



Sclerotinia sclerotiorum

Sclerotinia sclerotiorum is a plant pathogenic fungus and can cause a disease called **white mold** if conditions are conducive. *S. sclerotiorum* can also be known as cottony rot, watery soft rot, stem rot, drop, crown rot and blossom blight. A key characteristic of this pathogen is its ability to produce black resting structures known as sclerotia and white fuzzy growths of mycelium on the plant it infects. These sclerotia give rise to a fruiting body in the spring that produces spores in a sac which is why fungi in this class are called sac fungi (Ascomycota). This pathogen can occur on many continents and has a wide host range of plants. When *S. sclerotiorum* is onset in the field by favorable environmental conditions, losses can be great and control measures should be considered.

Hosts and symptoms

Sclerotinia sclerotiorum" (sometimes called white mold) is a fungal (Ascomycota) pathogen that infects a broad range of plants, affecting over 400 species across multiple families, including Brassicaceae (Cruciferae), Fabaceae (Leguminosae), Solanaceae, Asteraceae, and Apiaceae (Umbelliferae). Notably it is not known to infect grass species, and appears to be incapable of causing disease in clover. Common hosts of white mold are herbaceous, succulent plants, particularly flowers and vegetables. Sunflowers are common hosts for white mold. It can also affect woody ornamentals occasionally, usually on juvenile tissue. White mold can affect their hosts at any stage of growth, including seedlings, mature plants, and harvested products. It can usually be found on tissues with high water content and in close proximity to the soil. One of the first symptoms noticed is an obvious area of white, fluffy mycelial growth. Usually this is preceded by pale to dark brown lesions on the stem at the soil line. The mycelium then cover this necrotic area. Once the xylem is affected, other symptoms occur higher up in the plant. These can include chlorosis, wilting, leaf drop, and death quickly follows. On fruits, the initial dark lesions occur on the tissue that comes in contact with the soil. Next, white fungal mycelium covers the fruit and it decays. This can occur when the fruit is in the field or when in storage.^[1]

Sclerotinia sclerotiorum



Sclerotinia sclerotiorum on Phaseolus

Scientific classification

Kingdom:	<u>Fungi</u>
Division:	<u>Ascomycota</u>
Class:	<u>Leotiomycetes</u>
Order:	<u>Helotiales</u>
Family:	<u>Sclerotiniaceae</u>
Genus:	<u>Sclerotinia</u>
Species:	<u>S. sclerotiorum</u>

Binomial name

Sclerotinia sclerotiorum

(Lib.) de Bary (1884)

Synonyms

- *Hymenoscyphus sclerotiorum*
(Lib.) W.Phillips (1887)
- *Peziza sclerotiorum* Lib. (1837)
- *Sclerotinia libertiana* Fuckel
(1870)
- *Sclerotium varium* Pers. (1801)
- *Whetzelinia sclerotiorum* (Lib.)
Korf & Dumont (1972)

Importance

White mold affects a wide range of hosts and causes sclerotinia stem rot. It is known to infect 408 plant species. As a nonspecific plant pathogen,^[2] diverse host range and ability to infect plants at any stage of growth makes white mold a serious disease. The fungus can survive on infected tissues, in the soil, and on living plants. It affects young seedlings, mature plants, and fruit in the field or in storage. White mold can spread quickly in the field from plant to plant. It can also spread in a storage facility throughout the harvested crop. Some crops it affects commonly are soybeans,^[3] green beans, sunflowers, canola, and peanuts.^[4] White mold is the most common pathogen that affects sunflower and has been found to cause reduction in yield throughout the world including the United States, northern Europe, Great Britain and Russia.^[5] Economically significant hosts include *Vicia faba*, for which Lithourgidis et al. have done extensive work over the years.^[6]



