

# Genomic Modeling of Antimicrobial Resistance in Salmonella enterica: An Advanced Multi-Layer Perceptron Framework Applied to Chicken-Derived Isolates

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### Abstract :

This project presents an advanced framework for predicting antimicrobial resistance (AMR) in *Salmonella enterica* isolates from chicken sources using deep learning models. Starting with the collection and deduplication of *Salmonella* sequences, AMR gene identification was performed via the Resistance Gene Identifier (RGI) tool from the Comprehensive Antimicrobial Resistance Database. Data wrangling and t-SNE visualization enabled the integration and dimensional reduction of high-dimensional genomic data, facilitating better insight into AMR gene-antimicrobial relationships. The dataset was subsequently balanced using random over-sampling (ROSE) and filtered to exclude low-variance features. Finally, a Multi-Layer Perceptron (MLP) model was trained to predict resistance across 11 distinct drug classes, achieving an impressive accuracy of 86.5%. This model offers a promising approach to understanding and anticipating AMR patterns in poultry-related pathogens.

### Step 01 - Data Collection

Installing necessary library called BIOPYTHON for handling biological data has been Installed

```
In [1]: !pip install biopython

Collecting biopython
  Downloading biopython-1.84-cp312-cp312-win_amd64.whl.metadata (13 kB)
Requirement already satisfied: numpy in c:\users\dd\anaconda3\lib\site-packages (from biopython) (1.26.4)
Downloading biopython-1.84-cp312-cp312-win_amd64.whl (2.8 MB)
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Installing collected packages: biopython
Successfully installed biopython-1.84
```

Once done with the installation of BioPython, the complete genome sequences of chicken with salmonella enterica has been searched in NCBI nucleotide database and a total of 496 sequence has been found.

```
In [1]: from Bio import Entrez

# Set your email (required by NCBI)
Entrez.email = "rd97.dd@gmail.com"

# Define search query for Salmonella enterica with isolation source chicken
search_term = "Salmonella enterica[Organism] AND chicken[Isolation Source] AND complete genome[Title]"

# Perform search in the NCBI nucleotide database
handle = Entrez.esearch(db="nucleotide", term=search_term, retmax=0) # retmax=0 to only get the count
record = Entrez.read(handle)
handle.close()

# Get and print the total number of records (sequences) found
total_records = int(record["Count"])
print(f"Total sequences found: {total_records}")
```

Total sequences found: 496

A sample of 10 isolate files has been downloaded for verification purpose.

```
In [12]: from Bio import Entrez, SeqIO
import time
import os

# Set your email (required by NCBI)
Entrez.email = "rd97.dd@gmail.com"

# Define broader search queries
search_term = "Salmonella enterica[Organism] AND chicken[Isolation Source] AND complete genome[Title]"

# Define the download directory
output_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
os.makedirs(output_directory, exist_ok=True)

# Function to download a specific number of sequences
def download_sequences(search_term, output_directory, num_sequences=10):
    # Search the NCBI nucleotide database
    handle = Entrez.esearch(db="nucleotide", term=search_term, retmax=num_sequences)
    record = Entrez.read(handle)
    handle.close()

    total_records = int(record["Count"])
    print(f"Total sequences found: {total_records}")

    id_list = record["IdList"]

    if not id_list:
        print("No IDs found, stopping download.")
        return

    for seq_id in id_list:
        try:
            # Fetch the metadata
            handle = Entrez.efetch(db="nucleotide", id=seq_id, rettype="gb", retmode="text")
            metadata = handle.read()
            handle.close()

            # Save metadata
            metadata_file = os.path.join(output_directory, f"{seq_id}_metadata.gb")
            with open(metadata_file, "w") as metadata_handle:
                metadata_handle.write(metadata)

            # Fetch the FASTA sequence
            handle = Entrez.efetch(db="nucleotide", id=seq_id, rettype="fasta", retmode="text")
            seq_record = SeqIO.read(handle, "fasta")
            handle.close()

            # Check if the sequence is valid
            if not seq_record.seq or not len(seq_record.seq):
                print(f"Warning: Sequence {seq_id} is empty or undefined.")
                continue

            # Save the FASTA sequence
            fasta_file = os.path.join(output_directory, f"{seq_id}.fasta")
            with open(fasta_file, "w") as out_handle:
                SeqIO.write(seq_record, out_handle, "fasta")

            print(f"Downloaded and saved {fasta_file}")

        except Exception as e:
            print(f"Error processing {seq_id}: {e}")

    print("Download complete!")

# Download the first 10 sequences
download_sequences(search_term, output_directory, num_sequences=10)
```

Total sequences found: 496

Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791964398.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791964390.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791964384.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791964378.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791964373.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791959104.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791959093.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791348005.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791334406.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2791334263.fasta  
Download complete!

In next step, duplicates has been checked based of file names.

```
In [14]: from Bio import SeqIO
import os

def check_duplicates(fasta_directory):
    seen_sequences = set()
    duplicates = []

    for filename in os.listdir(fasta_directory):
        if filename.endswith(".fasta"):
            filepath = os.path.join(fasta_directory, filename)
            for record in SeqIO.parse(filepath, "fasta"):
                sequence = str(record.seq)
                if sequence in seen_sequences:
                    duplicates.append(filename)
                else:
                    seen_sequences.add(sequence)
```

```

if duplicates:
    print(f"Duplicate files found: {set(duplicates)}")
else:
    print("No duplicate sequences found.")

# Set the path to your FASTA files directory
fasta_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
check_duplicates(fasta_directory)

```

No duplicate sequences found.

Here, its been checked for duplicates with strain ids from metadata

```

In [3]: from Bio import Entrez
import os
import re

def extract_strain_info(metadata_directory):
    strain_info = {}

    for filename in os.listdir(metadata_directory):
        if filename.endswith("_metadata.gb"):
            filepath = os.path.join(metadata_directory, filename)
            with open(filepath, "r") as file:
                content = file.read()
                match = re.search(r"strain=(\S+)", content)
                if match:
                    strain = match.group(1)
                    if strain in strain_info:
                        strain_info[strain].append(filepath)
                    else:
                        strain_info[strain] = [filepath]

    # Print out strains and their corresponding files
    for strain, files in strain_info.items():
        if len(files) > 1:
            print(f"Strain {strain} found in files: {files}")

# Set the path to your metadata directory
metadata_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
extract_strain_info(metadata_directory)

```

Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded  
Unique strains only founded

Now, we have downloaded 496 sequences o along with meta data.

```

In [ ]: from Bio import Entrez, SeqIO
import time
import os

# Set your email (required by NCBI)
Entrez.email = "rd97.dd@gmail.com"

# Define broader search queries
search_term = "Salmonella enterica[Organism] AND chicken[Isolation Source] AND complete genome[Title]"

# Define the download directory
output_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
os.makedirs(output_directory, exist_ok=True)

# Function to download a specific number of sequences
def download_sequences(search_term, output_directory, num_sequences=496):
    # Search the NCBI nucleotide database
    handle = Entrez.esearch(db="nucleotide", term=search_term, retmax=num_sequences)
    record = Entrez.read(handle)
    handle.close()

    total_records = int(record["Count"])
    print(f"Total sequences found: {total_records}")

    id_list = record["IdList"]

    if not id_list:
        print("No IDs found, stopping download.")
        return

    for seq_id in id_list:
        try:
            # Fetch the metadata
            handle = Entrez.efetch(db="nucleotide", id=seq_id, rettype="gb", retmode="text")
            metadata = handle.read()
            handle.close()

            # Save metadata
            metadata_file = os.path.join(output_directory, f"{seq_id}_metadata.gb")
            with open(metadata_file, "w") as metadata_handle:
                metadata_handle.write(metadata)

            # Fetch the FASTA sequence
            handle = Entrez.efetch(db="nucleotide", id=seq_id, rettype="fasta", retmode="text")
            seq_record = SeqIO.read(handle, "fasta")
            handle.close()

            # Check if the sequence is valid
            if not seq_record.seq or not len(seq_record.seq):
                print(f"Warning: Sequence {seq_id} is empty or undefined.")
                continue

            # Save the FASTA sequence
            fasta_file = os.path.join(output_directory, f"{seq_id}.fasta")
            with open(fasta_file, "w") as out_handle:
                SeqIO.write(seq_record, out_handle, "fasta")

            print(f"Downloaded and saved {fasta_file}")

        except Exception as e:
            print(f"Error processing {seq_id}: {e}")

    print("Download complete!")

# Download the first 10 sequences
download_sequences(search_term, output_directory, num_sequences=496)

```

Total sequences found: 496

[illegible]

Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2484128728.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2482050684.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2476413604.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2475999171.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2459865826.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2459861217.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2459856665.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2459518531.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences\2333272181.fasta

```
In [1]: from Bio import Entrez, SeqIO
import os

# Set your email (required by NCBI)
Entrez.email = "rd97.dd@gmail.com"

# Define broader search queries
search_term = "Salmonella enterica[Organism] AND chicken[Isolation Source] AND complete genome[Title]"

# Define the download directory
output_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
os.makedirs(output_directory, exist_ok=True)

# Function to download a specific number of sequences
def download_sequences(search_term, output_directory, num_sequences=496):
    # Search the NCBI nucleotide database
    handle = Entrez.esearch(db="nucleotide", term=search_term, retmax=num_sequences)
    record = Entrez.read(handle)
    handle.close()

    total_records = int(record["Count"])
    print(f"Total sequences found: {total_records}")

    id_list = record["IdList"]

    # Track already downloaded sequences by checking existing files
    downloaded_ids = set([file.split('_')[0] for file in os.listdir(output_directory) if file.endswith('.fasta')])

    print(f"Already downloaded {len(downloaded_ids)} sequences, continuing from where it left off.")

    for seq_id in id_list:
        if seq_id in downloaded_ids:
            print(f"Skipping {seq_id}, already downloaded.")
            continue

        try:
            # Fetch the metadata
            handle = Entrez.efetch(db="nucleotide", id=seq_id, rettype="gb", retmode="text")
            metadata = handle.read()
            handle.close()

            # Save metadata
            metadata_file = os.path.join(output_directory, f"{seq_id}_metadata.gb")
            with open(metadata_file, "w") as metadata_handle:
                metadata_handle.write(metadata)

            # Fetch the FASTA sequence
            handle = Entrez.efetch(db="nucleotide", id=seq_id, rettype="fasta", retmode="text")
            seq_record = SeqIO.read(handle, "fasta")
            handle.close()

            # Check if the sequence is valid
            if not seq_record.seq or not len(seq_record.seq):
                print(f"Warning: Sequence {seq_id} is empty or undefined.")
                continue

            # Save the FASTA sequence
            fasta_file = os.path.join(output_directory, f"{seq_id}.fasta")
            with open(fasta_file, "w") as out_handle:
                SeqIO.write(seq_record, out_handle, "fasta")

            print(f"Downloaded and saved {fasta_file}")

        except Exception as e:
            print(f"Error processing {seq_id}: {e}")

    print("Download complete!")

# Resume downloading the remaining sequences
download_sequences(search_term, output_directory, num_sequences=496)
```

Total sequences found: 496

[illegible]



[illegible]

[illegible]

Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/605528405.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/605520020.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/605511148.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/605498156.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/601101465.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/563346448.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/548713695.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/529190224.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/332986951.fasta  
Downloaded and saved C:/Users/DD/Documents/Python Scripts/sequences/523806722.fasta  
Download complete!

