Acoustic Shadow Detection From Scanline Statistics of B-Mode or Radiofrequency Ultrasound Images

Ricky Hu^a, Rohit Singla¹, Farah Deeba^{a,*}

^aRobotics and Control Laboratory, Department of Electrical and Computer Engineering, University of British Columbia, Vancouver, Canada

Abstract

An acoustic shadow is an ultrasound image artifact that occurs in boundaries between significantly different material, resulting in a continuous dark region on the image. Shadows are important to detect as they identify anatomical features or may obscure regions of interests and images may be misinterpreted. A shadow detection method was developed by analyzing the statistics of patches of radiofrequency (RF) data. A second method was developed for brightness-mode (B-mode) images if RF data is unavailable by analyzing the cumulative entropy along interpolated scanline paths. Both methods utilized adaptive thresholding, needing only the pulse width of the transducer as an input parameter for easy utilization by different operators or equipment. Average Dice coefficients (\pm standard deviation) of 0.90 \pm 0.05 and 0.87 \pm 0.08 were obtained for the RF and B-mode methods, which is within range of the gold standard of manual detection on two different transducers and three different imaging locations. The results indicate that the methods are able to detect shadows with high versatility in different imaging scenarios. The method has the potential to aid interpretation of ultrasound images or serve as an important pre-processing step for machine learning methods.

Keywords: Acoustic Shadow, Ultrasound, Speckle

INTRODUCTION

11

12

13

14

15

16

18

19

21

22

23

24

Ultrasound devices have become increasingly affordable and portable, encouraging applications such as pointof-care ultrasound (Bouhemad et al., 2011), novice usage (Becker et al., 2016), and creating training data sets for machine learning (Ghose et al., 2013). However, ultrasound is susceptible to unique artifacts that increase the difficult of interpretation and processing of images. One artifact is an acoustic shadow, which occurs when an ultrasound wave propagates from the transducer to a boundary of two materials with high impedance differences (Kremkau and Taylor, 1986). The wave is almost completely reflected and beyond the boundary is a continuous dark region and a loss of anatomical features. Shadows occur in air-tissue, tissue-bone, and tissue-lesion interfaces. Shadows can aid interpretation, such as identifying the presence of a gall stones (Good et al., 1979) or spinal level (Galiano et al., 2005). However, shadows, such as from poor transducer contact, can lead to misinterpretation of anatomy, particularly by novice users and automated processing algorithms. Thus, the identification of shadows is an important preprocessing step in many applications.

Several methods have been used in literature to detect shadows. Geometric techniques model the path of an ultrasound signal for an expected image along the scanline using a random walk (Karamalis et al., 2012). Regions are then flagged as a shadow if a pixel is below a confidence threshold. However, geometric techniques require knowledge of the ultrasound transducer properties to assign weights to a random walk, such as the focal length, radius of curvature, and thickness. The technique would be cumbersome to implement across different ultrasound machines, especially if the source of the ultrasound images are unknown. This reduces the data available for machine learning applications and requires accurate transducer parameter labels for each image.

Pixel intensity methods ignore the properties of the transducer an analyze only the graphical properties of the image (Hellier et al., 2010). Shadows have been detected on brain images by analyzing the entropy along a scanline to flag pixels of sudden low intensity as a potential shadow. The technique achieved a comparable Dice similarity coefficient as geometric methods but require specific thresholding, window sizing, filtering, and image mask parameterization for different anatomy. This method would be unfeasible in a clinical setting, particularly for novice users or point-of-care applications, as parameterization requires image processing expertise.

Machine learning methods have gained significant interest in medical imaging analysis. To our knowledge, no machine learning method has demonstrated capability of detecting shadows from multiple anatomy. Deep learning methods have identified features in a common image set that contains a shadow, such as neuroanatomical regions

^{*}Corresponding Author: Ricky Hu, Robotics and Control Laboratory, University of British Columbia, Room 3090, 2332 Main Mall, Vnacouver, BC, Canada V6T 1Z4. Email: rhu@ece.ubc.ca

Table 1: Transducer properties for different imaging scenarios

	Anatomy	Frequency	Depth	Gain	99
Linear	Forearm	11.0MHz	5.0cm	50%	100
Transducer	Elbow	11.0MHz	5.0cm	40%	101
(L14-5/38)	Ribcage	5.0MHz	10.0cm	30%	102
Curvilinear	Forearm	4.0MHz	5.0cm	50%	103
Transducer	Elbow	4.0MHz	5.0cm	40%	104
(C5-2/60)	Ribcage	3.3MHz	10.0cm	30%	105

in cranial scan (Milletari et al., 2017) or spinal levels in a¹⁰⁸ posterior scan (Hetherington et al., 2017). Although ma-¹⁰⁹ chine learning has the potential of providing automated¹¹⁰ feature recognition in multiple applications, a large data¹¹¹ set is required for an algorithm to recognize certain fea-¹¹² tures. Ultrasound imaging is highly variable due to unique¹¹³ artifacts, operator technique, and equipment. In addition,¹¹⁴ shadows occur in various anatomy. Previous techniques¹¹⁵ focus on a single anatomical region and training data was¹¹⁶ from a consistent imaging scenario. However, it is diffi-¹¹⁷ cult to construct a training data set with the generality¹¹⁸ required to recognize shadows in different scenarios usable¹¹⁹ for a variety of ultrasound applications.

