

1 The error in Bayesian phylogenetic reconstruction  
2 when speciation co-occurs

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8 **Abstract**

9 The tools for reconstructing phylogenetic relationships between taxo-  
10 nomic units (e.g. species) have become very advanced in the last three  
11 decades.

12 Among the most popular tools are Bayesian approaches, such as  
13 BEAST, MrBayes and RevBayes, that use efficient tree sampling routines  
14 to create a posterior probability distribution of the phylogenetic tree. A  
15 feature of these approaches is the possibility to incorporate known or  
16 hypothesized structure of the phylogenetic tree through the tree prior. It  
17 has been shown that the effect of the prior on the posterior distribution  
18 of trees can be substantial.

19 Currently implemented tree priors assume that speciation events are  
20 independent, where we know that speciation can coincide, for example,  
21 when trigger by a larger geographic change.

Here we explore the effects of ignoring speciation co-occurrence with an extensive simulation study.

We compare the inferred tree to the simulated tree, and find that ....

**Keywords:** computational biology, evolution, phylogenetics, Bayesian analysis, tree prior

## 1 Introduction

The computational tools that are currently available to the phylogeneticists go beyond the wildest imagination of those living four decades ago. Advances in computational power allowed the first cladograms to be inferred from DNA alignments in 1981 (Felsenstein 1981), and the first Bayesian tools emerged in 1996 (Rannala & Yang 1996), providing unprecedented flexibility in the setup of a phylogenetic model.

Currently, the most popular Bayesian phylogenetics tools are BEAST (Drummond & Rambaut 2007) and its offshoot BEAST2 (Bouckaert *et al.* 2014), MrBayes (Huelsenbeck & Ronquist 2001) and RevBayes (Höhna *et al.* 2016). They allow to incorporate known or hypothesized structure of a phylogenetic tree-to-be-inferred through model priors. With these priors and an alignment of DNA, RNA or protein sequences, they create a sample of the posterior distribution of phylogenies and parameter estimates (of the models used as a prior), in which more probable combinations are represented more often. Each of these tools use efficient tree sampling routines to rapidly create an informative posterior.

The model priors in Bayesian phylogenetic reconstruction can be grouped into three categories: (1) site model, specifying nucleotide substitutions, (2) clock model, specifying the rate of mutation per lineage in time, and (3) tree model, constituting the speciation model underlying branching events (speciation) and branch termination (extinction). The choice of site model (Posada &

48 Buckley 2004), clock model (Baele *et al.* 2012) or tree prior (Möller *et al.* 2018;  
49 Yang & Rannala 2005) is known to affect the posterior.

50 Current phylogenetic tools assume that only a single speciation event can  
51 occur at any given time. While this assumption is useful to construct a wide  
52 variety of successful models (for example: Maddison *et al.* 2007, Valente *et al.*  
53 2015, Etienne *et al.* 2012, Etienne *et al.* 2014), they disallow for environmental  
54 changes that trigger speciations in multiple clades at a same point in time.

55 The (constant-rate) birth-death (BD) model embodies the common assump-  
56 tion that only a single speciation event can occur at any given time. The MBD  
57 model relaxes this assumption, allowing events in which large-scale environmen-  
58 tal changes lead to a great number of species in relatively short time intervals.  
59 Such hypothesis can be useful to describe, for example, systems like cichlid fish  
60 diversification in the African Great Lakes: Malawi, Tanganyika and Victoria  
61 (Janzen *et al.* 2016, Janzen *et al.* 2017).

62 In the MBD model, parameters  $\lambda$  and  $\mu$  correspond, respectively, to the  
63 usual per-species speciation and extinction rates. Additionally,  $\nu$  is the rate  
64 at which an environmental change is triggered. When such event is triggered,  
65 all species present in the phylogeny at that moment have a probability  $q$  to  
66 speciate (independent on  $\lambda$ ). The number of species that speciate due to this  
67 can also be zero. [RJCB: Is this correct?] [GL: this is the code used  
68 for the simulation: "stats::rbinom(n = 1, size = N, prob = q)". It  
69 can actually yield zero. Now I am wondering if it is correct or not... ]  
70 [RJCB: I read in the PDF describing the likelihood derivation 'The  
71  $L_m^\lambda$  term takes into account, being in the state  $Q_m^k(t)$ , all the possible  
72 ways to have at least one birth event from whichever pool'. I would  
73 suggested changing the code to "1 + stats::rbinom(n = 1, size =  
74 N, prob = q)" and test it somehow to be sure the sim matches the

75 **likelihood ]** .