Sclerotinia stem rot (or 'white stem rot',^[7]) can cause economically significant yield losses in temperate climates, especially during cool and moist growing seasons. An analysis of soybean yields from 1996 to 2009 in the United States found that sclerotinia stem rot reduced yields by over ten million bushels in half of the studied growing seasons.^{[8][9]} During particularly bad years, these soybean yield reductions caused producers to lose millions of dollars.^[10] Compared to 23 common soybean diseases, sclerotinia stem rot was the second most problematic disease in the United States from 1996 to 2009.^{[8][9]} For soybeans, crop yields are inversely correlated with the incidence of Sclerotinia stem rot; an estimated of 0.25 metric ton per ha is lost for each 10% increment of diseased plants.^[11]

Environment

The pathogenic fungus *Sclerotinia sclerotiorum* proliferates in moist and cool environments. Under moist field conditions, *S. sclerotiorum* is capable of completely invading a plant host, colonizing nearly all of the plant's tissues with mycelium. Optimal temperatures for growth range from 15 to 21 degrees Celsius. Under wet conditions, *S. sclerotiorum* will produce an abundance of mycelium and sclerotia. Like most fungi, *S. sclerotiorum* prefers darker, shadier conditions as opposed to direct exposure to sunlight. For soybeans specifically, optimal conditions include canopy temperatures less than 28 °C and plant surface wetness for 12–16 h on a daily basis or continuous surface wetness for 42–72 h.^[11]

Life cycle

The lifecycle of *Sclerotinia sclerotiorum* can be described as monocyclic, as there are no secondary inoculums produced. During late summer to early fall, the fungus will produce a survival structure called a sclerotium either on or inside the tissues of a host plant. *S. sclerotiorum* sclerotia can remain viable for at least three years^[12] and germinate to produce fruiting bodies called apothecia, which are small, thin stalks ending with a cup-like structure about 3–6 mm in diameter.^[13] The cup of the apothecium is lined with asci, in which the ascospores are contained. When the ascospores are released from the asci, they are carried by the wind until they land on a suitable host. The ascospores of *S. sclerotiorum* infect aboveground plant host tissue^[14] and begin

to invade the host's tissues via mycelium, causing infection. *S. sclerotiorum* is capable of invading nearly all tissue types including stems, foliage, flowers, fruits, and roots. Eventually white, fluffy mycelium will begin to grow on the surface of the infected tissues. At the end of the growing season, *S. sclerotiorum* will once again produce sclerotia. The sclerotia will then remain on the surface of the ground or in the soil, on either living or dead plant parts until the next season. The lifecycle will then continue respectively.

There are two theories contending to explain the majority of *S. sclerotiorum* virulence: The oxalate-dependent theory and the pH-dependent theory. The oxalate theory was very credible because ultraviolet mutants producing knockout of oxalic acid production do have drastically reduced virulence. Similar results have also obtained with *Botrytis cinerea*, similarly an oxalic acid producing pathogen, with similar knockouts. However Davidson et al. 2016 and others have created transgenic hosts for oxalate oxidase and oxalate decarboxylase and charted the results day by day. They find that *initial* infection is not noticeably dependent on oxalate (although lesion *expansion* does require it for pH reduction and chelation of calcium). This supports the pH theory, with oxalates being merely a part of pH.^[15]

Control

Control of white mold on crops can depend greatly on cultural, biological, and chemical practices. Cultural practices include planting disease resistant crops,^[16] planting at lower densities and higher row spacing to promote air circulation. This would allow for creation of microclimates that are less favorable for disease development.^{[17][12]} Besides that, excessive irrigation should be avoided until flowering (which is the most active period of infection) has ceased.^[11] Furthermore, in susceptible areas, crop rotations should include at least two to three years of non-host crops (for example cereals and corn).^[12] Good weed control can also limit the amount of host plants in a field and reduce white mold pressure. Fields with heavy disease pressure may also be flooded for a period of four to five weeks until the sclerotia lose their viability.^[17] Tillage reduction can also reduce the number of viable *S. sclerotiorum* spores.^[18]



This image captures a cluster of apothecia from a downward angle, so that only the tops are visible. The apothecia in this photo are circular, tan, and have a white lining near the edge of the structure. For size reference, a dime is also included.



