutate(
    op = case_when(op == "~ ~ ~ ~ ~" ~> "~ ~ ~ ~ ~",
                  op == "= ~ ~ ~ ~ ~" ~> "= ~ ~ ~ ~ ~"),
    Parameter =
      paste(lhs, op, rhs)
  ) |>
  select(Parameter,
         MI = mi,
         EPC = epc,
         "std EPC" = sepc.all
  ) |>
  kable(digits = 3,
        booktab = T,
        linesep = "",
```

Table 9: Selected modification indices for determining baseline model

	Parameter	MI	EPC	std EPC
Elementary level				
348	ITEM18 \leftrightarrow ITEM19	32.503	0.247	0.319
116	PA \rightarrow ITEM14	25.403	0.977	0.210
178	ITEM3 \leftrightarrow ITEM12	23.654	-0.248	-0.226
228	ITEM13 \leftrightarrow ITEM12	20.844	0.232	0.213
148	ITEM2 \leftrightarrow ITEM14	16.457	0.245	0.163
100	DP \rightarrow ITEM16	15.696	0.310	0.197
217	ITEM13 \leftrightarrow ITEM14	14.844	0.282	0.180
83	EE \rightarrow ITEM11	14.780	0.251	0.206
326	ITEM4 \leftrightarrow ITEM17	14.165	0.096	0.174
106	DP \rightarrow ITEM17	13.820	-0.181	-0.197
Secondary level				
339	ITEM9 \leftrightarrow ITEM19	42.687	0.351	0.355
247	ITEM16 \leftrightarrow ITEM20	28.275	0.223	0.199
345	ITEM17 \leftrightarrow ITEM18	21.951	0.148	0.221
335	ITEM7 \leftrightarrow ITEM21	21.602	0.248	0.192
346	ITEM17 \leftrightarrow ITEM19	20.837	-0.160	-0.218
84	EE \rightarrow ITEM22	20.306	0.321	0.225
98	DP \rightarrow ITEM14	20.142	-0.404	-0.210
147	ITEM2 \leftrightarrow ITEM14	19.895	0.239	0.155
149	ITEM2 \leftrightarrow ITEM20	18.463	-0.164	-0.155
333	ITEM7 \leftrightarrow ITEM18	18.163	-0.159	-0.202

Note:

Rows highlighted in red are of special concerns

```

caption =
  "Selected modification indices for determining baseline model") |>
kable_styling(
  latex_options = "striped"
) |>
row_spec(
  c(1:2, 11),
  color = "red"
) |>
footnote(general =
  "Rows highlighted in red are of special concerns") |>
pack_rows(index = c(
  "Elementary level" = 10,
  "Secondary level" = 10
))

```

(3) Re-specification of model 3 to model 4 (only for secondary teacher)

The parameter ITEM9 \sim ITEM19 was relaxed for estimation.

```
respecified4.sec <- 'ITEM9 ~~ ITEM19
                    '
model4.sec <- paste(model3.sec, respecified4.sec)
```

Then, the model fit were re-estimated for secondary group, only

```
cfa4.sec <-
  cfa(
    model4.sec,
    data = mbi.sec,
    estimator = "MLM",
    mimic = "Mplus"
  )
```

4.1.7 Examine Model 4

Note that at this point I had taken model 3 as the baseline model for elementary teachers, and model 4 was to achieve the baseline model for secondary teachers.

(1) Inspect fit indices of model4 (comparing to 3)

```
model4.sec.fit <-
  cfa.summary.mlm.a(
    cfa4.sec
  ) |>
  t() |>
  as.data.frame()

names(model4.sec.fit ) <- model4.sec.fit[1,]

model4.sec.fit <- model4.sec.fit [-1,]
model4.sec.fit <-
  model4.sec.fit |>
  mutate(Model = "Secondary level") |>
  select(Model, everything())

rownames(model4.sec.fit ) <- NULL

#combine model 1 and 2 tables
model3.both[1,1] <- "Elementary level†"
model4.sec.fit[1,1] <- "Secondary level†"
compare1234 <-
  rbind(initial.both,
        model2.both,
        model3.both,
        model4.sec.fit )

