

class 11: structural bioinformatics pt1

Rachel Galleta (A:16859649)

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AlphaFold Data Base (AFBD)

the EBI maintains the largest database of Alphafold structure prediction models at:
from last class (before Halloween) we saw that the PDB had 244,290 (Oct 2025)
the total number of proteins sequences in UniprotKB is 199,579,901.

Key point: This is a tiny fraction of sequence space that has structural coverage
(0.12%)

244290/199579901*100

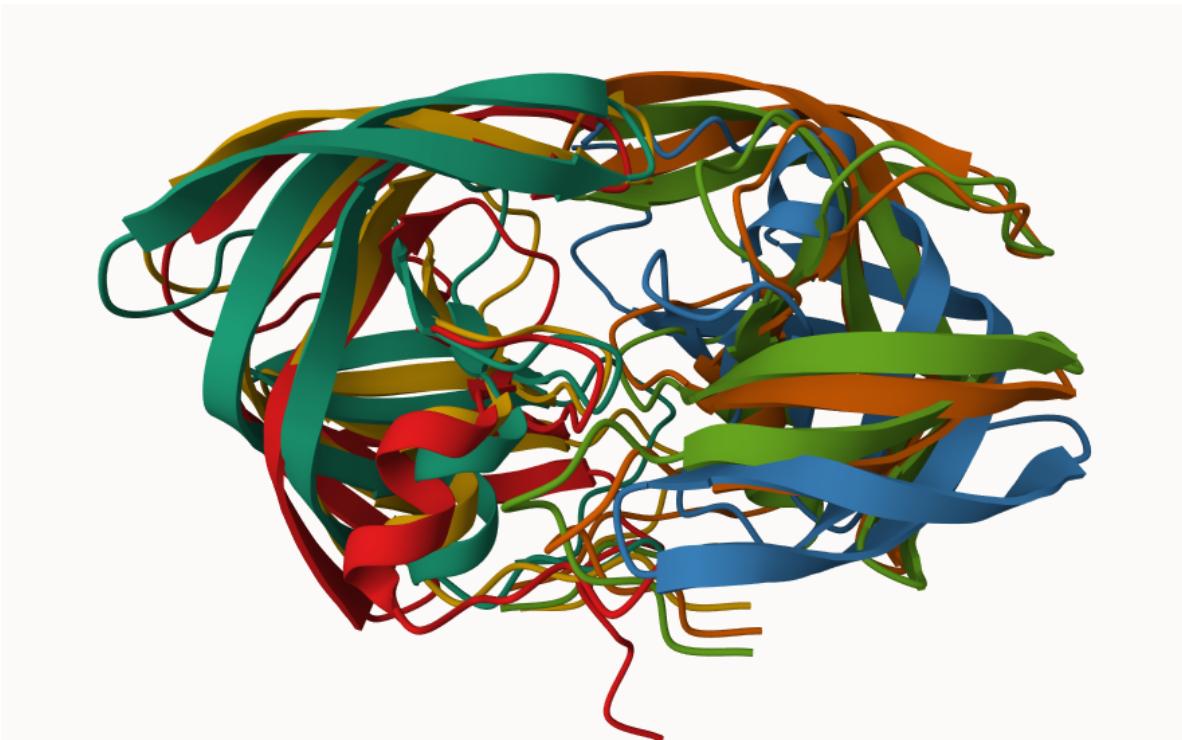
[1] 0.1224021

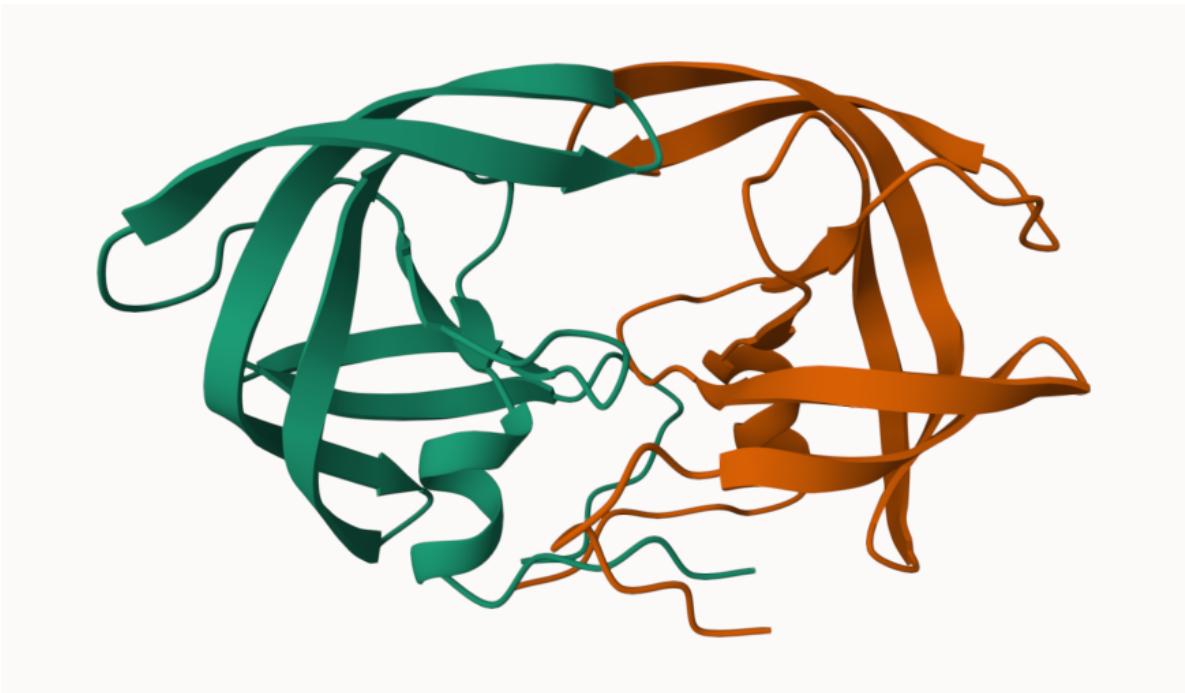
AFDB is attempting to address this gap..

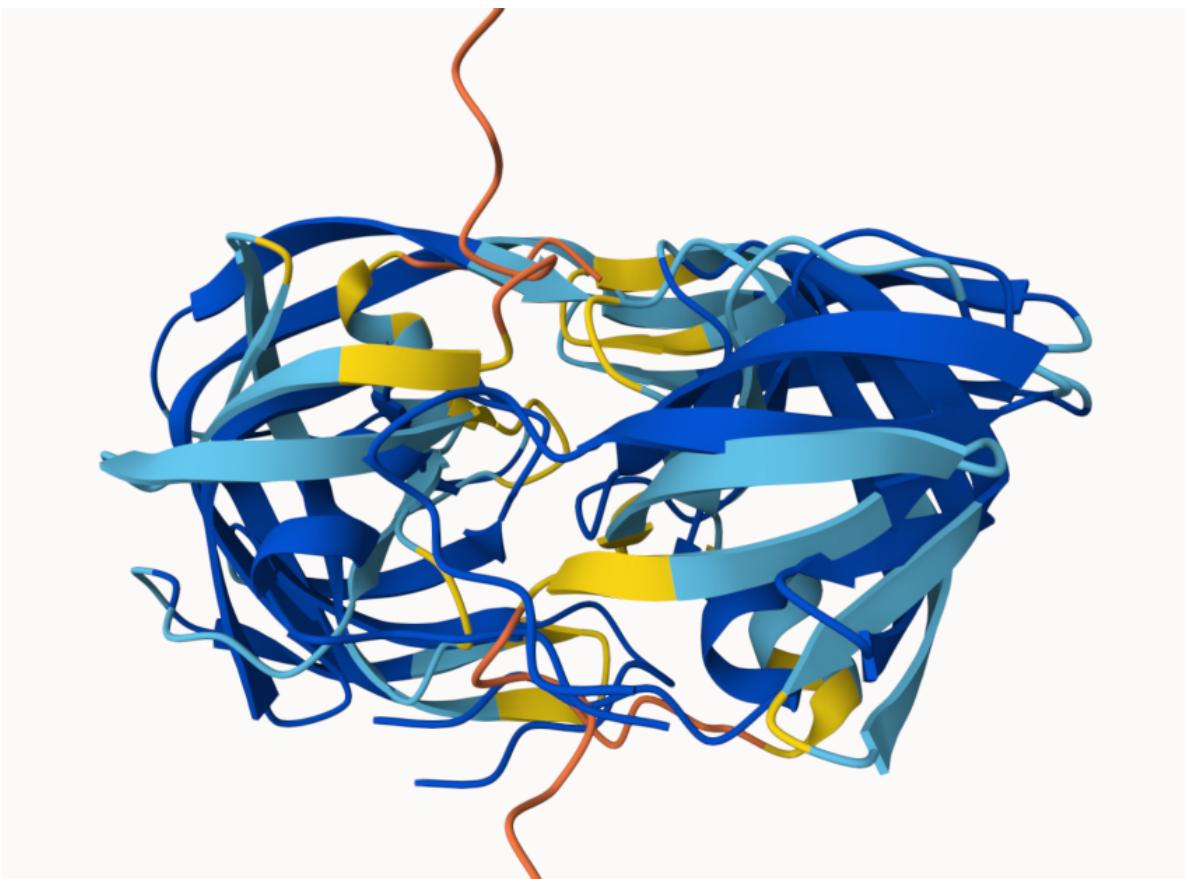
There are two “Quality Scores” from the AlphaFold one for residues (ie, each amino acid) called pLDDT score. The other PAE score that measures the confidence in the relative position of two residues(i.e a score for every)