Sclerotinia sclerotiorum on bushbean

Coniothyrium minitans, a coelomycete distributed worldwide, is a pathogen of *S. sclerotiorum*^{[19][20]} and is a commercial biocontrol agent for sclerotinia stem rot. Application of *C. minitans* should occur three months before *S. sclerotiorum* development and be incorporated into the soil.^[21] Correct use of *C. minitans* can reduce *S. sclerotiorum* by 95% and sclerotinia stem rot 10 to 70%.^{[22][23]}

Systemic and contact fungicides are registered for white molds. For instance, in soybeans, there are three classes of fungicides that are labeled for white mold control: methyl benzimidazole carbamates, succinate dehydrogenase inhibitors, and demethylation inhibitors.^[12] Additionally, herbicides containing lactofen have also been reported to indirectly control white mold.^{[24][25][26]} However, the use of lactofen herbicides can harm crops in years without high disease potential.^[26]

Potential as a bioherbicide

Despite its broad host range (and partly because of it), this species has been proposed as a potentially useful bioherbicide or mycoherbicide. Notably it is not known to infect common forage grass species like ryegrass (*Lolium* spp.), and appears to be incapable of causing disease symptoms in clover. Due to the ubiquitous nature of this fungus in temperate climates, research has shown that wild-type sclerotium-forming isolates would be safe to use in permanent grass/clover pastures in these regions of the world,^[27] and its application for weed control would **not** to lead unusual levels of inoculum (airborne ascospores and soil-borne sclerotia) in the environment beyond the application site or increased disease risk to non-target plants because of its limited dispersibility.^[28]

References

1. "Page 1 : USDA ARS" (<https://web.archive.org/web/20170531013637/https://www.ars.usda.gov/plains-area/docs/white-mold-research/about-sclerotinia/page-1/>). www.ars.usda.gov. Archived from the original (<https://www.ars.usda.gov/plains-area/docs/white-mold-research/about-sclerotinia/page-1/>) on May 31, 2017. Retrieved 2020-04-29.
2. Pressete, Carolina Girotto; Giannini, Laila Santos Vieira; de Paula, Daniela Aparecida Chagas; do Carmo, Mariana Araújo Vieira; Assis, Diego Magno; Santos, Mário Ferreira Conceição; Machado, José da Cruz; Marques, Marcos José; Soares, Marisi Gomes; Azevedo, Luciana (December 2019). "Sclerotinia Sclerotiorum (White Mold): Cytotoxic, Mutagenic, and Antimalarial Effects In Vivo and In Vitro" (<https://doi.org/10.1111/1750-3841.14910>). *Journal of Food Science*. **84** (12): 3866–3875. doi:[10.1111/1750-3841.14910](https://doi.org/10.1111%2F1750-3841.14910) (<https://doi.org/10.1111%2F1750-3841.14910>). ISSN 0022-1147 (<https://search.worldcat.org/issn/0022-1147>). PMID 31750949 (<https://pubmed.ncbi.nlm.nih.gov/31750949/>).
3. Bennett, J. Michael; Rhetic, Emeritus; Hicks, Dale R.; Naeve, Seth L.; Bennett, Nancy Bush (2014). *The Minnesota Soybean Field Book* (<https://web.archive.org/web/20130930151502/http://www1.extension.umn.edu/agriculture/soybean/docs/minnesota-soybean-field-book.pdf>) (PDF). St Paul, MN: University of Minnesota Extension. p. 81. Archived from the original (<http://www.extension.umn.edu/agriculture/soybean/docs/minnesota-soybean-field-book.pdf>) (PDF) on 30 September 2013. Retrieved 21 February 2016.
4. [reference 4]
5. Harveson, R. M.; Markell, S. G.; Block, C. C.; Gulya, T. J., eds. (January 2016). *Compendium of Sunflower Diseases and Pests* (<https://dx.doi.org/10.1094/9780890545096>). The American Phytopathological Society. doi:[10.1094/9780890545096](https://doi.org/10.1094/9780890545096) (<https://doi.org/10.1094%2F9780890545096>). ISBN 978-0-89054-509-6.

