

The bacterial density of clinical rectal swabs is highly variable,  
correlates with sequencing contamination, and predicts patient risk  
of extraintestinal infection

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```
# For ease of use, place the processed 16S data files of interest into the same directory as your current
# Basic processing with selection of OTUs
# I. Read in opti_mcc.shared
otu_good <- load_shared(shared = "vre.miseq.trim.contigs.good.unique.good.filter.unique.precluster.pick

# Trim _S from end of rownames(otu_good)
rownames(otu_good) <- str_remove(rownames(otu_good), "_S\\d+")

# II. Read in cons.taxonomy
otu_good_taxonomy <- load_tax("vre.miseq.trim.contigs.good.unique.good.filter.unique.precluster.pick.pi

# Create lables to check for real specimen
otu_df <- data.frame(decostand(otu_good, "total") * 100, Sample_name = row.names(otu_good), stringsAsFa
# Create Experiment from first string of characters prior to first "_"
mutate(experiment = factor(case_when(str_detect(Sample_name, "Woods_") ~ "Swab",
                                     str_detect(Sample_name, "^AE_") ~ "AE",
                                     str_detect(Sample_name, "^Empty_") ~ "Empty",
                                     str_detect(Sample_name, "^IsoCtrl_") ~ "IsoCtrl",
                                     str_detect(Sample_name, "^Water|^WATER") ~ "Water",
                                     str_detect(Sample_name, "^Zymo") ~ "Mock"),
                                     levels = c("Swab", "AE", "Empty", "IsoCtrl", "Water", "Mock"))
) %>%
# metadata columns at front, followed by all of the count data
mutate(experiment = str_extract(Sample_name, "^[:alpha:]+")) %>%
mutate(Sample_ID = str_replace(Sample_name, "^[:alpha:]+_", "")) %>%
mutate(experiment = if_else(experiment == "Woods",
                           str_replace(experiment, "Woods", "Swab"),
                           experiment)) %>%
dplyr::select(Sample_ID, experiment, everything(), -Sample_name)
# otu_df with rownames added for PCA analysis later
# note that the data is in wide format

# gather the OTU from wide to long
tidy_otu_df <- otu_df %>%
gather("Otu", "relative_abundance", -c(Sample_ID, experiment)) %>%
```

```

mutate(Otu = as_factor(Otu))

# link OTU to genus
otu_genus_link <- otu_good_taxonomy%>%
  dplyr::select(OTU, Genus)%>%
  mutate(Otu_genus = str_c(OTU, Genus, sep="-"))%>%
  rename("Otu"=OTU)%>%
  mutate(Otu = as_factor(Otu))# first create a new variable

tidy_genus_df<-inner_join(tidy_otu_df, otu_genus_link, by = "Otu")
rm(tidy_otu_df)

# this file is the results from Nicole for the ddPCR runs on the stoll swab specimens
ddPCR<- read_excel("16S_EvaGreen_WoodsRectalSwabs_Combined.xlsx",
  col_types = c("text", "text", "skip",
    "skip", "skip", "skip", "skip",
    "skip", "numeric", "skip", "skip",
    "skip", "skip", "skip", "skip",
    "skip"), sheet = 1)%>%
  mutate(Sample_ID = str_replace(Sample, "^Iso[:space:]Ctrl[:space:]", ""))%>%
  rename(swab_type = `Case/Control`)%>%
  mutate(swab_type = if_else(swab_type=="Case", "case", swab_type),
    swab_type = if_else(swab_type=="Control", "control", swab_type))%>%
  mutate(experiment = case_when(swab_type == "IsoCtrl" ~ "Isolation control",
    swab_type == "Water" ~ "Water",
    TRUE ~ "Rectal swab"))%>%
  rename(ddPCR_reads_per_sample = `Total 16S copies/isolation`)%>%
  filter(!is.na(ddPCR_reads_per_sample))%>%
  dplyr::select(Sample_ID, swab_type, experiment, ddPCR_reads_per_sample)%>%
  mutate(Sample_ID = if_else(experiment=="Water", "NEG", Sample_ID))%>%
  filter(!(ddPCR_reads_per_sample>18 & experiment=="Water"))%>%
  mutate(Sample_ID = if_else(experiment=="Water", paste(Sample_ID, LETTERS[1:25], sep=""), Sample_ID))# dp
# remove duplicate "Sample" variable, order variables to join
summary(ddPCR$ddPCR_reads_per_sample)

##      Min.    1st Qu.      Median      Mean    3rd Qu.      Max.
## 9.000e+00 4.975e+04 1.151e+06 8.532e+07 3.623e+07 3.234e+09

cases<-ddPCR %>% filter(swab_type == "case")%>%dplyr::pull(Sample_ID)%>%as_factor() # create a vector o
control<-ddPCR %>% filter(swab_type == "control")%>%dplyr::pull(Sample_ID)%>%as_factor()

# this file is the results from Miseq summary
Miseq_quant <- read_csv("workbook.csv")[, 1:2]%>%
  rename(Sample_ID = `Sample ID`, miseq_reads_per_sample = `Reads PF/Sample`)%>%
  mutate(experiment = str_extract(Sample_ID, "^[:alpha:]+"))%>%
  mutate(Sample_ID = str_replace(Sample_ID, "^[:alpha:]+_", "")) %>%
  mutate(experiment = if_else(experiment == "Woods",
    "Rectal swab",
    experiment))%>%
  mutate(swab_type = as_factor(case_when(Sample_ID %in% cases ~ "Case",
    Sample_ID %in% control ~ "Control",
    TRUE ~ "Isolation Control")))%>%
  dplyr::select(Sample_ID, swab_type, experiment, miseq_reads_per_sample)

```

```
## Warning: Missing column names filled in: 'X3' [3]
```

```
ddPCR_miseq_comparison<-left_join(ddPCR, Miseq_quant, by = "Sample_ID")%>%
  mutate(experiment.y = if_else(experiment.y== "AE", "Elution buffer", experiment.y),
         experiment.y = if_else(experiment.y=="IsoCtrl", "Isolation control", experiment.y),
         experiment.y = factor(experiment.y, levels=c("Water", "Isolation control", "Elution buffer", "Rectal swab")))
mutate(ddPCR_reads_per_sample=if_else(experiment.y=="Elution buffer", 0,
                                      ddPCR_reads_per_sample))%>%
mutate(ddPCR_reads_per_sample = na_if(ddPCR_reads_per_sample, 0))

# Table 1 ddPCR
ddPCR_miseq_comparison%>%
  group_by(experiment.x)%>%
  summarize(`Mean` = mean(ddPCR_reads_per_sample, na.rm=T),
            `Median` = median(ddPCR_reads_per_sample, na.rm=T),
            `Minimum` = min(ddPCR_reads_per_sample, na.rm=T),
            `Maximum` = max(ddPCR_reads_per_sample, na.rm=T),
            `Standard deviation` = sd(ddPCR_reads_per_sample, na.rm=T),
            `Interquartile range` = IQR(ddPCR_reads_per_sample, na.rm=T))%>%
  mutate_if(is.double, ~round(., digits=2))%>%
  pivot_longer(cols=-experiment.x, names_to="metric", values_to="value")%>%
  pivot_wider(names_from=experiment.x, values_from=value)%>%
  dplyr::select(metric, Water, `Isolation control`, `Rectal swab`)
```

```
## # A tibble: 6 x 4
##   metric      Water `Isolation control` `Rectal swab`
##   <chr>      <dbl>          <dbl>          <dbl>
## 1 Mean      13.4            3150.          96625320.
## 2 Median    13.6            3223.          3165086.
## 3 Minimum   12.2            1408.          11846.
## 4 Maximum   14.4            5209.          3234306771.
## 5 Standard deviation 1.1            1278.          341147947.
## 6 Interquartile range 1.09            984.          46339173.
```

```
# Table 1 miseq
ddPCR_miseq_comparison%>%
  group_by(experiment.x)%>%
  summarize(`Mean` = mean(miseq_reads_per_sample, na.rm=T),
            `Median` = median(miseq_reads_per_sample, na.rm=T),
            `Minimum` = min(miseq_reads_per_sample, na.rm=T),
            `Maximum` = max(miseq_reads_per_sample, na.rm=T),
            `Standard deviation` = sd(miseq_reads_per_sample, na.rm=T),
            `Interquartile range` = IQR(miseq_reads_per_sample, na.rm=T))%>%
  mutate_if(is.double, ~round(., digits=2))%>%
  pivot_longer(cols=-experiment.x, names_to="metric", values_to="value")%>%
  pivot_wider(names_from=experiment.x, values_from=value)%>%
  dplyr::select(metric, Water, `Isolation control`, `Rectal swab`)
```

```
## # A tibble: 6 x 4
##   metric      Water `Isolation control` `Rectal swab`
##   <chr>      <dbl>          <dbl>          <dbl>
## 1 Mean      73858.          53687.          73926.
## 2 Median    84455.          55708.          74505.
```

```
## 3 Minimum          44181.          23946.          83.7
## 4 Maximum          92939.          78621.         145353.
## 5 Standard deviation 26049.          17298.          22790.
## 6 Interquartile range 24379.          20939.          29514.
```

#### *# Statistical testing*

```
TukeyHSD(aov(log(ddPCR_reads_per_sample)~experiment.y,data=ddPCR_miseq_comparison))
```

```
## Tukey multiple comparisons of means
## 95% family-wise confidence level
##
## Fit: aov(formula = log(ddPCR_reads_per_sample) ~ experiment.y, data = ddPCR_miseq_comparison)
##
## $experiment.y
##
```

	diff	lwr	upr	p adj
Isolation control-Water	5.388694	-0.04311471	10.82050	0.0523715
Rectal swab-Water	12.242045	7.77865241	16.70544	0.0000000
Rectal swab-Isolation control	6.853351	3.67734340	10.02936	0.0000022

```
TukeyHSD(aov(miseq_reads_per_sample~experiment.y,data=ddPCR_miseq_comparison))
```

```
## Tukey multiple comparisons of means
## 95% family-wise confidence level
##
## Fit: aov(formula = miseq_reads_per_sample ~ experiment.y, data = ddPCR_miseq_comparison)
##
## $experiment.y
##
```

	diff	lwr	upr	p adj
Isolation control-Water	-24909.36441	-66290.248	16471.52	0.4052023
Elution buffer-Water	-15434.03756	-56814.921	25946.85	0.7695710
Rectal swab-Water	68.10432	-33935.141	34071.35	0.9999999
Elution buffer-Isolation control	9475.32685	-24312.023	43262.68	0.8868640
Rectal swab-Isolation control	24977.46873	781.845	49173.09	0.0400965
Rectal swab-Elution buffer	15502.14188	-8693.482	39697.77	0.3486703

```
just_swabs_otu<-otu_df %>%
  filter(experiment == "Swab")%>%
  dplyr::select(Sample_ID, everything())

all_data<-inner_join(ddPCR%>%dplyr::select(-experiment), otu_df)

swab_labels <- read_excel("case_control_samples_5_28_2019.xlsx") %>%
  unite("swab_label", c(swab_type, case_or_control), sep = "_", remove = FALSE)

ddPCR_labeled<-ddPCR %>% dplyr::select(-swab_type, -experiment)%>%
  inner_join(swab_labels, by = c("Sample_ID"))

all_data_swabs<-inner_join(ddPCR_labeled, just_swabs_otu, by =("Sample_ID"))
all_data_swabs<-all_data_swabs %>%
  mutate(above_threshold = as_factor(if_else(ddPCR_reads_per_sample >= 1e06, "above", "below")))
```

```

# Make all our character columns factor
all_data_swabs<-all_data_swabs %>%
  mutate(swab_label = as_factor(swab_label)) %>%
  mutate(swab_type = as_factor(swab_type)) %>%
  mutate(case_or_control = as_factor(case_or_control))%>%
  mutate(shannon=diversity(.[,8:(nrow(.)-1)]))

all_data_swabs<-as.data.frame(all_data_swabs) # Hadley Wickham won't let you set rownames! Gotta change
rownames(all_data_swabs) <- all_data_swabs$Sample_ID
# all_data_swabs_df with rownames added for PCA analysis later
# note that the data is in wide format

all_data_swabs_tidy<-all_data_swabs%>%gather("Otu", "relative_abundance", -c(Sample_ID,ddPCR_reads_per_
  mutate(Otu = as_factor(Otu))