#print the table
multi.fit.tab(compare1234,
              "Fit indices for two subgroups, model 4, comparing to preceding models",
              c("Baseline model for elementary teachers",
                "Baseline model for secondary teachers")) |>
```

Table 10: Fit indices for two subgroups, model 4, comparing to preceding models

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Initial model						
Elementary level	826.573(206, <0.001)	0.857	0.840	0.072(<0.001)	0.068	1.225
Secondary level	999.359(206, <0.001)	0.836	0.816	0.075(<0.001)	0.077	1.284
Model 2						
Elementary level	477.667(202, <0.001)	0.936	0.927	0.049(0.679)	0.050	1.224
Secondary level	587.538(202, <0.001)	0.920	0.909	0.053(0.168)	0.056	1.278
Model 3						
Elementary level†	451.061(201, <0.001)	0.942	0.934	0.046(0.876)	0.049	1.210
Secondary level	535.759(201, <0.001)	0.931	0.920	0.049(0.629)	0.053	1.275
Model 4						
Secondary level‡	505.831(200, <0.001)	0.937	0.927	0.047(0.859)	0.052	1.273

* Chi square scaling factor

† Baseline model for elementary teachers

‡ Baseline model for secondary teachers

```

pack_rows(index = c(
  "Initial model" = 2,
  "Model 2" = 2,
  "Model 3" = 2,
  "Model 4" = 1
)
) |>
row_spec(c(5,7),
  color = "red"
)

```

See table 10. Based on a moderately satisfactory goodness-of-fit (MLM $\chi^2_{200} = 505.831$; CFI = 0.937; RMSEA = 0.047; SRMR = 0.052) and to balance fit with parsimony, I consider Model 4 as the final baseline model for secondary teachers.

```

cfa3.elm <-
  cfa(
    model3.elm,
    data = mbi.elm,
    estimator = "MLM"
    #mimic = "Mplus"
  )

cfa4.sec <-
  cfa(
    model4.sec,
    data = mbi.elm,
    estimator = "MLM"
    #mimic = "Mplus"
  )

```

4.1.8 Visualize the final baseline models for each group

```
library(semPlot)
grps <- list(EE = c("ITEM1", "ITEM2", "ITEM3", "ITEM6", "ITEM8",
  "ITEM13", "ITEM14", "ITEM16", "ITEM20"),
  DP = c("ITEM5", "ITEM10", "ITEM11", "ITEM15", "ITEM22"),
  PA = c("ITEM4", "ITEM7", "ITEM9", "ITEM12",
    "ITEM17", "ITEM18", "ITEM19", "ITEM21"))

order.manifest <- c("ITEM4", "ITEM7", "ITEM9", "ITEM12",
  "ITEM17", "ITEM18", "ITEM19", "ITEM21",
  "ITEM5", "ITEM10", "ITEM11", "ITEM15", "ITEM22",
  "ITEM1", "ITEM2", "ITEM3", "ITEM6", "ITEM8",
  "ITEM13", "ITEM14", "ITEM16", "ITEM20")

order.latent <- c("PA", "DP", "EE")
par(mfrow=c(1,2))

semPaths(cfa3.elm,
  "col", #un-weighted edges
  "no", #edge label is standardized
  reorder = F,
  latents = order.latent,
  manifest = order.manifest,
  sizeLat = 8,
  sizeLat2 = 5,
  sizeMan = 6,
  sizeMan2 = 3,
  curveAdjacent = "cov", # if edge for adjacent nodes curly or not, "reg"
  shapeMan = "rectangle",
  style = "lisrel",
  group = "latent",
  curve = 0.3,
  curvature = 0.1, #theme = "colorblind", #cardinal = "lat cov",
  curvePivot = F, # curly edge or not
  rotation = 2,
  color = c("#c68642", "#58668b", "#8874a3"), #edge.color = "steelblue",
  shapeLat = "ellipse",
  label.font = 2,
  label.color = "white", #Label.scale = T,
  label.prop = 0.7
)

title(main = list("Elementary School Teachers",
  cex = 1, font = 1), outer = F, line = -3)

semPaths(cfa4.sec,
  "col", #un-weighted edges
  "no", #edge label is standardized
  reorder = F,
  latents = order.latent,
  manifest = order.manifest,
  sizeLat = 8,
  sizeLat2 = 5,
  sizeMan = 6,
```