Checked for duplicates based on file names after downloading the sequences

```
In [3]: from Bio import SeqIO
import os

def check_duplicates(fasta_directory):
    seen_sequences = set()
    duplicates = []

    for filename in os.listdir(fasta_directory):
        if filename.endswith(".fasta"):
            filepath = os.path.join(fasta_directory, filename)
            for record in SeqIO.parse(filepath, "fasta"):
                sequence = str(record.seq)
                if sequence in seen_sequences:
                    duplicates.append(filename)
                else:
                    seen_sequences.add(sequence)

    if duplicates:
        print(f"Duplicate files found: {set(duplicates)}")
    else:
        print("No duplicate sequences found.")

# Set the path to your FASTA files directory
fasta_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
check_duplicates(fasta_directory)
```

Duplicate files found: {'2791333611.fasta', '2791327646.fasta', '2711458674.fasta', '1128998525.fasta', '749298461.fasta', '2791328715.fasta', '2321041129.fasta', '2791327658.fasta', '1046220323.fasta', '2482324433.fasta', '1863657517.fasta', '2711462317.fasta', '1033136052.fasta', '998623498.fasta', '1864594723.fasta', '2791328192.fasta', '1864593957.fasta', '2321008513.fasta', '2791332598.fasta', '995922935.fasta', '2711462769.fasta', '2321043499.fasta', '2339477813.fasta', '2036914008.fasta', '2078699049.fasta', '2638491622.fasta', '2082554870.fasta', '1995427272.fasta', '2521002406.fasta', '2100237189.fasta', '2784851814.fasta', '2717387261.fasta', '1863661087.fasta', '2550213792.fasta', '2791328750.fasta', '1634456755.fasta', '983377086.fasta', '2711462605.fasta', '2791327776.fasta', '1982652612.fasta', '2711475867.fasta', '2638491693.fasta', '2791334100.fasta', '2638510809.fasta', '2791328742.fasta', '2791327275.fasta', '2791959104.fasta', '2321043586.fasta', '998647728.fasta', '2279079344.fasta', '2791328684.fasta', '1828945320.fasta', '2711466246.fasta', '2288537321.fasta', '2087863205.fasta', '2550225976.fasta', '2550213748.fasta', '2638491583.fasta', '2711469117.fasta', '2711475272.fasta', '2060646254.fasta', '674230814.fasta', '2791348005.fasta', '2710806370.fasta', '2258710042.fasta', '2265635730.fasta', '2476810552.fasta', '2791334406.fasta', '995907166.fasta', '2791964378.fasta', '1736666103.fasta', '1130930627.fasta', '749298420.fasta', '2100424222.fasta', '998642845.fasta', '2791328489.fasta', '2477330759.fasta', '2168004414.fasta', '2515152310.fasta', '2710805975.fasta', '2791964398.fasta', '2321014208.fasta', '2294922788.fasta', '1635401364.fasta', '2791327413.fasta', '2791328717.fasta', '2321042278.fasta', '2791327513.fasta', '2791959093.fasta', '2791326870.fasta', '1221158448.fasta', '2100896152.fasta', '995907817.fasta', '2550213647.fasta', '2279083905.fasta', '2711464918.fasta', '2791325747.fasta', '1951777374.fasta', '2550287957.fasta', '1829031305.fasta', '2711460722.fasta', '2100356487.fasta', '2791323978.fasta', '1546887966.fasta', '1531933061.fasta', '2273089235.fasta', '2711461464.fasta', '749297197.fasta', '2711488657.fasta', '1829022561.fasta', '1733272414.fasta', '2791327656.fasta', '2791329900.fasta', '2711459283.fasta', '2791327653.fasta', '2601133276.fasta', '2288538671.fasta', '2485141349.fasta', '915846032.fasta', '1046302332.fasta', '1196285509.fasta', '2100166053.fasta', '1797869977.fasta', '2791332929.fasta', '995923317.fasta', '2288530062.fasta', '2791328843.fasta', '1945020687.fasta', '2791964390.fasta', '1863662535.fasta', '2791964373.fasta', '2258485671.fasta', '2572860206.fasta', '2791334217.fasta', '2339534824.fasta', '2550255387.fasta', '2791326641.fasta', '749296449.fasta', '2550213764.fasta', '2740278012.fasta', '2711495709.fasta', '2791334263.fasta', '1736663574.fasta', '2288890305.fasta', '749314944.fasta', '2711458425.fasta', '1863660810.fasta', '771524686.fasta', '2791964384.fasta', '2321031274.fasta'}

In this step, we have checked for duplicate sequences based on strains

```
In [5]: from Bio import Entrez
import os
import re

def extract_strain_info(metadata_directory):
    strain_info = {}

    for filename in os.listdir(metadata_directory):
        if filename.endswith(".metadata.gb"):
            filepath = os.path.join(metadata_directory, filename)
            with open(filepath, "r") as file:
                content = file.read()
                match = re.search(r"strain=(\\S+)", content) # Adjust regex based on metadata format
                if match:
                    strain = match.group(1)
                    if strain in strain_info:
                        strain_info[strain].append(filename)
                    else:
                        strain_info[strain] = [filename]

    # Print out strains and their corresponding files
    for strain, files in strain_info.items():
        if len(files) > 1:
            print(f"Strain {strain} found in files: {files}")

# Set the path to your metadata directory
metadata_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
extract_strain_info(metadata_directory)
```