# link OTU to genus
otu_genus_link <- otu_good_taxonomy%>%
  dplyr::select(OTU,Genus)%>%
  mutate(Otu_genus = str_c(OTU,Genus, sep="-"))%>%
  rename("Otu"=OTU)%>%
  mutate(Otu = as_factor(Otu))# first create a new variable

all_data_swabs%>%
  dplyr::select(above_threshold, Otu0001)%>%
  mutate(contaminant_present = as.numeric(Otu0001>0))%>%
  # group_by(above_threshold)%>%
  summarize(percent_contaminated = mean(contaminant_present))

##   percent_contaminated
## 1                0.6239316

```

```

swab_initial<-all_data_swabs%>%
  filter(swab_type == "initial")%>%
  pull(Sample_ID)

samples_for_first_plot<-c(swab_initial,"1","2","3","4","5","6","NEGP","NEGQ","NEGR","NEGS","NEGT","NEGU",
  "NEGC","NEGD","NEGE","NEGF","NEGG","NEGH","NEGI","NEGJ","NEGK","NEGL","NEGM","NEGN","NEGO")

correlation_ddPCR_miseq<-ddPCR_miseq_comparison %>%
  filter(!is.na(experiment.y), Sample_ID %in% samples_for_first_plot)%>%
  ggplot(aes(x=ddPCR_reads_per_sample, y =miseq_reads_per_sample))+
  geom_point(aes(color= experiment.x),alpha=0.5)+
  scale_x_log10()+
  scale_y_log10()+
  labs(x="ddPCR: 16S gene copies per sample",
       y = "Illumina MiSeq 16S reads per sample") +
  coord_cartesian(ylim = c(10, max(Miseq_quant$miseq_reads_per_sample)))+
  theme_bw()+
  theme(panel.grid=element_blank(),
        legend.position = c(0.12,0.3),
        panel.border = element_blank(),
        axis.line = element_line())+

```

```

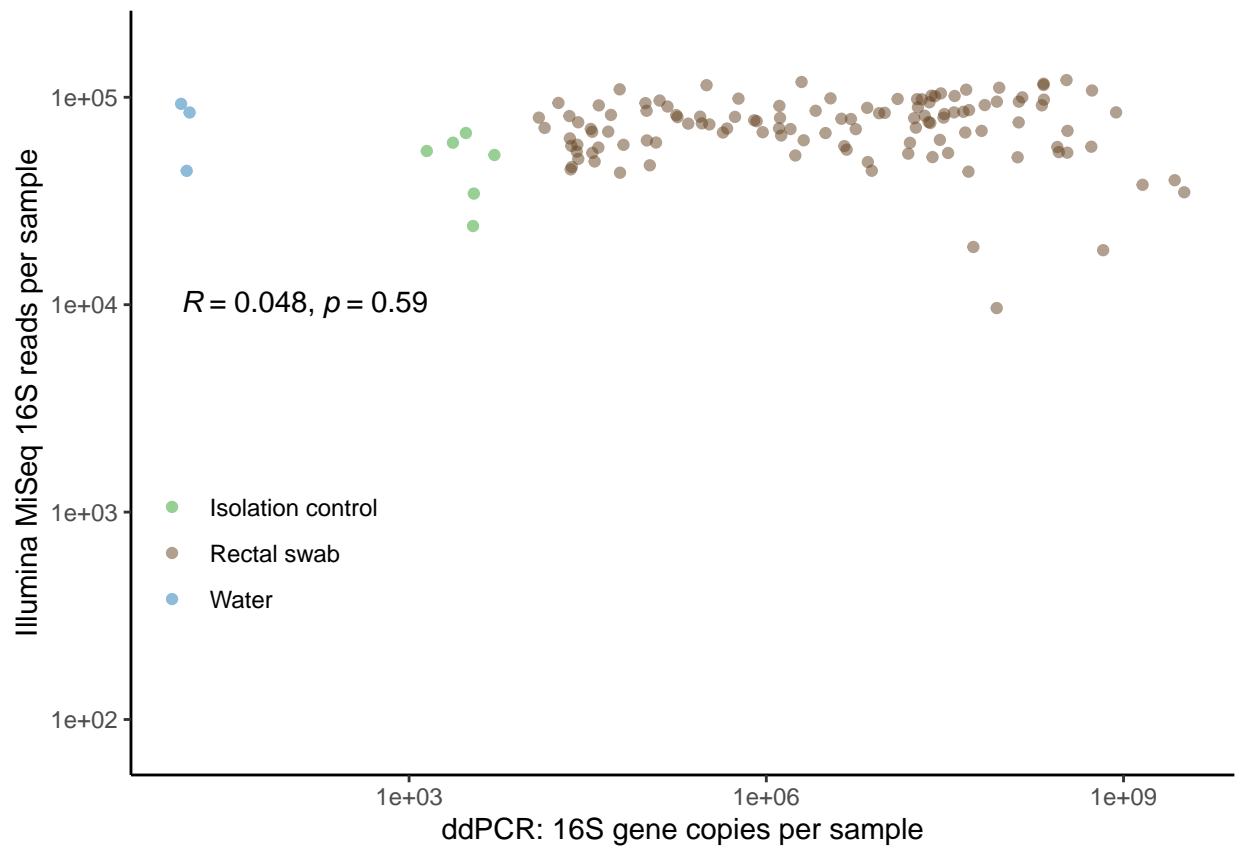
scale_color_manual(values=c("#33a02c", "#654321", "#1f78b4"))+
labs(color = NULL, cor=NULL)+
stat_cor(label.y=4, method="pearson")+
coord_cartesian(ylim=c(101.9, 105.25))

correlation_ddPCR_miseq

```

```
## Warning: Removed 6 rows containing non-finite values (stat_cor).
```

```
## Warning: Removed 6 rows containing missing values (geom_point).
```



```

comparisons<-list(c("Isolation\ncontrol", "Rectal\nswab"),
                  c("Elution\nbuffer", "Rectal\nswab"),
                  c("Water", "Rectal\nswab"))

negative_control_v_sample<-ddPCR_miseq_comparison %>%
  pivot_longer(cols=c(ddPCR_reads_per_sample, miseq_reads_per_sample),
               names_to="read_type",
               values_to="read_count")%>%
  mutate(experiment.x = case_when(experiment.y=="Elution buffer"~"Elution\nbuffer",
                                   experiment.y=="Isolation control"~"Isolation\ncontrol",
                                   experiment.y=="Rectal swab"~"Rectal\nswab",
                                   experiment.y=="Water"~"Water",
                                   is.na(experiment.y)~"Water"),

```

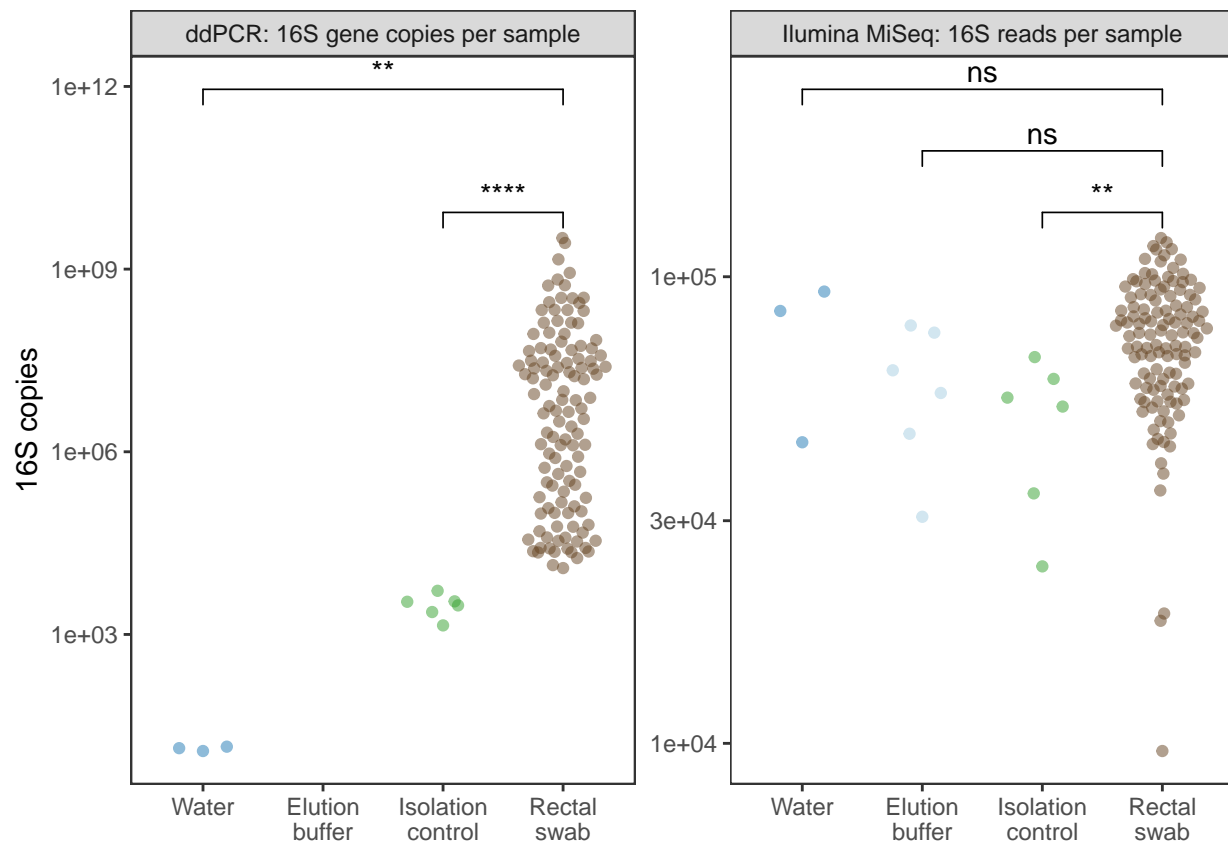
```

    experiment.x=factor(experiment.x, levels=c("Water", "Elution\nbuffer", "Isolation\ncontrol",
        "Rectal\nswab")),
    read_type = if_else(read_type=="ddPCR_reads_per_sample",
        "ddPCR: 16S gene copies per sample",
        "Illumina MiSeq: 16S reads per sample"))>%
filter(!is.na(experiment.y), Sample_ID %in% samples_for_first_plot)>%
ggplot(aes(x=experiment.x, y = read_count, color=experiment.x))+
ggbeeswarm::geom_quasirandom(alpha=0.5)+
facet_wrap(~read_type, scales="free")+
scale_y_log10()+
theme_bw()+
theme(panel.grid=element_blank(),
    legend.position = "none")+
labs(x=NULL, y = "16S copies")+
scale_color_manual(values=c("#1f78b4", "#a6cee3", "#33a02c", "#654321"))+
ggpubr::stat_compare_means(comparisons=comparisons, label = "p.signif", method="wilcox.test")
negative_control_v_sample

```

## Warning: Removed 6 rows containing non-finite values (stat\_signif).

## Warning: Removed 6 rows containing missing values (position\_quasirandom).