```

sizeMan2 = 3,
curveAdjacent = "cov", #if edge for adjacent nodes curly or not, "reg"
shapeMan = "rectangle",
style = "lisrel",
group = "latent",
curve = 0.3,
curvature = 0.1, #theme = "colorblind", #cardinal = "lat cov",
curvePivot = F, # curly edge or not
layout = "tree",
rotation = 2,
color = c("#c68642", "#58668b", "#8874a3"), #edge.color = "steelblue",
shapeLat = "ellipse",
label.font = 2,
label.color = "white", #Label.scale = T,
label.prop = 0.7
)
title(main = list("Secondary School Teachers",
                  cex = 1, font = 1), outer = F, line = -3)
mtext("Figure 1 Hypothesized configural model", cex = 1.5, side = 1, line = -5, outer = TRUE)

```

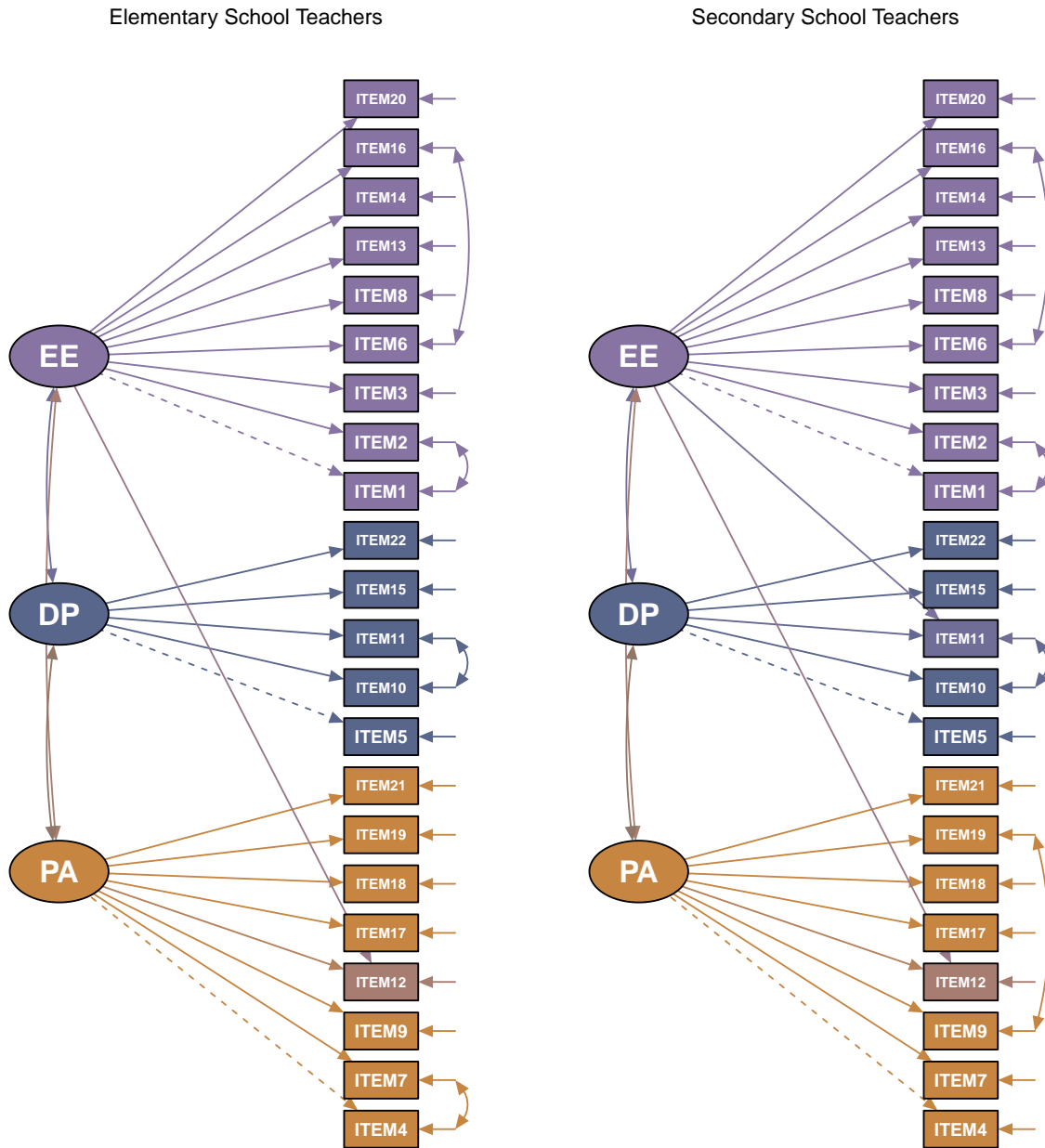



Figure 1 Hypothesized configural model

The the plot of baseline model for each group of teachers was created. See pciture 1 There are three

parameters (two residual co-variances [Item 4 with Item 7; Item 9 with Item 19] and one cross-loading [Item 11 on F1]) that were not part of the originally postulated model and that differ across the two groups of teachers. They have the same number of factors and the same factor-loading pattern (A question: Is there any standard for such a pattern? The current baseline models have one cross-loading that differs across the groups so they are not exactly same.)

5 Testing factorial equivalence of MBI between elementary and secondary school teachers

With the baseline models established through the previous steps, I could combine them to generate a common baseline model for both group, or configural model, which bridges the baseline model and the final task-testing factorial in-variance across groups.

5.1 Establish configural model (inv1.model)

5.1.1 Combine the datasets