Generating your own structure predictions

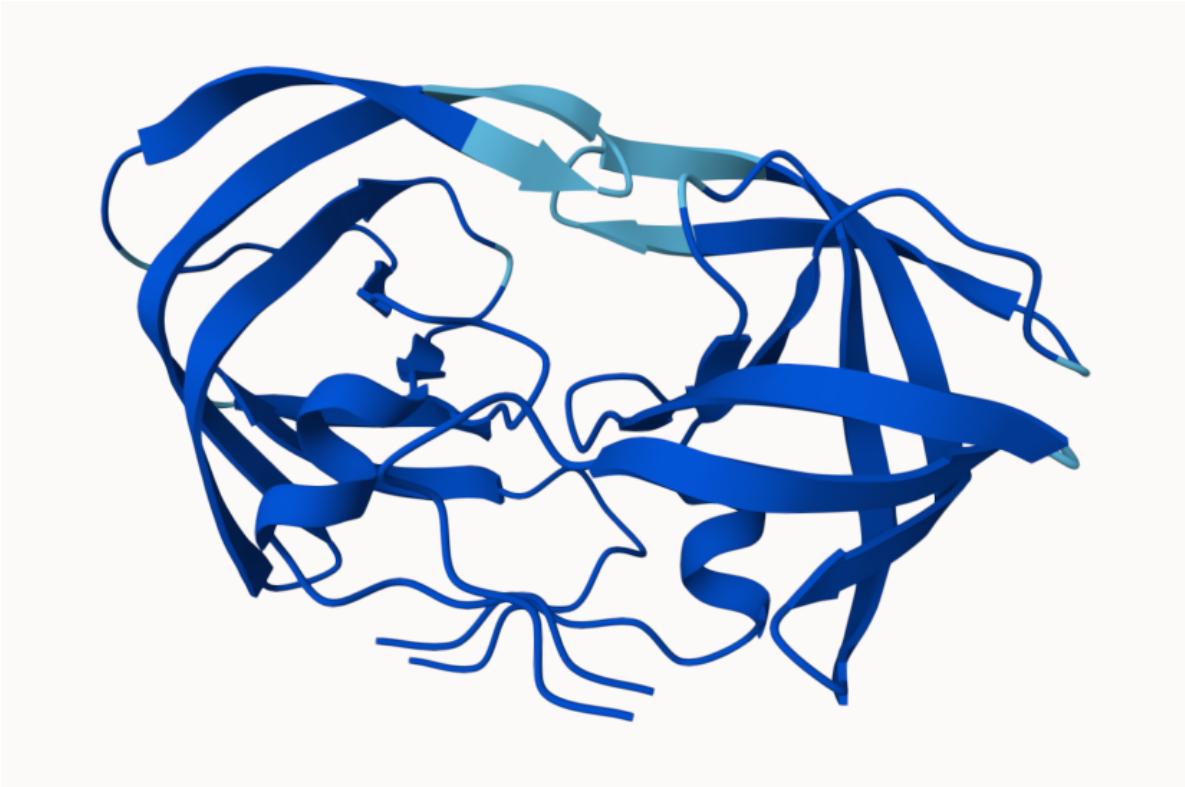
figure of 5 generated HIV PR







pLDDT score for model 1



Custom analysis of resulting models in R

Read Key result files into R, The first thing I need to know is what my results directory /folder is called.

```
results_dir <- "HIVPR_dimer_23119"  
pdb_files <- list.files(path=results_dir,  
                         pattern="*.pdb",  
                         full.names = TRUE)  
  
# Print our PDB file names  
basename(pdb_files)
```

```
[1] "HIVPR_dimer_23119_unrelaxed_rank_001_alphaFold2_multimer_v3_model_4_seed_000.pdb"  
[2] "HIVPR_dimer_23119_unrelaxed_rank_002_alphaFold2_multimer_v3_model_1_seed_000.pdb"  
[3] "HIVPR_dimer_23119_unrelaxed_rank_003_alphaFold2_multimer_v3_model_5_seed_000.pdb"  
[4] "HIVPR_dimer_23119_unrelaxed_rank_004_alphaFold2_multimer_v3_model_2_seed_000.pdb"  
[5] "HIVPR_dimer_23119_unrelaxed_rank_005_alphaFold2_multimer_v3_model_3_seed_000.pdb"
```

```
library(bio3d)
```

Warning: package 'bio3d' was built under R version 4.4.3

```
m1<- read.pdb(pdb_files[1])  
m1
```

Call: read.pdb(file = pdb_files[1])

Total Models#: 1

Total Atoms#: 1514, XYZs#: 4542 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 0 (residues: 0)

Non-protein/nucleic resid values: [none]

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWP  
KPKMIGGIGGF  
IKVRQYD  
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP  
QITLWQRPLVTIKIGGQLKE  
ALLDTGADDTVLEEMSLPGRWP  
KPKMIGGIGGF  
IKVRQYDQILIEICGHKAIGTVLVGPTP  
VNIIGRNLLTQIGCTLNF
```

+ attr: atom, xyz, calpha, call

```
m1_A <- trim.pdb(m1, chain = "A")  
m1_A
```

Call: trim.pdb(pdb = m1, chain = "A")

Total Models#: 1

Total Atoms#: 757, XYZs#: 2271 Chains#: 1 (values: A)

Protein Atoms#: 757 (residues/Calpha atoms#: 99)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 0 (residues: 0)

Non-protein/nucleic resid values: [none]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEMSLPGRWPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF

+ attr: atom, helix, sheet, seqres, xyz,
calpha, call

```
unique(m1$atom$chain)
```

```
[1] "A" "B"
```

```
pdb5 <- pdbaln(pdb_files, fit=TRUE, exefile="msa")
```

Reading PDB files:

HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_001_alphaFold2_multimer_v3_model_4_seed_00