"SA20095440" found in files: ['1000369143\_metadata.gb', '998647728\_metadata.gb']  
Strain "SA19960848" found in files: ['1001551848\_metadata.gb', '995907166\_metadata.gb']  
Strain "SA19970769" found in files: ['1001552415\_metadata.gb', '995922935\_metadata.gb']  
Strain "SA19970510" found in files: ['1001554681\_metadata.gb', '995907817\_metadata.gb']  
Strain "SA19980677" found in files: ['1001555507\_metadata.gb', '995923317\_metadata.gb']  
Strain "SA20094352" found in files: ['1002985512\_metadata.gb', '998642845\_metadata.gb']  
Strain "CG29" found in files: ['1030103469\_metadata.gb', '1033136052\_metadata.gb']  
Strain "SA020709004001" found in files: ['1045707385\_metadata.gb', '1046220323\_metadata.gb']  
Strain "SA01AB09084001" found in files: ['1045716295\_metadata.gb', '1046302332\_metadata.gb']  
Strain "NS5391" found in files: ['1127388936\_metadata.gb', '1128998525\_metadata.gb']  
Strain "87" found in files: ['1129908976\_metadata.gb', '1130930627\_metadata.gb']  
Strain "FORC\_038" found in files: ['1190123157\_metadata.gb', '1196285509\_metadata.gb']  
Strain "D90" found in files: ['1219800280\_metadata.gb', '1221158448\_metadata.gb']  
Strain "ATCC" found in files: ['1528951495\_metadata.gb', '1531933061\_metadata.gb', '605520020\_metadata.gb', '605528405\_metadata.gb', '674281104\_metadata.gb', '820762957\_metadata.gb', '914828066\_metadata.gb', '914835508\_metadata.gb', '914845112\_metadata.gb']  
Strain "S" found in files: ['1543688490\_metadata.gb', '1546887966\_metadata.gb']  
Strain "JT" found in files: ['1633748201\_metadata.gb', '1634456755\_metadata.gb']  
Strain "CD-SL01" found in files: ['1634015463\_metadata.gb', '1635401364\_metadata.gb']  
Strain "SJTUF87912v2" found in files: ['1695497099\_metadata.gb', '1736663574\_metadata.gb']  
Strain "SJTUF13520v2" found in files: ['1695513588\_metadata.gb', '1736666103\_metadata.gb']  
Strain "SL-312" found in files: ['1732125948\_metadata.gb', '1733272414\_metadata.gb']  
Strain "SCSM4.1" found in files: ['1788239996\_metadata.gb', '1797869977\_metadata.gb']  
Strain "SE124" found in files: ['1828351404\_metadata.gb', '1828945320\_metadata.gb']  
Strain "SI85" found in files: ['1828458697\_metadata.gb', '1829031305\_metadata.gb']  
Strain "SI67" found in files: ['1828463287\_metadata.gb', '1829022561\_metadata.gb']  
Strain "CVM" found in files: ['1836411705\_metadata.gb', '1842989566\_metadata.gb', '1843494016\_metadata.gb', '1843498400\_metadata.gb', '1843507307\_metadata.gb', '1843512211\_metadata.gb', '1843516861\_metadata.gb', '1843531062\_metadata.gb', '1843540331\_metadata.gb', '1843544971\_metadata.gb', '1843576148\_metadata.gb', '1843580775\_metadata.gb', '1843585404\_metadata.gb', '1843589994\_metadata.gb', '1843599223\_metadata.gb', '1843608483\_metadata.gb', '1843613175\_metadata.gb', '1843618362\_metadata.gb', '1843629339\_metadata.gb', '1843633944\_metadata.gb', '1843643169\_metadata.gb', '1843647868\_metadata.gb', '1843652501\_metadata.gb', '1846455821\_metadata.gb', '1951535343\_metadata.gb', '1951777374\_metadata.gb', '2090918790\_metadata.gb', '2090956231\_metadata.gb', '2090974716\_metadata.gb', '2091165092\_metadata.gb', '2091169855\_metadata.gb', '2091174439\_metadata.gb', '2091178941\_metadata.gb', '2091183720\_metadata.gb', '2091229717\_metadata.gb', '2091239126\_metadata.gb', '2091243740\_metadata.gb', '2091253165\_metadata.gb', '2091264036\_metadata.gb', '2091277794\_metadata.gb', '2091323346\_metadata.gb', '2091333084\_metadata.gb', '2091346897\_metadata.gb', '2091356744\_metadata.gb', '2091370368\_metadata.gb', '2091375002\_metadata.gb', '2091402345\_metadata.gb', '2091418357\_metadata.gb', '2091422800\_metadata.gb', '2091431810\_metadata.gb', '2091446076\_metadata.gb', '2091455043\_metadata.gb', '2091469148\_metadata.gb', '2091473540\_metadata.gb', '2091478163\_metadata.gb', '2091482867\_metadata.gb', '2136510056\_metadata.gb', '2518923471\_metadata.gb', '2521002406\_metadata.gb', '696581462\_metadata.gb', '696593549\_metadata.gb', '744788240\_metadata.gb', '767833982\_metadata.gb', '767838360\_metadata.gb', '767842734\_metadata.gb']  
Strain "VNSEC031" found in files: ['1863533289\_metadata.gb', '1863660810\_metadata.gb']  
Strain "VNSEC023" found in files: ['1863537930\_metadata.gb', '1863661087\_metadata.gb']  
Strain "VNSEC013" found in files: ['1863542555\_metadata.gb', '1863657517\_metadata.gb']  
Strain "VNSEC003" found in files: ['1863547333\_metadata.gb', '1863662535\_metadata.gb']  
Strain "VNSEC002" found in files: ['1863552396\_metadata.gb', '1864594723\_metadata.gb']  
Strain "VNSEC001" found in files: ['1863557100\_metadata.gb', '1864593957\_metadata.gb']  
Strain "R16.1424" found in files: ['1943036513\_metadata.gb', '1945020687\_metadata.gb']  
Strain "GSJ/2016-Sa1-017" found in files: ['1979358710\_metadata.gb', '1982652612\_metadata.gb']  
Strain "R16.0556" found in files: ['1993496820\_metadata.gb', '1995427272\_metadata.gb']  
Strain "KUFSE-SAL0043" found in files: ['2034574262\_metadata.gb', '2036914008\_metadata.gb']  
Strain "CFSAN008081" found in files: ['2049354170\_metadata.gb', '2082554870\_metadata.gb']  
Strain "Y220MC516" found in files: ['2060222687\_metadata.gb', '2060646254\_metadata.gb']  
Strain "SA18578" found in files: ['2077453074\_metadata.gb', '2078699049\_metadata.gb']  
Strain "S16" found in files: ['2087468524\_metadata.gb', '2087863205\_metadata.gb']  
Strain "ZC-S1" found in files: ['2093998598\_metadata.gb', '2100166053\_metadata.gb']  
Strain "S90" found in files: ['2094623483\_metadata.gb', '2100356487\_metadata.gb']  
Strain "SP" found in files: ['2094623485\_metadata.gb', '2100424222\_metadata.gb']  
Strain "S146" found in files: ['2094642037\_metadata.gb', '2100237189\_metadata.gb']  
Strain "07Q015" found in files: ['2095951993\_metadata.gb', '2100896152\_metadata.gb']  
Strain "Y221MC54" found in files: ['2167845084\_metadata.gb', '2168004414\_metadata.gb']  
Strain "Y220MC56" found in files: ['2258055768\_metadata.gb', '2258485671\_metadata.gb']  
Strain "Y220MC514" found in files: ['2258066866\_metadata.gb', '2258710042\_metadata.gb']  
Strain "SE006" found in files: ['2263926704\_metadata.gb', '2265635730\_metadata.gb']  
Strain "0134" found in files: ['2271345254\_metadata.gb', '2273089235\_metadata.gb']  
Strain "KCID6" found in files: ['2277353412\_metadata.gb', '2279079344\_metadata.gb']  
Strain "BCID6" found in files: ['2277362348\_metadata.gb', '2279083905\_metadata.gb']  
Strain "AH19MC58" found in files: ['2288173494\_metadata.gb', '2288530062\_metadata.gb']  
Strain "Z214C1328" found in files: ['2288320740\_metadata.gb', '2288890305\_metadata.gb']  
Strain "MRS17\_00712" found in files: ['2294456242\_metadata.gb', '2294922788\_metadata.gb']  
Strain "190821\_1" found in files: ['2320729955\_metadata.gb', '2321014208\_metadata.gb']  
Strain "190819\_2" found in files: ['2320734399\_metadata.gb', '2321042278\_metadata.gb']  
Strain "190807\_1" found in files: ['2320738824\_metadata.gb', '2321031274\_metadata.gb']  
Strain "190729\_8" found in files: ['2320748473\_metadata.gb', '2321043499\_metadata.gb']  
Strain "190729\_4" found in files: ['2320752890\_metadata.gb', '2321043586\_metadata.gb']  
Strain "190704\_2" found in files: ['2320757305\_metadata.gb', '2321008513\_metadata.gb']  
Strain "190610\_1" found in files: ['2320761748\_metadata.gb', '2321041129\_metadata.gb']  
Strain "XM3104" found in files: ['2333213671\_metadata.gb', '2339534824\_metadata.gb']  
Strain "CVCC" found in files: ['2333272181\_metadata.gb', '2339477813\_metadata.gb']  
Strain "R22.0044" found in files: ['2475999171\_metadata.gb', '2476810552\_metadata.gb']  
Strain "013" found in files: ['2476413604\_metadata.gb', '2477330759\_metadata.gb']  
Strain "S1467" found in files: ['2482050684\_metadata.gb', '2482324433\_metadata.gb']  
Strain "CRIN508879" found in files: ['2484128728\_metadata.gb', '2485141349\_metadata.gb']  
Strain "CRSE-01" found in files: ['2514088622\_metadata.gb', '2515152310\_metadata.gb']  
Strain "SECMV-15" found in files: ['2548245062\_metadata.gb', '2550287957\_metadata.gb']  
Strain "SECMV-17" found in files: ['2548250070\_metadata.gb', '2550213647\_metadata.gb']  
Strain "SE" found in files: ['2548260632\_metadata.gb', '2550213764\_metadata.gb']  
Strain "SECMV-13" found in files: ['2548275539\_metadata.gb', '2550225976\_metadata.gb']  
Strain "SECMV-10" found in files: ['2548291250\_metadata.gb', '2550255387\_metadata.gb']  
Strain "SECMV-14" found in files: ['2548296188\_metadata.gb', '2550213748\_metadata.gb']  
Strain "SECMV-5" found in files: ['2548306062\_metadata.gb', '2550213792\_metadata.gb']  
Strain "Z1323CSL0015" found in files: ['2570622740\_metadata.gb', '2572860206\_metadata.gb', '2703404023\_metadata.gb', '2711459283\_metadata.gb']  
Strain "LAS" found in files: ['2595857884\_metadata.gb', '2601133276\_metadata.gb']  
Strain "405987R1\_550" found in files: ['2636846570\_metadata.gb', '2638491583\_metadata.gb']  
Strain "401964R1\_549" found in files: ['2636851041\_metadata.gb', '2638491693\_metadata.gb']  
Strain "180121R1S\_548" found in files: ['2636855593\_metadata.gb', '2638491622\_metadata.gb']  
Strain "SSSE-01" found in files: ['2679318990\_metadata.gb', '2710805975\_metadata.gb']  
Strain "SSSE-03" found in files: ['2679323484\_metadata.gb', '2710806370\_metadata.gb']  
Strain "Z1323CSL0016" found in files: ['2703413299\_metadata.gb', '2711458674\_metadata.gb']  
Strain "Z1323CSL0014" found in files: ['2703417940\_metadata.gb', '2711458425\_metadata.gb']  
Strain "Z1323CSL0027" found in files: ['2703422591\_metadata.gb', '2711461464\_metadata.gb']  
Strain "Z1323CSL0034" found in files: ['2703436556\_metadata.gb', '2711469117\_metadata.gb']  
Strain "Z1323CSL0017" found in files: ['2703441194\_metadata.gb', '2711464918\_metadata.gb']  
Strain "Z1323CSL0002" found in files: ['2703445817\_metadata.gb', '2711466246\_metadata.gb']  
Strain "Z1323CSL0006" found in files: ['2703450442\_metadata.gb', '2711460722\_metadata.gb']  
Strain "Z1323CSL0062" found in files: ['2704865509\_metadata.gb', '2711462317\_metadata.gb']  
Strain "Z1323CSL0057" found in files: ['2704874751\_metadata.gb', '2711475867\_metadata.gb']  
Strain "Z1323CSL0053" found in files: ['2704891984\_metadata.gb', '2711495709\_metadata.gb']  
Strain "Z1323CSL0045" found in files: ['2704905772\_metadata.gb', '2711488657\_metadata.gb']  
Strain "Z1323CSL0047" found in files: ['2704910405\_metadata.gb', '2711475272\_metadata.gb']  
Strain "Z1323CSL0052" found in files: ['2704915029\_metadata.gb', '2711462605\_metadata.gb']  
Strain "Z1323CSL0051" found in files: ['2704924300\_metadata.gb', '2711462769\_metadata.gb']  
Strain "ST.G536" found in files: ['2717024100\_metadata.gb', '2717387261\_metadata.gb']  
Strain "IJC55-22" found in files: ['2739185197\_metadata.gb', '2740278012\_metadata.gb']  
Strain "H4" found in files: ['2784324973\_metadata.gb', '2784851814\_metadata.gb']  
Strain "Z01320SL0044" found in files: ['2790142919\_metadata.gb', '2791959093\_metadata.gb']  
Strain "Z01320SL0045" found in files: ['2790153796\_metadata.gb', '2791964378\_metadata.gb']  
Strain "Z01320SL0042" found in files: ['2790167917\_metadata.gb', '2791959104\_metadata.gb']  
Strain "Z01320SL0040" found in files: ['2790172290\_metadata.gb', '2791964398\_metadata.gb']  
Strain "Z01320SL0041" found in files: ['2790176664\_metadata.gb', '2791964390\_metadata.gb']  
Strain "Z01320SL0038" found in files: ['2790181112\_metadata.gb', '2791964373\_metadata.gb']  
Strain "Z01320SL0043" found in files: ['2790191447\_metadata.gb', '2791964384\_metadata.gb']  
Strain "Z01320SL0030" found in files: ['2790219725\_metadata.gb', '2791325747\_metadata.gb']  
Strain "Z01320SL0033" found in files: ['2790226660\_metadata.gb', '2791348005\_metadata.gb']  
Strain "Z01320SL0034" found in files: ['2790232908\_metadata.gb', '2791323978\_metadata.gb']  
Strain "Z01320SL0031" found in files: ['2790238422\_metadata.gb', '2791328742\_metadata.gb']  
Strain "Z01320SL0036" found in files: ['2790243305\_metadata.gb', '2791327776\_metadata.gb']  
Strain "Z01320SL0037" found in files: ['2790247716\_metadata.gb', '2791329900\_metadata.gb']