To address existing needs for versatile detection, a method was developed utilizing radiofrequency (RF) or brightness-¹²¹ mode (B-mode) data that can detect shadows from multiple anatomy or transducers with minimum user configu-¹²² ration required.

METHODS

57

60

61

65

67

71

74

77

78

81

82

91

Data Collection

Ultrasound RF and B-mode data was acquired by scan
ning 37 adult participants with informed written consent,

approved by the University of British Columbia Research

approved by the University of British Columbia Research

Ethics Board (Study ID: H18-01199). The scans included a

forearm scan near the distal end of the pronator quadratus,

an elbow scan near the cubital fossa, and a rib scan on the

anterior surface of right ribs 11-12. Each scan was taken

with both a curvilinear (C5-2/60, Ultrasonix, Canada) and

linear (L14-5/38, Ultrasonix, Canada) transducer. Dif
ferent transducer settings were used for each anatomical

region and transducer, summarized in Table 1. The ex
periment was designed to generate a dataset from various

and imaging scenarios to validate the versatility of the shadow

detection method.

Radiofrequency Speckle Analysis

To detect shadows, patches of speckle was analyzed ¹⁴⁵ on the RF signal. Speckle occurs due to multiplicative ¹⁴⁶ scattering of acoustic waves in a material, resulting in a ¹⁴⁷ granular patch on the image. B-mode data commonly at ¹⁴⁸ tempts to remove speckle, but speckle contains information ¹⁴⁹ of the acoustic interactions in tissue (Burckhardt, 1978). ¹⁵⁰

Speckle can then characterize different regions, such as a region of tissue or a region of signal loss in a shadow. In addition, B-mode data can be manipulated by an operator to visually enhance an image, such as adjusting time-gain compensation or dynamic range. Thus, speckle analysis can provide a more robust shadow detection usable across different machines and operators.

One of the first models for speckle is with a one-parameter Rayleigh distribution to model the probability density of a random walk (Burckhardt, 1978). The Rayleigh is capable for modeling fully developed speckle, which does not occur in limited scattering (Tuthill et al., 1988). More generalized models have been applied such as the Rician, Homodyned-K, and Nakagami distributions to characterize speckle (Destrempes and Cloutier, 2010). Speckle has been leveraged to analyze features such as classifying tumorigenicity of breast lesions (Byra et al., 2016) or levels of liver fibrosis (Ho et al., 2012). Shadow detection presents a simpler problem as a shadow and non-shadow region contain significantly different speckle patterns. Thus, the Nakagami distribution expressed in Eq. 1 was chosen to model speckle. The Nakagami distribution provides greater generality than the Rayleigh distribution while being more computationally efficient than the Rician or Homodyned K distributions.

$$\Phi(x,\mu,\omega) = 2\left(\frac{\mu}{\omega}\right)^{\mu} \frac{1}{\Gamma(\mu)} x^{(2\mu-1)} e^{\frac{-\mu}{\omega}x^2}$$
 (1)

Where x is the RF intensity, μ is the shape parameter, ω is the scale parameter and $\Gamma(\mu)$ is the gamma distribution.

To detect shadows, the raw RF data was first processed by computing the echo envelope of each scanline with a Hilbert transform. An absolute logarithmic scale of the echo envelope was taken to generate an "RF Image", visually similar to B-mode but without filtering to remove speckle. Next, the RF image was divided into overlapped patches with a width of a single RF data point and a length of three times the pulse width. This patch size was demonstrated in literature to be sufficiently large to capture multiple wavelengths and scattering events while being small enough to be useful in differentiating different regions on the millimeter scale (Byra et al., 2016). Next, the RF data in each patch was fit to a Nakagami distribution using a maximum likelihood estimate to compute the fitted Nakagami shape and scale parameters μ and ω , producing a map of Nakagami parameter values for an image as shown in Fig. 1.

Since the Nakagami distributions are significantly different for shadow regions with abundant speckle and non-shadow regions with minimal speckle, a simple automated thresholding scheme was used. Otsu's method was applied on the entire image to compute a threshold for the Nakagami parameters. Then, for each scanline, the deepest data point that is above the threshold is labeled as the shadow boundary and all data points below are labeled as a shadow.

