

class 11: structural bioinformatics pt1

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AlphaFold Data Base (AFBD)

the EBI maintains the largest database of Alphafold structure prediction models at:

from last class (before Halloween) we saw that the PDB had 244,290 (Oct 2025)

the total number of proteins sequences in UniprotKB is 199,579,901.

Key point: This is a tiny fraction of sequence space that has structural coverage (0.12%)

244290/199579901*100

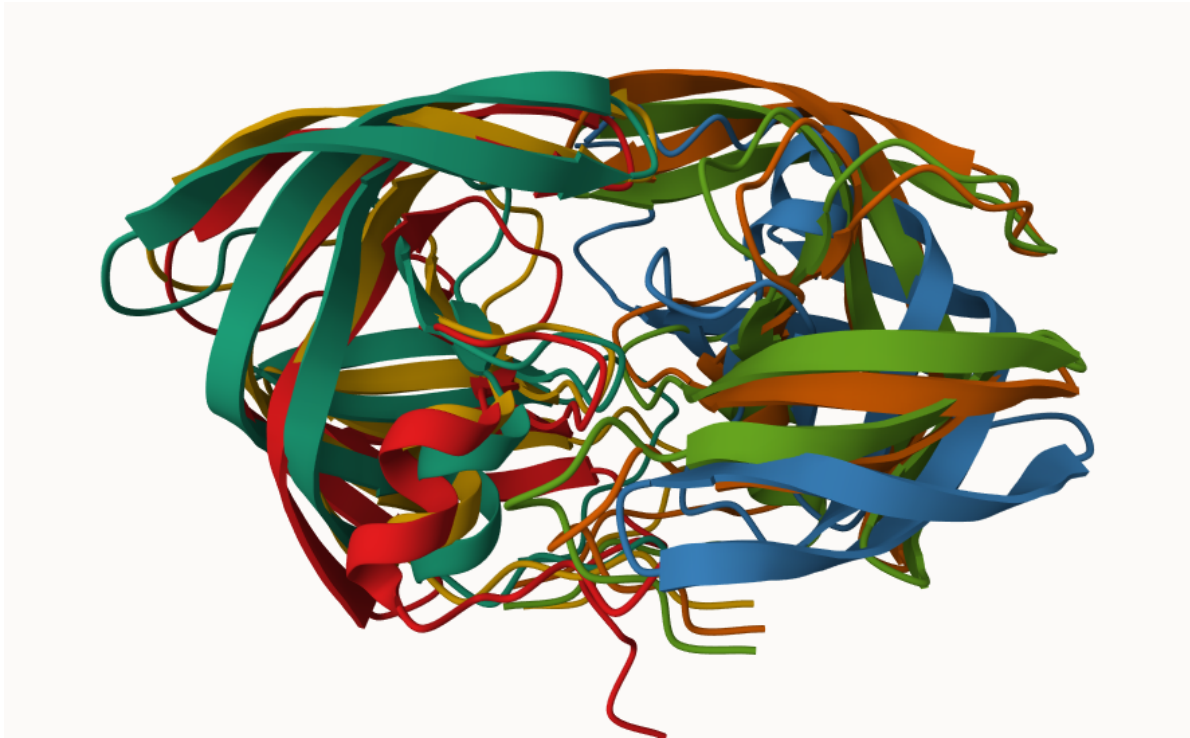
[1] 0.1224021

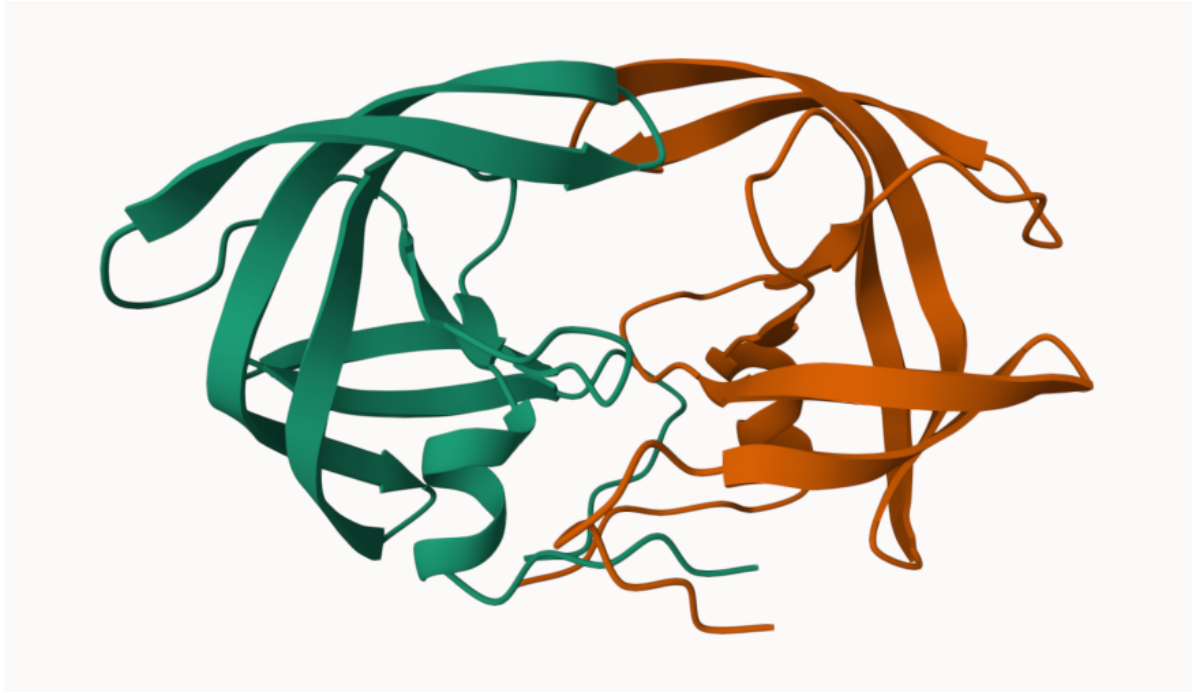
AFDB is attempting to address this gap..

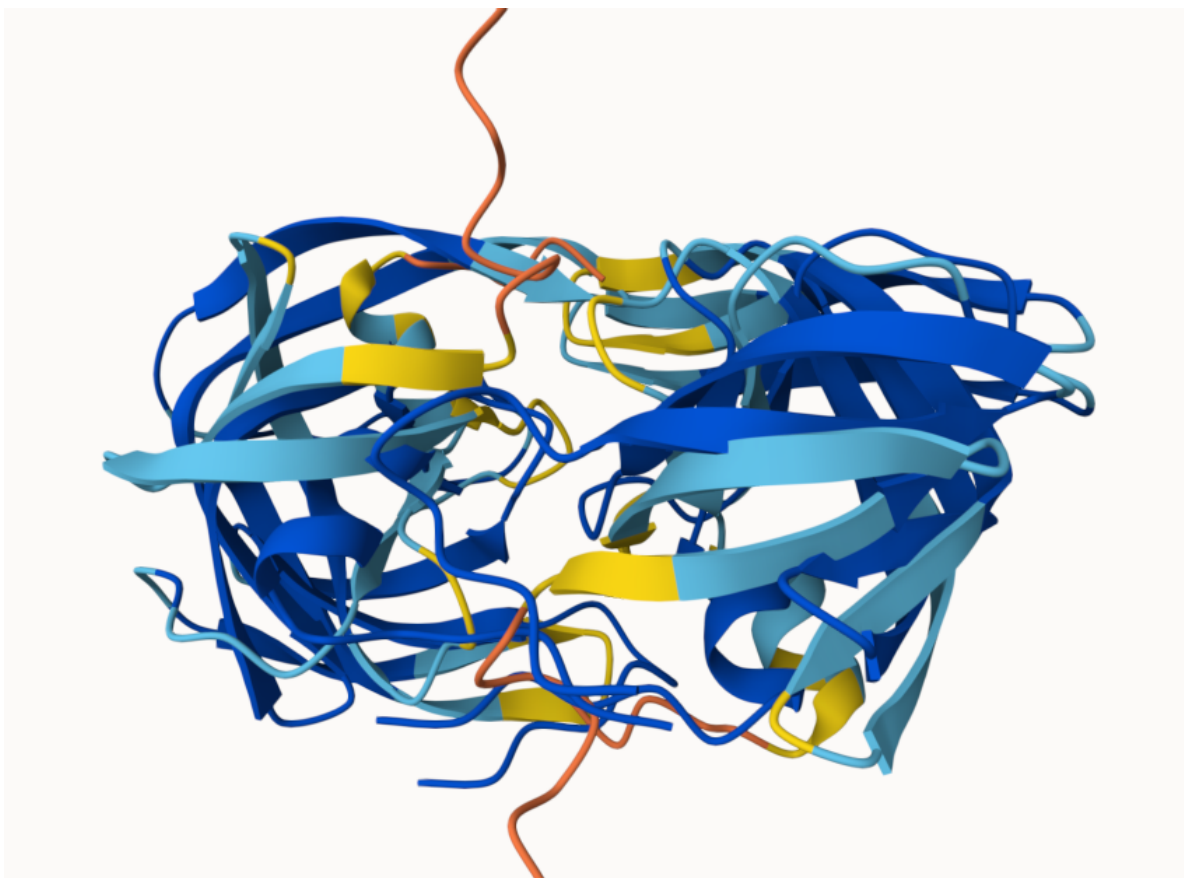
There are two “Quality Scores” from the Alphafold one for residues (ie, each amino acid) called pLDDT score. The other PAE score that measures the confidence in the relative position of two residues (i.e a score for every)