Strain "Z01320SL0035" found in files: ['2790252121\_metadata.gb', '2791327646\_metadata.gb']  
 Strain "Z01320SL0032" found in files: ['2790256505\_metadata.gb', '2791332598\_metadata.gb']  
 Strain "Z01319SL0021" found in files: ['2790261062\_metadata.gb', '2791326641\_metadata.gb']  
 Strain "Z01319SL0015" found in files: ['2790265499\_metadata.gb', '2791328717\_metadata.gb']  
 Strain "Z01319SL0020" found in files: ['2790270044\_metadata.gb', '2791327275\_metadata.gb']  
 Strain "Z01320SL0029" found in files: ['2790274631\_metadata.gb', '2791332929\_metadata.gb']  
 Strain "Z01319SL0016" found in files: ['2790279127\_metadata.gb', '2791327653\_metadata.gb']  
 Strain "Z01319SL0017" found in files: ['2790283963\_metadata.gb', '2791328843\_metadata.gb']  
 Strain "Z01319SL0022" found in files: ['2790292938\_metadata.gb', '2791327656\_metadata.gb']  
 Strain "Z01320SL0026" found in files: ['2790297568\_metadata.gb', '2791334100\_metadata.gb']  
 Strain "Z01320SL0027" found in files: ['2790302094\_metadata.gb', '2791328684\_metadata.gb']  
 Strain "Z01319SL0009" found in files: ['2790306333\_metadata.gb', '2791327658\_metadata.gb']  
 Strain "Z01319SL0019" found in files: ['2790311342\_metadata.gb', '2791328750\_metadata.gb']  
 Strain "Z01319SL0010" found in files: ['2790316599\_metadata.gb', '2791326870\_metadata.gb']  
 Strain "Z01320SL0025" found in files: ['2790321214\_metadata.gb', '2791333611\_metadata.gb']  
 Strain "Z01320SL0024" found in files: ['2790326108\_metadata.gb', '2791328192\_metadata.gb']  
 Strain "Z01319SL0008" found in files: ['2790330984\_metadata.gb', '2791327513\_metadata.gb']  
 Strain "Z01319SL0006" found in files: ['2790335951\_metadata.gb', '2791328715\_metadata.gb']  
 Strain "Z01319SL0001" found in files: ['2790340519\_metadata.gb', '2791328489\_metadata.gb']  
 Strain "Z01319SL0004" found in files: ['2790345631\_metadata.gb', '2791334217\_metadata.gb']  
 Strain "Z01319SL0003" found in files: ['2790350405\_metadata.gb', '2791334406\_metadata.gb']  
 Strain "Z01319SL0005" found in files: ['2790355281\_metadata.gb', '2791334263\_metadata.gb']  
 Strain "Z01320SL0023" found in files: ['2790361485\_metadata.gb', '2791327413\_metadata.gb']  
 Strain "EC20111175" found in files: ['604278585\_metadata.gb', '749297197\_metadata.gb']  
 Strain "EC20111174" found in files: ['604290478\_metadata.gb', '749296449\_metadata.gb']  
 Strain "EC20100101" found in files: ['604336542\_metadata.gb', '749298461\_metadata.gb']  
 Strain "EC20090698" found in files: ['604362088\_metadata.gb', '749298420\_metadata.gb']  
 Strain "C500" found in files: ['674188659\_metadata.gb', '749314944\_metadata.gb']  
 Strain "CMCC" found in files: ['747132831\_metadata.gb', '747137039\_metadata.gb']  
 Strain "EC20090641" found in files: ['896682452\_metadata.gb', '915846032\_metadata.gb']  
 Strain "SA02D710168701" found in files: ['949867589\_metadata.gb', '983377086\_metadata.gb']  
 Strain "EC20121178" found in files: ['995901180\_metadata.gb', '998623498\_metadata.gb']

In [9]:

```
from Bio import SeqIO
import os

def find_duplicate_files(fasta_directory):
    seen_sequences = {}
    duplicates_count = {}

    # Traverse all files in the given directory
    for filename in os.listdir(fasta_directory):
        if filename.endswith(".fasta"):
            filepath = os.path.join(fasta_directory, filename)
            for record in SeqIO.parse(filepath, "fasta"):
                sequence = str(record.seq)
                if sequence in seen_sequences:
                    # Increment count for the file that has this sequence
                    if seen_sequences[sequence] in duplicates_count:
                        duplicates_count[seen_sequences[sequence]].append(filepath)
                    else:
                        duplicates_count[seen_sequences[sequence]] = [filepath]
                else:
                    # Store the file in which this sequence was first seen
                    seen_sequences[sequence] = filepath

    # Flatten the duplicates_count dictionary and count duplicates
    flat_duplicates = [file for files in duplicates_count.values() for file in files]
    unique_duplicates = set(flat_duplicates)

    print(f"Number of duplicate files found: {len(unique_duplicates)}")

    # Print all duplicate file paths
    if unique_duplicates:
        print("Duplicate files:")
        for dup_file in unique_duplicates:
            print(dup_file)

# Set the path to your FASTA files directory
fasta_directory = "C:/Users/DD/Documents/Python Scripts/sequences"

# Call the function to find and count duplicate files
find_duplicate_files(fasta_directory)
```

Number of duplicate files found: 150

Duplicate files:

C:/Users/DD/Documents/Python Scripts/sequences\998623498.fasta  
C:/Users/DD/Documents/Python Scripts/sequences\2740278012.fasta  
C:/Users/DD/Documents/Python Scripts/sequences\1046302332.fasta  
C:/Users/DD/Documents/Python Scripts/sequences\1829031305.fasta  
C:/Users/DD/Documents/Python Scripts/sequences\2288537321.fasta  
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C:/Users/DD/Documents/Python Scripts/sequences\2100424222.fasta

Checked duplicates based on metadata downloaded

```
In [11]: import os
import re
from collections import defaultdict

def extract_strain_info(metadata_directory):
    strain_info = defaultdict(list)

    # Iterate through all metadata files
    for filename in os.listdir(metadata_directory):
        if filename.endswith("_metadata.gb"):
            filepath = os.path.join(metadata_directory, filename)
            with open(filepath, "r") as file:
                content = file.read()
                # Extract strain information using regular expression
                match = re.search(r"strain=(\S+)", content)
                if match:
                    strain = match.group(1)
                    strain_info[strain].append(filename)

    # Count and print strains found in multiple files
    duplicate_strain_count = 0
    for strain, files in strain_info.items():
        if len(files) > 1:
            duplicate_strain_count += 1
            print(f"Strain {strain} found in {len(files)} files: {files}")

    # Print total count of duplicate strains
    print(f"\nTotal number of strains found in multiple files: {duplicate_strain_count}")

# Set the path to your metadata directory
metadata_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
extract_strain_info(metadata_directory)
```

