



Metagenome Overview

MG-RAST ID 4633498.3 Download Analyze Search

Metagenome Name ssr_13603__R1__L001

PI

Organization

Visibility Private

Static Link <http://metagenomics.anl.gov/linkin.cgi?metagenome=4633498.3>

NCBI Proj

GOLD ID

PubMed ID

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METAGENOME SUMMARY

Dataset ssr_13603__R1__L001 was uploaded on 05/18/2015 and contains 43,577 sequences totaling 6,477,024 basepairs with an average length of 148 bps. The piechart below breaks down the uploaded sequences into 3 distinct categories.

2,769 sequences (6.4%) failed to pass the QC pipeline. Of the sequences that passed QC, 42,078 sequences (96.6%) contain ribosomal RNA genes. 0 (0.0%) of the sequences that passed QC have no rRNA genes.

The analysis results shown on this page are computed by MG-RAST. Please note that authors may upload data that they have published their own analysis for, in such cases comparison within the MG-RAST framework can not be done.



DOWNLOAD data and annotations



ANALYZE annotations in detail.



SEARCH through annotations.

Sequence Breakdown

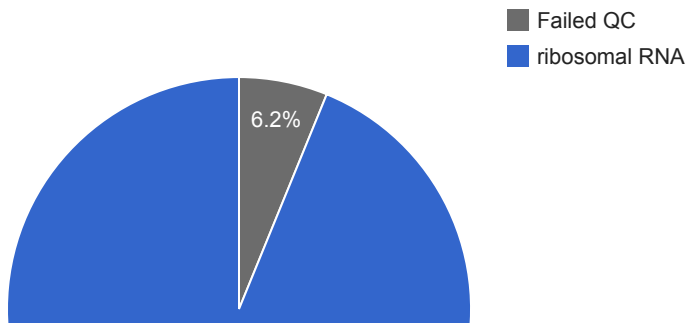


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*Note: Sequences containing multiple predicted features are only counted in one category.
Currently downloading of sequences via chart slices is not enabled.*

PROJECT INFORMATION

This dataset is part of project [Sprague-Jan2015-13603](#).
There are 0 other metagenomes in this project

- » find metagenomes within this project
- » find metagenomes within this biome
- » find metagenomes within this country
- » find metagenomes within 10 | 30 | 100 kilometers

ANALYSIS FLOWCHART

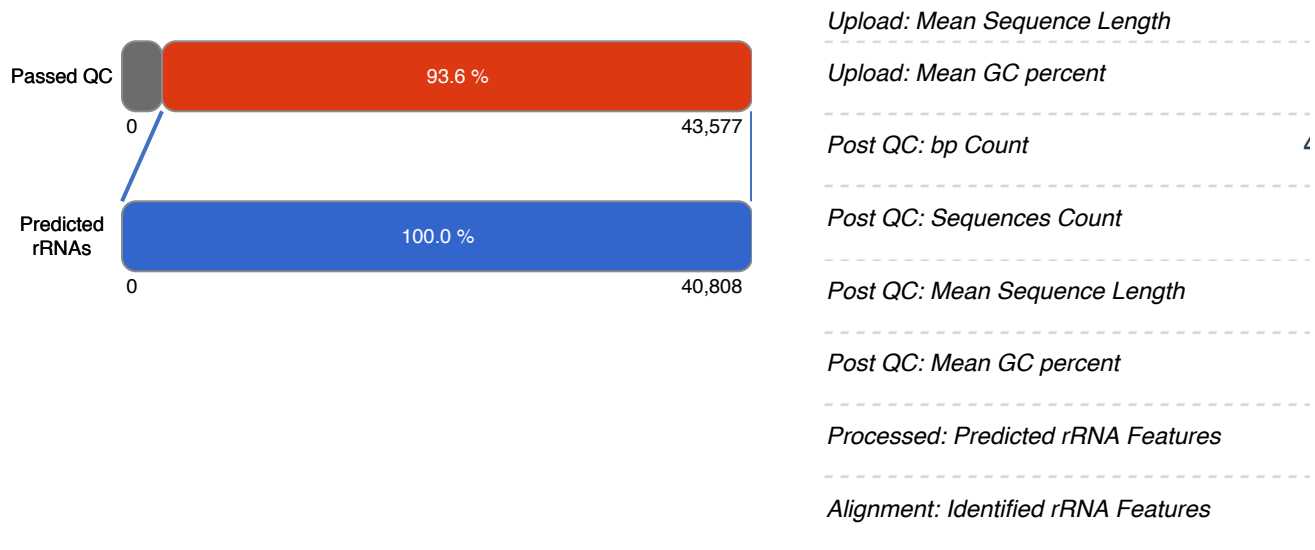
2,769 sequences failed quality control. Of the 40,808 sequences (totaling 4,753,862 bps) that passed quality control, 42,078 (103.1%) produced a total of 705 identified ribosomal RNAs.