```
mbi.both <-  
  merge(  
    data.frame(  
      mbi.elm,  
      group = "elementary"  
    ),  
    data.frame(  
      mbi.sec,  
      group = "secondary"  
    ),  
    all = TRUE,  
    sort = FALSE  
  )
```

5.1.2 Define the configural model

No equality constraints are imposed for this model.

```
inv1.model <- '  
  EE =~ 1*ITEM1 + ITEM2 + ITEM3 + ITEM6 + ITEM8 + ITEM13 + ITEM14 + ITEM16 + ITEM20  
  DP =~ 1*ITEM5 + ITEM10 + ITEM11 + ITEM15 + ITEM22  
  PA =~ 1*ITEM4 + ITEM7 + ITEM9 + ITEM12 + ITEM17 + ITEM18 + ITEM19 + ITEM21  
  
# Common modifications (from baseline models built above)  
  EE =~ ITEM12 # common cross-loading  
  ITEM1 ~~ ITEM2 # common residual covariances (3)  
  ITEM6 ~~ ITEM16  
  ITEM10 ~~ ITEM11  
  
# Group-specific parameters for elementary teachers:  
  ITEM4 ~~ c(NA, 0)*ITEM7 # specific residual covariance
```

Table 11: Fit indices for the configural model (inv1.model)

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Configural (inv1)	939.696(401, <0.001)	0.939	0.929	0.046(0.975)	0.051	1.266

* Chi square scaling factor

```
# Group-specific parameters for secondary teachers:
  EE =~ c(0, NA)*ITEM11 # specific cross-loading
  ITEM9 ~~ c(0, NA)*ITEM19 # specific residual covariance
,
```

5.1.3 Estimate the configural model

The model fit results derived from this model represent a multi-group version of the combined baseline models for elementary and secondary teacher.