76       Unfortunately, a tree prior according to this model, providing the probabil-  
77 ity of a species tree under the MBD model, is unavailable in current Bayesian  
78 phylogenetic tools. Whilst a likelihood equation has been derived (Laudanno  
79 2018), it has not been implemented as tree prior yet. There are various rea-  
80 sons for this. First, the computation of the MBD likelihood involves solving a  
81 set of non-linear differential equations **[GL: are they actually non-linear?]**  
82 **[RJCB: Definitely, I see an exponentiation and combinatorial term,**  
83 **both are non-linear]**, and while this computation is quite fast, it still takes  
84 much more time than the corresponding probability of the BD model which is a  
85 simple analytical formula. In a Bayesian MCMC chain, the tree prior probabil-  
86 ity must be calculated many times, and hence the total computation will take  
87 considerably longer with a PBD tree prior.

88       Here we aim to explore the effect of using the BD prior on MBD simulated  
89 phylogenies. In brief, we simulate phylogenies with co-occurring speciation events  
90 using the MBD process. Given this species tree, we simulate a DNA sequence  
91 alignment. Then, we use BEAST2 on these alignments to infer a posterior of  
92 phylogenies, using a BD prior. We quantify the difference between the (BD)  
93 posterior phylogenies and the simulated (MBD) species tree. Furthermore, while  
94 we evidently know the clock and site models used in the simulation, using a  
95 different clock and/or site model prior in inference may compensate or increase  
96 this difference between inferred and simulated tree. To study this, we also  
97 explore the effect of a different clock and site model prior in inference.

98       **[RJCB: This setup is wrong, discuss described setup by Gio today]**

99       The MBD model has 4 parameters, depicted in table 2. We pick values of  $\nu$  in  
100 such a way we expect a multiple speciation event to be triggered zero ( $\nu = 0$ ),  
101 once, twice , four and eight times **[GL: One thing Rampal and I discussed**

102 to do was to use an equivalent  $\lambda$  in BD to have a similar amount  
 103 of mutations for both kind of simulated trees. In the code there  
 104 should be already something like that (it might be not completely  
 105 accurate though). I explain myself better: a) you simulate with some  
 106 MBD par setup; b) you count the amount of mutations you had;  
 107 c) from this number you decide what *lambda* to use to simulate the  
 108 BD process to have the (expected) same amount of mutations. It's  
 109 probably much much easier than going from BD to MBD. ] . For  
 110 each expected number of triggered events, we only keep those phylogenies that  
 111 actually realized the expected number of triggered events. We pick values of  $q$   
 112 that are 0.0 (a speciation barrier at the triggered event), 0.25, 0.5 and 1.0. We  
 113 set our extinction rate  $\mu$  to 0.1 in all simulation. As we select our phylogenies on  
 114 their number of lineages, we calculate  $\lambda$  in a such a way that the mean expected  
 115 number of lineages equals the desired numbers of taxa of 50, 100 and 200. For  
 116  $\nu = 0$ , the model falls back to a standard BD model. Note that the  $\lambda$  and  $q$   
 117 have different units and it is a misconception to think that for  $\lambda = q$  (already  
 118 impossible due to their units) the MBD model would reduce to a BD model.

119 We simulate protracted birth-death trees, using the MBD package (Lau-  
 120 danno 2018) in the R programming language (R Core Team 2013). The first  
 121 tree has a random number generator seed of 1, which is incremented by 1 for  
 122 each simulated tree. For each combination of  $\lambda, \mu, \nu$  and  $q$ , we generate species  
 123 trees with a crown age of 15 million years Only trees with the desired number  
 124 of good taxa are kept.

125 From an (MBD) species tree, we create a BEAST2 posterior using the 'pirou-  
 126 ette' (Bilderbeek 2018) R package: 'pirouette' first simulates a DNA align-  
 127 ment that has the same history as the species tree, using the **phangorn** package  
 128 (Schliep 2011). The DNA sequence of the root ancestor consists of four equally

129 sized single-nucleotide blocks of adenine, cytosine, guanine and thymine respec-  
 130 tively (for example, for a DNA sequence length of 12, this would be AAACC-  
 131 CGGGTTT). Throughout evolutionary time, we use equal mutation rates be-  
 132 tween the four DNA nucleotides, also called the Jukes-Cantor (Jukes *et al.* 1969)  
 133 nucleotide substitution model. The neat separation of the nucleotides is for vi-  
 134 sualization and debugging purposes and has no effect in any other way. The  
 135 equal amount of nucleotides does matter, assuring any nucleotide mutation is  
 136 equally likely to be observed.