```
ddpcr_v_miseq<-ggarrange(negative_control_v_sample,
  correlation_ddPCR_miseq,
  nrow=2, align="h", labels = c("A", "B"))
```

```
## Warning: Removed 6 rows containing non-finite values (stat_signif).
```

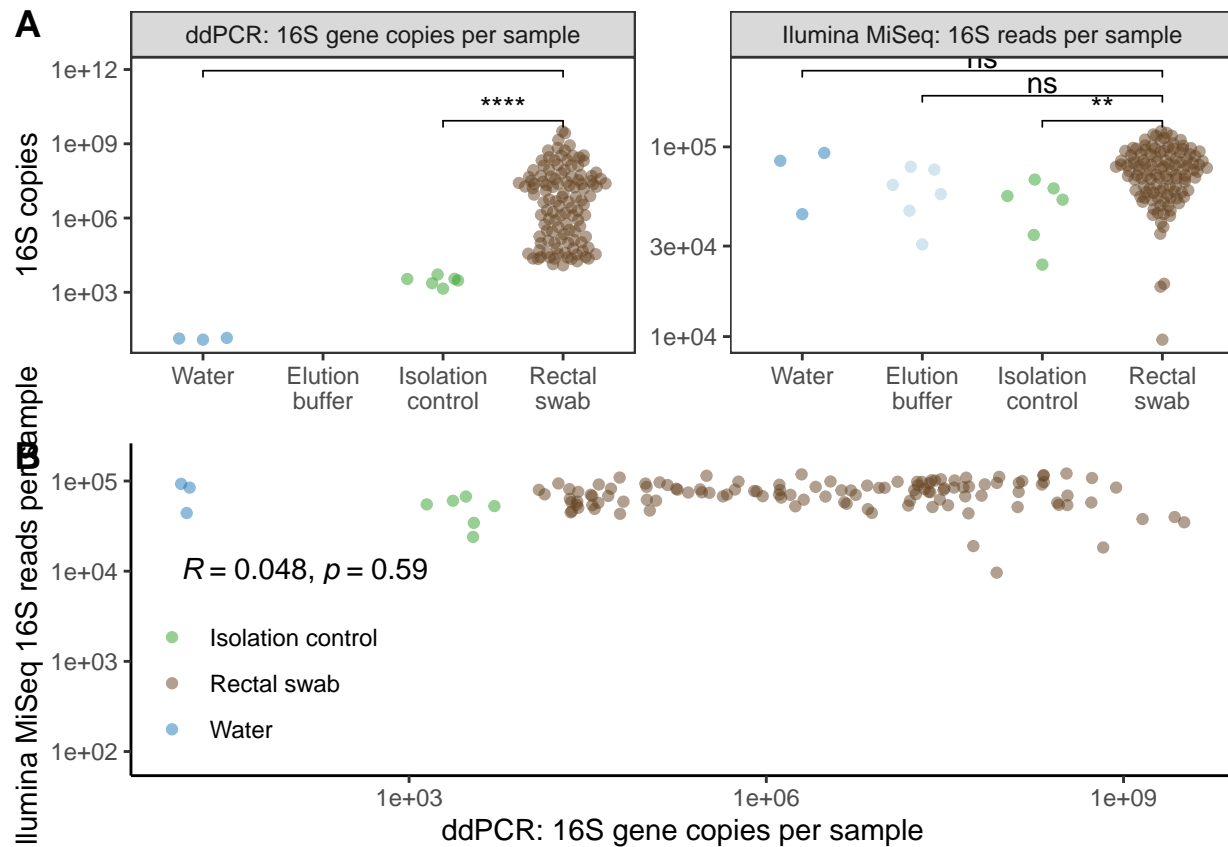
```
## Warning: Removed 6 rows containing missing values (position_quasirandom).
```

```
## Warning: Removed 6 rows containing non-finite values (stat_cor).
```

```
## Warning: Removed 6 rows containing missing values (geom_point).
```

```
## Warning: Graphs cannot be horizontally aligned unless the axis parameter is set.
## Placing graphs unaligned.
```

```
# ddpcr_v_miseq
# ggeexport(ddpcr_v_miseq, filename="ddpcr_v_miseq.pdf")
ddpcr_v_miseq
```



```
contaminant_abundance<-tidy_genus_df %>%
  filter(experiment %in% c("AE", "Empty", "WaterNeg", "IsoCtrl") , relative_abundance > 1) %>%
  group_by(Otu_genus)%>%
  summarize(mean_abundance=mean(relative_abundance, na.rm = TRUE))%>%
```

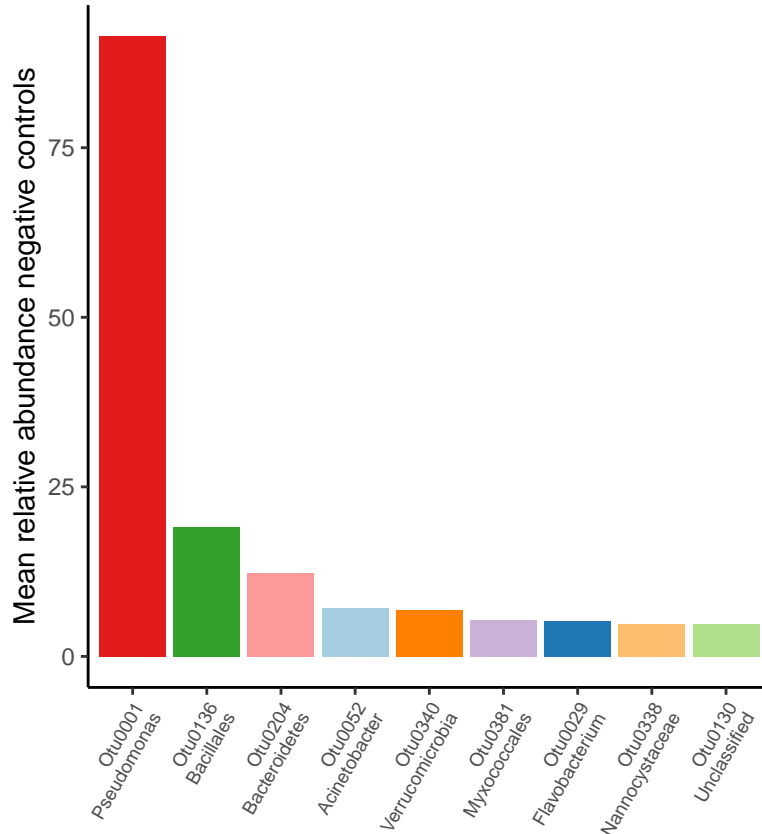


```

ungroup()%>%
mutate(Otu_genus=str_replace(Otu_genus,"_unclassified",""),
      Otu_genus=str_replace(Otu_genus,"Bacteria","Unclassified"),
      Otu_genus=str_replace(Otu_genus,"-","\n"),
      Otu_genus=factor(Otu_genus))%>%
filter(mean_abundance>4.5)%>%
ggplot(aes(x = fct_reorder(Otu_genus, mean_abundance, .desc = TRUE),
      y = mean_abundance,
      fill = Otu_genus)) +
geom_bar(stat = "identity") +
labs(x = "Otu", y = "Mean relative abundance negative controls") +
theme_bw()+
theme(axis.text.x= element_text(angle=60, hjust=1, size=7),
      legend.position="none",
      panel.grid = element_blank(),
      aspect.ratio = 1,
      panel.border = element_blank(),
      axis.line = element_line())+
labs(x=NULL)+
  scale_fill_manual(values = c("#e31a1c", "#1f78b4", "#a6cee3",
                              "#b2df8a", "#33a02c", "#fb9a99",
                              "#fdbf6f", "#ff7f00",
                              "#cab2d6", "#6a3d9a"))

```

contaminant\_abundance



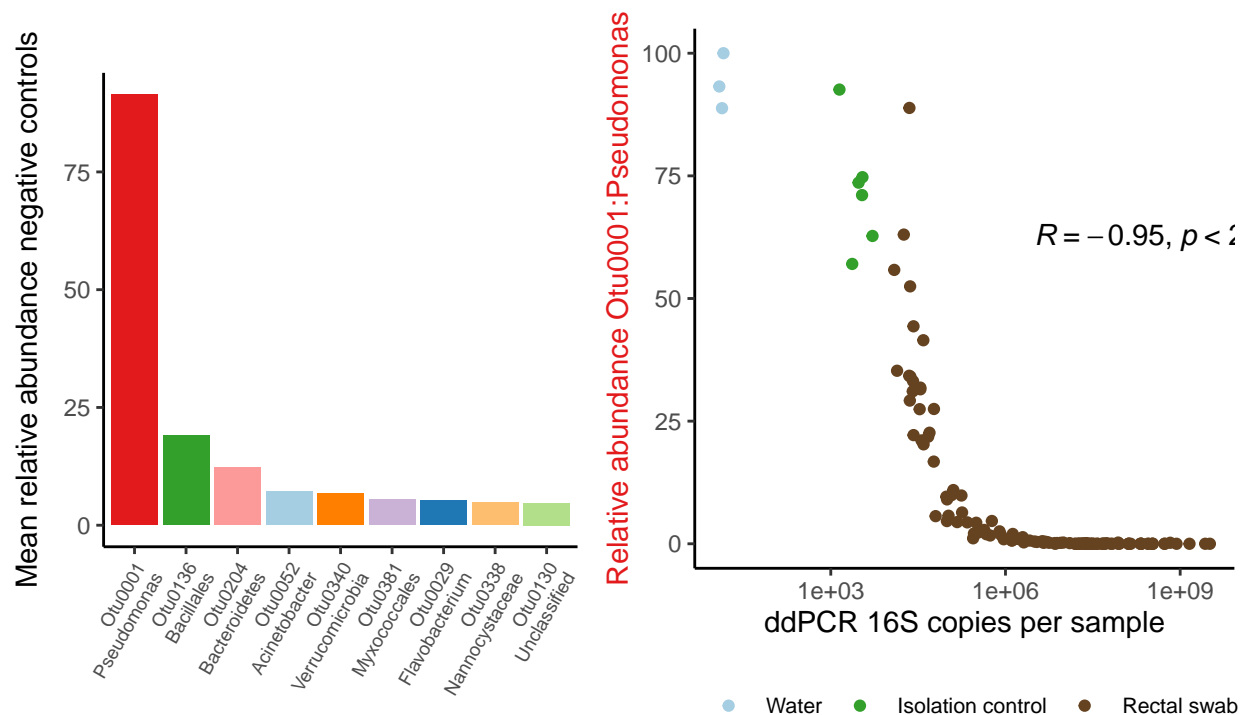
```

pseudo_v_abundance<-all_data%>%
  mutate(experiment=case_when(experiment == "AE" ~ "Elution buffer",
                              experiment == "IsoCtrl" ~ "Isolation control",
                              experiment == "Swab" ~ "Rectal swab",
                              experiment == "Water" ~ "Water"),
         experiment=factor(experiment,
                           levels=c("Water", "Elution buffer",
                                     "Isolation control", "Rectal swab")))%>%
  filter(Sample_ID %in% samples_for_first_plot,
         experiment != "Elution buffer")%>%
  ggplot(aes(x=ddPCR_reads_per_sample, y=Otu0001))+
  geom_point(aes(color=experiment))+
  scale_x_log10()+
  theme_bw()+
  theme(panel.grid = element_blank(),
        panel.border = element_blank(),
        axis.line = element_line(),
        legend.position = "bottom",
        axis.title.y = element_text(color="#e31a1c"),
        legend.text = element_text(size=7.5),
        aspect.ratio = 1)+
  labs(color=NULL, x="ddPCR 16S copies per sample", y = "Relative abundance Otu0001:Pseudomonas")+
  scale_color_manual(values=rev(c("#654321", "#33a02c", "#a6cee3")))+
  stat_cor(method="spearman", show.legend = FALSE, label.y=62.5, label.x=6.5)

joined_contamination<-ggarrange(contaminant_abundance, pseudo_v_abundance, nrow=1,
                                widths=c(0.9,1),align="v")

joined_contamination

```



```
# ggelexport(joined_contamination, filename="joined_contamination_edited.pdf")
```

```
just_initial<-all_data_swabs%>%
  filter(swab_type=="initial")

otu.swab.hel<- just_initial %>%
  dplyr::select(contains("Otu"))%>%
  decostand("hellinger")

tidy_swab_pca <- prcomp(otu.swab.hel)

loadings<-data.frame(tidy_swab_pca$rotation)%>%
  mutate(Sample_ID = rownames(.))%>%
  dplyr::select(Sample_ID,PC1, PC2)%>%
  mutate(size=sqrt(PC1^2+PC2^2))%>%
  arrange(desc(size))%>%
  head(5)%>%
  mutate(above_threshold="loading")

plot_by_burden<-data.frame(tidy_swab_pca$x)%>%
  mutate(Sample_ID = just_initial$Sample_ID)%>%
  dplyr::select(Sample_ID, PC1, PC2)%>%
```

```
mutate(size=sqrt(PC1^2+PC2^2))%>%
inner_join(all_data_swabs%>%dplyr::select(Sample_ID, above_threshold))%>%
mutate(above_threshold = case_when(above_threshold == "above"~"High biomass",
                                   above_threshold == "below" ~"Low biomass"))
```

```
## Joining, by = "Sample_ID"
```

```
find_centroids<-plot_by_burden%>%
  group_by(above_threshold)%>%
  summarize(PC1_mean = mean(PC1),
            PC2_mean = mean(PC2))
```