GSC MIXS INFO

Investigation Type	mimark
Project Name	Sprague-Jan20
Latitude and Longitude	
Country and/or Sea, Location	
Collection Date	
Environment (Biome)	
Environment (Feature)	
Environment (Material)	
Environmental Package	
Sequencing Method	
More Metadata	

ANALYSIS STATISTICS

Upload: bp Count	6
Upload: Sequences Count	



DRISEE [?]

Duplicate Read Inferred Sequencing Error Estimation (Keegan et al., PLoS Computational Biology, 2012)

DRISEE could not produce a profile, this is an Amplicon dataset.

DRISEE is a tool that utilizes artificial duplicate reads (ADRs) to provide a platform independent assessment of sequencing data. DRISEE is designed to consider shotgun data. Currently, it is not appropriate for amplicon data.

Note that DRISEE is designed to examine sequencing error in raw whole genome shotgun sequence data. It has been removed, but that the sequence data have not been modified in any additional way. (e.g.) Assembly or merging DRISEE's ability to provide an accurate assessment of error by removing error before it is analyzed.

KMER PROFILES [?] [HIDE](#)

Redraw the below plot using the following kmer-plot type:

kmer rank abundance ▼

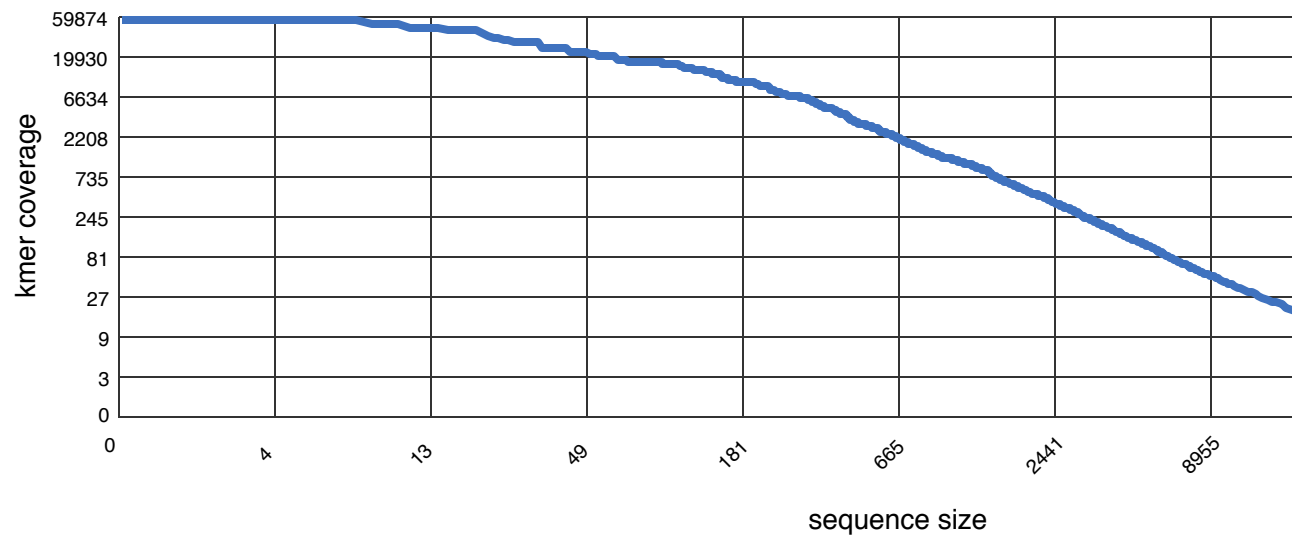
15-mer ▼

The kmer abundance spectra are tools to summarize the redundancy (repetitiveness) of sequence datasets by counting sequences.