137 In our Bayesian inference (see below) we use the same site model as the  
 138 (obviously correct) site model prior, but we also explore the effect of assuming a  
 139 more complex site model prior. We predict with the more complex substitution  
 140 model, that there will be more noise and hence our inference error will increase.  
 141 On the other hand, we dare not rule out that the inference error will decrease,  
 142 due to more flexibility in the more complex prior. We set the mutation rate in  
 143 such a way to maximize the information contained in the alignment. To do so,  
 144 we set the mutation rate such that we expect on average one (possibly silent)  
 145 mutation per nucleotide between crown age and present, which equates to  $\frac{1}{15}$   
 146 mutations per million years. The DNA sequence length is chosen to provide a  
 147 resolution of  $10^3$  years, that is, to have one expected nucleotide change per  $10^3$   
 148 years per lineage on average. As one nucleotide is expected to have on average  
 149 one (possibly silent) mutation per 15 million years,  $15 \cdot 10^3$  nucleotides result  
 150 in 1 mutation per alignment per  $10^3$  years (which is coincidentally the same  
 151 as Möller *et al.* 2018). The simulation of these DNA alignments follows a strict  
 152 clock model, which we will specify as one of the two clock models assumed in  
 153 the Bayesian inference (see below).

154 From here, the 'babette' R package (Bilderbeek & Etienne 2018) takes over  
 155 and converts the DNA alignment to a BEAST2 posterior. We set up the

156 BEAST2 analysis to assume either a Jukes-Cantor or GTR nucleotide sub-  
 157 stitution model. The Jukes-Cantor model is the correct one, as it is used for  
 158 simulating that alignment, where the GTR model is the site model that is picked  
 159 as a default by most users. For our clock model, we assume either a strict or  
 160 relaxed log-normal clock model. Also here, the strict clock model is the correct  
 161 one, as it is used for simulating the alignment, but the relaxed log-normal clock  
 162 model is the one most commonly used. We set the BD model as a tree prior,  
 163 as gauging the effect of this incorrect assumption is the goal of this study. We  
 164 assume an MRCA prior with a tight normal distribution around the crown age,  
 165 by choosing the crown age as mean, and a standard deviation of  $0.5 \cdot 10^{-3}$  time  
 166 units, resulting in 95% of the crown ages inferred have the same resolution (of  
 167  $10^{-3}$  time units) as the alignment. We ran the MCMC chain to generate 1111  
 168 states, of which we remove the first 10% (also called the 'burn-in'). Of the  
 169 remaining 1000 MCMC states, the Effective Sample Size (ESS) of the posterior  
 170 must at least be 200 for a strong enough inference (Drummond & Bouckaert  
 171 2015). An ESS can be increased by increasing the number of samples or decreas-  
 172 ing the autocorrelation between samples. If the ESS is less than 200, we decrease  
 173 autocorrelation by doubling the MCMC sampling interval of that simulation,  
 174 until the ESS exceeds 200.

175 We compare each posterior phylogeny to the (sampled) species tree using the  
 176 nLTT statistic (Janzen *et al.* 2015), from the nLTT package (Janzen 2015). The  
 177 nLTT statistic equals the area between the normalized lineages-through-time-  
 178 plots of two phylogenies, which has a range from zero (for identical phylogenies)  
 179 to one. We use inference error and nLTT statistic interchangeably. Compar-  
 180 ing the simulated species tree with each of the posterior species trees yields a  
 181 distribution of nLTT statistics.

182 The input trees generated with a  $\nu = 0$ , in which all BEAST2's assumptions

183 are met, allow us to measure the noise of the experiment.

184 We produce one data set as a comma-separated file. The general data set  
185 has 144 [RJCB: recalc] different combinations of parameter combinations.  
186 The experiment is computationally intensive: pilot experiments show that the  
187 experiment takes roughly 100 days of CPU time and 20 days of wall clock time  
188 (which includes the queued waiting for computational resources) per replicate.  
189 Due to this, we choose to perform ten replicates, so that the complete experiment  
190 will take an acceptable time of roughly seven months.

191 We display the data set as an nLTT statistics distribution per parameter  
192 combination as a faceted violin plot, showing the effect of the number of species  
193 (a proxy for the amount of information), the number of triggered events and  
194 the intensity of such a triggered event. We only show the nLTT distributions  
195 that were generated under the (correct) assumptions of a Jukes-Cantor site  
196 model and a strict clock model, separated per sampling method used. We  
197 display the nLTT statistic distributions separated per site or clock model in the  
198 supplementary information.