Strain "SA20095440" found in 2 files: ['1000369143\_metadata.gb', '998647728\_metadata.gb']  
Strain "SA19960848" found in 2 files: ['1001551848\_metadata.gb', '995907166\_metadata.gb']  
Strain "SA19970769" found in 2 files: ['1001552415\_metadata.gb', '995922935\_metadata.gb']  
Strain "SA19970510" found in 2 files: ['1001554681\_metadata.gb', '995907817\_metadata.gb']  
Strain "SA19980677" found in 2 files: ['1001555507\_metadata.gb', '995923317\_metadata.gb']  
Strain "SA20094352" found in 2 files: ['1002985512\_metadata.gb', '998642845\_metadata.gb']  
Strain "CG29" found in 2 files: ['1030103469\_metadata.gb', '1033136052\_metadata.gb']  
Strain "SA020T09004001" found in 2 files: ['1045707385\_metadata.gb', '1046220323\_metadata.gb']  
Strain "SA01AB09084001" found in 2 files: ['1045716295\_metadata.gb', '1046302332\_metadata.gb']  
Strain "NS5391" found in 2 files: ['1127388936\_metadata.gb', '1128998525\_metadata.gb']  
Strain "87C" found in 2 files: ['1129908976\_metadata.gb', '1130930627\_metadata.gb']  
Strain "FORC\_038" found in 2 files: ['1190123157\_metadata.gb', '1196285509\_metadata.gb']  
Strain "D90" found in 2 files: ['1219800280\_metadata.gb', '1221158448\_metadata.gb']  
Strain "ATCC" found in 9 files: ['1528951495\_metadata.gb', '1531933061\_metadata.gb', '605520020\_metadata.gb', '605528405\_metadata.gb', '674281104\_metadata.gb', '820762957\_metadata.gb', '914820066\_metadata.gb', '914835508\_metadata.gb', '914845112\_metadata.gb']  
Strain "S" found in 2 files: ['1543688490\_metadata.gb', '1546887966\_metadata.gb']  
Strain "JT" found in 2 files: ['1633748201\_metadata.gb', '1634456755\_metadata.gb']  
Strain "CD-SL01" found in 2 files: ['1634015463\_metadata.gb', '1635401364\_metadata.gb']  
Strain "SJTUF87912v2" found in 2 files: ['1695497099\_metadata.gb', '1736663574\_metadata.gb']  
Strain "SJTUF13520v2" found in 2 files: ['1695513588\_metadata.gb', '1736666103\_metadata.gb']  
Strain "SL-312" found in 2 files: ['1732125948\_metadata.gb', '1733272414\_metadata.gb']  
Strain "SCSM4.1" found in 2 files: ['1788239996\_metadata.gb', '1797869977\_metadata.gb']  
Strain "SE124" found in 2 files: ['1828351404\_metadata.gb', '1828945320\_metadata.gb']  
Strain "SI85" found in 2 files: ['1828458697\_metadata.gb', '1829031305\_metadata.gb']  
Strain "SI67" found in 2 files: ['1828463287\_metadata.gb', '1829022561\_metadata.gb']  
Strain "CVM" found in 67 files: ['1836411705\_metadata.gb', '1842989566\_metadata.gb', '1843494016\_metadata.gb', '1843498400\_metadata.gb', '1843507307\_metadata.gb', '1843512211\_metadata.gb', '1843516861\_metadata.gb', '1843531062\_metadata.gb', '1843540331\_metadata.gb', '1843544971\_metadata.gb', '1843576148\_metadata.gb', '1843580775\_metadata.gb', '1843585404\_metadata.gb', '1843589994\_metadata.gb', '1843599223\_metadata.gb', '1843608483\_metadata.gb', '1843613175\_metadata.gb', '1843618362\_metadata.gb', '1843629339\_metadata.gb', '1843633944\_metadata.gb', '1843643169\_metadata.gb', '1843647868\_metadata.gb', '1843652501\_metadata.gb', '1846455821\_metadata.gb', '1951535343\_metadata.gb', '1951777374\_metadata.gb', '2090918790\_metadata.gb', '2090956231\_metadata.gb', '20909774716\_metadata.gb', '2091165092\_metadata.gb', '2091169855\_metadata.gb', '2091174439\_metadata.gb', '2091178941\_metadata.gb', '2091183720\_metadata.gb', '2091229717\_metadata.gb', '2091239126\_metadata.gb', '2091243740\_metadata.gb', '2091253165\_metadata.gb', '2091264036\_metadata.gb', '2091277794\_metadata.gb', '2091323346\_metadata.gb', '2091333084\_metadata.gb', '2091346897\_metadata.gb', '2091356744\_metadata.gb', '2091370368\_metadata.gb', '2091375002\_metadata.gb', '2091402345\_metadata.gb', '2091418357\_metadata.gb', '2091422800\_metadata.gb', '2091431810\_metadata.gb', '2091446076\_metadata.gb', '2091455843\_metadata.gb', '2091469184\_metadata.gb', '2091473540\_metadata.gb', '2091478163\_metadata.gb', '2091482867\_metadata.gb', '2136510056\_metadata.gb', '2518923471\_metadata.gb', '2521002406\_metadata.gb', '696581462\_metadata.gb', '696593549\_metadata.gb', '744788240\_metadata.gb', '767833982\_metadata.gb', '767838360\_metadata.gb', '767842734\_metadata.gb', '767847001\_metadata.gb', '771524686\_metadata.gb']  
Strain "VNSEC031" found in 2 files: ['1863533289\_metadata.gb', '1863660810\_metadata.gb']  
Strain "VNSEC023" found in 2 files: ['1863537930\_metadata.gb', '1863661087\_metadata.gb']  
Strain "VNSEC013" found in 2 files: ['1863542555\_metadata.gb', '1863657517\_metadata.gb']  
Strain "VNSEC003" found in 2 files: ['1863547333\_metadata.gb', '1863662535\_metadata.gb']  
Strain "VNSEC002" found in 2 files: ['1863552396\_metadata.gb', '1864594723\_metadata.gb']  
Strain "VNSEC001" found in 2 files: ['1863557100\_metadata.gb', '1864593957\_metadata.gb']  
Strain "R16.1424" found in 2 files: ['1943036513\_metadata.gb', '1945020687\_metadata.gb']  
Strain "GSJ/2016-SaI-017" found in 2 files: ['1979358710\_metadata.gb', '1982652612\_metadata.gb']  
Strain "R16.0556" found in 2 files: ['1993496820\_metadata.gb', '1995427272\_metadata.gb']  
Strain "KUFSE-SAL0043" found in 2 files: ['2034574262\_metadata.gb', '2036914008\_metadata.gb']  
Strain "CFSAN008081" found in 2 files: ['2049354170\_metadata.gb', '2082554870\_metadata.gb']  
Strain "Y220MC516" found in 2 files: ['2060222687\_metadata.gb', '2060646254\_metadata.gb']  
Strain "SA18578" found in 2 files: ['2077453074\_metadata.gb', '2078699049\_metadata.gb']  
Strain "SI6" found in 2 files: ['2087468524\_metadata.gb', '2087863205\_metadata.gb']  
Strain "ZC-S1" found in 2 files: ['2093998598\_metadata.gb', '2100166053\_metadata.gb']  
Strain "S90" found in 2 files: ['2094623483\_metadata.gb', '2100356487\_metadata.gb']  
Strain "SP" found in 2 files: ['2094623485\_metadata.gb', '2100424222\_metadata.gb']  
Strain "SI146" found in 2 files: ['2094642037\_metadata.gb', '2100237189\_metadata.gb']  
Strain "07Q015" found in 2 files: ['2095951993\_metadata.gb', '2100896152\_metadata.gb']  
Strain "Y221MC54" found in 2 files: ['2167845084\_metadata.gb', '2168004414\_metadata.gb']  
Strain "Y220MC56" found in 2 files: ['2258055768\_metadata.gb', '2258485671\_metadata.gb']  
Strain "Y220MC514" found in 2 files: ['2258066866\_metadata.gb', '2258710042\_metadata.gb']  
Strain "SE006" found in 2 files: ['2263926704\_metadata.gb', '2265635730\_metadata.gb']  
Strain "0134" found in 2 files: ['2271345254\_metadata.gb', '2273089235\_metadata.gb']  
Strain "KCID6" found in 2 files: ['2277353412\_metadata.gb', '2279079344\_metadata.gb']  
Strain "BCID6" found in 2 files: ['2277362348\_metadata.gb', '2279083905\_metadata.gb']  
Strain "AH19MC58" found in 2 files: ['2288173494\_metadata.gb', '2288530062\_metadata.gb']  
Strain "Y214C1328" found in 2 files: ['2288320740\_metadata.gb', '2288890305\_metadata.gb']  
Strain "MR517\_00712" found in 2 files: ['2294456242\_metadata.gb', '2294922788\_metadata.gb']  
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Strain "190819\_2" found in 2 files: ['2320734399\_metadata.gb', '2321042278\_metadata.gb']  
Strain "190807\_1" found in 2 files: ['2320738824\_metadata.gb', '2321031274\_metadata.gb']  
Strain "190729\_8" found in 2 files: ['2320748473\_metadata.gb', '2321043499\_metadata.gb']  
Strain "190729\_4" found in 2 files: ['2320752890\_metadata.gb', '2321043586\_metadata.gb']  
Strain "190704\_2" found in 2 files: ['2320757305\_metadata.gb', '2321008513\_metadata.gb']  
Strain "190610\_1" found in 2 files: ['2320761748\_metadata.gb', '2321041129\_metadata.gb']  
Strain "XM3104" found in 2 files: ['2333213671\_metadata.gb', '2339534824\_metadata.gb']  
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Strain "R22.0044" found in 2 files: ['2475999171\_metadata.gb', '2476810552\_metadata.gb']  
Strain "013" found in 2 files: ['2476413604\_metadata.gb', '2477330759\_metadata.gb']  
Strain "SI1467" found in 2 files: ['2482050684\_metadata.gb', '2482324433\_metadata.gb']  
Strain "CRIN508879" found in 2 files: ['2484128728\_metadata.gb', '2485141349\_metadata.gb']  
Strain "CRSE-01" found in 2 files: ['2514088622\_metadata.gb', '2515152310\_metadata.gb']  
Strain "SECVM-15" found in 2 files: ['2548245062\_metadata.gb', '2550287957\_metadata.gb']  
Strain "SECVM-17" found in 2 files: ['2548250070\_metadata.gb', '2550213647\_metadata.gb']  
Strain "SE" found in 2 files: ['2548260632\_metadata.gb', '2550213764\_metadata.gb']  
Strain "SECVM-13" found in 2 files: ['2548275539\_metadata.gb', '2550225976\_metadata.gb']  
Strain "SECVM-10" found in 2 files: ['2548291250\_metadata.gb', '2550255387\_metadata.gb']  
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Strain "Z1323CSL0015" found in 4 files: ['2570622740\_metadata.gb', '2572860206\_metadata.gb', '2703404023\_metadata.gb', '2711459283\_metadata.gb']  
Strain "LAS" found in 2 files: ['2595857884\_metadata.gb', '2601133276\_metadata.gb']  
Strain "40S987R1\_S50" found in 2 files: ['2636846570\_metadata.gb', '2638491583\_metadata.gb']  
Strain "40I964R1\_S49" found in 2 files: ['2636851041\_metadata.gb', '2638491693\_metadata.gb']  
Strain "180121R1S\_S48" found in 2 files: ['2636855593\_metadata.gb', '2638491622\_metadata.gb']  
Strain "SSSE-01" found in 2 files: ['2679318990\_metadata.gb', '2710805975\_metadata.gb']  
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Strain "Z1323CSL0047" found in 2 files: ['2704910405\_metadata.gb', '2711475272\_metadata.gb']  
Strain "Z1323CSL0052" found in 2 files: ['2704915029\_metadata.gb', '2711462605\_metadata.gb']  
Strain "Z1323CSL0051" found in 2 files: ['2704924300\_metadata.gb', '2711462769\_metadata.gb']  
Strain "ST.6536" found in 2 files: ['2717024100\_metadata.gb', '2717387261\_metadata.gb']  
Strain "IJC55-22" found in 2 files: ['2739185197\_metadata.gb', '2740278012\_metadata.gb']  
Strain "H4" found in 2 files: ['2784324973\_metadata.gb', '2784851814\_metadata.gb']  
Strain "Z01320SL0044" found in 2 files: ['2790142919\_metadata.gb', '2791959093\_metadata.gb']  
Strain "Z01320SL0045" found in 2 files: ['2790153796\_metadata.gb', '2791964378\_metadata.gb']  
Strain "Z01320SL0042" found in 2 files: ['2790167917\_metadata.gb', '2791959104\_metadata.gb']  
Strain "Z01320SL0040" found in 2 files: ['2790172290\_metadata.gb', '2791964398\_metadata.gb']  
Strain "Z01320SL0041" found in 2 files: ['2790176664\_metadata.gb', '2791964390\_metadata.gb']  
Strain "Z01320SL0038" found in 2 files: ['2790181112\_metadata.gb', '2791964373\_metadata.gb']  
Strain "Z01320SL0043" found in 2 files: ['2790191447\_metadata.gb', '2791964384\_metadata.gb']  
Strain "Z01320SL0030" found in 2 files: ['2790219725\_metadata.gb', '2791325747\_metadata.gb']  
Strain "Z01320SL0033" found in 2 files: ['2790226660\_metadata.gb', '2791348005\_metadata.gb']  
Strain "Z01320SL0034" found in 2 files: ['2790232908\_metadata.gb', '2791323978\_metadata.gb']  
Strain "Z01320SL0031" found in 2 files: ['2790238422\_metadata.gb', '2791328742\_metadata.gb']  
Strain "Z01320SL0036" found in 2 files: ['2790243305\_metadata.gb', '2791327776\_metadata.gb']  
Strain "Z01320SL0037" found in 2 files: ['2790247716\_metadata.gb', '2791329900\_metadata.gb']



```

Strain "Z01320S10035" found in 2 files: ['2790252121_metadata.gb', '2791327646_metadata.gb']
Strain "Z01320S10032" found in 2 files: ['2790256505_metadata.gb', '2791332598_metadata.gb']
Strain "Z01319S10021" found in 2 files: ['2790261062_metadata.gb', '2791326641_metadata.gb']
Strain "Z01319S10015" found in 2 files: ['2790265499_metadata.gb', '2791328717_metadata.gb']
Strain "Z01319S10020" found in 2 files: ['2790270044_metadata.gb', '2791327275_metadata.gb']
Strain "Z01320S10029" found in 2 files: ['2790274631_metadata.gb', '2791332929_metadata.gb']
Strain "Z01319S10016" found in 2 files: ['2790279127_metadata.gb', '2791327653_metadata.gb']
Strain "Z01319S10017" found in 2 files: ['2790283963_metadata.gb', '2791328843_metadata.gb']
Strain "Z01319S10022" found in 2 files: ['2790292938_metadata.gb', '2791327656_metadata.gb']
Strain "Z01320S10026" found in 2 files: ['2790297568_metadata.gb', '2791334100_metadata.gb']
Strain "Z01320S10027" found in 2 files: ['2790302094_metadata.gb', '2791328684_metadata.gb']
Strain "Z01319S10009" found in 2 files: ['2790306333_metadata.gb', '2791327658_metadata.gb']
Strain "Z01319S10019" found in 2 files: ['2790311342_metadata.gb', '2791328750_metadata.gb']
Strain "Z01319S10010" found in 2 files: ['2790316599_metadata.gb', '2791326870_metadata.gb']
Strain "Z01320S10025" found in 2 files: ['2790321214_metadata.gb', '2791333611_metadata.gb']
Strain "Z01320S10024" found in 2 files: ['2790326108_metadata.gb', '2791328192_metadata.gb']
Strain "Z01319S10008" found in 2 files: ['2790330984_metadata.gb', '2791327513_metadata.gb']
Strain "Z01319S10006" found in 2 files: ['2790335951_metadata.gb', '2791328715_metadata.gb']
Strain "Z01319S10001" found in 2 files: ['2790340519_metadata.gb', '2791328489_metadata.gb']
Strain "Z01319S10004" found in 2 files: ['2790345631_metadata.gb', '2791334217_metadata.gb']
Strain "Z01319S10003" found in 2 files: ['2790350405_metadata.gb', '2791334406_metadata.gb']
Strain "Z01319S10005" found in 2 files: ['2790355281_metadata.gb', '2791334263_metadata.gb']
Strain "Z01320S10023" found in 2 files: ['2790361485_metadata.gb', '2791327413_metadata.gb']
Strain "EC20111175" found in 2 files: ['604278585_metadata.gb', '749297197_metadata.gb']
Strain "EC20111174" found in 2 files: ['604290478_metadata.gb', '749296449_metadata.gb']
Strain "EC20100101" found in 2 files: ['604336542_metadata.gb', '749298461_metadata.gb']
Strain "EC20090698" found in 2 files: ['604362088_metadata.gb', '749298420_metadata.gb']
Strain "CS00" found in 2 files: ['674188659_metadata.gb', '749314944_metadata.gb']
Strain "CMCC" found in 2 files: ['747132831_metadata.gb', '747137039_metadata.gb']
Strain "EC20090641" found in 2 files: ['896682452_metadata.gb', '915846032_metadata.gb']
Strain "SA02D10168701" found in 2 files: ['949867589_metadata.gb', '983377086_metadata.gb']
Strain "EC20121178" found in 2 files: ['995901180_metadata.gb', '998623498_metadata.gb']