The kmer spectrum plots the number of distinct N-bp sequences as a function of coverage level, placing low-coverage repetitive sequences at right. The kmer rank abundance graph plots the kmer coverage as a function of abundance. The ranked kmer consumed graph shows the fraction of the dataset that is explained by the most abundant kmers.

[Download chart data](#)

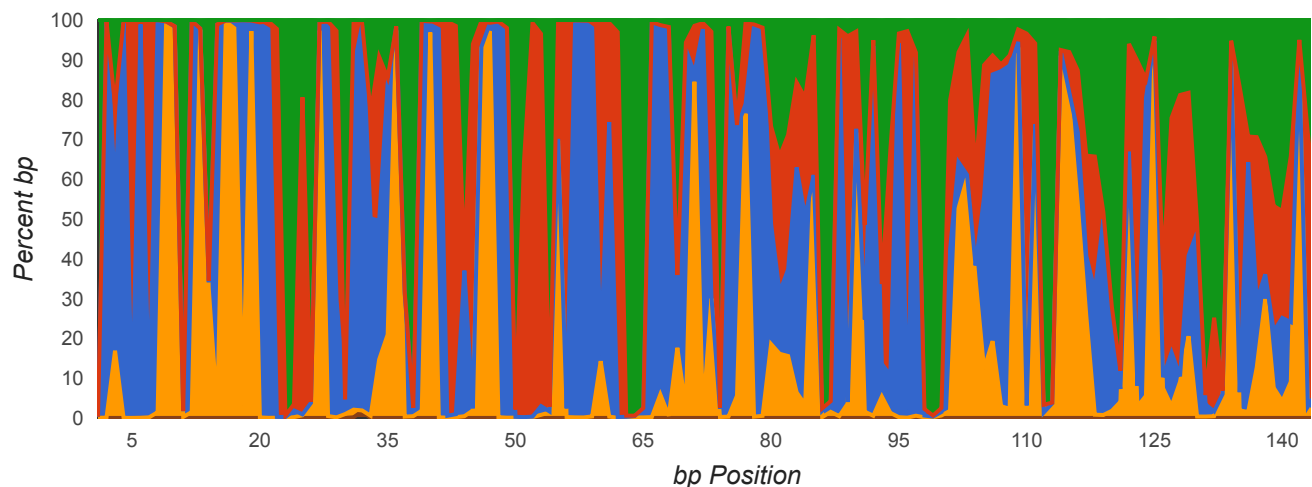
The image is currently dynamic. To be able to right-click/save the image, please click the static button [static](#)



NUCLEOTIDE POSITION HISTOGRAM [\[?\]](#) [HIDE](#)

These graphs show the fraction of base pairs of each type (A, C, G, T, or ambiguous base "N") at each position starting from the first 151 base pairs. Amplicon datasets should show consensus sequences; shotgun datasets should have roughly equal proportions of each base type.

[Download chart data](#)



SOURCE HITS DISTRIBUTION [\[?\]](#) [HIDE](#)

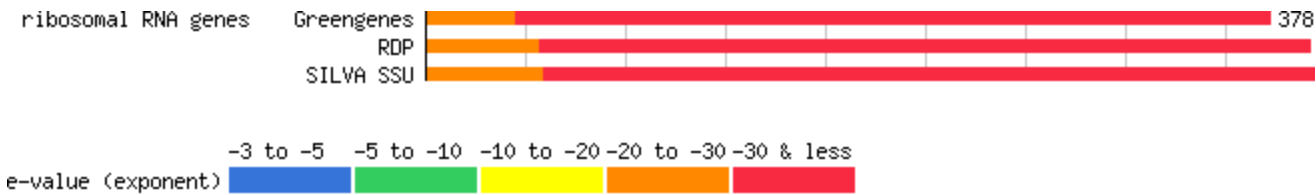
42,078 (96.6%) of reads had similarity to ribosomal RNA genes.

The graph below displays the number of features in this dataset that were annotated by the different databases belonging to different functional hierarchies, and ribosomal RNA databases. The bars representing annotated ribosomal RNA databases have different numbers of hits, but can also have different types of annotation data.

There are 15,945,780 sequences in the M5NR protein database and 309,342 sequences in the M5RNA ribosomal database. The unique sequences from the below protein databases and the M5RNA ribosomal database contain all the unique sequences.

databases.