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_002_alphaFold2_multimer_v3_model_1_seed_00
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_003_alphaFold2_multimer_v3_model_5_seed_00
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_004_alphaFold2_multimer_v3_model_2_seed_00
HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_005_alphaFold2_multimer_v3_model_3_seed_00
.....

Extracting sequences

```
pdb/seq: 1 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_001_alphafold2_multime  
pdb/seq: 2 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_002_alphafold2_multime  
pdb/seq: 3 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_003_alphafold2_multime  
pdb/seq: 4 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_004_alphafold2_multime  
pdb/seq: 5 name: HIVPR_dimer_23119/HIVPR_dimer_23119_unrelaxed_rank_005_alphafold2_multime
```

pdbs

	1	50
[Truncated_Name:1]	HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI				
[Truncated_Name:2]	HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI				
[Truncated_Name:3]	HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI				
[Truncated_Name:4]	HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI				
[Truncated_Name:5]	HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGI				

	1	50
	51	100
[Truncated_Name:1]	HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:2]	HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:3]	HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:4]	HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:5]	HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				

	51	100
	101	150
[Truncated_Name:1]	HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIG				
[Truncated_Name:2]	HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIG				
[Truncated_Name:3]	HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIG				
[Truncated_Name:4]	HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIG				
[Truncated_Name:5]	HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIG				

	101	150
	151	198
[Truncated_Name:1]	HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:2]	HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:3]	HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:4]	HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:5]	HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				

	151	198

Call:

