Figure 1

Riccardo Mosca

2025-05-12

This markdown show how to generate Figure 1B and Figure S1E-F

Figure 1B Loading packages and qPCR fold changes values

```
library(dplyr)
library(ggplot2)
library(ggpubr)
qPCR <- read.table(file =</pre>
"/Users/riccardomosca/Desktop/RAPseq PAPER/FIGUREs/FIGURE1/NEW/HUR qPCR",
    header = T, stringsAsFactors = F)
qPCR means <- qPCR %>%
    group_by(Substrate, Target) %>%
    summarise(Mean Fold Change = mean(Fold Change))
qPCR means <- as.data.frame(qPCR means)</pre>
head(qPCR_means)
##
       Substrate Target Mean_Fold_Change
## 1 Full Length
                     NT
                                    2.385
## 2 Full Length
                     T1
                                    5.500
## 3 Full Length
                     T3
                                    0.000
## 4 fragmented
                     NT
                                    1.210
## 5 fragmented
                     T1
                                   47.500
## 6 fragmented
                     T3
                                   81.000
```

plotting

dev.off() Fig1B

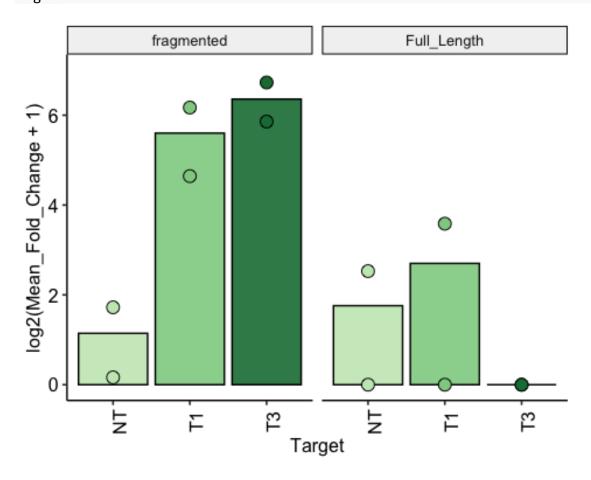


Figure S1E Loading packages and annotated peak files, keeping the unique targets

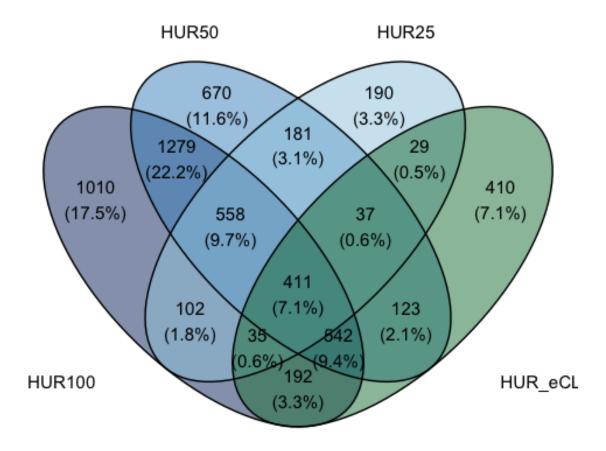
```
library(eulerr)
library(tidyverse)
library(clusterProfiler)
library(org.Hs.eg.db)
library(reshape2)
library(ggvenn)
library(dplyr)

path <-
"/Users/riccardomosca/Desktop/RAPseq_PAPER/FIGUREs/FIGURE1/NEW/HUR_hnRNPA1_ti
tration_revision_231016/DEEPER_SEQ/PEAKs"

# Different HUR titrations
types <- c("HUR100", "HUR50", "HUR25")

for (t in types) {
    # build the filename and read it in</pre>
```

```
infile <- file.path(path, paste0(t, " annotated.txt"))</pre>
    peaks <- read.delim(infile, sep = "\t", header = TRUE)</pre>
    # unique RNAs
    df <- peaks %>%
        dplyr::select(gene name, Gene BS, gene ID, positive fa, negative fa)
%>%
        filter(!duplicated(gene_name)) %>%
        as.data.frame()
    # rename first column to 'gene'
    colnames(df)[1] <- "gene"</pre>
    assign(paste0("", t), df)
}
# Reading HUR eCLIP bed file from ENCODE
# https://www.encodeproject.org/experiments/ENCSR090LNQ/
eCLIP_HUR_peaks <-
read.delim("/Users/riccardomosca/Desktop/RAPseq PAPER/FIGURE5/FIGURE1/NEW/HUR
_hnRNPA1_titration_revision_231016/PEAKs/HUR_eCLIP_annotated_peaks.txt",
    sep = "\t", header = T)
HUR_eCLIP <- eCLIP_HUR_peaks %>%
    dplyr::select(gene name)
HUR_eCLIP <- HUR_eCLIP[!duplicated(HUR_eCLIP$gene_name), ]</pre>
HUR eCLIP <- as.data.frame(HUR eCLIP)</pre>
colnames(HUR eCLIP)[1] <- "gene"</pre>
# Venn diagram
gene_HUR100 <- HUR100$gene
gene_HUR50 <- HUR50$gene
gene HUR25 <- HUR25$gene
gene_HUR_eCLIP <- HUR_eCLIP$gene</pre>
ABCD <- list(HUR100 = gene_HUR100, HUR50 = gene_HUR50, HUR25 = gene_HUR25,
HUR eCLIP = gene HUR eCLIP)
ggvenn(ABCD, fill color = c("#09316A", "#4292C6", "#9ECAE0", "#187D41"),
stroke_size = 0.5,
set_name_size = 4)
```