6. Saharan, G. S.; Mehta, Naresh (2008). *Sclerotinia Diseases of Crop Plants: Biology, Ecology and Disease Management*. Dordrecht London: Springer. ISBN 978-1-4020-8407-2. OCLC 288440265 (<https://search.worldcat.org/oclc/288440265>). ISBN 978-1-4020-8408-9. ISBN 1402084072. ISBN 2008924858.
7. McLoughlin, Austein G.; Wytinck, Nick; Walker, Philip L.; Girard, Ian J.; Rashid, Khalid Y.; Kievit, Teresa de; Fernando, W. G. Dilantha; Whyard, Steve; Belmonte, Mark F. (9 May 2018). "Identification and application of exogenous dsRNA confers plant protection against Sclerotinia sclerotiorum and Botrytis cinerea" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5943259>). *Sci Rep.* **8** (1): 7320. Bibcode:2018NatSR...8.7320M (<https://ui.adsabs.harvard.edu/abs/2018NatSR...8.7320M>). doi:10.1038/s41598-018-25434-4 (<https://doi.org/10.1038%2Fs41598-018-25434-4>). PMC 5943259 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5943259>). PMID 29743510 (<https://pubmed.ncbi.nlm.nih.gov/29743510>).
8. Wrather, Allen; Koenning, Steve (January 2009). "Effects of Diseases on Soybean Yields in the United States 1996 to 2007" (<https://apsjournals.apsnet.org/doi/10.1094/PHP-2009-0401-01-RS>). *Plant Health Progress.* **10** (1): 24. doi:10.1094/PHP-2009-0401-01-RS (<https://doi.org/10.1094%2FPHP-2009-0401-01-RS>). ISSN 1535-1025 (<https://search.worldcat.org/issn/1535-1025>).
9. Koenning, Stephen R.; Wrather, J. Allen (January 2010). "Suppression of Soybean Yield Potential in the Continental United States by Plant Diseases from 2006 to 2009" (<https://doi.org/10.1094%2FPHP-2010-1122-01-RS>). *Plant Health Progress.* **11** (1): 5. doi:10.1094/PHP-2010-1122-01-RS (<https://doi.org/10.1094%2FPHP-2010-1122-01-RS>). ISSN 1535-1025 (<https://search.worldcat.org/issn/1535-1025>).
10. "USDA - National Agricultural Statistics Service Homepage" (<https://www.nass.usda.gov/>). www.nass.usda.gov. Retrieved 2020-04-29.
11. Hartman, G.L.; Rupe, J.C.; Sikora, E.J.; Domier, L.L.; Davis, J.A.; Steffey, K.L. (2015). *Compendium of Soybean Diseases and Pests, Fifth Edition*. St. Paul, Minnesota U.S.: The American Phytopathological Society. pp. 285–297. ISBN 978-0-89054-473-0.
12. Peltier, Angelique J.; Bradley, Carl A.; Chilvers, Martin I.; Malvick, Dean K.; Mueller, Daren S.; Wise, Kiersten A.; Esker, Paul D. (2012-06-01). "Biology, Yield loss and Control of Sclerotinia Stem Rot of Soybean" (<https://doi.org/10.1603%2FIPM11033>). *Journal of Integrated Pest Management.* **3** (2): 1–7. doi:10.1603/IPM11033 (<https://doi.org/10.1603%2FIPM11033>).
13. Abawi, G. S. (1979). "Epidemiology of Diseases Caused by *Sclerotinia* Species" (http://www.apsnet.org/publications/phytopathology/backissues/Documents/1979Abstracts/Phyto69_899.htm). *Phytopathology.* **69** (8): 899. doi:10.1094/Phyto-69-899 (<https://doi.org/10.1094%2FPhyto-69-899>).
14. "White mold (Sclerotinia)" (<https://www.apsnet.org/edcenter/disandpath/funglasco/pdlessons/Pages/WhiteMold.aspx>). *White mold (Sclerotinia)*. Retrieved 2023-06-20.
15. Xu, Liangsheng; Li, Guoqing; Jiang, Daohong; Chen, Weidong (2018-08-25). "Sclerotinia sclerotiorum: An Evaluation of Virulence Theories" (<https://doi.org/10.1146%2Fannurev-phyto-080417-050052>). *Annual Review of Phytopathology.* **56** (1). Annual Reviews: 311–338. doi:10.1146/annurev-phyto-080417-050052 (<https://doi.org/10.1146%2Fannurev-phyto-080417-050052>). ISSN 0066-4286 (<https://search.worldcat.org/issn/0066-4286>). PMID 29958073 (<https://pubmed.ncbi.nlm.nih.gov/29958073>). S2CID 49615444 (<https://api.semanticscholar.org/CorpusID:49615444>).
16. Conrad, Audrey Marie (2022-04-21). *Management of Sclerotinia sclerotiorum in soybean using the biofungicides Bacillus amyloliquefaciens and Coniothyrium minitans* (https://hammer.purdue.edu/articles/thesis/Management_of_Sclerotinia_sclerotiorum_in_soybean_using_the_biofungicides_Bacillus_amyloliquefaciens_and_Coniothyrium_minitans/19630221/1) (thesis thesis). Purdue University Graduate School. doi:10.25394/pgs.19630221.v1 (<https://doi.org/10.25394%2Fpgs.19630221.v1>).
17. Pohronezny, Ken, 1946- (2003). *Compendium of tomato diseases* (<http://worldcat.org/oclc/451534013>). American phytopathological Society. ISBN 0-89054-300-3. OCLC 451534013 (<https://search.worldcat.org/oclc/451534013>).