Download chart data

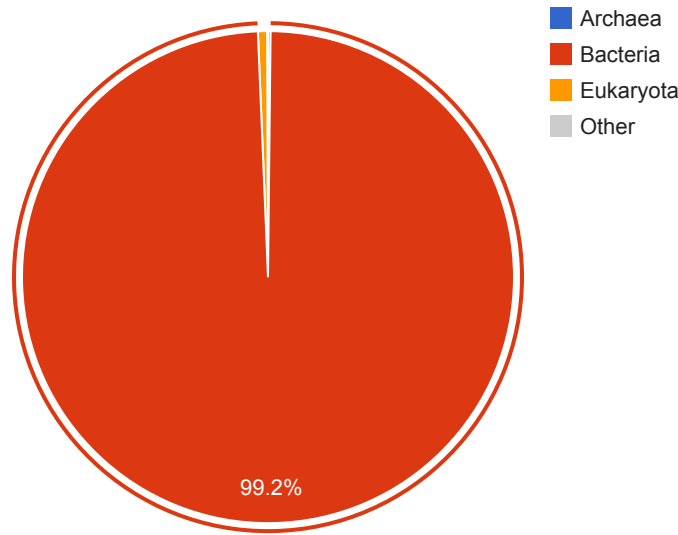


TAXONOMIC HITS DISTRIBUTION [HIDE](#)

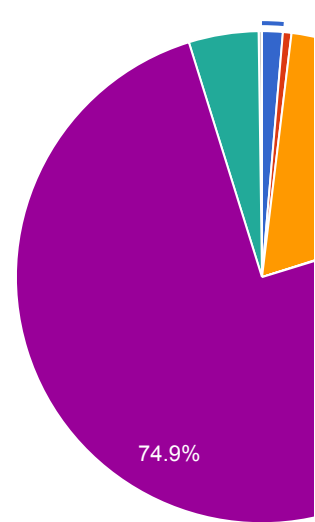
The pie charts below illustrate the distribution of taxonomic domains, phyla, and orders for the annotations. Each slice represents the percentage of proteins and ribosomal RNA genes annotated to the indicated taxonomic level. This information is based on all the annotations. An interactive [Krona](#) chart of the full taxonomy is also available. Click on a slice or legend to view all sequences annotated with the indicated taxonomic level in the analysis page.

View taxonomic interactive chart

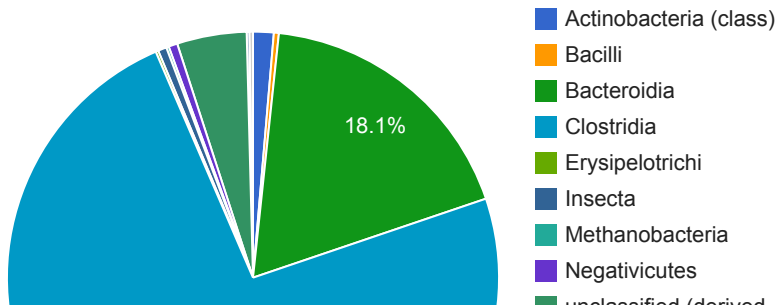
domain [Download chart data](#)



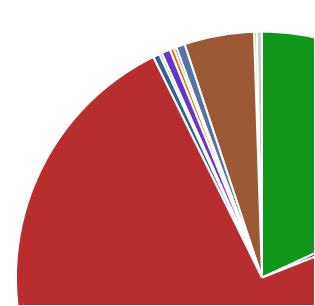
phylum [Download chart data](#)



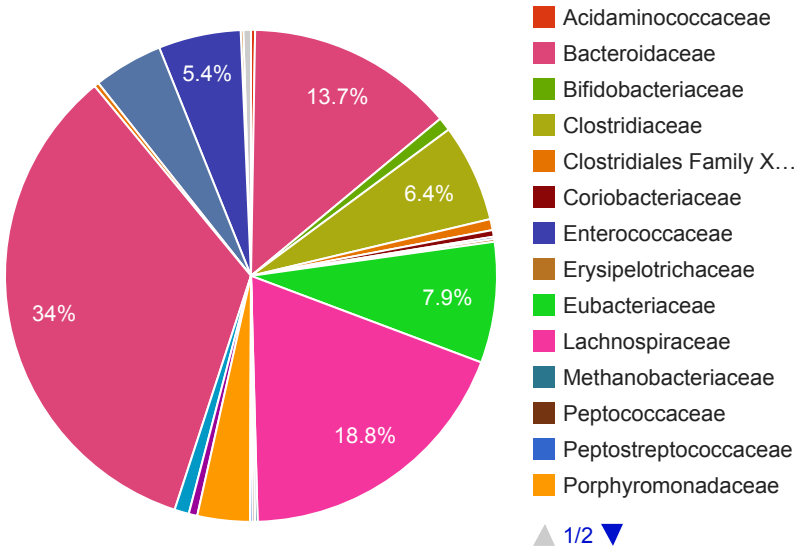
class [Download chart data](#)