```
inv1.fit <-
  cfa(
    inv1.model,
    data = mbi.both,
    estimator = "MLM",
    group = "group"
  )
```

5.1.4 Summarize the results

```
#extract the key indicators
inv1.fit.indices <-
  cfa.summary.mlm.a(inv1.fit) |>
  t() |>
  as.data.frame()

#define column and row names for the indicator table
names(inv1.fit.indices) <- inv1.fit.indices[1,]
inv1.fit.indices <- inv1.fit.indices[-1,]
rownames(inv1.fit.indices) <- NULL
inv1.fit.indices$Model <- "Configural (inv1)"

#print the table
multi.fit.tab(
  inv1.fit.indices,
  "Fit indices for the configural model (inv1.model)"
)
```

See table 11. Results for this configural model (inv1.model) were as follows: MLM chi-square(401) = 939.696, CFI = 0.939, RMSEA = 0.046, and SRMR = 0.051.

5.2 Impose equality constraints on factor loadings of configural model

5.2.1 Constrain common factor loadings equal step by step(inv2.model)

- (1) Define and inspect the fit indices of model 2 (inv2.model)

All the common factor loadings were constrained equal across groups. If the results show significant improvement from configural model, we get the evidence about multi-group in-variance. If not, we need to further explore which parameter(s) bring about the difference observed.

```
inv2.fit <-  
  cfa(inv1.model,  
      data = mbi.both,  
      estimator = "MLM",  
      group = "group",  
      group.equal = c("loadings"),  
      group.partial = c("EE =~ ITEM11")  
  )  
  
#extract the key indicators  
inv2.fit.indices <-  
  cfa.summary.mlm.a(inv2.fit) |>  
  t() |>  
  as.data.frame()  
  
#define column and row names for the indicator table  
names(inv2.fit.indices) <- inv2.fit.indices[1,]  
inv2.fit.indices <- inv2.fit.indices[-1,]  
rownames(inv2.fit.indices) <- NULL  
inv2.fit.indices$Model <- "Model12 (inv2)†"  
  
#merge configural model and inv2.model.  
fit.indices.12 <- # 12 is for inv1 and inv 2  
  rbind(  
    inv1.fit.indices,  
    inv2.fit.indices  
  )  
  
#print the table  
multi.fit.tab(  
  fit.indices.12,  
  "Comparison Fit indices between the configural model (inv1.model) and model 2 (inv2.model)",  
  "Configural model + 20 common factor loadings constrained equal across groups"  
)
```

See table 12. As indicated by the very slightly higher MLM chi-square value (939.696→995.433) and lower CFI value (0.939→0.935), compared with the configural model, results suggest that the model does not fit the data quite as well as it did with no factor-loading constraints imposed. Thus, we explore further to find out the parameter(s) brought about the non-invariance.

- (2) Examine the modification indices for inv2.model

MIIs of inv2.model were calculated. In seeking evidence of non-invariance, we focus only on the factor loadings that were constrained equal across the groups. In addition, in testing for invariance, only those

Table 12: Comparison Fit indices between the configural model (inv1.model) and model 2 (inv2.model)

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Configural (inv1)	939.696(401, <0.001)	0.939	0.929	0.046(0.975)	0.051	1.266
Model2 (inv2) [†]	995.433(421, <0.001)	0.935	0.928	0.046(0.967)	0.057	1.263

* Chi square scaling factor

[†] Configural model + 20 common factor loadings constrained equal across groups

parameters that were constrained equal, are of relevance. Hence, only the parameter statement that meets these requirements were extracted.

First, to simplify the searching for relevant parameters, I defined an object including all parameters were relevant, so that what parameters were shown in MI table were automatically controlled.

