## Class09

## Ralph Goguanco

```
DataExp.df <- read.csv("E:\\Downloads\\DataExportSummary.csv")
DataExp.df</pre>
```

```
##
                                              EM Multiple.methods Neutron Other
              Molecular.Type X.ray
                                        NMR
## 1
              Protein (only) 144616 11881 6759
                                                               185
                                                                         70
## 2 Protein/Oligosaccharide
                                                                 5
                                                                          0
                                                                                0
                                8551
                                         31 1133
                  Protein/NA
                                7623
                                        274 2183
                                                                 3
                                                                                0
         Nucleic acid (only)
                                                                 8
                                                                          2
## 4
                                2396 1399
                                              61
                                                                                1
## 5
                        Other
                                 154
                                         31
                                               3
                                                                          0
                                                                                0
                                                                          0
                                          6
                                               0
                                                                                4
## 6
     Oligosaccharide (only)
                                  11
      Total
##
## 1 163543
## 2
       9720
## 3
     10083
## 4
       3867
## 5
        188
## 6
         22
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

## (163351+10139)/187423

### ## [1] 0.9256601

Q2: What proportion of structures in the PDB are protein?

#### 163543/187423

#### ## [1] 0.8725877

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

There are 4496 total structures in the current PDB

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

It would be too crowded if we added three atoms per water molecule since there's a lot of them.

Q5: There is a conserved water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have (see note below)?

The residue number of the atom associated with the water molecule is ALA28:CA

##3. Introduction to BIO3D in R

```
library(bio3d)
read.pdb(file = "1hsg")

## Note: Accessing on-line PDB file

##
## Call: read.pdb(file = "1hsg")
##
## Total Models#: 1
## Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
##
```

Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)

Non-protein/nucleic Atoms#: 172 (residues: 128)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP

VNIIGRNLLTQIGCTLNF

## + attr: atom, xyz, seqres, helix, sheet,
## calpha, remark, call

Q7: How many amino acid residues are there in this pdb object?

There are 198 amino acid residues in this pdb object.

Q8: Name one of the two non-protein residues?

Water and  ${\rm MK1}$ 

Q9: How many protein chains are in this structure?

There are 2 protein chains in this structure

Q10. Which of the packages above is found only on BioConductor and not CRAN?

msa

##

##

## ##

## ## ##

##

##

##

## ##

Q11. Which of the above packages is not found on BioConductor or CRAN?:

#### bio3d-view

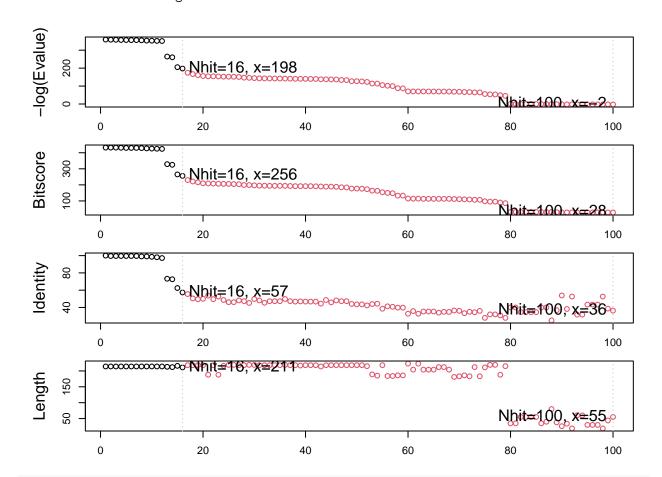
Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?

#### TRUE

```
library(bio3d)
aa <- get.seq("1ake_A")</pre>
## Warning in get.seq("lake_A"): Removing existing file: seqs.fasta
## Fetching... Please wait. Done.
##
                                                                                60
  pdb|1AKE|A
                \tt MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLVT
##
##
                61
                                                                                120
                DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
  pdb|1AKE|A
##
                                                                                120
##
##
               121
                                                                                180
  pdb|1AKE|A
                 VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
##
              121
                                                                                180
##
                                                    214
##
                 YYSKEAEAGNTKYAKVDGTKPVAEVRADLEKILG
## pdb|1AKE|A
##
               181
##
## Call:
##
     read.fasta(file = outfile)
##
## Class:
##
     fasta
##
## Alignment dimensions:
##
     1 sequence rows; 214 position columns (214 non-gap, 0 gap)
##
## + attr: id, ali, call
     Q13. How many amino acids are in this sequence, i.e. how long is this sequence?
214
# Blast or hmmer search
b <- blast.pdb(aa)</pre>
    Searching ... please wait (updates every 5 seconds) RID = 1FYHBM5D016
##
## Reporting 100 hits
```

# # Plot a summary of search results hits <- plot(b)</pre>

## \* Possible cutoff values: 197 -3
## Yielding Nhits: 16 100
##
## \* Chosen cutoff value of: 197
## Yielding Nhits: 16



hits <- NULL hits\$pdb.id <- c('1AKE\_A','6S36\_A','6RZE\_A','3HPR\_A','1E4V\_A','5EJE\_A','1E4Y\_A','3X2S\_A','6HAP\_A','6HAM

```
#Download related PDB files
files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)</pre>
```

## Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 1AKE.pdb exists. Skipping download

## Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6S36.pdb exists. Skipping download

## Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6RZE.pdb exists. Skipping download

```
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 3HPR.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 1E4V.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 5EJE.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 1E4Y.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 3X2S.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6HAP.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 6HAM.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4K46.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 3GMT.pdb exists. Skipping download
## Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE): pdbs/
## 4PZL.pdb exists. Skipping download
##
# Align releated PDBs
#pdbs <- pdbaln(files, fit = TRUE), exefile="msa")</pre>
# Vector containing PDB codes for figure axis
#ids <- basename.pdb(pdbs$id)</pre>
# Draw schematic alignment
#(pdbs, labels=ids)
```

What do you note about this plot? Are the black and colored lines similar or different? Where do you think they differ most and why?

The black and colored lines are different, they differe in fluctuation heights but the patterns are almost similar.