Generating your own structure predictions

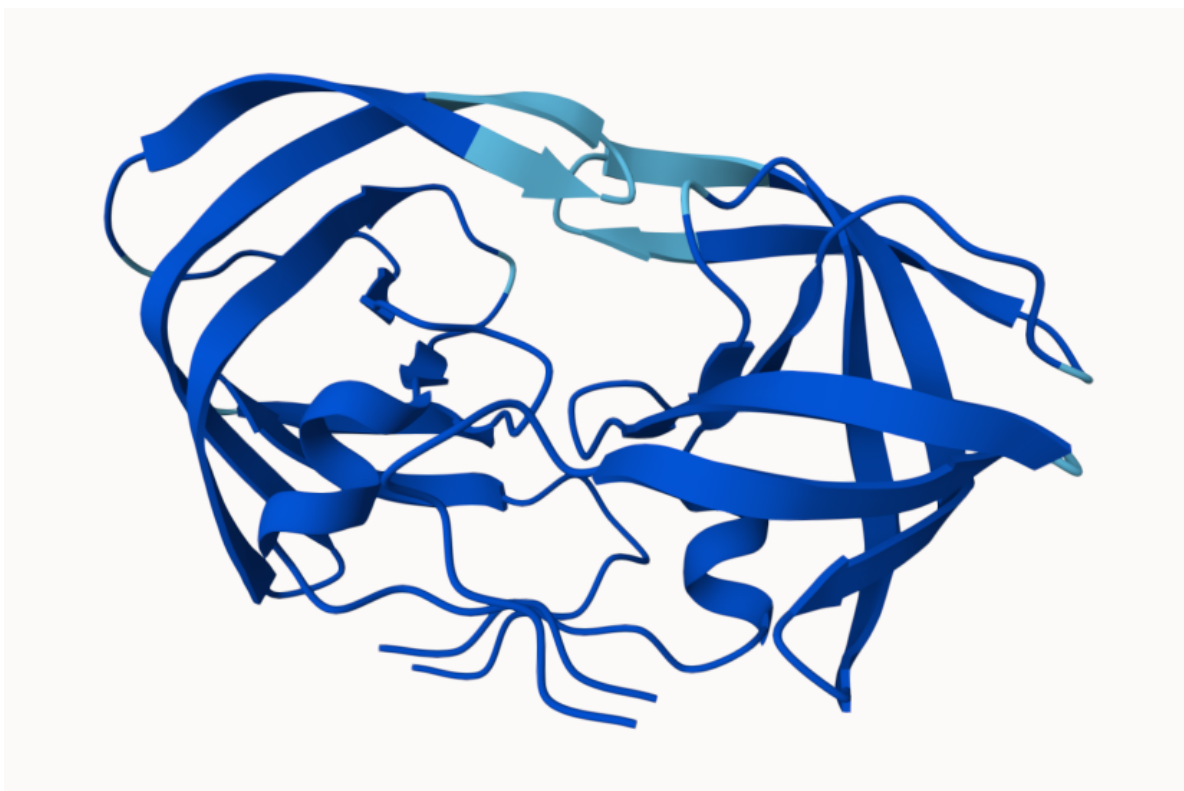
figure of 5 generated HIV PR







pLDDT score for model 1



Custom analysis of resulting models in R

Read Key result files into R, The first thing I need to know is what my results directory /folder is called.

```
results_dir <-"HIVPR_dimer_23119"  
pdb_files <- list.files(path=results_dir,  
                        pattern="*.pdb",  
                        full.names = TRUE)
```

```
# Print our PDB file names  
basename(pdb_files)
```

```
[1] "HIVPR_dimer_23119_unrelaxed_rank_001_alphafold2_multimer_v3_model_4_seed_000.pdb"  
[2] "HIVPR_dimer_23119_unrelaxed_rank_002_alphafold2_multimer_v3_model_1_seed_000.pdb"  
[3] "HIVPR_dimer_23119_unrelaxed_rank_003_alphafold2_multimer_v3_model_5_seed_000.pdb"  
[4] "HIVPR_dimer_23119_unrelaxed_rank_004_alphafold2_multimer_v3_model_2_seed_000.pdb"  
[5] "HIVPR_dimer_23119_unrelaxed_rank_005_alphafold2_multimer_v3_model_3_seed_000.pdb"
```

```
library(bio3d)
```