```
#create the parameter statements of relevancy
itemset1 <- "ITEM1 + ITEM2 + ITEM3 + ITEM6 + ITEM8 + ITEM13 + ITEM14 + ITEM16 + ITEM20 + ITEM12"
itemset2 <- "ITEM5 + ITEM10 + ITEM11 + ITEM15 + ITEM22"
itemset3 <- "ITEM4 + ITEM7 + ITEM9 + ITEM12 + ITEM17 + ITEM18 + ITEM19 + ITEM21"
#create relevant statement for EE
relevant.items1 <-
  stringr::str_replace_all(
    stringr::str_split_1(itemset1, "\\+" ),
    " ",
    ""
  )
relevant.items1 <-
  paste("EE", "→", relevant.items1)
#create relevant statement for DP
relevant.items2 <-
  stringr::str_replace_all(
    stringr::str_split_1(itemset2, "\\+" ),
    " ",
    ""
  )
relevant.items2 <-
  paste("DP", "→", relevant.items2)
#create relevant statement for PA
relevant.items3 <-
  stringr::str_replace_all(
    stringr::str_split_1(itemset3, "\\+" ),
    " ",
    ""
  )
relevant.items3 <-
  paste("PA", "→", relevant.items3)
#combine the above into one
relevant.items <- c(relevant.items1, relevant.items2, relevant.items3)
```

Next, I extract MI table with relevant parameters.

```
inv2.model.MI <- modindices(inv2.fit,
  standardized = TRUE,
  minimum.value = 3.84,
```

```

      free.remove = FALSE,
      op = "~",
      sort. = TRUE) |>
mutate(Parameter = paste(lhs, "→", rhs)) |>
filter(Parameter %in% relevant.items) |>
arrange(group)

```

Then, MI tables with relevant parameters was printed in descending order of MI. Potential parameters that are very possibly undermining equivalence across elementary and secondary teachers were highlighted in red.

```

inv2.model.MI.elm <-
  inv2.model.MI |>
  filter(group == 1) |>
  select(Parameter,
    MI = mi,
    EPC = epc,
    "std EPC" = sepc.all
  )

inv2.model.MI.sec <-
  inv2.model.MI |>
  filter(group == 2) |>
  select(Parameter,
    MI = mi,
    EPC = epc,
    "std EPC" = sepc.all
  )

rbind(inv2.model.MI.elm, inv2.model.MI.sec) |>
  kable(digits = 3,
    booktab = T,
    linesep = "",
    caption =
      "Selected modification indices for inv2.model") |>
  kable_styling(
    latex_options = "striped"
  ) |>
  pack_rows(index = c("Elementary teachers" = nrow(inv2.model.MI.elm),
    "Secondary teachers" = nrow(inv2.model.MI.sec)
  )
  ) |>
  row_spec(
    c(1,3),
    color = "red"
  ) |>
  footnote(general =
    "Rows highlighted in red are of special concerns")

```

See table 13. Of all the eligible parameters, the factor loading of Item 11 on DP appears to be the most problematic in terms of its equivalence across elementary and secondary teachers. I relaxed this factor loading (Item 11 by DP) for establishing the the next model, model3 (inv3.model)

(3) Re-specify model2 to fit model 3 (inv3.model)

Table 13: Selected modification indices for inv2.model

Parameter	MI	EPC	std EPC
Elementary teachers			
DP → ITEM11	11.949	0.195	0.122
DP → ITEM5	5.708	0.248	0.155
Secondary teachers			
DP → ITEM11	8.742	-0.142	-0.084
DP → ITEM5	5.708	-0.248	-0.159
DP → ITEM15	4.514	0.116	0.081
PA → ITEM7	4.381	0.202	0.079

Note:

Rows highlighted in red are of special concerns

```
inv3.fit <-
  cfa(inv1.model,
    data = mbi.both,
    estimator = "MLM",
    group = "group",
    group.equal = c("loadings"),
    group.partial = c("#EE =~ ITEM11",
                      "DP =~ ITEM11"
                    )
  )
```

5.2.2 Constrain common factor loadings equal step by step(inv3.model)

(1) Inspect fit indices of model 3 (inv3.model)

```
#extract the key indicators
inv3.fit.indices <-
  cfa.summary.mlm.a(inv3.fit) |>
  t() |>
  as.data.frame()

#define column and row names for the indicator table
names(inv3.fit.indices) <- inv3.fit.indices[1,]
inv3.fit.indices <- inv3.fit.indices[-1,]
rownames(inv3.fit.indices) <- NULL
inv3.fit.indices$Model <- "Model3 (inv3)†"

#merge configural model and inv2.model.
fit.indices.123 <- # 123 is for inv1, 2 and 3 models
  rbind(
    inv1.fit.indices,
    inv2.fit.indices,
    inv3.fit.indices
  )