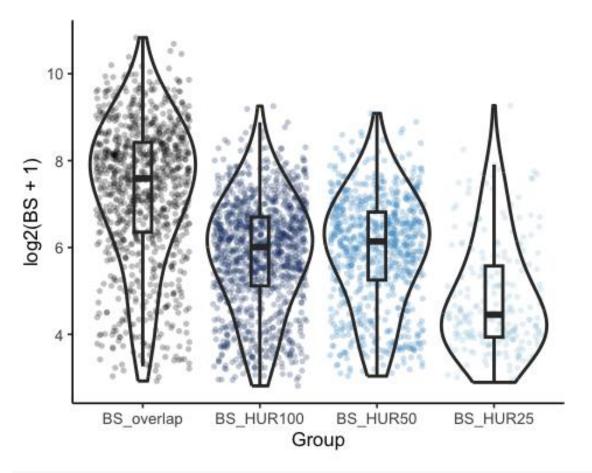
Computing the mean of Gene BS for the three titrations overlap, and each specific one.

```
overlap_HUR100_HUR50 <- subset(HUR100, gene %in% HUR50$gene)</pre>
overlap_HUR50_HUR25 <- subset(HUR50, gene %in% HUR25$gene)</pre>
overlap_HUR100_HUR50_HUR25 <- subset(overlap_HUR100_HUR50, gene %in%
overlap HUR50 HUR25$gene)
specific_HUR100 <- HUR100[!as.character(HUR100$gene) %in%</pre>
as.character(HUR50$gene),
specific_HUR100 <- specific_HUR100[!as.character(specific_HUR100$gene) %in%</pre>
as.character(HUR25$gene),
specific_HUR50 <- HUR50[!as.character(HUR50$gene) %in%</pre>
as.character(HUR100$gene),
specific_HUR50 <- specific_HUR50[!as.character(specific_HUR50$gene) %in%</pre>
as.character(HUR25$gene),
specific_HUR25 <- HUR25[!as.character(HUR25$gene) %in%</pre>
as.character(HUR100$gene),
specific_HUR25 <- specific_HUR25[!as.character(specific_HUR25$gene) %in%</pre>
as.character(HUR50$gene),
```

```
# Determine the maximum length of the vectors to combine
max length <- max(length(overlap HUR100 HUR50 HUR25$Gene BS),
length(specific_HUR100$Gene_BS),
    length(specific_HUR50$Gene_BS), length(specific_HUR25$Gene_BS))
# Create vectors of equal length by recycling shorter vectors
Gene BS HUR100 50 25 <- c(overlap HUR100 HUR50 HUR25 Gene BS, rep(NA,
max length -
    length(overlap_HUR100_HUR50_HUR25$Gene_BS)))
Gene BS HUR100 <- c(specific HUR100$Gene BS, rep(NA, max length -
length(specific_HUR100$Gene_BS)))
Gene_BS_HUR50 <- c(specific_HUR50$Gene_BS, rep(NA, max_length -</pre>
length(specific HUR50$Gene BS)))
Gene_BS_HUR25 <- c(specific_HUR25$Gene_BS, rep(NA, max_length -</pre>
length(specific HUR25$Gene BS)))
# Making an unique df
combined BS <- data.frame(BS overlap = Gene BS HUR100 50 25, BS HUR100 =
Gene BS HUR100,
    BS HUR50 = Gene BS HUR50, BS HUR25 = Gene BS HUR25)
combined BS <- melt(combined BS, value.name = "BS", variable.name = "Group")</pre>
combined_BS <- na.omit(combined_BS)</pre>
head(combined_BS)
##
          Group
                        BS
## 1 BS overlap 252.89770
## 2 BS overlap 424.43825
## 3 BS_overlap 623.91807
## 4 BS overlap 674.67210
## 5 BS overlap 52.20435
## 6 BS_overlap 109.50906
combined BS %>%
    group_by(Group) %>%
    summarise(n = n())
## # A tibble: 4 × 2
##
     Group
##
     <fct>
                <int>
## 1 BS overlap
                  969
## 2 BS_HUR100
                 1202
## 3 BS_HUR50
                  793
## 4 BS HUR25
                  219
aa \leftarrow ggplot(data = combined BS, aes(x = Group, y = log2(BS + 1))) +
geom_jitter(aes(color = Group),
```

```
pch = 16, alpha = 0.25) + scale_colour_manual(values = c("#000000",
"#0A316A",
    "#4292C6", "#9ECAE0")) + theme_classic(base_size = 12.5) +
theme(legend.position = "none")

FigS1E <- aa + geom_violin(trim = T, bw = 0.75, scale = "width", lwd = 1,
fill = NA) +
    geom_boxplot(outlier.shape = NA, width = 0.15, lwd = 1, fill = NA)</pre>
FigS1E
```



```
# collect specific Gene BS vectors into a named list
specific_list <- list(HUR100 = specific_HUR100$Gene_BS, HUR50 =
specific_HUR50$Gene_BS,
    HUR25 = specific_HUR25$Gene_BS)

# run wilcox.test() for each, comparing to the overlap vector
wt <- lapply(names(specific_list), function(nm) {
    y <- specific_list[[nm]]
    test <- wilcox.test(overlap_HUR100_HUR50_HUR25$Gene_BS, y)
    data.frame(comparison = paste0("overlap_vs_", nm), W =
as.numeric(test$statistic),
    p.value = test$p.value)</pre>
```

```
# combine into one df
wt_df <- do.call(rbind, wt)</pre>
```