## 199 2 Results

## 200 3 Glossary

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### Distribution of inference errors

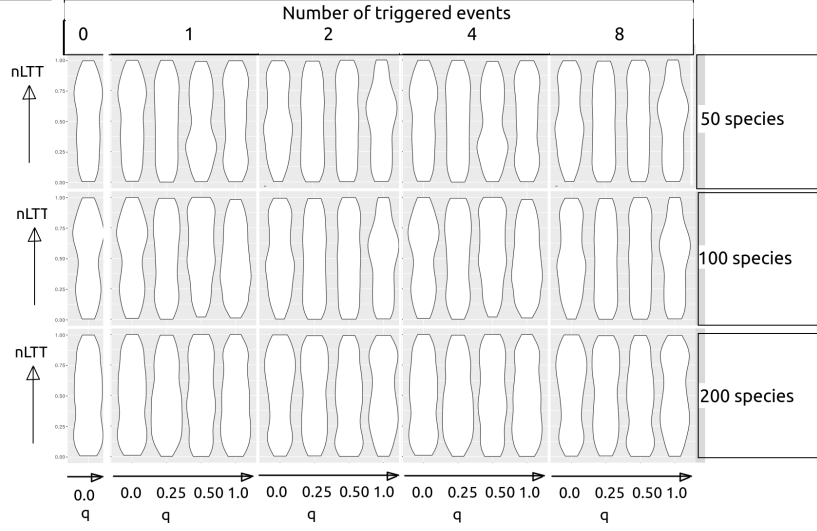


Figure 1: nLTT statistic distribution per setup, under the (correct) assumptions of a strict clock and Jukes-Cantor site model.

Term	Definition
Phylogenetics	The inference of evolutionary relationships of groups of organisms using genetics
Model prior	Knowledge or assumptions about the ontogeny of evolutionary histories
Posterior	A collection of phylogenies and parameter estimates, in which more probable combinations (determined by the data and the model prior) are presented more frequently

Table 1: Glossary

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## 267 **A Acknowledgements**

268 **[RJCB: put this section here, as the journal does not request for this]**

269 We would like to thank the Center for Information Technology of the University  
 270 of Groningen for their support and for providing access to the Peregrine high  
 271 performance computing cluster.

## 272 **B Authors' contributions**

273 **[RJCB: put this section here, as the journal does not request for this]**

274 RSE conceived the idea for this experiment. GL created and tested the MBD  
 275 package. RJCB created and tested the experiment. GL and RJCB wrote the  
 276 first draft of the manuscript. RSE contributed substantially to revisions.

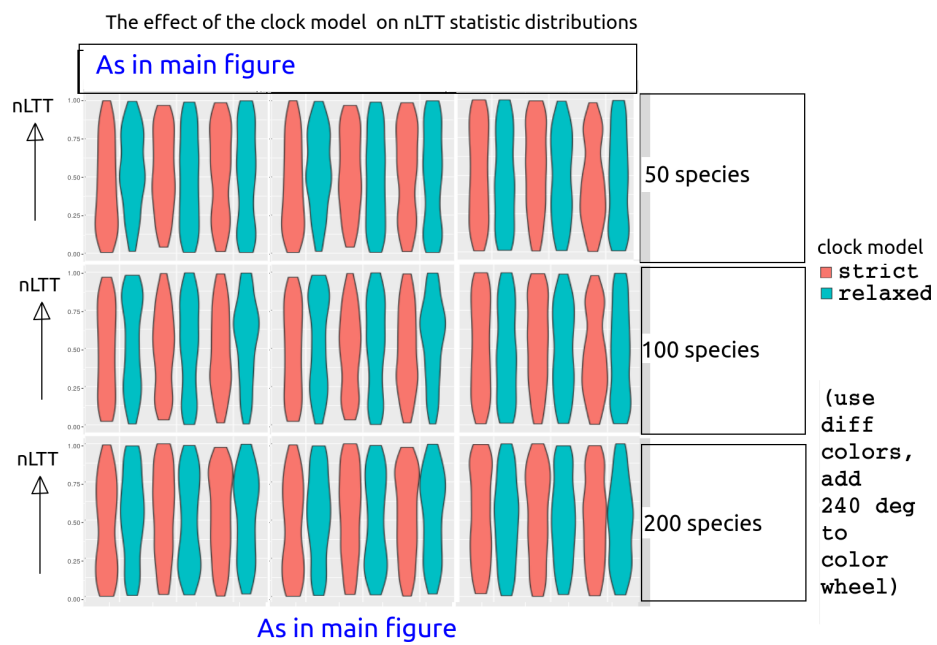


Figure 2: nLTT statistic distribution per biological parameter set per clock model, using the general data set, under the (correct) assumption of a Jukes-Cantor site model.