#print the table
```

Table 14: Comparison Fit indices across the inv1.model through inv3.model

Model	Chi square (df, p)	Δ Chi-square(df,p)*	CFI	TLI	RMSEA(p)	SRMR
Configural (inv1)	939.696(401, <0.001)	—	0.939	0.929	0.046(0.975)	0.051
Model2 (inv2) [†]	995.433(421, <0.001)	56.14(20, 0)	0.935	0.928	0.046(0.967)	0.057
Model3 (inv3) [‡]	1020.224(421, <0.001)	80.621(20, 0)	0.932	0.925	0.047(0.910)	0.056

* Chi square difference statistics of model of the row compared to configural model

[†] Configural model + 20 common factor loadings constrained equal across groups

[‡] Inv2.model + a parameter(DP By Item11) set relaxed

```
delta.fit.tab(fit.indices.123,
              "Comparison Fit indices across the inv1.model through inv3.model",
              c("Configural model + 20 common factor loadings constrained equal across groups",
                "Inv2.model + a parameter(DP By Item11) set relaxed"))
```

The fit statistics of inv3.model: MLM chi-square[420] 969.990, CFI 0.937, RMSEA 0.045, SRMR 0.054. The difference in model fit between InvModel 3 and the configural model (InvModel 1) is, though not significant, approaching to significance at 0.05 level ($p = 0.057$). Hence, I decide to make further improvement to the model

In this step, my results were substantially different from the slides. Here in the slides the p value should be 0.048, base on which the decision was that we should make further improvement. Yet, I got a insignificant p already. I tried several different possibilities. I found if I set "group.partial = c("DP = ITEM11")" instead of "group.partial = c("EE = ITEM11", "DP = ITEM11")", I would have seen a significant p value very close to the slides. Yet the degree of freedom would turn to 421 instead of 420. I suppose there should be something I have done wrong.

(2) Examine the modification indices for inv3.model

I extract MI table with relevant parameters.

```
inv3.model.MI <- modindices(inv3.fit,
                             standardized = TRUE,
                             minimum.value = 3.84,
                             free.remove = FALSE,
                             op = "=",
                             sort. = TRUE) |>
mutate(Parameter = paste(lhs, "<math>\rightarrow</math>", rhs)) |>
filter(Parameter %in% relevant.items) |>
arrange(group)
```

Then, MI tables with relevant parameters was printed in descending order of MI.

```
inv3.model.MI.elm <-
  inv3.model.MI |>
  filter(group == 1) |>
  select(Parameter,
          MI = mi,
          EPC = epc,
          "std EPC" = sepc.all
  )
```


Table 15: Selected modification indices for inv3.model

Parameter	MI	EPC	std EPC
Elementary teachers			
DP → ITEM5	8.806	0.316	0.190
Secondary teachers			
DP → ITEM5	8.807	-0.316	-0.198
PA → ITEM7	4.394	0.202	0.079

Note:

Rows highlighted in red are of special concerns

```
inv3.model.MI.sec <-
  inv3.model.MI |>
  filter(group == 2) |>
  select(Parameter,
    MI = mi,
    EPC = epc,
    "std EPC" = sepc.all
  )

rbind(inv3.model.MI.elm, inv3.model.MI.sec) |>
  kable(digits = 3,
    booktab = T,
    linesep = "",
    caption =
      "Selected modification indices for inv3.model") |>
  kable_styling(
    latex_options = "striped"
  ) |>
  pack_rows(index = c("Elementary teachers" = nrow(inv3.model.MI.elm),
    "Secondary teachers" = nrow(inv3.model.MI.sec)
  )
  )|>
  row_spec(
    c(1:2),
    color = "red"
  ) |>
  footnote(general =
    "Rows highlighted in red are of special concerns")
```

Potential parameters that are very possibly undermining equivalence across elementary and secondary teachers were highlighted in red. Actually, other than DP measured by ITEM5, none of the other parameters were both of concern and present simultaneously in two groups. I only had one candidate.

In this step AGAIN, my results were dramatically inconsistent with the slides, I was not able to get the parameter "DP measured by Item15" in the MI table.

(3) Re-specify model2 to fit model 3 (inv3.model)