```
principal_component_plot<-
  ggplot(data=plot_by_burden)+
  geom_point(aes(x=PC1, y=PC2,color=above_threshold),show.legend = FALSE, alpha=0.5)+
  geom_point(data = find_centroids, aes(x=PC1_mean, y =PC2_mean, color=above_threshold),size=7)+
  stat_ellipse(aes(x=PC1, y=PC2,color=above_threshold),type = "norm", linetype = 2, show.legend = FALSE)+
  theme_bw()+
  theme(panel.grid=element_blank(),
        legend.title = element_blank(),
        legend.position = "none",
        axis.title = element_text(size=9))+
  scale_color_manual(values=c("#1f78b4","#e31a1c"))+
  labs(x="PC1 (11.8%) explained", y="PC2 (8.6%) explained")
```

```
myControl <- trainControl(
  method="cv",
  number=10,
  verboseIter = TRUE
)
gridsearch_burden<-data.frame(
  mtry=seq(1:10),
  splitrule="gini",
  min.node.size=5
)

set.seed(4763)
otu_burden_model <- train(
  above_threshold~.,
  method = "ranger",
  importance="permutation",
  oob.error = TRUE,
  seed = 4763,
  trControl = myControl,
  tuneGrid = gridsearch_burden,
  data = all_data_swabs%>%
  dplyr::select(- nearZeroVar(.),-Sample_ID,
                -ddPCR_reads_per_sample, -swab_type,
                -swab_label)%>%
  mutate(pair_ID=factor(pair_ID),
         case_or_control=factor(case_or_control),
         above_threshold=factor(above_threshold))
```

[illegible]



[illegible]

```

## - Fold08: mtry=10, splitrule=gini, min.node.size=5
## + Fold09: mtry= 1, splitrule=gini, min.node.size=5
## - Fold09: mtry= 1, splitrule=gini, min.node.size=5
## + Fold09: mtry= 2, splitrule=gini, min.node.size=5
## - Fold09: mtry= 2, splitrule=gini, min.node.size=5
## + Fold09: mtry= 3, splitrule=gini, min.node.size=5
## - Fold09: mtry= 3, splitrule=gini, min.node.size=5
## + Fold09: mtry= 4, splitrule=gini, min.node.size=5
## - Fold09: mtry= 4, splitrule=gini, min.node.size=5
## + Fold09: mtry= 5, splitrule=gini, min.node.size=5
## - Fold09: mtry= 5, splitrule=gini, min.node.size=5
## + Fold09: mtry= 6, splitrule=gini, min.node.size=5
## - Fold09: mtry= 6, splitrule=gini, min.node.size=5
## + Fold09: mtry= 7, splitrule=gini, min.node.size=5
## - Fold09: mtry= 7, splitrule=gini, min.node.size=5
## + Fold09: mtry= 8, splitrule=gini, min.node.size=5
## - Fold09: mtry= 8, splitrule=gini, min.node.size=5
## + Fold09: mtry= 9, splitrule=gini, min.node.size=5
## - Fold09: mtry= 9, splitrule=gini, min.node.size=5
## + Fold09: mtry=10, splitrule=gini, min.node.size=5
## - Fold09: mtry=10, splitrule=gini, min.node.size=5
## + Fold10: mtry= 1, splitrule=gini, min.node.size=5
## - Fold10: mtry= 1, splitrule=gini, min.node.size=5
## + Fold10: mtry= 2, splitrule=gini, min.node.size=5
## - Fold10: mtry= 2, splitrule=gini, min.node.size=5
## + Fold10: mtry= 3, splitrule=gini, min.node.size=5
## - Fold10: mtry= 3, splitrule=gini, min.node.size=5
## + Fold10: mtry= 4, splitrule=gini, min.node.size=5
## - Fold10: mtry= 4, splitrule=gini, min.node.size=5
## + Fold10: mtry= 5, splitrule=gini, min.node.size=5
## - Fold10: mtry= 5, splitrule=gini, min.node.size=5
## + Fold10: mtry= 6, splitrule=gini, min.node.size=5
## - Fold10: mtry= 6, splitrule=gini, min.node.size=5
## + Fold10: mtry= 7, splitrule=gini, min.node.size=5
## - Fold10: mtry= 7, splitrule=gini, min.node.size=5
## + Fold10: mtry= 8, splitrule=gini, min.node.size=5
## - Fold10: mtry= 8, splitrule=gini, min.node.size=5
## + Fold10: mtry= 9, splitrule=gini, min.node.size=5
## - Fold10: mtry= 9, splitrule=gini, min.node.size=5
## + Fold10: mtry=10, splitrule=gini, min.node.size=5
## - Fold10: mtry=10, splitrule=gini, min.node.size=5
## Aggregating results
## Selecting tuning parameters
## Fitting mtry = 8, splitrule = gini, min.node.size = 5 on full training set

```

```

set.seed(4763)
important_features_burden<-ranger::importance_pvalues(
  otu_burden_model$finalModel,
  method = "altmann",
  formula = above_threshold~.,
  data = all_data_swabs%>%
  dplyr::select(- nearZeroVar(.),-Sample_ID,
    -ddPCR_reads_per_sample, -swab_type,
    -swab_label)%>%

```



```
mutate(pair_ID=factor(pair_ID),
       case_or_control=factor(case_or_control),
       above_threshold=factor(above_threshold)))%>%
  as.data.frame()%>%
  rownames_to_column()%>%
  rename(features="rowname")
```

```
## Warning in cbind(x$variable.importance, pval): number of rows of result is not a
## multiple of vector length (arg 2)
```

```
otus_for_burden<-important_features_burden%>%
  filter(importance>0,
         pvalue<0.05)%>%
  inner_join(otu_good_taxonomy, by = c("features"="OTU"))%>%
  mutate(name=str_c(features, Genus, sep = " "))%>%
  rename(`Mean Decrease in Accuracy`="importance")%>%
  mutate(ci=`Mean Decrease in Accuracy`/qnorm(pvalue, lower.tail = FALSE),
         lower_ci=`Mean Decrease in Accuracy`-1.96*ci,
         upper_ci=`Mean Decrease in Accuracy`+1.96*ci,
         name=str_remove(name, "_unclassified"),
         name=str_remove(name, " incertae sedis"),
         name=str_replace(name, "_", " "),
         name=factor(name),
         name=fct_reorder(name, `Mean Decrease in Accuracy`)
         )%>%
  arrange(desc(lower_ci))

important_otus<-otus_for_burden%>%
  arrange(desc(`Mean Decrease in Accuracy`))%>%
  head(15)%>%
  pull(features)

feature_importance_plot<-ggplot(otus_for_burden, aes(x=name, y = `Mean Decrease in Accuracy`))+
  geom_segment(aes(x=name, xend=name, y=lower_ci, yend=upper_ci))+
  geom_point(color="red")+
  coord_flip()+
  theme_bw()+
  theme(panel.grid=element_blank(),
        axis.text.y = element_text(size=7),
        axis.title.x = element_text(size=9))+
  labs(x=NULL,y="Mean Decrease in Accuracy")
```

```
forest_identified_otus<-just_initial%>%
  dplyr::select(Sample_ID,Otu0001,Otu0029,Otu0008,Otu0016,Otu0026,Otu0042,Otu0058,Otu0006,Otu0005,
               ddPCR_reads_per_sample)%>%
  pivot_longer(cols=contains("Otu"), names_to="OTU", values_to="rel_abund")%>%
  mutate(bacterial_type = case_when(
    OTU %in% c("Otu0001","Otu0008", "Otu0029") ~ "Common sequencing contaminant",
    OTU %in% c("Otu0016","Otu0042")~ "Common skin bacteria",
    OTU %in% c("Otu0026","Otu0058","Otu0006","Otu0005") ~"Common gut bacteria"))%>%
  inner_join(otu_good_taxonomy)%>%
  mutate(present = if_else(rel_abund >0,"Present","Absent"),
         name=str_c(OTU, Genus, sep = " "),
```

```

    name=str_remove(name,"_unclassified"),
    name=str_replace(name, "_", " "),
    name=factor(name, levels=c("Otu0001 Pseudomonas", "Otu0008 Pseudomonas",
                              "Otu0029 Flavobacterium","Otu0016 Staphylococcus",
                              "Otu0042 Corynebacterium","Otu0026 Lactobacillus",
                              "Otu0058 Bacillus","Otu0006 Bacteroides","Otu0005 Akkermansia")))

## Joining, by = "OTU"

otu_burden<-forest_identified_otus%>%
  ggplot(aes(x=present, y=ddPCR_reads_per_sample))+
  geom_quasirandom(alpha=0.4,aes(color=bacterial_type))+
  facet_wrap(~name)+
  stat_compare_means(label = "p.signif", label.x = 1.5, label.y=8.9)+
  geom_boxplot(alpha=0, show.legend = FALSE,aes(color=bacterial_type))+
  scale_y_log10()+
  scale_x_discrete(labels = c("Absent","Present"))+
  theme_bw()+
  theme(panel.grid=element_blank(),
        legend.position = "none")+
  labs(y="16S gene copies per sample",
       x=NULL,
       color=NULL)+
  scale_color_manual(values=c("#654321","#e31a1c", "#33a02c"))

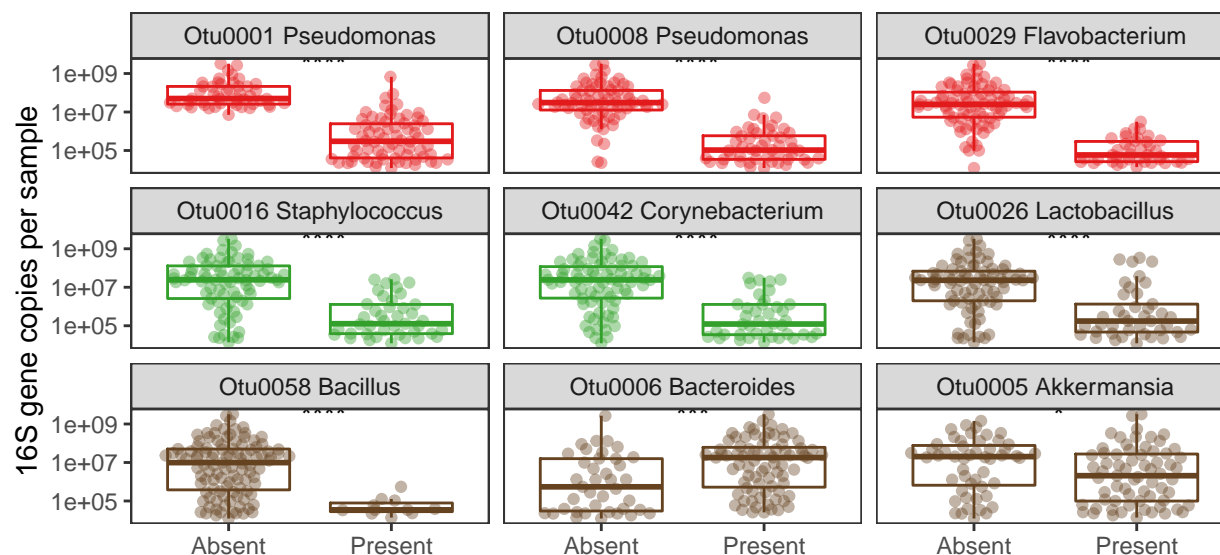
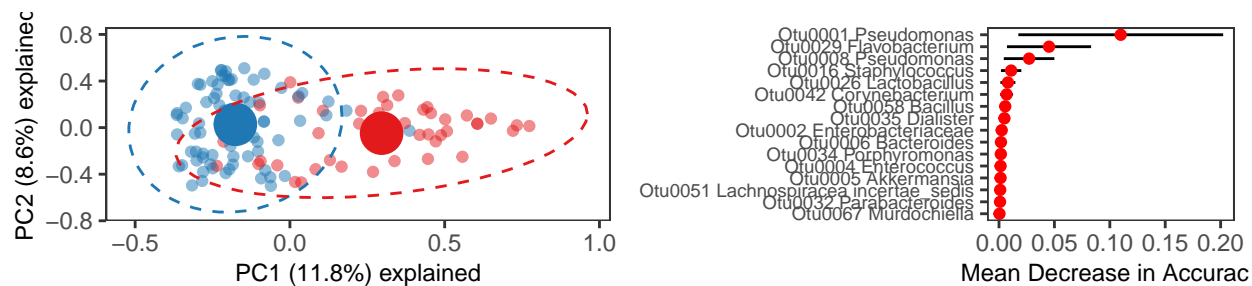
multi.plot<-ggarrange(ggarrange(principal_component_plot, feature_importance_plot),
  otu_burden,align="hv",nrow=2,heights = c(1,2))