18. Pethybridge, Sarah J.; Brown, Bryan J.; Kikkert, Julie R.; Ryan, Matthew R. (2019-05-31). "Rolled–crimped cereal rye residue suppresses white mold in no-till soybean and dry bean" (https://www.cambridge.org/core/product/identifier/S174217051900022X/type/journal_article). *Renewable Agriculture and Food Systems*. **35** (6): 599–607. doi:10.1017/S174217051900022X (<https://doi.org/10.1017%2FS174217051900022X>). ISSN 1742-1705 (<https://search.worldcat.org/issn/1742-1705>). S2CID 191660928 (<https://api.semanticscholar.org/CorpusID:191660928>).
19. Campbell, W. A. (March 1947). "A New Species of Coniothyrium Parasitic on Sclerotia" (<https://dx.doi.org/10.2307/3755006>). *Mycologia*. **39** (2): 190–195. doi:10.2307/3755006 (<https://doi.org/10.2307%2F3755006>). ISSN 0027-5514 (<https://search.worldcat.org/issn/0027-5514>). JSTOR 3755006 (<https://www.jstor.org/stable/3755006>).
20. Arakere, Udayashankar C.; Konappa, Narasimhamurthy (2022). "12.9 Coniothyrium minitans as biopesticide". *Biopesticides*.
21. "CDMS Home" (<http://www.cdms.net/>). www.cdms.net. Retrieved 2020-04-29.
22. Boland, G.J. (May 1997). "Stability Analysis for Evaluating the Influence of Environment on Chemical and Biological Control of White Mold (Sclerotinia sclerotiorum) of Bean" (<https://dx.doi.org/10.1006/bcon.1997.0515>). *Biological Control*. **9** (1): 7–14. Bibcode:1997BiolC...9....7B (<https://ui.adsabs.harvard.edu/abs/1997BiolC...9....7B>). doi:10.1006/bcon.1997.0515 (<https://doi.org/10.1006%2Fbcon.1997.0515>). ISSN 1049-9644 (<https://search.worldcat.org/issn/1049-9644>).
23. Zeng, Wenting; Kirk, William; Hao, Jianjun (February 2012). "Field management of Sclerotinia stem rot of soybean using biological control agents" (<https://dx.doi.org/10.1016/j.biocontrol.2011.09.012>). *Biological Control*. **60** (2): 141–147. Bibcode:2012BiolC..60..141Z (<https://ui.adsabs.harvard.edu/abs/2012BiolC..60..141Z>). doi:10.1016/j.biocontrol.2011.09.012 (<https://doi.org/10.1016%2Fj.biocontrol.2011.09.012>). ISSN 1049-9644 (<https://search.worldcat.org/issn/1049-9644>).
