BIGGIE: A Distributed Pipeline for Genomic Variant Calling

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Motivation: faster, open-source genome variant calling tools

Impact:

Human genome variation is being used more and more to impact disease diagnosis and treatment, however:

- current tools frequently disagree on variant calls
- different types of variation require specialized tools

Current Tools:

- ► GATK [2]: slow and difficult to use
- ► CASAVA [1]: fast, but not free
- samtools mpileup [3]: slow and some accuracy issues

Our Goal:

- fast, distributed variant caller
- separate the genome into regions of high and low complexity and use the right tool for the right region

Aligned Reads (BAM) BIGGIE Distributed Sort and Index Classify into Low- and Highcomplexity High-Complexity Simple SNP Caller SNPs and SVs

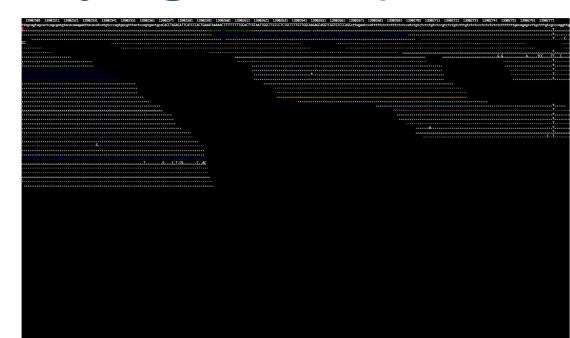
Figure 1: BIGGIE pipeline

Per-base SNP caller

Main idea:

- Distrubted pipeline for variant calling using Spark [4]
- Assign a complexity score to each base
- ▶ Use a simple SNP caller at bases with a low complexity score
- ▶ Use more robust structural variant callers at high complexity bases

Complexity region examples:



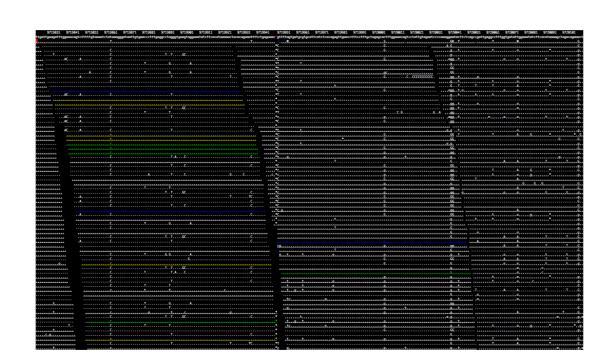


Figure 2: Different variant calling tools should be used for regions of the genome.

Complexity score features:

Name	Weight	Description
Substitution	3	Number of aligned reads showing a sub-
		stitution with respect to the reference.
Insertion	10	Number of aligned reads showing an in-
		sertion with respect to the reference.
Deletion	10	Number of aligned reads showing a dele-
		tion with respect to the reference.
Low Quality	3	Number of reads aligned with low map
		quality (a common indicator of a repeti-
		tive region).
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Table 1: Relative weight of features for computing complexity.

Incorporating high complexity regions

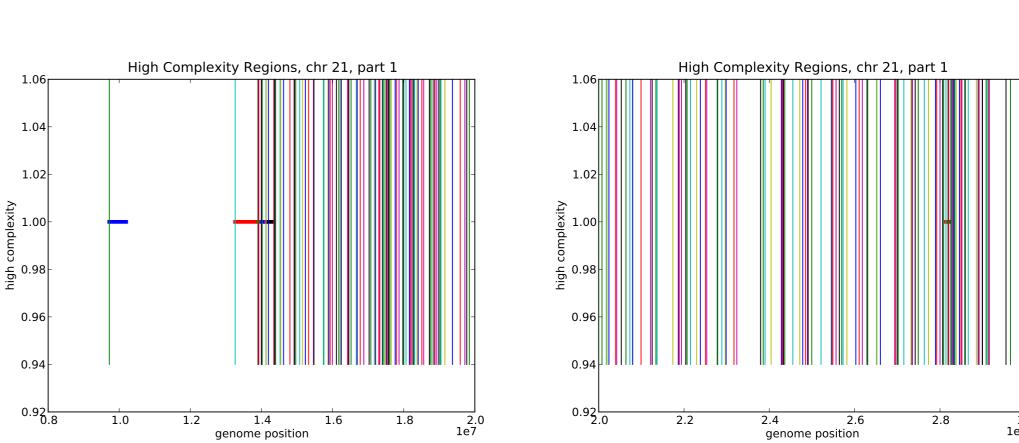


Figure 4: Regions are fairly uniformly distributed, except near the chromosome ends.

- We group bases into a high-complexity region in a greedy fashion, maintaining that the overall high-complexity base density is > t
- \blacktriangleright We filter out regions that are < 500 bases long

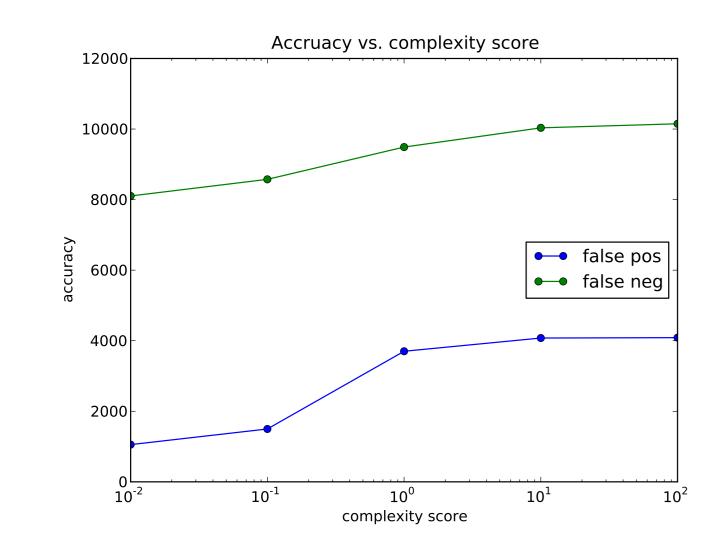
Stats, $t=5\%$	
Number of high complexity regions	3603
Percentage of genome is high complexity regions	16.6%