## Warning: Graphs cannot be vertically aligned unless the axis parameter is set.
## Placing graphs unaligned.

## Warning: Graphs cannot be horizontally aligned unless the axis parameter is set.
## Placing graphs unaligned.

multi.plot

```



```
sofa_metadata <- read_csv("clinical_metadata_1.csv")
```

```
##
## -- Column specification -----
## cols(
##   .default = col_double(),
##   Sample_ID = col_character(),
##   culture_type = col_character(),
##   organism = col_character(),
##   above_threshold = col_character()
## )
## i Use `spec()` for the full column specifications.
```

```
sofa_metadata%>%
  dplyr::select(Augmentin:Zosyn)%>%
  pivot_longer(cols=c(Augmentin:Zosyn),names_to="abx", values_to="dose")%>%
  mutate(dose=as.numeric(dose>0))%>%
  group_by(abx)%>%
  summarize(total_doses=sum(dose))%>%
  arrange(desc(total_doses))
```

```
## # A tibble: 12 x 2
##   abx          total_doses
##   <chr>          <dbl>
## 1 Vancomycin      35
```

```
## 2 Flagyl          22
## 3 Zosyn           20
## 4 Cefepime       18
## 5 Ceftazidime     4
## 6 Augmentin       2
## 7 Oral_Vanco      2
## 8 Meropenem       1
## 9 Doxycycline     0
## 10 Rectal_Vanco   0
## 11 Rifaximin      0
## 12 Unasyn         0
```

```
sofa_metadata%>%
  dplyr::select(ddPCR_reads_per_sample, Augmentin:Zosyn)%>%
  pivot_longer(cols=c(Augmentin:Zosyn), names_to="antibiotic", values_to="dot")%>%
  nest(dot, ddPCR_reads_per_sample)%>%
  mutate(correlation=map(data, ~cor.test(. $dot, log(. $ddPCR_reads_per_sample))%>%
    tidy()))%>%
  unnest(correlation)%>%
  filter(!is.na(estimate),
    antibiotic %in% c("Cefepime", "Flagyl", "Vancomycin", "Zosyn"))%>%
  dplyr::select(antibiotic, estimate, p.value)%>%
  mutate_if(is.numeric, ~round(., 3))
```

```
## Warning: All elements of `...` must be named.
## Did you want `data = c(dot, ddPCR_reads_per_sample)`?
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```
## # A tibble: 4 x 3
##   antibiotic estimate p.value
##   <chr>          <dbl>   <dbl>
## 1 Cefepime      -0.023   0.805
## 2 Flagyl        0.061   0.512
## 3 Vancomycin   -0.088   0.347
## 4 Zosyn        -0.257   0.005
```

```
nested<-sofa_metadata%>%
  dplyr::select(Cefepime, Flagyl, Meropenem, Vancomycin, Zosyn:rheumd, total_sofa, -MRN, -aids, -msld, -di)
  mutate_if(is.character, ~as.numeric(.x))%>%
  mutate(log_burden = log(ddPCR_reads_per_sample))%>%
  dplyr::select(-ddPCR_reads_per_sample)%>%
  pivot_longer(cols=c(Cefepime:total_sofa), names_to="variable", values_to="value")%>%
  nest(-variable)%>%
  mutate(binary_data = map(data, ~mutate(., value=if_else(value>0, 1, 0))))%>%
  mutate(mean_binary = map(binary_data, ~summarize(., mean=mean(value))),
```

```

    mean_continous = map(data, ~summarize(., mean=mean(value))),
    se_cont=map(data, ~summarize(., se=sd(value)/(sqrt(n())))),
    standard_error_binary=map(binary_data, ~summarize(., se=sd(value)/(sqrt(n()))))%>%
  unnest(mean_continous, se_cont, mean_binary, standard_error_binary)%>%
  mutate(stat = if_else(variable %in% c("age", "total_sofa", "charlson_score"), mean1, mean))

## Warning: All elements of `...` must be named.
## Did you want `data = c(log_burden, value)`?

## Warning: unnest() has a new interface. See ?unnest for details.
## Try `df %>% unnest(c(mean_continous, se_cont, mean_binary, standard_error_binary))`, with `mutate()`

comorbidities<-nested%>%
  filter(!(variable %in% c("Cefepime", "Flagyl", "Vancomycin", "Zosyn", "Meropenem")))%>%
  mutate(real_se = if_else(variable %in% c("age", "total_sofa", "charlson_score"), se, se1))%>%
  dplyr::select(variable, stat, real_se, binary_data, data)%>%
  mutate(number=if_else(variable %in% c("age", "total_sofa", "charlson_score"), 116, stat*116),
    stat=round(stat, 2),
    real_se=round(real_se, 2),
    descriptor = if_else(variable %in% c("age", "total_sofa", "charlson_score"), real_se, number))%>%
  dplyr::select(variable, stat, descriptor, binary_data, data)

binary_comorbidities <-comorbidities%>%
  filter(!(variable %in% c("age", "total_sofa", "charlson_score")))

continous_comorbidities<-comorbidities%>%
  filter((variable %in% c("age", "total_sofa", "charlson_score")))

tested_binary_comorbidities<-binary_comorbidities%>%
  mutate(testing = map(data, ~t.test(log_burden~value, data=.)%>%
    tidy()%>%
    dplyr::select(p.value)))%>%
  unnest(testing)%>%
  mutate(compare_means = map(binary_data, ~group_by(., value)%>%
    summarize(mean_burden = mean(log_burden),
      se_burden = sd(log_burden)/sqrt(n())))%>%
  unnest(compare_means)%>%
  mutate(value = if_else(value>0, "received", "did_not_receive"))%>%
  mutate(mean_burden = round(mean_burden, 2),
    se_burden = round(se_burden*1.96, 2))%>%
  dplyr::select(-data, -binary_data)%>%
  pivot_wider(names_from=value, values_from=c(mean_burden, se_burden))%>%
  unite("without_comorbidity", mean_burden_did_not_receive, se_burden_did_not_receive, sep="±")%>%
  unite("with_comorbidity", mean_burden_received, se_burden_received, sep="±")

tested_binary_comorbidities

## # A tibble: 20 x 6
##   variable    stat descriptor p.value without_comorbidity with_comorbidity
##   <chr>      <dbl>      <dbl>   <dbl> <chr>                <chr>
## 1 ami        0.16        18    0.0254 14.83±0.7            16.31±1.03
## 2 bmt        0.17        20    0.114  15.28±0.68          14.01±1.37

```

## 3	canc	0.7	81	0.835	15.17±1.22	15.02±0.71
## 4	cdiff	0.09	10	0.721	15.03±0.65	15.42±1.96
## 5	cevd	0.21	24	0.195	14.88±0.72	15.78±1.13
## 6	chf	0.33	38	0.484	14.91±0.74	15.39±1.12
## 7	copd	0.46	53	0.621	15.21±0.87	14.9±0.86
## 8	dementia	0.03	4	0.0266	14.97±0.63	17.59±1.42
## 9	diabwc	0.41	47	0.0428	14.53±0.75	15.85±1.02
## 10	GenderCode	0.45	52	0.718	15.17±0.82	14.94±0.93
## 11	hp	0.09	10	0.668	15.03±0.66	15.4±1.5
## 12	leukemia	0.26	30.	0.583	15.17±0.7	14.76±1.28
## 13	lymphoma	0.12	14	0.608	15±0.64	15.57±2.05
## 14	metacanc	0.47	54	0.0850	14.56±0.82	15.64±0.9
## 15	mld	0.1	12	0.615	15.12±0.65	14.55±2.08
## 16	pud	0.14	16	0.614	15.12±0.68	14.72±1.35
## 17	pvd	0.06	7	0.315	14.99±0.64	16.23±2.15
## 18	RaceCode	0.15	17	0.423	15.17±0.67	14.46±1.57
## 19	rend	0.4	46	0.149	14.7±0.79	15.63±0.97
## 20	rheumd	0.04	5	0.649	15.03±0.63	15.81±3.06

```
antibiotics<-nested%>%
  filter(variable %in% c("Cefepime","Flagyl","Vancomycin","Zosyn"))%>%
  mutate(num = map(binary_data, ~summarize(.,number = sum(value))))%>%
  unnest(num)%>%
  mutate(variable = if_else(variable == "Flagyl", "Metronidazole",variable),
         variable = if_else(variable == "Zosyn", "Piperacillin-Tazobactam",variable),
         stat = round(stat, 2))%>%
  arrange(desc(number))%>%
  rename(number_received="number")%>%
  mutate(compare_means = map(binary_data, ~t.test(log_burden~value, data=.)%>%
    tidy())%>%
    mutate(p.value=round(p.value,digits=3))%>%
    dplyr::select(p.value)))%>%
  unnest(compare_means)%>%
  mutate(compare_means = map(binary_data, ~group_by(.,value)%>%
    summarize(mean_burden = mean(log_burden),
              se_burden = sd(log_burden)/sqrt(n()))))%>%
  unnest(compare_means)%>%
  mutate(value = if_else(value>0,"received","did_not_receive"))%>%
  mutate(mean_burden = round(mean_burden, 2),
         se_burden = round(se_burden,2))%>%
  dplyr::select(-data,-binary_data, -mean,-mean1,-se,-se1)%>%
  pivot_wider(names_from=value,values_from=c(mean_burden, se_burden))%>%
  unite("did_not_recieve_burden",mean_burden_did_not_receive, se_burden_did_not_receive, sep="+")%>%
  unite("did_receive_burden",mean_burden_received, se_burden_received, sep="+")
```

antibiotics

##	#	A tibble:	4 x 6
##		variable	stat number_received p.value did_not_recieve_b~ did_receive_bur~
##		<chr>	<dbl> <dbl> <dbl> <chr> <chr>
##	1	Vancomycin	0.3 35 0.580 15.18±0.39 14.81±0.53
##	2	Metronidazo~	0.19 22 0.791 15.03±0.37 15.21±0.55
##	3	Piperacilli~	0.17 20 0.006 15.46±0.34 13.15±0.7
##	4	Cefepime	0.16 18 0.352 14.95±0.35 15.7±0.72

```

nested<-sofa_metadata%>%
  dplyr::select(Cefepime, Flagyl, Meropenem, Vancomycin, Zosyn:rheumd, total_sofa)%>%
  mutate_if(is.character, ~as.numeric(.x))%>%
  mutate(Sample_ID = sofa_metadata$Sample_ID)%>%
  pivot_longer(cols=-Sample_ID, names_to="variable", values_to="value"%>%
  inner_join(all_data_swabs%>%dplyr::select(Sample_ID, ddPCR_reads_per_sample))%>%
  mutate(log_reads=log(ddPCR_reads_per_sample))%>%
  dplyr::select(-Sample_ID,-ddPCR_reads_per_sample)%>%
  filter(!(variable %in% c("MRN", "Augmentin","Unasyn",
                          "Opiate_use","Doxycycline","Rifaxamin",
                          "Oral_Vanco","Rectal_Vanco","Cefoxitin")))%>%

  nest(-variable)%>%
  mutate(cor_result = map(data, ~cor.test(.$value, .$log_reads, data=.)%>%
                                tidy()))%>%

  unnest(cor_result)%>%
  dplyr::select(-statistic,-parameter,-method,-alternative)%>%
  mutate_if(is.numeric, ~round(.,3))%>%
  rename(`Pearson r`="estimate")%>%
  mutate(conf.low = as.character(conf.low),
         conf.high = as.character(conf.high),
         `(` = "(",
         `)` = ")")%>%
  unite(col="lower_ci",`(`, conf.low, sep="")%>%
  unite(col="upper_ci",conf.high,`)`, sep="")%>%
  unite(col = "95% CI", lower_ci, upper_ci, sep="-")%>%
  filter(!is.na(`Pearson r`))

```