143

106

107

124

125

126

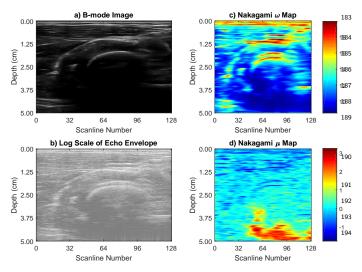


Figure 1: The Nakagami parameter maps computed and compared to the a) B-mode image and d) echo envelope. The echo envelope contains the unfiltered speckle that can be analyzed by statistical distributions

B-mode Scanline Analysis

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

176

177

178

180

181

Many ultrasound machines do not provide access to RF data for speckle analysis. Thus, a shadow detection method on B-mode images was developed to address the need for usability across different equipment with minimum configuration. First, the cumulative scanline entropy is computed for each pixel, which is similar to the "Rupture Criterion" (Hellier et al., 2010), with the window size fixed as three times the pulse width, η , as defined in Eq. ¹⁹⁵

$$S_{i,j} = \sum_{i=1}^{3\eta} I(i-1)log_2 \frac{I(i-1)}{I(i+1)} + I(i+1)log_2 \frac{I(i+1)}{I(i-1)} (2) \frac{1}{1} \frac{1$$

Where $S_{i,j}$ is the cumulative entropy at pixel i on scan-line j, η is the pulse width, I(i) is the pixel intensity of i.

Note that for linear images, tracking the scanlines is simple. For curvilinear images, the scanline paths were tracked by first capturing the slope of the lateral edges by following the ring-down regions. Ring-down is a consistent artifact that produces bright bands at the top of an image due to the fluid in the transducer reflecting a continuous signal. The scanlines were then interpolated between the slopes of the lateral edges.

Next, Otsu's method is applied similarly to compute a threshold entropy value. The intuition of the threshold is different than in RF analysis. In RF analysis, the threshold separates patches of intense and minimal speckle. In B-mode analysis, the threshold separates pixels of a shadow boundary, which has high entropy, and pixels not of a shadow boundary, which include shadow and non-shadow regions. Thus, shadows can be identified by finding the last pixel on a scanline with an entropy higher than the threshold, representing a bright shadow boundary.

Validation

A trained annotator manually outlined shadow regions on B-mode images. The manual regions were used as a gold standard, as manual identification is common in clinical practice and has been used in previous literature for comparison. A dice coefficient was computed to compare similarity of manual and automated shadow detection.

Results

Examples of detected shadows from both methods are highlighted in gray in Fig. 2 in different imaging scenarios. The Dice coefficients for both methods for different anatomy and trasducers are shown in Table 2.

Table 2: Average Dice coefficients for different imaging scenarios \pm standard deviation

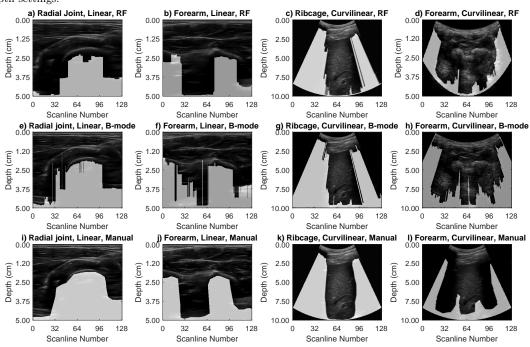
		RF	B-Mode
Linear (L14-5/38)	Forearm	0.90 ± 0.05	$0.92 {\pm} 0.07$
	Elbow	0.95 ± 0.10	$0.86 {\pm} 0.07$
	Ribcage	0.83 ± 0.09	$0.81 {\pm} 0.05$
Curvilinear (C5-2/60)	Forearm	0.85 ± 0.04	$0.86 {\pm} 0.10$
	Elbow	0.89 ± 0.06	0.92 ± 0.09
	Ribcage	$0.85 {\pm} 0.05$	0.90 ± 0.11

Discussion

Previous studies reported that the Dice coefficient between manual annotators recorded a mean of 0.86 ± 0.09 (Hellier et al., 2010) . Every scenario detected from both methods achieved a Dice coefficient within the range of manual detection. This supports the versatility of the detection method as both methods are able to identify shadows across different anatomy and transducers with minimum configuration.

However, the inconsistency of manual detection presents two problems. One is that by using a subjective gold standard with Dice upper bound below 1.0, it is difficult to compare the performance of different algorithms. Secondly, the inconsistency of manually identified shadows reflect on the lack of a clear definition for a shadow. In images with a shadow, such as in Fig. 2 a), the shadow boundary of the radial joint appears as a bright arc. Looking closely at the scanline of the boundary, the pixels first increase in brightness and then decrease before becoming a clear, dark shadow. There is no consensus as to where the shadow begins, whether it is the brightness point on the scanline or when the signal drops below a threshold. Thus, there a limitation on validation and this work focused on improving the versatility and usability of shadow detection while maintaining similar accuracy as previous methods.