```
pdbaln(files = pdb_files, fit = TRUE, exefile = "msa")
```

Class:

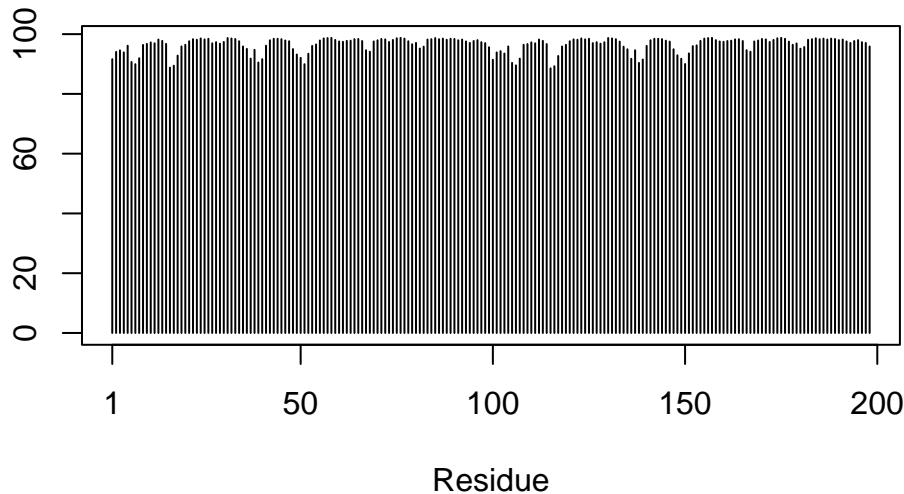
```
pdb, fasta
```

Alignment dimensions:

```
5 sequence rows; 198 position columns (198 non-gap, 0 gap)
```

```
+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

```
plotb3(m1$atom$b[m1$calpha])
```



Residue conservation from alignment file

```
aln_file <- list.files(path=results_dir,
                        pattern=".a3m$",
                        full.names = TRUE)
aln_file
```

```
[1] "HIVPR_dimer_23119/HIVPR_dimer_23119.a3m"
```

```
aln <- read.fasta(aln_file[1], to.upper = TRUE)
```

```
[1] " ** Duplicated sequence id's: 101 **"
[2] " ** Duplicated sequence id's: 101 **"
```

```
dim(aln$ali)
```

```
[1] 5397 132
```

```

sim <- conserv(aln)

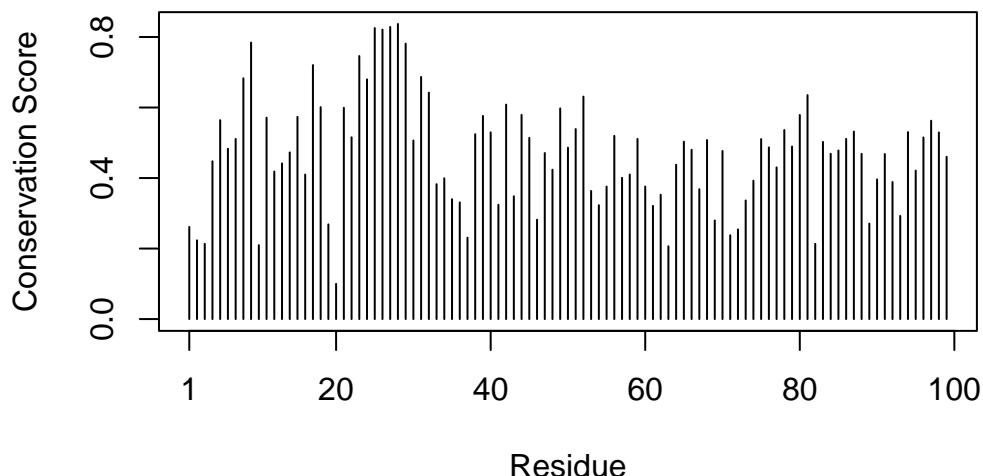
plotb3(sim[1:99], (m1$atom$b[m1$calpha]) ,
       sse=m1_A,
       ylab="Conservation Score")

```

Warning in plotb3(sim[1:99], (m1\$atom\$b[m1\$calpha]), sse = m1_A, ylab = "Conservation Score"): Length of input 'resno' does not equal the length of input 'x'; Ignoring 'resno'

Warning in pdb2sse(sse): No helix and sheet defined in input 'sse' PDB object: try using dssp()

Warning in plotb3(sim[1:99], (m1\$atom\$b[m1\$calpha]), sse = m1_A, ylab = "Conservation Score"): Length of input 'sse' does not equal the length of input 'x'; Ignoring 'sse'



```

con <- consensus(aln, cutoff = 0.9)
con$seq

```