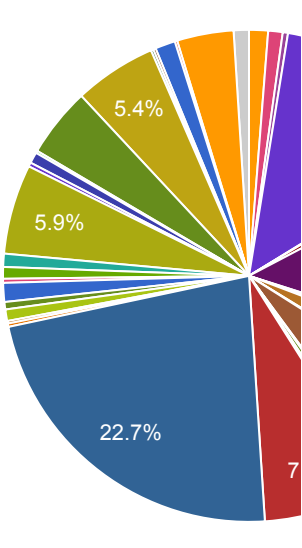
order [Download chart data](#)



family [Download chart data](#)



genus [Download chart data](#)



RANK ABUNDANCE PLOT [HIDE](#)

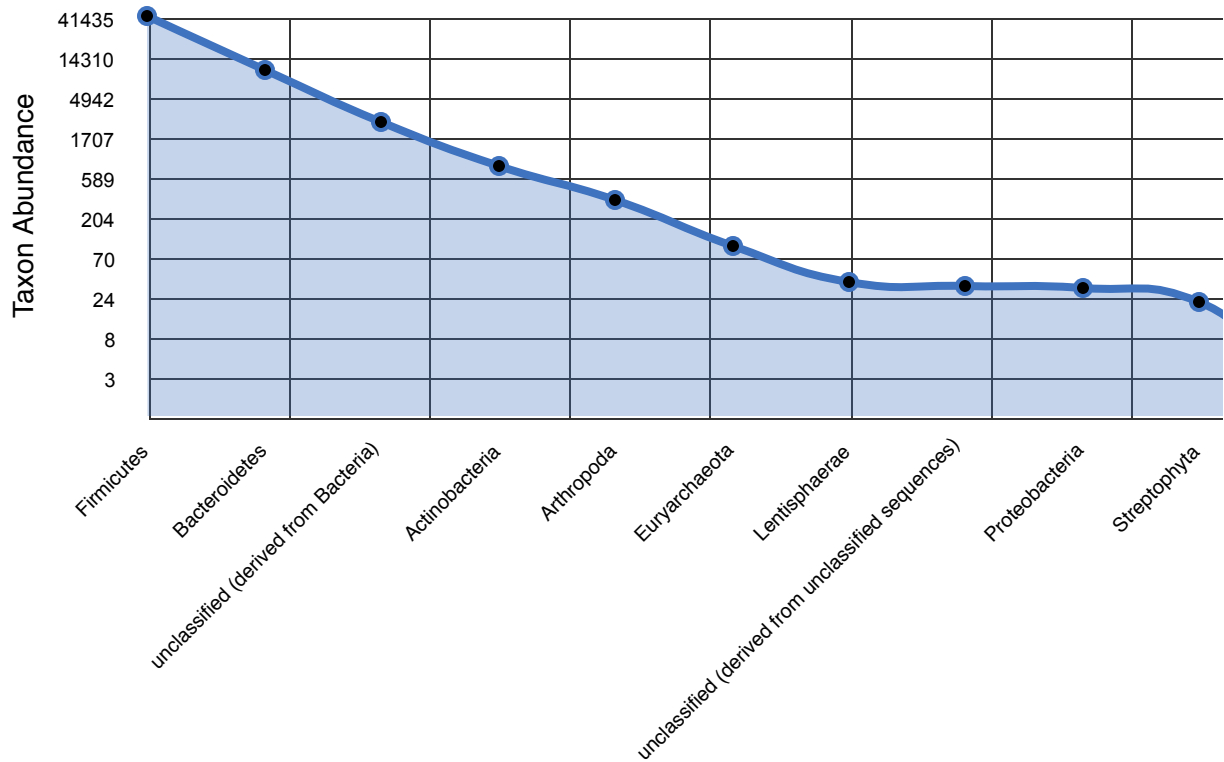
Redraw the below plot using the following taxonomic level: phylum

The plot below shows the phylum abundances ordered from the most abundant to least abundant. Only the top 50 abundances of annotations in each phylum on a log scale.