```
## Joining, by = "Sample_ID"
```

```
## Warning: All elements of `...` must be named.
## Did you want `data = c(value, log_reads)`?
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```
## Warning in cor(x, y): the standard deviation is zero
```

```

nested%>%
  dplyr::select(-data)

```

```

## # A tibble: 28 x 4
##   variable   `Pearson r` p.value `95% CI`
##   <chr>         <dbl>   <dbl> <chr>
## 1 Cefepime    -0.023   0.805 (-0.205-0.16)
## 2 Flagyl       0.061   0.512 (-0.122-0.241)
## 3 Meropenem    0.042   0.652 (-0.141-0.223)
## 4 Vancomycin  -0.088   0.347 (-0.266-0.096)
## 5 Zosyn       -0.257   0.005 (-0.419--0.078)
## 6 age         0.288   0.002 (0.112-0.447)
## 7 ami         0.159   0.088 (-0.024-0.332)
## 8 bmt        -0.143   0.126 (-0.317-0.041)

```

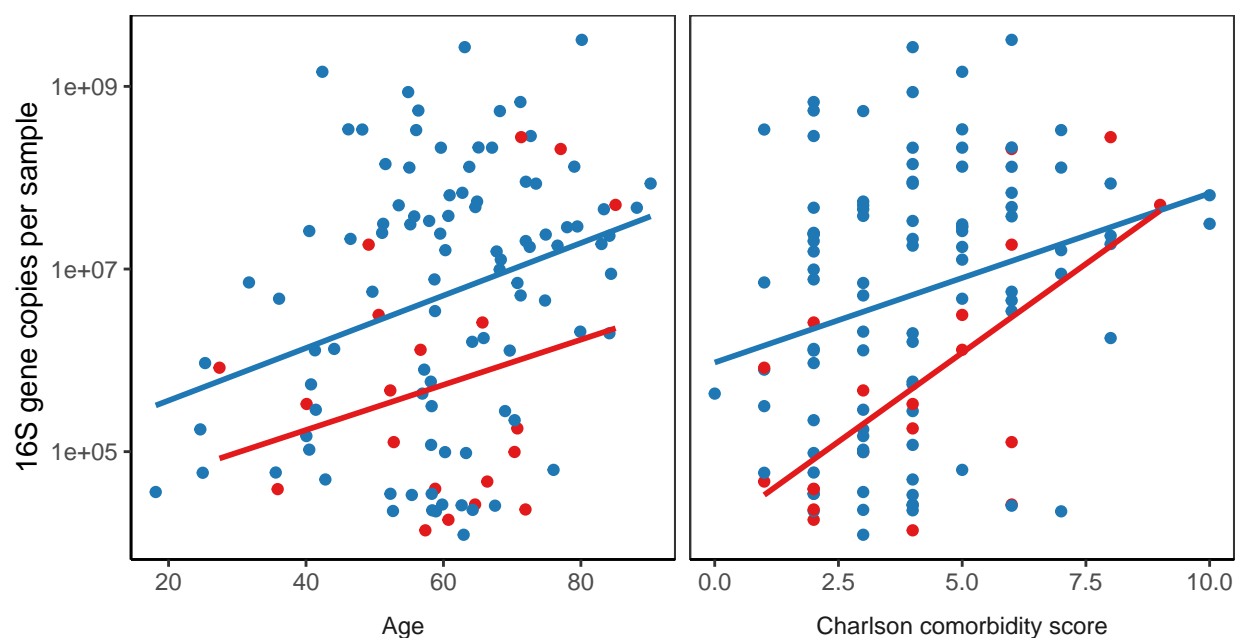
```
## 9  canc          -0.021   0.827 (-0.202-0.162)
## 10 cdiff          0.032   0.732 (-0.151-0.213)
## # ... with 18 more rows
```

```
clinical_predictors_initial<-sofa_metadata%>%
  mutate(`Piperacillin-Tazobactam use` = factor(if_else(Zosyn>0,"Recieved in Emergency Department","Did
dplyr::select(ddPCR_reads_per_sample, age, charlson_score,`Piperacillin-Tazobactam use`)%>%
  rename(Age="age",
         `Charlson comorbidity score` = "charlson_score"))%>%
  pivot_longer(cols=-c(ddPCR_reads_per_sample,`Piperacillin-Tazobactam use`),names_to="vars",values_to=
  ggplot(aes(x=values,y=ddPCR_reads_per_sample,color=`Piperacillin-Tazobactam use`))+
  geom_point()+
  scale_y_log10()+
  facet_wrap(~vars,scales="free_x",ncol=3,strip.position = "bottom")+
  theme_bw()+
  theme(panel.grid = element_blank(),
        panel.background = element_blank(),
        axis.line = element_line(),
        legend.position = "bottom",
        aspect.ratio = 1,strip.placement = "outside",
        strip.background = element_blank())+
  labs(x=NULL, y="16S gene copies per sample")+
  geom_smooth(se=FALSE,method="lm",show.legend = FALSE)+
  scale_color_manual(values = c("#1f78b4","#e31a1c","#000000"))
```

```
clinical_predictors_initial
```

```
## `geom_smooth()` using formula 'y ~ x'
```

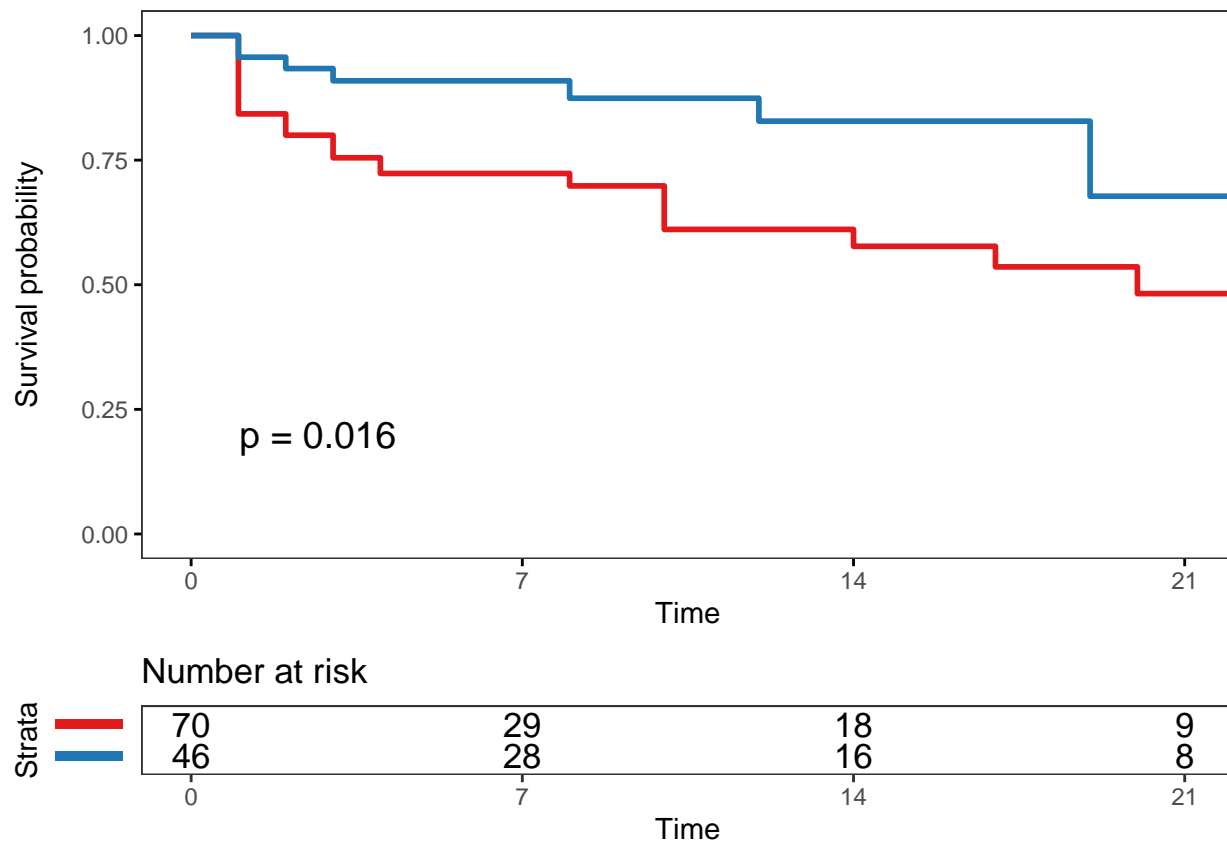




Piperacillin–Tazobactam use    ● Did not receive    ● Recieved in Emergency Department

```
kaplan_meier_fit <- survfit(Surv(survival_time, infection) ~ t, data = sofa_metadata%>%
  # filter(survival_time<=25)%>%
  rename(t="above_threshold"))
survp<-ggsurvplot(kaplan_meier_fit,conf.int = FALSE, # Add confidence interval
  pval = TRUE,
  risk.table = T,
  xlim=c(0,21),
  tables.y.text = FALSE,
  break.time.by = 7,
  censor = F,
  legend = "none",
  palette = c("#e31a1c","#1f78b4"),
  ggtheme = theme_bw()+theme(panel.grid = element_blank()))

survp
```



```
cleantime<-read_csv("cleantime.csv")
```

```
##
## -- Column specification -----
## cols(
##   MRN = col_double(),
##   admit = col_character(),
##   time = col_character(),
##   Sample_ID = col_character(),
##   case_or_control = col_character(),
##   pair_ID = col_double(),
##   swab_type = col_character()
## )
```

```
Single_Patient_Summary <- read_csv("VRE-Single-Patient-Summary.csv")%>%
  inner_join(cleantime%>%
    dplyr::select(Sample_ID, time)%>%
    mutate(time = mdy_hm(time)),
    by = c("swab_1_time"="time"))%>%
  inner_join(sofa_metadata)%>%
  inner_join(all_data_swabs)%>%dplyr::select(Sample_ID, case_or_control,pair_ID)
```

```
##
## -- Column specification -----
## cols(
```

```
## hosp_id = col_double(),
## APS_Score = col_double(),
## Comorbidity_Score = col_double(),
## APACHE_Score = col_double(),
## days_between_swabs = col_double(),
## swab_1_time = col_datetime(format = ""),
## swab_2_time = col_datetime(format = ""),
## spo2_count = col_double(),
## pao2_count = col_double(),
## fio2_count = col_double(),
## pao2_count_24 = col_double(),
## invasive = col_double(),
## supl = col_double(),
## ra = col_double(),
## hfnc = col_double(),
## noninvasive = col_double()
## )
```

```
## Joining, by = "Sample_ID"
## Joining, by = "Sample_ID"
```

```
reason_for_admit <- read_csv("reason_for_admit.csv")%>%
  inner_join(cleantime)%>%
    dplyr::select(MRN, Sample_ID))%>%
  dplyr::select(-MRN)
```

```
##
## -- Column specification -----
## cols(
##   MRN = col_double(),
##   ProblemDescription = col_character(),
##   Category = col_character()
## )
```

```
## Joining, by = "MRN"
```

```
admit_dx<-
  inner_join(Single_Patient_Summary, reason_for_admit)%>%
  distinct()%>%
  mutate(Category = as.character(Category))%>%
  mutate(reason_for_admit =case_when(
    Category == "connective_tissue"~"baseline",
    Category %in% c("cardiac","respiratory_failure","dehydration")~
      "cardio_pulm",
    Category %in% c("gi_bleed","GI_anatomic")~"gastro",
    Category %in% c("pain","lymphoma","solid_malignancy",
      "acute_leukemia")~"malignancy",
    Category == "transplant"~"transplant",
    Category == "neurologic"~"neurologic",
    Category == "trauma"~"trauma",
    str_detect(Category, "infection")~"sepsis",
  ),
```

```

reason_for_admit=factor(reason_for_admit),
reason_for_admit=fct_reorder(reason_for_admit, ddPCR_reads_per_sample),
cardio_pulm = if_else(reason_for_admit=="cardio_pulm",1,0),
gastro=if_else(reason_for_admit=="gastro",1,0),
malignancy=if_else(reason_for_admit=="malignancy",1,0),
trauma=if_else(reason_for_admit=="trauma",1,0),
sepsis =if_else(str_detect(reason_for_admit, "sepsis"),1,0),
neuro = if_else(reason_for_admit=="neurologic",1,0),
transplant=if_else(reason_for_admit=="transplant",1,0))

```