Figure 2: A comparison of the detected shadows from the RF method, B-mode method, and manual detection. It is important to differentiate between a shadow and attentuation. In subfigure b), the RF method performs more robustly in identifying scanlines 32-64 as attenuation, rather than shadow. This is likely due to pixel intensity methods previously being dependent on multiple filtering kernels tuned for different anatomy and depth settings.



252

267 268

269

Conclusions

222

223

224

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

248 249

250

251

Two methods were developed for acoustic shadow de-²⁵⁴ tection requiring only the transducer pulse width as the ²⁵⁵ input parameter. When comparing to manual detection, ²⁵⁶ the methods achieved a Dice similarity coefficient above X ²⁵⁸ for three different anatomical locations and two different ²⁵⁹ transducers, indicating high similarity. The versatility of ²⁶⁰ the shadow detection method has potential to improve the ²⁶² interpretation of ultrasound images with shadow artifacts ²⁶³ or to serve as a pre-processing step for machine learning ²⁶⁴ methods in the future.

References

Becker DM, Tafoya CA, Becker SL, Kruger GH, Tafoya MJ, Becker TK. The use of portable ultrasound devices in low- and middle-income countries: a systematic review of the literature. Tropical Medicine & International Health, 2016;21:294–311.

Bouhemad B, Brisson H, Le-Guen M, Arbelot C, Lu Q, Rouby₂₇₅
JJ. Bedside ultrasound assessment of positive end-expiratory₂₇₆
pressure-induced lung recruitment. American Journal of Respi-₂₇₇
ratory and Critical Care Medicine, 2011;183:341–347.

Burckhardt CB. Speckle in ultrasound B-mode scans, 1978.

Byra M, Nowicki A, Wróblewska-Piotrzkowska H, Dobruch-Sobczak₂₈₀ K. Classification of breast lesions using segmented quantitative₂₈₁ ultrasound maps of homodyned K distribution parameters. Med.₂₈₂ Phys., 2016;43:5561–5569.

Destrempes F, Cloutier G. A critical review and uniformized repre-284 sentation of statistical distributions modeling the ultrasound echo envelope. Ultrasound Med. Biol., 2010;36:1037–1051.

Galiano K, Obwegeser AA, Bodner G, Freund M, Maurer H, Kamelger FS, Schatzer R, Ploner F. Ultrasound guidance for facet joint injections in the lumbar spine: A computed tomography-controlled feasibility study. Anesthesia and Analgesia, 2005;101:579–583.

Ghose S, Oliver A, Mitra J, Martí R, Lladó X, Freixenet J, Sidibé D, Vilanova JC, Comet J, Meriaudeau F. A supervised learning framework of statistical shape and probability priors for automatic prostate segmentation in ultrasound images. Medical Image Analysis, 2013;17:587–600.

Good LI, Edell SL, Soloway RD, Trotman BW, Mulhern C, Arger Pa. Ultrasonic properties of gallstones. Effect of stone size and composition. Gastroenterology, 1979;77:258–263.

Hellier P, Coupé P, Morandi X, Collins DL. An automatic geometrical and statistical method to detect acoustic shadows in intraoperative ultrasound brain images. Medical Image Analysis, 2010;14:195–204.

Hetherington J, Lessoway V, Gunka V, Abolmaesumi P, Rohling R. SLIDE: automatic spine level identification system using a deep convolutional neural network. International Journal of Computer Assisted Radiology and Surgery, 2017;12:1189–1198.

Ho MC, Lin JJ, Shu YC, Chen CN, Chang KJ, Chang CC, Tsui PH. Using ultrasound Nakagami imaging to assess liver fibrosis in rats. Ultrasonics, 2012;52:215–222.

Karamalis A, Wein W, Klein T, Navab N. Ultrasound confidence maps using random walks. Medical Image Analysis, 2012;16:1101–1112.

Kremkau FW, Taylor KJ. Artifacts in ultrasound imaging. Journal of Ultrasound in Medicine, 1986;5:227–237.

Milletari F, Ahmadi SA, Kroll C, Plate A, Rozanski V, Maiostre J, Levin J, Dietrich O, Ertl-Wagner B, Bötzel K, Navab N. Hough-CNN: Deep learning for segmentation of deep brain regions in MRI and ultrasound. Computer Vision and Image Understanding, 2017;164:92–102.

Tuthill TA, Sperry RH, Parker KJ. Deviations from rayleigh statistics in ultrasonic speckle. Ultrasonic Imaging, 1988;10:81–89.