Results

Simulating data:

- ▶ Used reads simulated from the consensus sequence for Venter's genome
- ► Better approximates the true pattern of SNPs, indels, and structural variants found in a true genome; reads were aligned using BWA and SNAP

Effect of thresholds:



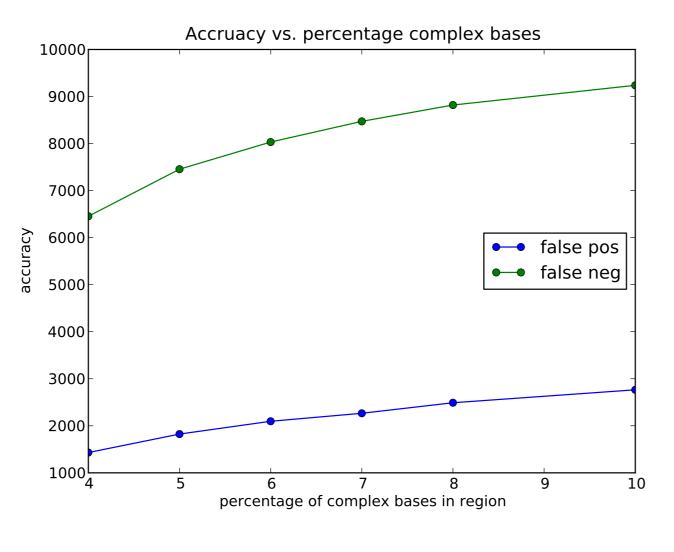


Figure 3: On the left are the per-base results, measuring false negatives only on the regions we called. Both accuracy measures increase as the threshold increases, but the number of correct calls increases as well. We see a similar pattern on the right for the region results, where the number of false positives and false negatives increase with the density of complex bases in high-complexity regions, but the number of true calls increases as well.

Results

Timing Results:

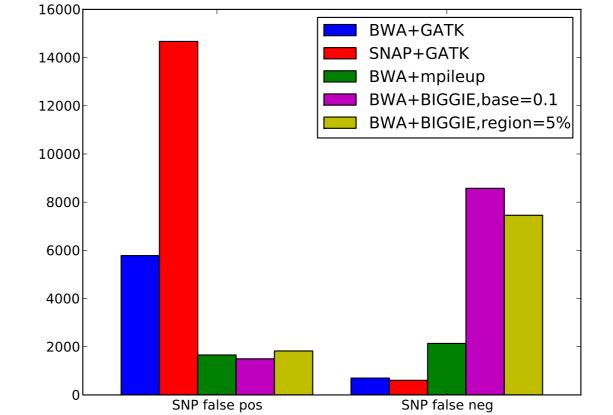
Algorithm Runtime
GATK 35m 17s
mpileup 49m 53s
BIGGIE 4m 38s

Table 2: Timing results for GATK, mpileup, and BIGGIE. The runtime is not significantly impacted by the complexity threshold.

Low vs. High Complexity:

region type false pos false neg correct low-complexity high-complexity 2289 2788 13046

Table 3: Our performance degrades in the high complexity regions, which is why a special purpose variant caller should be used.



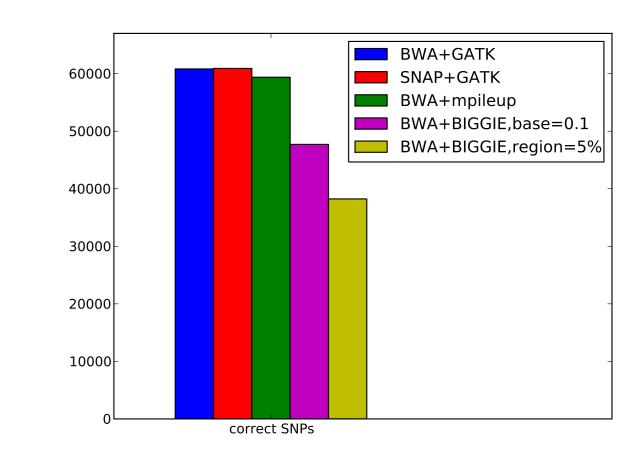


Figure 5: Accuracy comparison of BIGGIE with mpileup and GATK. False positives in BIGGIE are often associated with alignment errors or confusion with a small indel. For each algorithm, a very small percentage of correct SNP bases actually have the incorrect (unphased) genotype.

Future Work: Use the high and low complexity regions to distribute the reads across machines, then call variants using appropriate algorithms.

References

- [1] CASAVA. (2012) http://support.illumina.com/sequencing/sequencing_software/casava.ilmn.
- [2] DePristo M. *et al*, "A framework for variation discovery and genotyping using next-generation DNA sequencing data." *Nature Genetics* (2011), 43:491-498.
 - [3] Li H. et al and 1000 Genome Project Data Processing Subgroup, "The Sequence alignment/map (SAM) format and SAMtools."
 - Bioinformatics (2009), 25: 2078-9.

 [4] Zaharia M. et al, "Resilient Distributed Datasets: A Fault-Tolerant Abstraction for In-Memory Cluster Computing." NSDI (2012).