```
## Joining, by = "Sample_ID"
```

```

survival<-coxph(Surv(survival_time, infection) ~ log(ddPCR_reads_per_sample)+
  total_sofa+Comorbidity_Score+case_or_control+
  +Zosyn+sepsis+
  frailty(factor(pair_ID), distribution = "gaussian",
    sparse = FALSE, method = "reml"),
  data = admit_dx)

```

```

conf_int_survival<-survival%>%
  confint()%>%
  data.frame()%>%
  mutate(across(is.numeric, ~exp(.)))%>%
  rownames_to_column(var="variable")%>%
  filter(!str_detect(variable,"gauss"))%>%
  rename(lower_ci="X2.5..",
    upper_ci="X97.5..",
  )%>%
  mutate(range = (upper_ci-lower_ci)/2)

```

```
## Warning: Predicate functions must be wrapped in `where()`.
```

```

##
## # Bad
## data %>% select(is.numeric)
##
## # Good
## data %>% select(where(is.numeric))
##
## i Please update your code.
## This message is displayed once per session.

```

```

coef_survival<-survival%>%
  summary()%>%
  coef()%>%
  data.frame()%>%
  rownames_to_column(var="variable")%>%
  mutate(variable = if_else(variable=="log(ddPCR_reads_per_sampl", "log(ddPCR_reads_per_sample)", variable))
  mutate(coef=exp(coef),
    se.coef.=exp(se.coef.),
    se2=exp(se2))

```

```

mixed_effects_survival<-inner_join(coef_survival, conf_int_survival, by ="variable")%>%
  mutate(variable = str_replace(variable, "case_or_controlcase", "vre_colonized"))%>%
  dplyr::select(variable, coef, p, lower_ci, upper_ci, range)%>%
  mutate(across(where(is.numeric), ~round(.,3)))%>%
  mutate(lower_ci = as.character(lower_ci),
         upper_ci = as.character(upper_ci))%>%
  mutate(left_parenth="(",
         right_parenth=")")%>%
  unite(left_side, c("left_parenth", "lower_ci"), sep="")%>%
  unite(right_side, c("upper_ci", "right_parenth"), sep="")%>%
  unite(ninety_five_conf, c("left_side", "right_side"), sep="-")

```

```
mixed_effects_survival
```

```

##           variable  coef      p ninety_five_conf range
## 1 log(ddPCR_reads_per_sample) 1.213 0.003   (1.067-1.378) 0.155
## 2           total_sofa 0.987 0.845   (0.866-1.125) 0.129
## 3      Comorbidity_Score 1.017 0.650   (0.947-1.092) 0.072
## 4       vre_colonized 0.632 0.211   (0.308-1.298) 0.495
## 5             Zosyn 2.320 0.082   (0.899-5.986) 2.543
## 6             sepsis 2.203 0.056   (0.981-4.944) 1.981

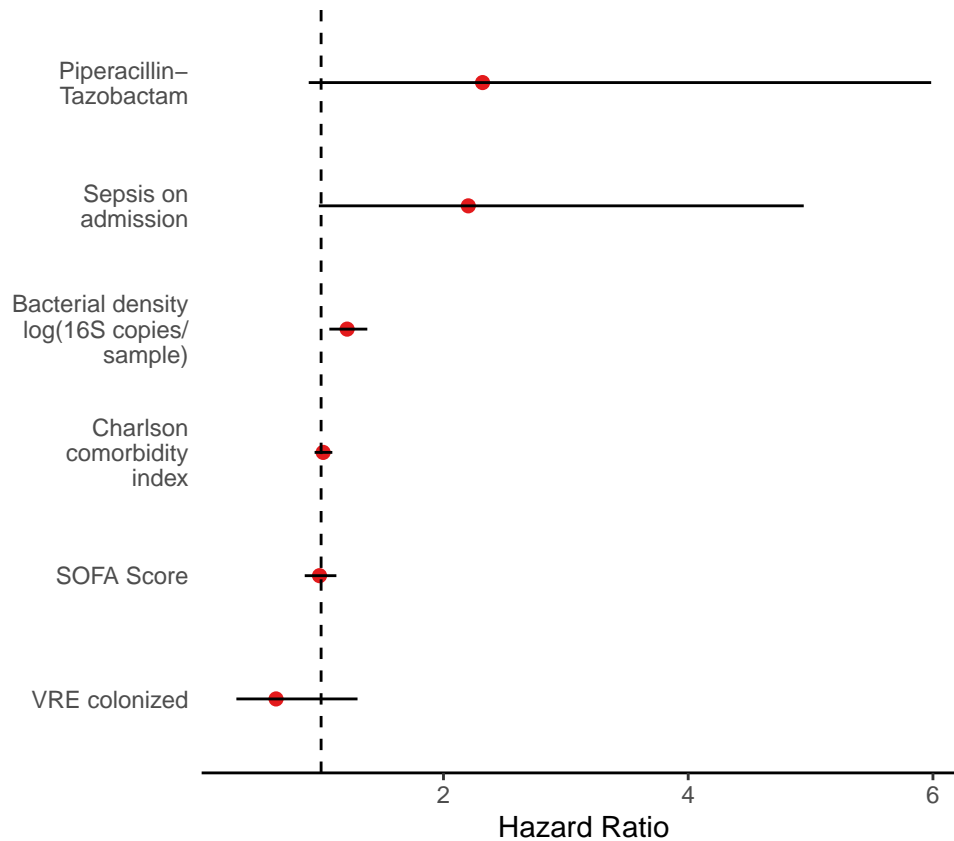
```

```

forest_plot<-inner_join(coef_survival, conf_int_survival, by ="variable")%>%
  mutate(variable = factor(variable),
         variable = fct_reorder(variable, coef))%>%
  mutate(variable = case_when(
    variable=="Zosyn"~"Piperacillin-\nTazobactam",
    variable=="sepsis"~"Sepsis on\nadmission",
    variable=="log(ddPCR_reads_per_sample)"~"Bacterial density\nlog(16S copies/\nsample)",
    variable=="Comorbidity_Score"~"Charlson\ncomorbidity\nindex",
    variable=="total_sofa"~"SOFA Score",
    variable=="case_or_controlcase"~"VRE colonized"
  ))%>%
  mutate(variable = factor(variable),
         variable = fct_reorder(variable, coef))%>%
  ggplot(aes(x=variable))+
  geom_point(aes(y=coef), color="#e31a1c", size=2)+
  geom_segment(aes(y=lower_ci, yend=upper_ci, x=variable, xend=variable))+
  theme_bw()+
  theme(panel.grid=element_blank(),
        panel.border = element_blank(),
        axis.ticks.y = element_blank(),
        axis.line.x = element_line(),
        aspect.ratio = 1)+
  coord_flip()+
  geom_hline(aes(yintercept=1), linetype="dashed")+
  labs(y="Hazard Ratio",
       x=NULL)

```

```
forest_plot
```



```
lmer_model<-lmerTest::lmer(log(ddPCR_reads_per_sample) ~
  (1|pair_ID)+total_sofa+Zosyn+
  case_or_control+age+cardio_pulm+
  +neuro+trauma+charlson_score+
  sepsis+transplant+gastro+
  malignancy,
  data = admit_dx%>%
  mutate(pair_ID=factor(pair_ID)))
```

```
## Registered S3 methods overwritten by 'lme4':
##   method                      from
##   cooks.distance.influence.merMod car
##   influence.merMod              car
##   dfbeta.influence.merMod       car
##   dfbetas.influence.merMod      car
```

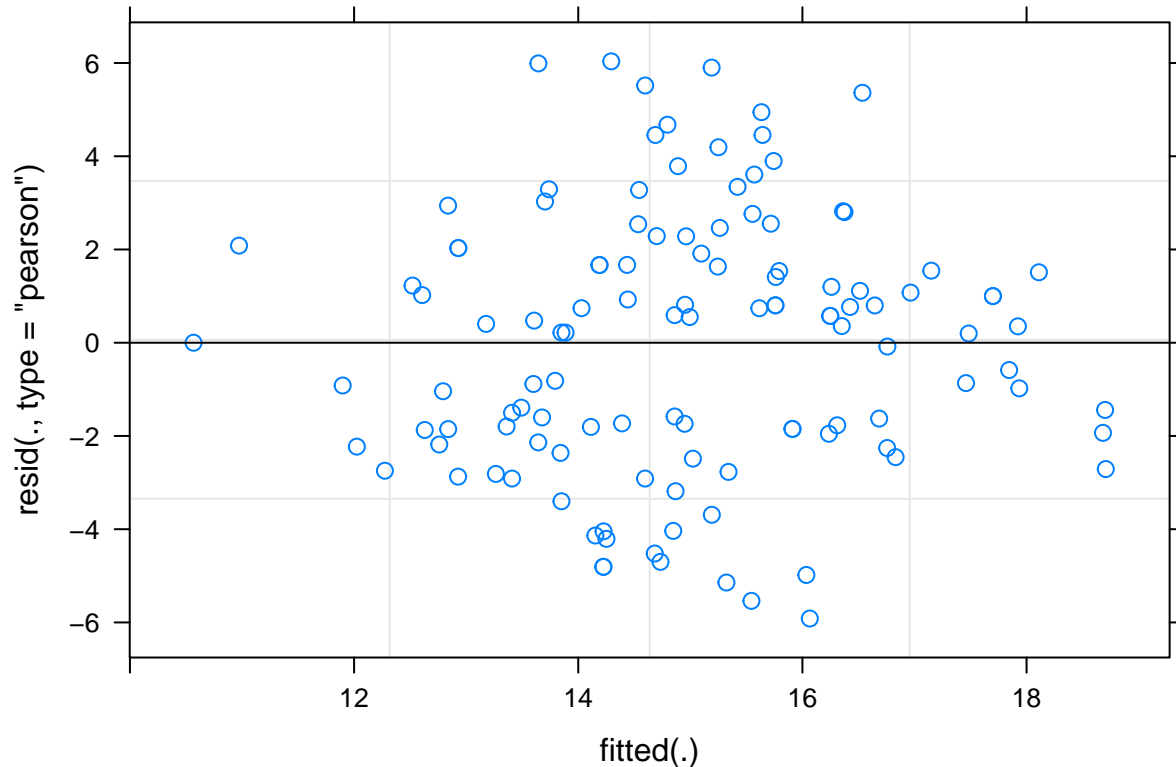
```
summary(lmer_model)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
## lmerModLmerTest]
## Formula: log(ddPCR_reads_per_sample) ~ (1 | pair_ID) + total_sofa + Zosyn +
##   case_or_control + age + cardio_pulm + +neuro + trauma + charlson_score +
##   sepsis + transplant + gastro + malignancy
## Data: admit_dx %>% mutate(pair_ID = factor(pair_ID))
```

```
##
## REML criterion at convergence: 582.1
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -1.9731 -0.6483  0.1191  0.5972  2.0130
##
## Random effects:
##   Groups   Name      Variance Std.Dev.
## pair_ID   (Intercept) 0.2783   0.5275
## Residual                8.9892   2.9982
## Number of obs: 119, groups: pair_ID, 59
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)    9.44783    3.33395 105.84727   2.834  0.00551 **
## total_sofa      0.07026    0.10712 101.21330   0.656  0.51339
## Zosyn          -1.84047    0.76022 102.08869  -2.421  0.01725 *
## case_or_controlcase -0.07260    0.59477  74.33851  -0.122  0.90318
## age             0.04270    0.02084 105.03598   2.049  0.04294 *
## cardio_pulm     0.67448    3.23880 105.74564   0.208  0.83543
## neuro           2.40806    3.68542 105.92980   0.653  0.51491
## trauma          1.35297    3.46025 105.79873   0.391  0.69658
## charlson_score  0.45054    0.15258  95.62554   2.953  0.00396 **
## sepsis          1.44459    3.21480 105.69914   0.449  0.65409
## transplant     -0.52552    3.62682 105.99196  -0.145  0.88507
## gastro          2.30090    3.29014 102.87953   0.699  0.48592
## malignancy      0.40270    3.19721 105.54270   0.126  0.90001
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