```

Total number of strains found in multiple files: 144

```

In [13]: import os
import re
from collections import defaultdict

def extract_strain_info(metadata_directory):
    strain_info = defaultdict(list)

    # Iterate through all metadata files
    for filename in os.listdir(metadata_directory):
        if filename.endswith("_metadata.gb"):
            filepath = os.path.join(metadata_directory, filename)
            with open(filepath, "r") as file:
                content = file.read()
                # Extract strain information using regular expression
                match = re.search(r"Strain=(\S+)", content)
                if match:
                    strain = match.group(1)
                    strain_info[strain].append(filename)

    # Count strains and files
    strain_file_counts = defaultdict(int)
    for files in strain_info.values():
        for file in files:
            strain_file_counts[file] += 1

    # Print strains and file counts
    for file, count in strain_file_counts.items():
        print(f"File {file} contains {count} strains")

    # Print the number of duplicate strains
    duplicate_strain_count = sum(1 for files in strain_info.values() if len(files) > 1)
    print(f"\nTotal number of strains found in multiple files: {duplicate_strain_count}")

# Set the path to your metadata directory
metadata_directory = "C:/Users/DD/Documents/Python Scripts/sequences"
extract_strain_info(metadata_directory)

```

File 1000369143\_metadata.gb contains 1 strains  
File 998647728\_metadata.gb contains 1 strains  
File 1001427523\_metadata.gb contains 1 strains  
File 1001551848\_metadata.gb contains 1 strains  
File 995907166\_metadata.gb contains 1 strains  
File 1001552415\_metadata.gb contains 1 strains  
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File 2790311342\_metadata.gb contains 1 strains  
File 2791328750\_metadata.gb contains 1 strains  
File 2790316599\_metadata.gb contains 1 strains  
File 2791326870\_metadata.gb contains 1 strains  
File 2790321214\_metadata.gb contains 1 strains  
File 2791333611\_metadata.gb contains 1 strains  
File 2790326108\_metadata.gb contains 1 strains  
File 2791328192\_metadata.gb contains 1 strains  
File 2790330984\_metadata.gb contains 1 strains  
File 2791327513\_metadata.gb contains 1 strains  
File 2790335951\_metadata.gb contains 1 strains  
File 2791328715\_metadata.gb contains 1 strains  
File 2790340519\_metadata.gb contains 1 strains  
File 2791328489\_metadata.gb contains 1 strains  
File 2790345631\_metadata.gb contains 1 strains  
File 2791334217\_metadata.gb contains 1 strains  
File 2790350405\_metadata.gb contains 1 strains  
File 2791334406\_metadata.gb contains 1 strains  
File 2790355281\_metadata.gb contains 1 strains  
File 2791334263\_metadata.gb contains 1 strains  
File 2790361485\_metadata.gb contains 1 strains  
File 2791327413\_metadata.gb contains 1 strains  
File 332986951\_metadata.gb contains 1 strains  
File 523806722\_metadata.gb contains 1 strains  
File 523815970\_metadata.gb contains 1 strains  
File 523821078\_metadata.gb contains 1 strains  
File 529190224\_metadata.gb contains 1 strains  
File 548713695\_metadata.gb contains 1 strains  
File 559187652\_metadata.gb contains 1 strains  
File 563346448\_metadata.gb contains 1 strains  
File 564743501\_metadata.gb contains 1 strains  
File 601101465\_metadata.gb contains 1 strains  
File 602804670\_metadata.gb contains 1 strains  
File 602808998\_metadata.gb contains 1 strains  
File 602813325\_metadata.gb contains 1 strains  
File 602817697\_metadata.gb contains 1 strains  
File 602822031\_metadata.gb contains 1 strains  
File 602826365\_metadata.gb contains 1 strains  
File 604190314\_metadata.gb contains 1 strains  
File 604257961\_metadata.gb contains 1 strains  
File 604268692\_metadata.gb contains 1 strains  
File 604278585\_metadata.gb contains 1 strains  
File 749297197\_metadata.gb contains 1 strains  
File 604290478\_metadata.gb contains 1 strains  
File 749296449\_metadata.gb contains 1 strains  
File 604307148\_metadata.gb contains 1 strains  
File 604322487\_metadata.gb contains 1 strains  
File 604336542\_metadata.gb contains 1 strains  
File 749298461\_metadata.gb contains 1 strains  
File 604350507\_metadata.gb contains 1 strains  
File 604362088\_metadata.gb contains 1 strains  
File 749298420\_metadata.gb contains 1 strains  
File 605498156\_metadata.gb contains 1 strains  
File 605511148\_metadata.gb contains 1 strains  
File 605537200\_metadata.gb contains 1 strains  
File 605546632\_metadata.gb contains 1 strains  
File 605555317\_metadata.gb contains 1 strains  
File 630832964\_metadata.gb contains 1 strains  
File 674188659\_metadata.gb contains 1 strains  
File 749314944\_metadata.gb contains 1 strains  
File 674230814\_metadata.gb contains 1 strains  
File 682036555\_metadata.gb contains 1 strains  
File 682049973\_metadata.gb contains 1 strains  
File 682058852\_metadata.gb contains 1 strains  
File 682063065\_metadata.gb contains 1 strains  
File 682067343\_metadata.gb contains 1 strains  
File 682080448\_metadata.gb contains 1 strains  
File 682084826\_metadata.gb contains 1 strains  
File 682089197\_metadata.gb contains 1 strains  
File 682093565\_metadata.gb contains 1 strains  
File 686507741\_metadata.gb contains 1 strains  
File 747132831\_metadata.gb contains 1 strains  
File 747137039\_metadata.gb contains 1 strains  
File 754295078\_metadata.gb contains 1 strains  
File 754295112\_metadata.gb contains 1 strains  
File 820758584\_metadata.gb contains 1 strains  
File 896682452\_metadata.gb contains 1 strains  
File 915846032\_metadata.gb contains 1 strains  
File 896682995\_metadata.gb contains 1 strains  
File 901905107\_metadata.gb contains 1 strains  
File 901911536\_metadata.gb contains 1 strains  
File 924635423\_metadata.gb contains 1 strains  
File 924639146\_metadata.gb contains 1 strains  
File 924643612\_metadata.gb contains 1 strains  
File 930813018\_metadata.gb contains 1 strains  
File 930817479\_metadata.gb contains 1 strains  
File 933889448\_metadata.gb contains 1 strains  
File 933894153\_metadata.gb contains 1 strains  
File 949867589\_metadata.gb contains 1 strains  
File 983377086\_metadata.gb contains 1 strains  
File 949872475\_metadata.gb contains 1 strains  
File 949877344\_metadata.gb contains 1 strains  
File 953768973\_metadata.gb contains 1 strains  
File 953773649\_metadata.gb contains 1 strains  
File 971178605\_metadata.gb contains 1 strains  
File 995901180\_metadata.gb contains 1 strains  
File 998623498\_metadata.gb contains 1 strains  
File 995915826\_metadata.gb contains 1 strains  
File 995918924\_metadata.gb contains 1 strains  
File 995920397\_metadata.gb contains 1 strains

File 995923765\_metadata.gb contains 1 strains  
File 998623489\_metadata.gb contains 1 strains

Total number of strains found in multiple files: 144

### Data collection summary :

complete salmonella sequences derived from chicken isolates has been searched in NCBI (National Center for Biotechnology Information) Pathogen detection database. It has been founded that 496 sequences are present. A through examination of presence of duplicates has been done based of sequence IDs and strains.