Warning: package 'bio3d' was built under R version 4.4.3

```
m1<- read.pdb(pdb_files[1])
m1
```

Call: read.pdb(file = pdb_files[1])

Total Models#: 1
Total Atoms#: 1514, XYZs#: 4542 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 0 (residues: 0)
Non-protein/nucleic resid values: [none]

Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, calpha, call

```
m1_A <- trim.pdb(m1, chain = "A")
m1_A
```

Call: trim.pdb(pdb = m1, chain = "A")

Total Models#: 1
Total Atoms#: 757, XYZs#: 2271 Chains#: 1 (values: A)

Protein Atoms#: 757 (residues/Calpha atoms#: 99)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 0 (residues: 0)

Non-protein/nucleic resid values: [none]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF

+ attr: atom, helix, sheet, seqres, xyz,
calpha, call

```
unique(m1$atom$chain)
```

```
[1] "A" "B"
```

```
pdbbs <- pdbaln(pdb_files, fit=TRUE, exefile="msa")
```

Reading PDB files:

HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_001_alphafold2_multimer_v3_model_4_seed_001.pdb
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_002_alphafold2_multimer_v3_model_1_seed_001.pdb
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_003_alphafold2_multimer_v3_model_5_seed_001.pdb
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_004_alphafold2_multimer_v3_model_2_seed_001.pdb
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_005_alphafold2_multimer_v3_model_3_seed_001.pdb
.....

Extracting sequences

pdb/seq: 1 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_001_alphafold2_multimer_v3_model_4_seed_001.pdb
pdb/seq: 2 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_002_alphafold2_multimer_v3_model_1_seed_001.pdb
pdb/seq: 3 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_003_alphafold2_multimer_v3_model_5_seed_001.pdb
pdb/seq: 4 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_004_alphafold2_multimer_v3_model_2_seed_001.pdb
pdb/seq: 5 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_005_alphafold2_multimer_v3_model_3_seed_001.pdb

```
pdbbs
```

```
1 . . . 50
[Truncated_Name:1]HIVPR_dime PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI
[Truncated_Name:2]HIVPR_dime PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI
[Truncated_Name:3]HIVPR_dime PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI
[Truncated_Name:4]HIVPR_dime PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI
[Truncated_Name:5]HIVPR_dime PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI
*****
```

```

1 . . . . 50

51 . . . . 100
[Truncated_Name:1]HIVPR_dime GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP
[Truncated_Name:2]HIVPR_dime GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP
[Truncated_Name:3]HIVPR_dime GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP
[Truncated_Name:4]HIVPR_dime GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP
[Truncated_Name:5]HIVPR_dime GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP
*****
51 . . . . 100

101 . . . . 150
[Truncated_Name:1]HIVPR_dime QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMKMIGGIG
[Truncated_Name:2]HIVPR_dime QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMKMIGGIG
[Truncated_Name:3]HIVPR_dime QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMKMIGGIG
[Truncated_Name:4]HIVPR_dime QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMKMIGGIG
[Truncated_Name:5]HIVPR_dime QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMKMIGGIG
*****
101 . . . . 150

151 . . . . 198
[Truncated_Name:1]HIVPR_dime GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF
[Truncated_Name:2]HIVPR_dime GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF
[Truncated_Name:3]HIVPR_dime GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF
[Truncated_Name:4]HIVPR_dime GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF
[Truncated_Name:5]HIVPR_dime GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF
*****
151 . . . . 198