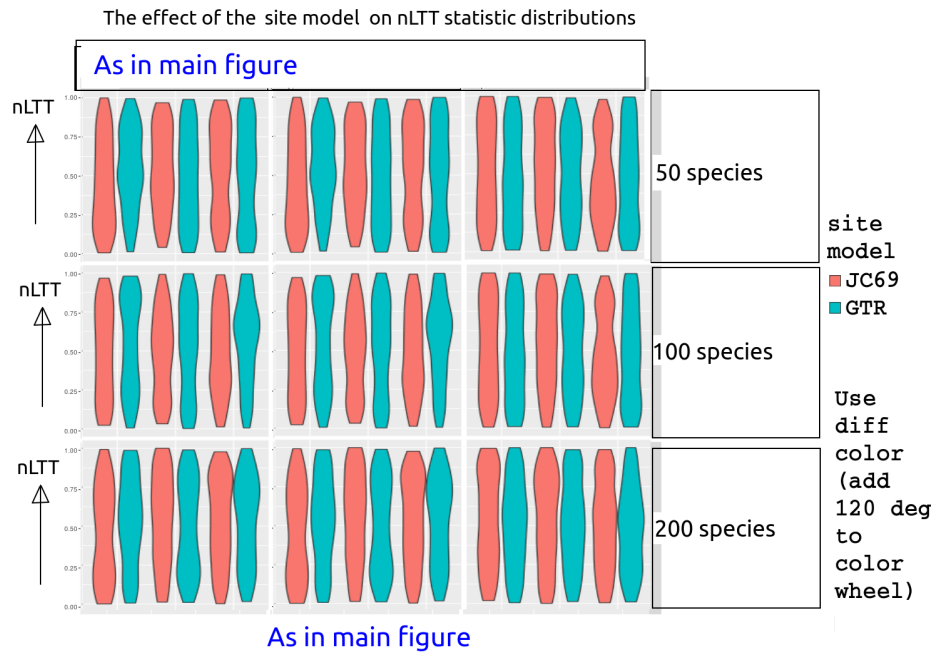


Figure 3: nLTT statistic distribution per biological parameter set per site model, using the general data set, under the (correct) assumption of a strict clock model.

	Description	Values
$\lambda$	Per-species speciation rate	calculated
$\mu$	Per-species extinction rate	0.0, 0.1
$\nu$	Multiple speciation trigger rate	occurs never, once, twice, four and eight times
$q$	Per-species probability of multiple speciation	0, 0.25, 0.5, 1.0
$n$	Number of good taxa	50, 100, 200
$t_c$	Crown age	15
$\sigma_c$	Standard deviation around crown age	0.001
$M_c$	Clock model	S, RLN
$M_t$	Site model	JC69, GTR
$r$	Mutation rate	$\frac{1}{15}$
$l_a$	DNA alignment length	15K
$f_i$	MCMC sampling interval	1K or more
$R_i$	RNG seed MBD tree generation	1, 2, etc.
$R_a$	RNG seed alignment simulation	$R_i$
$R_b$	RNG seed BEAST2	$R_i$

Table 2: Overview of the simulation parameters. Above the horizontal line are the MBD model’s parameters. The RNG seed  $R_i$  is 1 for the first simulation, 2 for the next, and so on. The clock models are abbreviated as ‘S’ for a strict and ‘RLN’ for a relaxed log-normal model. The site models are abbreviated as ‘JC69’ for Jukes-Cantor (Jukes *et al.* 1969) and ‘GTR’ for the generalized time-reversible model (Tavaré 1986).

$n$	Description
12 <b>[RJCB: recalc]</b>	simulation parameters, see table 2
1000	nLTT statistic values
11	ESSes of all parameters estimated by BEAST2 (see specs below)

Table 3: Specification of the data sets. Each row will contain one experiment, where the columns contain parameters, measurements and diagnostics. This table displays the content of the columns.  $n$  denotes the number of columns a certain item will occupy, resulting in a table of 1023 **[RJCB: recalc]** columns and 20K rows.

#	Description
1	posterior
2	likelihood
3	prior
4	treeLikelihood
5	TreeHeight
6	BirthDeath
7	BDBirthRate
8	BDDeathRate
9	logP.mrca
10	mrcatime
11	clockRate

Table 4: Overview of the 11 parameters estimated by BEAST2