```
inv4.fit <-
  cfa(inv1.model,
    data = mbi.both,
    estimator = "MLM",
    group = "group",
    group.equal = c("loadings"),
    group.partial = c("#EE =~ ITEM11",
                      "DP =~ ITEM11",
                      "DP =~ ITEM5"
                    ))
```

5.2.3 Constrain common factor loadings equal step by step(inv4.model)

- (1) Inspect fit indices of model 3 (inv4.model)

```
#extract the key indicators
inv4.fit.indices <-
  cfa.summary.mlm.a(inv4.fit) |>
  t() |>
  as.data.frame()

#define column and row names for the indicator table
names(inv4.fit.indices) <- inv4.fit.indices[1,]
inv4.fit.indices <- inv4.fit.indices[-1,]
rownames(inv4.fit.indices) <- NULL
inv4.fit.indices$Model <- "Model4 (inv4)**"

#merge configural model and inv2.model.
fit.indices.1234 <- # 123 is for inv1, 2, 3 and 4 models
  rbind(
    inv1.fit.indices,
    inv2.fit.indices,
    inv3.fit.indices,
    inv4.fit.indices
  )

#print the table
delta.fit.tab(fit.indices.1234,
  "Comparison Fit indices across the inv1.model through inv3.model",
  c("Configural model + 20 common factor loadings constrained equal across groups",
    "Inv2.model + a parameter(DP By Item11) set relaxed",
    "Inv3.model + a parameter(DP By Item5) set relaxed"))
```

- (1) Inspect fit indices of model 3 (inv3.model)
- (2) Examine the modification indices for inv3.model
- (3) Re-specify model2 to fit model 3 (inv3.model)

xie

```
inv1.fit.indices <-
  cfa.summary.mlm.a(inv1.fit) |>
```

Table 16: Comparison Fit indices across the inv1.model through inv3.model

Model	Chi square (df, p)	Δ Chi-square(df,p)*	CFI	TLI	RMSEA(p)	SRMR
Configural (inv1)	939.696(401, <0.001)	—	0.939	0.929	0.046(0.975)	0.051
Model2 (inv2) [†]	995.433(421, <0.001)	56.14(20, 0)	0.935	0.928	0.046(0.967)	0.057
Model3 (inv3) [‡]	1020.224(421, <0.001)	80.621(20, 0)	0.932	0.925	0.047(0.910)	0.056
Model4 (inv4) ^{**}	1020.224(421, <0.001)	80.621(20, 0)	0.932	0.925	0.047(0.910)	0.056

* Chi square difference statistics of model of the row compared to configural model

[†] Configural model + 20 common factor loadings constrained equal across groups

[‡] Inv2.model + a parameter(DP By Item11) set relaxed

[§] Inv3.model + a parameter(DP By Item5) set relaxed

Table 17: Fit indices for two subgroups, baseline models

Model	Chi square (df, p)	CFI	TLI	RMSEA(p)	SRMR	CSF*
Elementary level	826.573(206, <0.001)	0.857	0.840	0.072(<0.001)	0.068	1.225
Secondary level	999.359(206, <0.001)	0.836	0.816	0.075(<0.001)	0.077	1.284

* Chi square scaling factor

```

t() |>
as.data.frame()

initial.sec.fit <-
  cfa.summary.mlm.a(cfa.sec) |>
  t() |>
  as.data.frame()

initial.both <-
  rbind(
    initial.elm.fit[2,],
    initial.sec.fit[2,]
  )

names(initial.both) <-
  initial.elm.fit[1,]

rownames(initial.both) <- NULL

initial.both <-
  initial.both |>
  mutate(Model = c("Elementary level",
    "Secondary level")) |>
  select(Model, everything())

#print the table
multi.fit.tab(initial.both, "Fit indices for two subgroups, baseline models")

```

““