##
## Correlation matrix not shown by default, as p = 13 > 12.
## Use print(x, correlation=TRUE) or
##      vcov(x)          if you need it

plot(lmer_model)
```



```

coefficients<-summary(lmer_model)%>%
  coef()%>%
  data.frame()%>%
  rownames_to_column(var="variable")%>%
  rename(p_value="Pr...t..")

confidence_interval<-confint(lmer_model)%>%
  data.frame()%>%
  rownames_to_column(var="variable")%>%
  rename(lower_ci="X2.5..",
         upper_ci="X97.5..",
         )%>%
  mutate(range = (upper_ci-lower_ci)/2)

```

```
## Computing profile confidence intervals ...
```

```

mixed_effects<-inner_join(coefficients, confidence_interval, by = "variable")%>%
  mutate(variable=str_remove(variable, "reason_for_admit"),
         variable = str_replace(variable, "case_or_controlcase","vre_colonized"))%>%
  dplyr::select(variable, Estimate, p_value, lower_ci, upper_ci, range)%>%
  mutate(across(where(is.numeric), ~round(.,3)))%>%
  mutate(lower_ci = as.character(lower_ci),
         upper_ci = as.character(upper_ci))%>%
  mutate(left_parenth="(",

```



```

      right_parenth=")")%>%
unite(left_side, c("left_parenth", "lower_ci"), sep="")%>%
unite(right_side, c("upper_ci", "right_parenth"), sep="")%>%
unite(ninety_five_conf, c("left_side", "right_side"), sep="-")

mixed_effects

```

```

##      variable Estimate p_value ninety_five_conf range
## 1  (Intercept)    9.448   0.006   (3.241-15.69) 6.224
## 2    total_sofa    0.070   0.513  (-0.129-0.27) 0.200
## 3      Zosyn    -1.840   0.017  (-3.266--0.423) 1.421
## 4  vre_colonized  -0.073   0.903  (-1.183-1.056) 1.120
## 5        age     0.043   0.043   (0.004-0.083) 0.040
## 6   cardio_pulm   0.674   0.835  (-5.421-6.707) 6.064
## 7        neuro   2.408   0.515  (-4.534-9.273) 6.903
## 8      trauma    1.353   0.697  (-5.104-7.798) 6.451
## 9 charlson_score  0.451   0.004   (0.161-0.74) 0.290
##10      sepsis    1.445   0.654  (-4.582-7.428) 6.005
##11   transplant  -0.526   0.885  (-7.275-6.276) 6.776
##12      gastro    2.301   0.486  (-3.944-8.433) 6.188
##13   malignancy   0.403   0.900  (-5.607-6.357) 5.982

```

```

microbiome<-sofa_metadata%>%
  dplyr::select(Sample_ID, infection, culture_type,
                organism, Opiate_use:mld, above_threshold)%>%
  dplyr::select(-age)%>%
  mutate(across(where(is.numeric), ~if_else(>0, 1, 0)))%>%
  inner_join(all_data_swabs%>%
    dplyr::select(Sample_ID,
                  contains("Otu"),
                  ddPCR_reads_per_sample))%>%
  # dplyr::select(-Sample_ID)%>%
  mutate(contaminant = if_else(Otu0001>0, 1, 0),
         above_threshold= if_else(ddPCR_reads_per_sample>10^6, 1, 0))%>%
  inner_join(all_data_swabs%>%
    dplyr::select(pair_ID, Sample_ID))

```

```

## Joining, by = "Sample_ID"
## Joining, by = "Sample_ID"

```

```

micro.hel<-microbiome%>%
  dplyr::select(contains("Otu"))%>%
  decostand(., method="hellinger")

micro<-microbiome%>%
  dplyr::select(-contains("Otu"))

permanova<-adonis(micro.hel~infection, strata=micro$pair_ID,
                  data=micro, by="terms", permutations = 9999)

```

```

head<-coefficients(permanova)%>%
  as.data.frame()%>%
  rownames_to_column("variable")%>%
  filter(variable == "infection")%>%
  pivot_longer(cols=contains("Otu"),names_to="Otu",
               values_to = "importance")%>%
  arrange(importance)%>%
  inner_join(otu_genus_link)%>%
  head()

```

```
## Joining, by = "Otu"
```

```

tail<-coefficients(permanova)%>%
  as.data.frame()%>%
  rownames_to_column("variable")%>%
  filter(variable == "infection")%>%
  pivot_longer(cols=contains("Otu"),names_to="Otu",
               values_to = "importance")%>%
  arrange(importance)%>%
  inner_join(otu_genus_link)%>%
  tail()

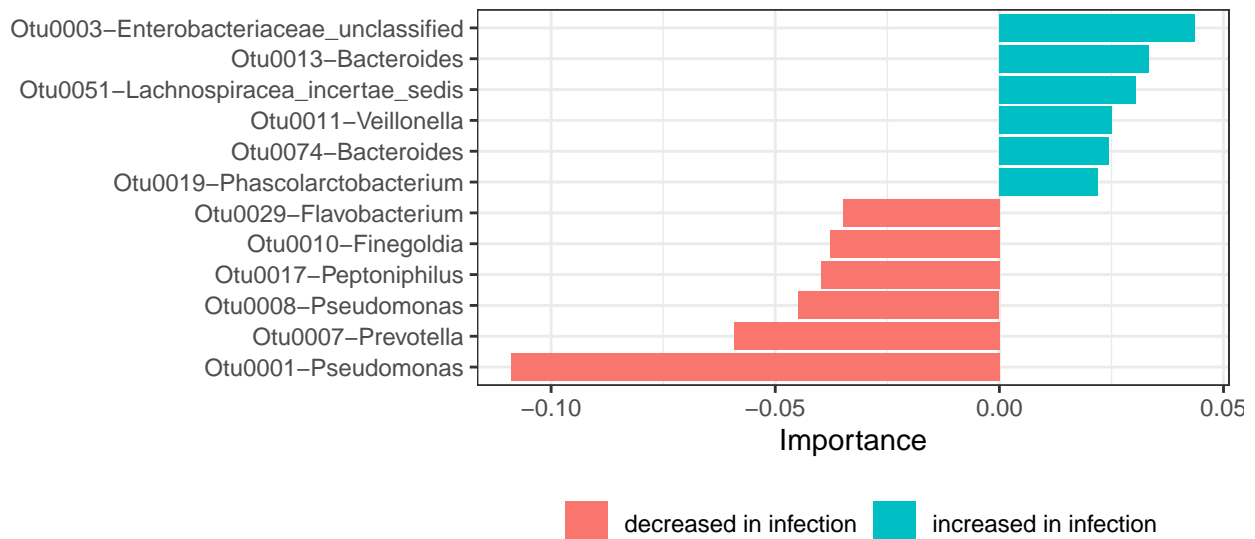
```

```
## Joining, by = "Otu"
```

```

rbind(head, tail)%>%
  mutate(protective_or_not=if_else(importance>0, "increased in infection",
                                   "decreased in infection"))%>%
  mutate(Otu_genus=factor(Otu_genus))%>%
  mutate(Otu_genus = fct_reorder(Otu_genus,importance))%>%
  ggplot(aes(x=Otu_genus, y = importance, fill=factor(protective_or_not)))+
  geom_col()+
  coord_flip()+
  theme_bw()+
  theme(aspect.ratio = .5,
        legend.position = "bottom")+
  labs(fill=NULL, x=NULL, y ="Importance")

```



```
infections_df<-all_data_swabs%>%
  dplyr::select(Sample_ID, swab_type,Otu0002,Otu0003)%>%
  inner_join(sofa_metadata)%>%
  mutate(
    staph = as.numeric(str_detect(organism, "Staph")),
    strep = as.numeric(str_detect(organism, "Strep")),
    pseudo = as.numeric(str_detect(organism, "Pseudo")),
    enterococcus = as.numeric(str_detect(organism, "Enterococcus")),
    klebsiella = as.numeric(str_detect(organism, "Klebsiella")),
    ecoli = as.numeric(str_detect(organism, "Escherichia")),
    none = as.numeric(str_detect(organism, "none")),
    enterobacter =as.numeric(str_detect(organism, "Enterobacter"))
  )%>%
  pivot_longer(cols=contains("Otu"), names_to = "Otu",
               values_to = "relative_abundance")%>%
  filter(Otu %in% c("Otu0002","Otu0003"))%>%
  group_by(Sample_ID, Otu)%>%
  summarize(ecoli = max(ecoli),
            klebsiella=max(klebsiella),
            enterobacter = max(enterobacter),
            abundance = sum(relative_abundance))%>%
  pivot_longer(
    cols=c(ecoli, klebsiella),
    names_to="organism",
    values_to="infection")
```

```
## Joining, by = "Sample_ID"
```

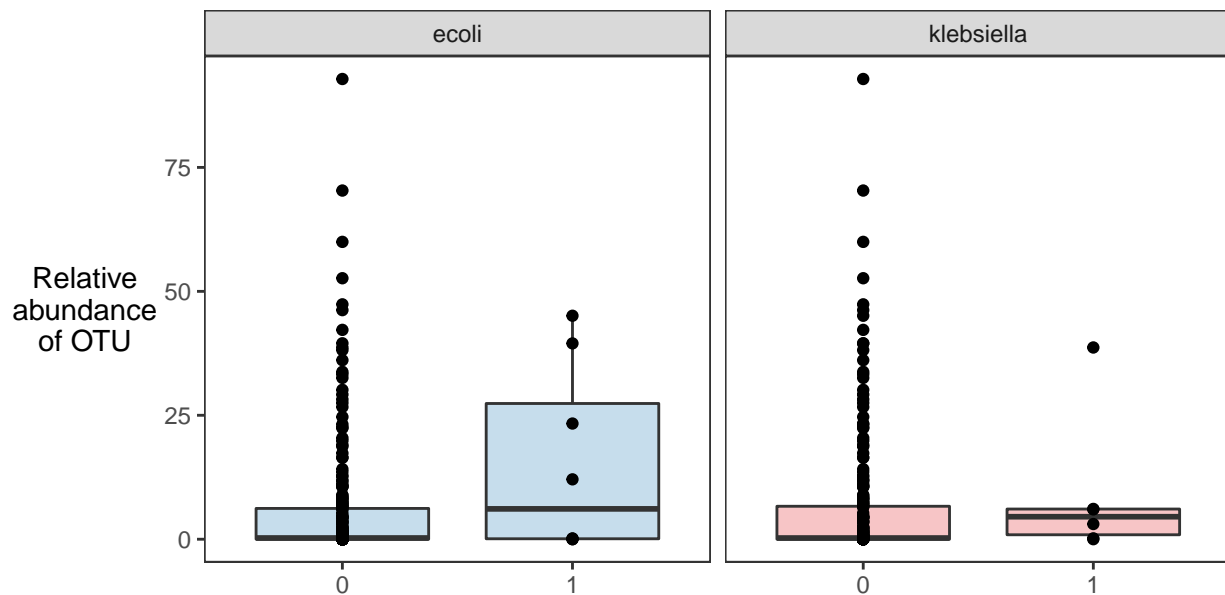
```
## `summarise()` has grouped output by 'Sample_ID'. You can override using the `.groups` argument.
```

```
sig<-infections_df%>%  
  mutate(infection = factor(infection))%>%  
  nest(-organism,-Otu)%>%  
  mutate(wilcox = map(data, ~wilcox.test(abundance~infection, data=.)%>%  
    tidy()))
```

```
## Warning: All elements of `...` must be named.
```

```
## Did you want `data = c(Sample_ID, enterobacter, abundance, infection)`?
```

```
infections_df%>%  
  ggplot(aes(x=factor(infection), y = abundance, fill=factor(organism)))+  
  geom_boxplot(alpha=0.25)+  
  geom_point()+  
  theme_bw()+  
  theme(legend.position = "none",  
        panel.grid = element_blank(),  
        axis.title.y=element_text(angle=0, vjust=0.5),  
        aspect.ratio = 1)+  
  facet_wrap(~organism)+  
  labs(y="Relative\nabundance\nof OTU",  
       x=NULL)+  
  scale_fill_manual(values=c("#1f78b4", "#e31a1c"))
```



```
ggsave("figure_7.pdf")
```

```
## Saving 6.5 x 4.5 in image
```