```

Call:

```
pdbaln(files = pdb_files, fit = TRUE, exefile = "msa")
```

Class:

```
pdbs, fasta
```

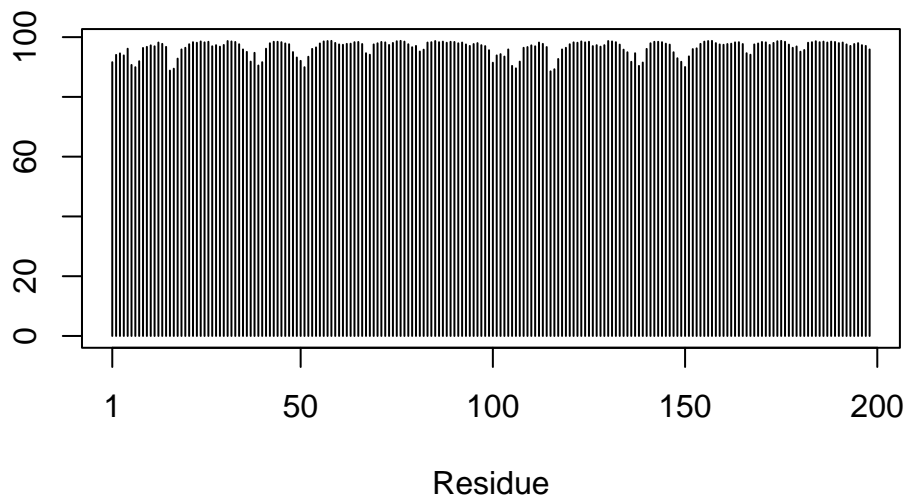
Alignment dimensions:

```
5 sequence rows; 198 position columns (198 non-gap, 0 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```



```
plotb3(m1$atom$b[m1$calpha])
```



Residue conservation from alignment file

```
aln_file <- list.files(path=results_dir,  
                      pattern=".a3m$",  
                      full.names = TRUE)  
aln_file
```

```
[1] "HIVPR_dimer_23119/HIVPR_dimer_23119.a3m"
```

```
aln <- read.fasta(aln_file[1], to.upper = TRUE)
```

```
[1] " ** Duplicated sequence id's: 101 **"  
[2] " ** Duplicated sequence id's: 101 **"
```

```
dim(aln$ali)
```

```
[1] 5397 132
```

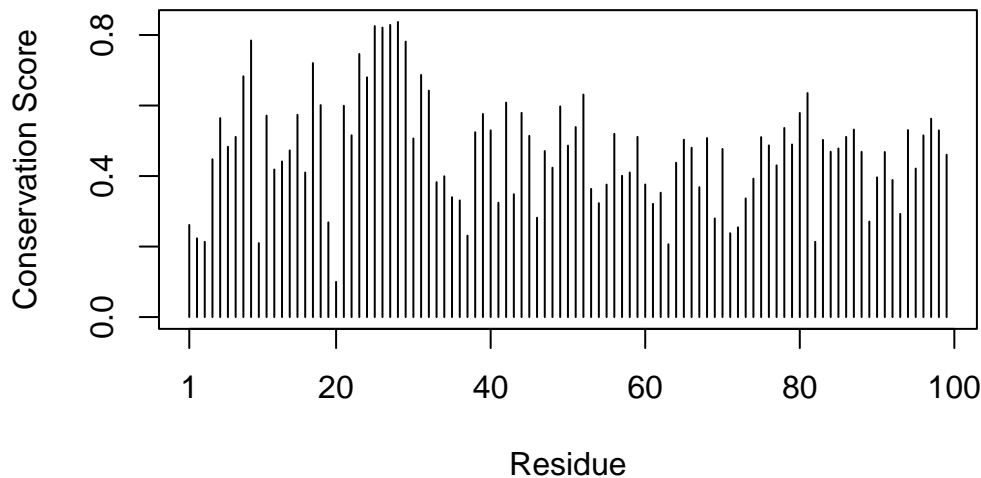
```
sim <- conserv(aln)

plotb3(sim[1:99],(m1$atom$b[m1$calpha]) ,
      sse=m1_A,
      ylab="Conservation Score")
```

Warning in plotb3(sim[1:99], (m1\$atom\$b[m1\$calpha]), sse = m1_A, ylab = "Conservation Score"): Length of input 'resno' does not equal the length of input 'x'; Ignoring 'resno'

Warning in pdb2sse(sse): No helix and sheet defined in input 'sse' PDB object: try using dssp()

Warning in plotb3(sim[1:99], (m1\$atom\$b[m1\$calpha]), sse = m1_A, ylab = "Conservation Score"): Length of input 'sse' does not equal the length of input 'x'; Ignoring 'sse'



```
con <- consensus(aln, cutoff = 0.9)
con$seq
```

```

[1] "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-"
[19] "-" "-" "-" "-" "-" "-" "D" "T" "G" "A" "-" "-" "-" "-" "-" "-" "-" "-"
[37] "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-"
[55] "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-"
[73] "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-"
[91] "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-"
[109] "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-" "-"
[127] "-" "-" "-" "-" "-" "-"

```