Finally we have obtained 299 unique sequences with their information stored in an excel file and that has been considered for further analysis.

## Step 02 - AMR gene identification using CARD- Resistance Gene Identifier

In this step AMR genes along with the drug classes for the acquired data has been detected using Resistance Gene Identifier tool provided by CARD database

Each and every sequences of 299 salmonella sequences has been fed into the RGI tool and AMR gene information has been collected in an excel file sepeartely.

As this Bioinformatics tools works in Linux enviroment, this has been seperately done in Linux OS by the means of iterating over the folder of salmonella seqeinces into RGI.

Finally the results has been stored in a separate file.

Step -3 Data Wrangling

In this stage Data wrangling carried out with the isolate information and the RGI results

In [1]:

```
import pandas as pd
df1 = pd.read_csv("C:/VIT/Semester 3/SET-03/isolates.csv")
df1
```

Out[1]:

	#Organism group	Strain	Isolate identifiers	Serovar	Isolate	Create date	Location	Isolation source	Isolation type	Food origin	SNP cluster	Min-same	Min-diff	BioSample	A
0	Salmonella enterica	WAPHL_SAL-A00031	"SRS469289","WAPHL_SAL-A00031"	Montevideo	PDT000000399.3	2015-02-11T06:08:33Z	USA:WA	chicken	environmental/other	NaN	PDS000172527.57	3.0	3.0	SAMN02182894	GCA_0114
1	Salmonella enterica	WAPHL_SAL-A00052	"SRS479789","WAPHL_SAL-A00052"	Ohio	PDT000000661.3	2015-02-11T06:08:35Z	USA:WA	chicken	environmental/other	NaN	PDS000027300.3	0.0	NaN	SAMN02182915	GCA_0106
2	Salmonella enterica	WAPHL_SAL-A00149	"SRS515215","WAPHL_SAL-A00149"	NaN	PDT000001787.3	2015-02-11T17:10:01Z	USA:WA	chicken	environmental/other	NaN	PDS000179566.108	7.0	13.0	SAMN02182983	GCA_0068
3	Salmonella enterica	MDH-2013-00166	"MDH-2013-00166","SRS523520"	Kentucky	PDT000002088.3	2015-02-11T17:10:02Z	USA:MN	chicken feces	environmental/other	NaN	PDS000117429.168	0.0	27.0	SAMN02378171	GCA_0106
4	Salmonella enterica	MDH-2013-00167	"MDH-2013-00167","SRS523566"	Thompson	PDT000002114.3	2015-02-11T17:10:02Z	USA:MN	chicken feces	environmental/other	NaN	PDS000032705.1262	3.0	1.0	SAMN02378172	
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	Salmonella enterica	YAH-CH5	"SRS21539871","YAH-CH5"	NaN	PDT002189990.1	2024-06-06T22:10:06Z	USA: Kentucky	chicken	environmental/other	NaN	PDS000050977.104	0.0	5.0	SAMN41714676	
295	Salmonella enterica	YAH-CH4	"SRS21539872","YAH-CH4"	Thompson	PDT002189991.1	2024-06-06T22:10:07Z	USA: Kentucky	chicken	environmental/other	NaN	PDS000032705.1262	0.0	3.0	SAMN41714675	
296	Salmonella enterica	YAH-CH3	"SRS21539869","YAH-CH3"	Agona	PDT002189992.1	2024-06-06T22:10:07Z	USA: Kentucky	chicken	environmental/other	NaN	PDS000050977.104	1.0	6.0	SAMN41714674	
297	Salmonella enterica	YAH-CH2	"SRS21539870","YAH-CH2"	Liverpool	PDT002189993.1	2024-06-06T22:10:07Z	USA: Kentucky	chicken	environmental/other	NaN	PDS000001378.421	30.0	8.0	SAMN41714673	
298	Salmonella enterica	YAH-CH1	"SRS21539868","YAH-CH1"	Agona	PDT002189994.1	2024-06-06T22:10:08Z	USA: Kentucky	chicken	environmental/other	NaN	PDS000050977.104	0.0	5.0	SAMN41714672	

299 rows × 17 columns

In [3]:

```
df2 = pd.read_excel("C:/VIT/Semester 3/SET-03/Genes by antimicrobial agents and classes.xlsx")
df2
```

Out[3]:

	Class	Antimicrobial Agent	Gene
0	Aminoglycosides	Gentamicin	aac(3)-Ia
1	Aminoglycosides	Gentamicin	aac(3)-IIa
2	Aminoglycosides	Gentamicin	aac(3)-IIIa
3	Aminoglycosides	Gentamicin	aac(3)-IV
4	Aminoglycosides	Gentamicin	aac(3)-IVa
...	...	...	...
219	Tetracyclines	Tetracycline	tet(D)
220	Tetracyclines	Tetracycline	tet(G)
221	Tetracyclines	Tetracycline	tet(M)
222	Tetracyclines	Tetracycline	tet(O)
223	Tetracyclines	Tetracycline	tet(X5)

224 rows × 3 columns

In [5]:

```
# Creating df3 with the 'Isolate' column from df1
df3 = df1[['Isolate']]
df3
```

Out[5]:

	Isolate
0	PDT000000399.3
1	PDT000000661.3
2	PDT000001787.3
3	PDT000002088.3
4	PDT000002114.3
...	...
294	PDT002189990.1
295	PDT002189991.1
296	PDT002189992.1
297	PDT002189993.1
298	PDT002189994.1

299 rows × 1 columns

In [7]:

```
genes = df2['Gene'].tolist()

# Create a DataFrame with the new columns
new_columns = pd.DataFrame({gene: None for gene in genes}, index=df3.index)

# Concatenate the new columns to the original DataFrame
df3 = pd.concat([df3, new_columns], axis=1)

df3
```

Out[7]:

	Isolate	aac(3)-Ia	aac(3)-IIa	aac(3)-IIIa	aac(3)-IV	aac(3)-IVa	aac(3)-VIa	aac(6')-Ib	aac(6')-Ib4	aac(6')-IIa	...	tet	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)
0	PDT000000399.3	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
1	PDT000000661.3	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
2	PDT000001787.3	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
3	PDT000002088.3	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
4	PDT000002114.3	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	PDT002189990.1	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
295	PDT002189991.1	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
296	PDT002189992.1	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
297	PDT002189993.1	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None
298	PDT002189994.1	None	None	None	None	None	None	None	None	None	...	None	None	None	None	None	None	None	None	None	None

299 rows × 187 columns

In [9]:

```
# Replace None with 0 or 1 based on the condition
for i, row in df3.iterrows():
    isolate = row['Isolate']
    matching_row = df1[df1['Isolate'] == isolate]
    if not matching_row.empty:
        amr_genotypes = matching_row.iloc[0]['AMR genotypes']
        for gene in genes:
            if gene in amr_genotypes:
                df3.at[i, gene] = 1
            else:
                df3.at[i, gene] = 0
```

In [10]:

```
# Check if any value in df3 (excluding the first column) is 1
has_ones = (df3.iloc[:, 1:] == 1).any().any()

if has_ones:
    print("df3 has at least one '1' in columns other than the first column.")
else:
    print("df3 does not have any '1's in columns other than the first column.")
```

df3 has at least one '1' in columns other than the first column.

In [13]:

df3

Out[13]:

	Isolate	aac(3)-Ia	aac(3)-IIa	aac(3)-IIIa	aac(3)-IV	aac(3)-IVa	aac(3)-VIa	aac(6')-Ib	aac(6')-Ib4	aac(6')-IIa	...	tet	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)
0	PDT000000399.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
1	PDT000000661.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
2	PDT000001787.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
3	PDT000002088.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
4	PDT000002114.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	PDT002189990.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
295	PDT002189991.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
296	PDT002189992.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
297	PDT002189993.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
298	PDT002189994.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0

299 rows × 187 columns

In [18]:

```
df3.to_csv("df3.csv", index=0)
```

In [20]:

```
# Verification step
verification_results = []
for i, row in df1.iterrows():
    isolate = row['Isolate']
    amr_genotypes = row['AMR genotypes']
    matching_row = df3[df3['Isolate'] == isolate]
    if not matching_row.empty:
        all_marked = all(gene not in genes or matching_row.iloc[0][gene] == 1 for gene in amr_genotypes)
        verification_results.append((isolate, all_marked))

# Print verification results
for isolate, result in verification_results:
    print(f"Isolate {isolate}: {'All genes marked as 1' if result else 'Some genes not marked as 1'})
```



[illegible]

[illegible]

Isolate PDT001197074.1: All genes marked as 1  
Isolate PDT001197075.1: All genes marked as 1  
Isolate PDT001197076.1: All genes marked as 1  
Isolate PDT001197077.1: All genes marked as 1  
Isolate PDT001197079.1: All genes marked as 1  
Isolate PDT001197080.1: All genes marked as 1  
Isolate PDT001197086.1: All genes marked as 1  
Isolate PDT001197087.1: All genes marked as 1  
Isolate PDT001197089.1: All genes marked as 1  
Isolate PDT001197090.1: All genes marked as 1  
Isolate PDT001197091.1: All genes marked as 1  
Isolate PDT001197094.1: All genes marked as 1  
Isolate PDT001197096.1: All genes marked as 1  
Isolate PDT001197097.1: All genes marked as 1  
Isolate PDT001197098.1: All genes marked as 1  
Isolate PDT001197099.1: All genes marked as 1  
Isolate PDT001197101.1: All genes marked as 1  
Isolate PDT001242435.1: All genes marked as 1  
Isolate PDT001608224.1: All genes marked as 1  
Isolate PDT001608225.1: All genes marked as 1  
Isolate PDT001608226.1: All genes marked as 1  
Isolate PDT001608227.1: All genes marked as 1  
Isolate PDT001608230.1: All genes marked as 1  
Isolate PDT001608231.1: All genes marked as 1  
Isolate PDT001608232.1: All genes marked as 1  
Isolate PDT001608233.1: All genes marked as 1  
Isolate PDT001608234.1: All genes marked as 1  
Isolate PDT001608235.1: All genes marked as 1  
Isolate PDT001608236.1: All genes marked as 1  
Isolate PDT001608237.1: All genes marked as 1  
Isolate PDT001608238.1: All genes marked as 1  
Isolate PDT001608242.1: All genes marked as 1  
Isolate PDT001608243.1: All genes marked as 1  
Isolate PDT001616280.1: All genes marked as 1  
Isolate PDT001616281.1: All genes marked as 1  
Isolate PDT001616283.1: All genes marked as 1  
Isolate PDT001616284.1: All genes marked as 1  
Isolate PDT001616285.1: All genes marked as 1  
Isolate PDT001616286.1: All genes marked as 1  
Isolate PDT001616352.1: All genes marked as 1  
Isolate PDT001616353.1: All genes marked as 1  
Isolate PDT001616361.1: All genes marked as 1  
Isolate PDT001616362.1: All genes marked as 1  
Isolate PDT001616363.1: All genes marked as 1  
Isolate PDT001616364.1: All genes marked as 1  
Isolate PDT002189985.1: All genes marked as 1  
Isolate PDT002189986.1: All genes marked as 1  
Isolate PDT002189987.1: All genes marked as 1  
Isolate PDT002189988.1: All genes marked as 1  
Isolate PDT002189989.1: All genes marked as 1  
Isolate PDT002189990.1: All genes marked as 1  
Isolate PDT002189991.1: All genes marked as 1  
Isolate PDT002189992.1: All genes marked as 1  
Isolate PDT002189993.1: All genes marked as 1  
Isolate PDT002189994.1: All genes marked as 1