The rank abundance curve is a tool for visually representing taxonomic richness and evenness.

[Download chart data](#)

The image is currently dynamic. To be able to right-click/save the image, please click the static button static



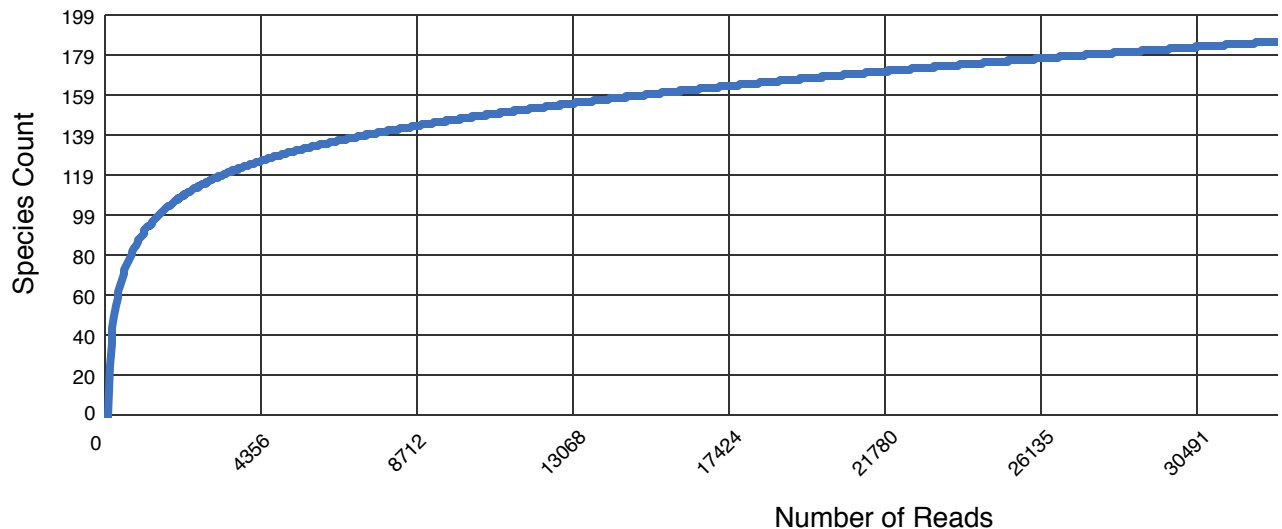
RAREFACTION CURVE [HIDE](#)

The plot below shows the rarefaction curve of annotated species richness. This curve is a plot of the total number of sequences sampled. On the left, a steep slope indicates that a large fraction of the species diversity remains to be sampled. On the right, a reasonable number of individuals is sampled: more intensive sampling is likely to yield only few additional species.

Sampling curves generally rise very quickly at first and then level off towards an asymptote as fewer new species are discovered. Rarefaction curves are calculated from the table of species abundance. The curves represent the average number of species expected to be found in a sample of the complete dataset.

[Download chart data](#)

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ALPHA DIVERSITY [\[?\]](#) [HIDE](#)

α -Diversity = 31.330 species



The above image shows the range of α -diversity values in project Sprague-Jan2015-13603. The min, max, and mean ranges (σ and 2σ) in different shades. The α -diversity of this metagenome is shown in red.

Alpha diversity summarizes the diversity of organisms in a sample with a single number. The alpha diversity of an distribution of the species-level annotations.

Annotated species richness is the number of distinct species annotations in the combined MG-RAST dataset. Share the logarithm of the relative abundances of annotated species. The species-level annotations are from all the anno

[Download source data](#)