24. Nelson, Kelly A.; Renner, Karen A.; Hammerschmidt, Ray (November 2002). "Cultivar and Herbicide Selection Affects Soybean Development and the Incidence of Sclerotinia Stem Rot" (<https://dx.doi.org/10.2134/agronj2002.1270>). *Agronomy Journal*. **94** (6): 1270–1281. Bibcode:2002AgrJ...94.1270N (<https://ui.adsabs.harvard.edu/abs/2002AgrJ...94.1270N>). doi:10.2134/agronj2002.1270 (<https://doi.org/10.2134%2Fagronj2002.1270>). ISSN 0002-1962 (<https://search.worldcat.org/issn/0002-1962>).
25. Nelson, Kelly A.; Renner, Karen A.; Hammerschmidt, Ray (April 2002). "Effects of Protoporphyrinogen Oxidase Inhibitors on Soybean (Glycine max L.) Response, Sclerotinia sclerotiorum Disease Development, and Phytoalexin Production by Soybean" ([https://dx.doi.org/10.1614/0890-037x\(2002\)016%5b0353:eopoio%5d2.0.co;2](https://dx.doi.org/10.1614/0890-037x(2002)016%5b0353:eopoio%5d2.0.co;2)). *Weed Technology*. **16** (2): 353–359. doi:10.1614/0890-037x(2002)016[0353:eopoio]2.0.co;2 (<https://doi.org/10.1614%2F0890-037x%282002%29016%5B0353%3Aeopoio%5D2.0.co%3B2>). ISSN 0890-037X (<https://search.worldcat.org/issn/0890-037X>). S2CID 83664788 (<https://api.semanticscholar.org/CorpusID:83664788>).
26. Dann, E. K.; Diers, B. W.; Hammerschmidt, R. (July 1999). "Suppression of Sclerotinia Stem Rot of Soybean by Lactofen Herbicide Treatment". *Phytopathology*. **89** (7): 598–602. doi:10.1094/phyto.1999.89.7.598 (<https://doi.org/10.1094%2Fphyto.1999.89.7.598>). ISSN 0031-949X (<https://search.worldcat.org/issn/0031-949X>). PMID 18944696 (<https://pubmed.ncbi.nlm.nih.gov/18944696>).
27. Bourdôt, Graeme W; Casonato, Seona G (January 2024). "Broad host-range pathogens as bioherbicides: managing nontarget plant disease risk" (<https://scijournals.onlinelibrary.wiley.com/doi/10.1002/ps.7410>). *Pest Management Science*. **80** (1): 28–34. doi:10.1002/ps.7410 (<https://doi.org/10.1002%2Fps.7410>). ISSN 1526-498X (<https://search.worldcat.org/issn/1526-498X>). Retrieved 3 February 2025.
28. Bourdôt, Graeme W.; Baird, David; Hurrell, Geoff A.; De Jong, Meindert D. (February 2006). "Safety zones for a Sclerotinia sclerotiorum-based mycoherbicide: Accounting for regional and yearly variation in climate" (<https://www.tandfonline.com/doi/full/10.1080/09583150500531966>). *Biocontrol Science and Technology*. **16** (4): 345–358. doi:10.1080/09583150500531966 (<https://doi.org/10.1080%2F09583150500531966>). ISSN 0958-3157 (<https://search.worldcat.org/issn/0958-3157>). Retrieved 3 February 2025.

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