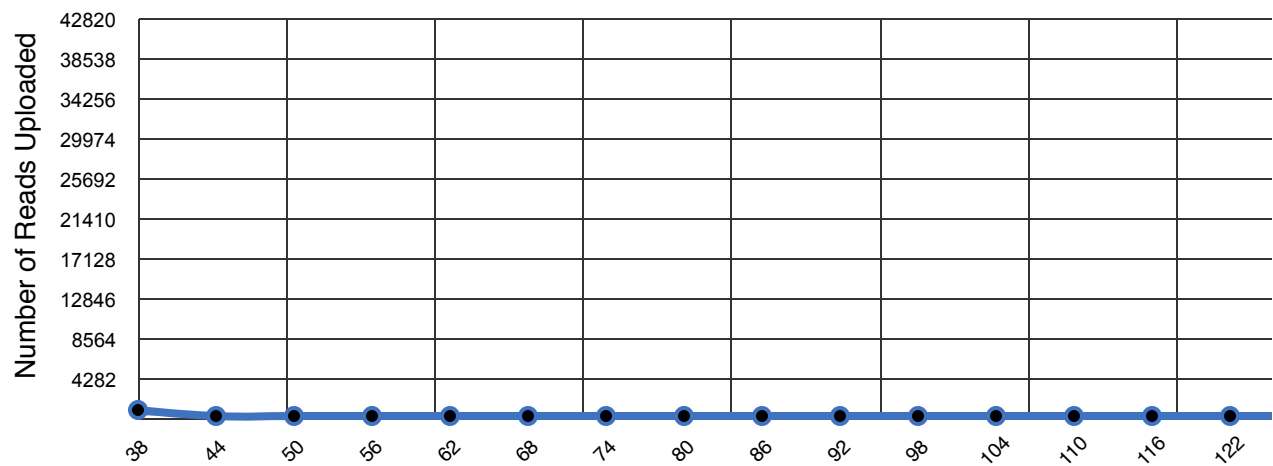
SEQUENCE LENGTH HISTOGRAM [HIDE](#)

The histograms below show the distribution of sequence lengths in basepairs for this metagenome. Each position represents a bp range.

The data used in these graphs are based on raw upload and post QC sequences.

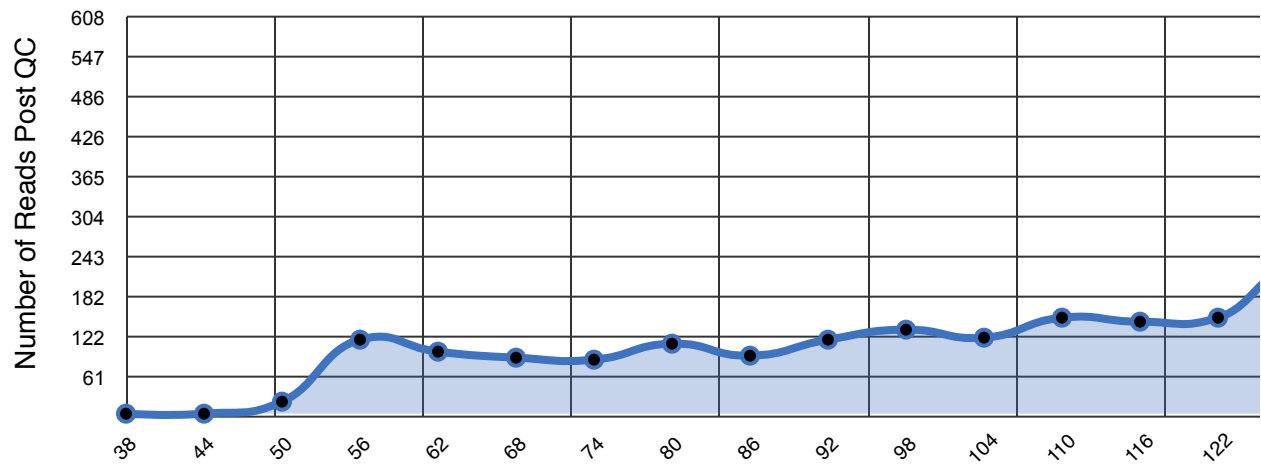
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[Download chart data](#)

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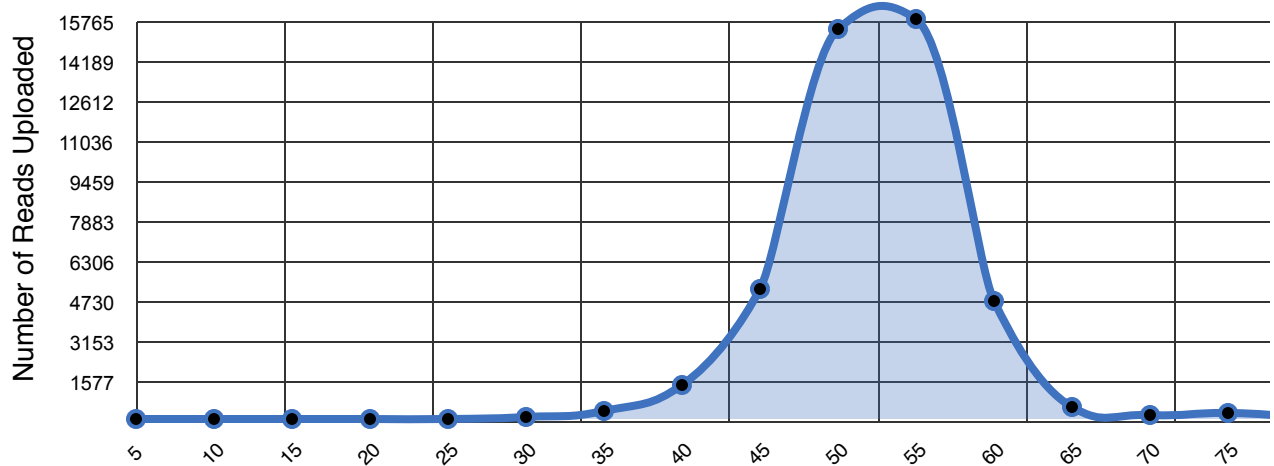


SEQUENCE GC DISTRIBUTION [HIDE](#)

The histograms below show the distribution of the GC percentage for this metagenome. Each position represents the range. The data used in these graphs is based on raw upload and post QC sequences.

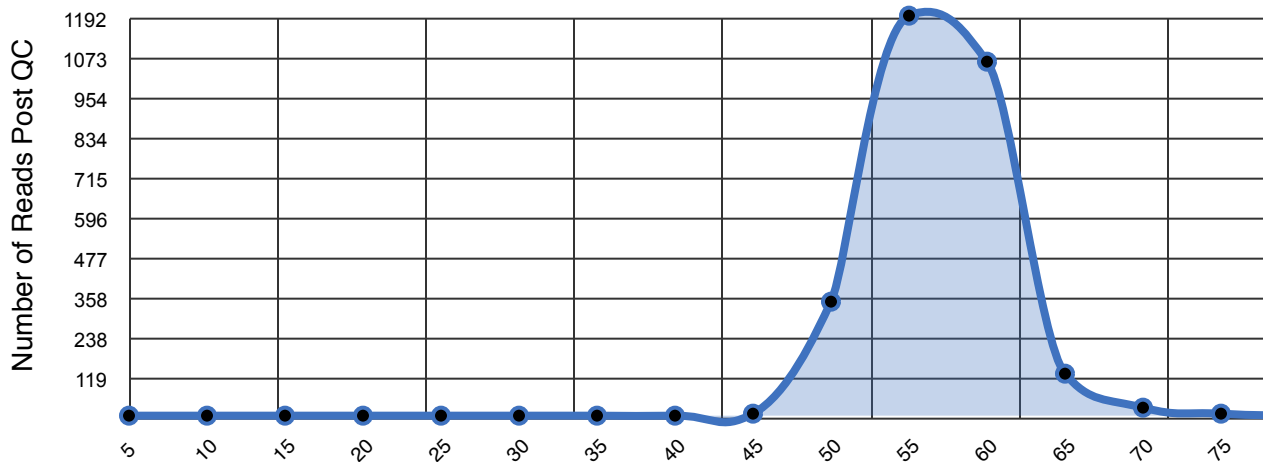
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DOWNLOAD THIS METAGENOME

We provide download capabilities for the submitted sequences, metadata, and all files with results that are produced. [Download page for this metagenome](#). This includes fasta files with annotations using the [M5NR](#).

We also provide access to the blat alignment summaries underlying our sequence analysis work on the [download page](#).

Please note: The graphs on this page allow downloading the underlying information as tables. The search results allow you to download sequences in an element to work with in the [workbench](#) feature on the [analysis page](#).

ANALYZE THIS METAGENOME

The [analysis page](#) provides access to analysis and comparative tools including tables, bar charts, trees, principle component analysis (including FASTA and QIIME). The [workbench](#) feature allows sub-selections of data to be used e.g. select all E. coli present just in E. coli reads across multiple data sets.

SEARCH THIS METAGENOME

Below searches return all predicted functions or organisms that contain the input text.

METADATA [\[?\]](#) [SHOW](#)