In [22]: df3

Out[22]:

	Isolate	aac(3)-Ia	aac(3)-IIa	aac(3)-IIla	aac(3)-IV	aac(3)-IVa	aac(3)-VIa	aac(6')-Ib	aac(6')-Ib4	aac(6')-IIa	...	tet	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)
0	PDT000000399.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
1	PDT000000661.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
2	PDT000001787.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
3	PDT000002088.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
4	PDT000002114.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	PDT002189990.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
295	PDT002189991.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
296	PDT002189992.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
297	PDT002189993.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0
298	PDT002189994.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	0

299 rows x 187 columns

In [24]: df3.to\_csv("df3.csv",index=0)

In [26]: df3['AMR Results'] = df3.iloc[:, 2:].apply(lambda row: 'Positive' if 1 in row.values else 'Negative', axis=1)

In [28]: df3

Out[28]:

	Isolate	aac(3)-Ia	aac(3)-IIa	aac(3)-IIla	aac(3)-IV	aac(3)-IVa	aac(3)-VIa	aac(6')-Ib	aac(6')-Ib4	aac(6')-IIa	...	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)	AMR Results
0	PDT000000399.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
1	PDT000000661.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
2	PDT000001787.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
3	PDT000002088.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
4	PDT000002114.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	PDT002189990.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
295	PDT002189991.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
296	PDT002189992.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
297	PDT002189993.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative
298	PDT002189994.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Negative

299 rows x 188 columns

In [30]: df3.to\_csv("df3.csv",index=0)

In [32]: # Add the drug classes column  
df3['drug classes'] = ''  
  
for i, row in df3.iterrows():  
 if row['AMR Results'] == 'Positive':

```
drug_classes = []
for gene in genes:
    if row[gene] == 1:
        matching_agent = df2[df2['Gene'] == gene]['Antimicrobial Agent'].values
        if len(matching_agent) > 0:
            drug_classes.append(matching_agent[0])
df3.at[i, 'drug_classes'] = ', '.join(drug_classes)

df3
```

Out[32]:

	Isolate	aac(3)-la	aac(3)-lla	aac(3)-llla	aac(3)-IV	aac(3)-IVa	aac(3)-VIa	aac(6')-lb	aac(6')-lb4	aac(6')-lla	...	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)	AMR Results	drug classes	
0	PDT000000399.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
1	PDT000000661.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
2	PDT000001787.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
3	PDT000002088.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
4	PDT000002114.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	PDT002189990.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
295	PDT002189991.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
296	PDT002189992.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
297	PDT002189993.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		
298	PDT002189994.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	Negative		

299 rows × 189 columns

In [34]: df3.to\_csv("df3.csv",index=0)

In [1]: `import pandas as pd
df=pd.read_csv("finaldata.csv")
df`

Out[1]:

	Isolate	aadA1	aadA2	aadA3	aadA4	aadA5	aadA6	aadA7	aadA8	aadA12	...	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)	drug classes
0	PDT000000399.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
1	PDT000000661.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
2	PDT000001787.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
3	PDT000002088.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
4	PDT000002114.3	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
294	PDT002189990.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
295	PDT002189991.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
296	PDT002189992.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
297	PDT002189993.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA
298	PDT002189994.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	NA

299 rows × 165 columns

In [3]: `# Strip any extra whitespace and remove rows where 'drug classes' column contains "NA"
df['drug classes'] = df['drug classes'].str.strip() # Remove any surrounding whitespace
df4 = df[df['drug classes'] != "NA"]
df4`

Out[3]:

	Isolate	aadA1	aadA2	aadA3	aadA4	aadA5	aadA6	aadA7	aadA8	aadA12	...	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)	drug classes
5	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin, Streptomycin, Amoxicillin-Clavul...
10	PDT000002530.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin, Tetracycline, Tetracycline
11	PDT000002532.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin, Tetracycline, Tetracycline
14	PDT000002538.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin, Amoxicillin-Clavulanic Acid, Amo...
19	PDT000002547.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Tetracycline, Tetracycline
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
285	PDT001616361.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Streptomycin, Sulfamethoxazole-Sulfisoxazole, ...
286	PDT001616362.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Streptomycin, Sulfamethoxazole-Sulfisoxazole, ...
287	PDT001616363.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Streptomycin, Sulfamethoxazole-Sulfisoxazole, ...
288	PDT001616364.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Streptomycin, Sulfamethoxazole-Sulfisoxazole, ...
291	PDT002189987.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Streptomycin

188 rows × 165 columns

In [5]: `# First, split the 'drug classes' column by commas, creating lists in each cell
df4['drug classes'] = df4['drug classes'].str.split(',')

# Expand the rows based on the split values in 'drug classes'
df5 = df4.explode('drug classes').reset_index(drop=True)

# Optional: Strip any whitespace around drug names after exploding
df5['drug classes'] = df5['drug classes'].str.strip()

# Display or save the resulting dataframe
df5.head() # This will display the first few rows`

C:\Users\DD\AppData\Local\Temp\ipykernel\_13416\3902397106.py:2: SettingWithCopyWarning:  
A value is trying to be set on a copy of a slice from a DataFrame.  
Try using .loc[row\_indexer,col\_indexer] = value instead

See the caveats in the documentation: [https://pandas.pydata.org/pandas-docs/stable/user\\_guide/indexing.html#returning-a-view-versus-a-copy](https://pandas.pydata.org/pandas-docs/stable/user_guide/indexing.html#returning-a-view-versus-a-copy)  
df4['drug\_classes'] = df4['drug\_classes'].str.split(',')

Out[5]:

	Isolate	aadA1	aadA2	aadA3	aadA4	aadA5	aadA6	aadA7	aadA8	aadA12	...	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)	drug_classes
0	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin
1	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin
2	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Amoxicillin-Clavulanic Acid
3	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Amoxicillin-Clavulanic Acid
4	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Amoxicillin-Clavulanic Acid

5 rows × 165 columns

In [7]:

df5.shape

Out[7]:

(1306, 165)

In [9]:

```
# Save df21 as a CSV file
df5.to_csv('finaldata_processed.csv', index=False)
```

In [13]:

```
# Get unique values in the 'drug_classes' column
unique_drug_classes = df5['drug_classes'].unique()

# Get the number of unique values
num_unique_drug_classes = df5['drug_classes'].nunique()

# Print the results
print(f"Number of unique drug classes: {num_unique_drug_classes}")
print("Unique drug classes:")
for drug_class in unique_drug_classes:
    print(drug_class)
```

Number of unique drug classes: 11  
Unique drug classes:  
Streptomycin  
Amoxicillin-Clavulanic Acid  
Tetracycline  
Ampicillin  
Sulfamethoxazole-Sulfisoxazole  
Chloramphenicol  
Ciprofloxacin  
Nalidixic Acid  
Trimethoprim-Sulfamethoxazole  
Gentamicin  
Ceftriaxone

In [35]:

```
import pandas as pd

# Replace 'your_csv_file.csv' with the path to your CSV file
df51 = pd.read_csv('finaldata_processed.csv')
df51
```

Out[35]:

	Isolate	aadA1	aadA2	aadA3	aadA4	aadA5	aadA6	aadA7	aadA8	aadA12	...	tet(31)	tet(A)	tet(B)	tet(C)	tet(D)	tet(G)	tet(M)	tet(O)	tet(X5)	drug_classes
0	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin
1	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Streptomycin
2	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Amoxicillin-Clavulanic Acid
3	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Amoxicillin-Clavulanic Acid
4	PDT000002344.3	0	0	0	0	0	0	0	0	0	...	0	0	1	0	0	0	0	0	0	Amoxicillin-Clavulanic Acid
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
1301	PDT001616364.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Sulfamethoxazole-Sulfisoxazole
1302	PDT001616364.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Nalidixic Acid
1303	PDT001616364.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Tetracycline
1304	PDT001616364.1	1	0	0	0	0	0	0	0	0	...	0	1	0	0	0	0	0	0	0	Tetracycline
1305	PDT002189987.1	0	0	0	0	0	0	0	0	0	...	0	0	0	0	0	0	0	0	0	Streptomycin

1306 rows × 165 columns

In [17]:

```
# Check the data type of the 'drug_classes' column
print("Data type of 'drug_classes':", df51['drug_classes'].dtype)

# Display the first few rows to inspect the formatting
print("\nFirst few rows of the 'drug_classes' column:")
print(df51['drug_classes'].head(10))
```

Data type of 'drug\_classes': object

First few rows of the 'drug\_classes' column:  
0 Streptomycin  
1 Streptomycin  
2 Amoxicillin-Clavulanic Acid  
3 Amoxicillin-Clavulanic Acid  
4 Amoxicillin-Clavulanic Acid  
5 Amoxicillin-Clavulanic Acid  
6 Amoxicillin-Clavulanic Acid  
7 Tetracycline  
8 Tetracycline  
9 Streptomycin  
Name: drug\_classes, dtype: object

In [19]:

```
# Convert 'drug_classes' to string to avoid NaN conversion
df51['drug_classes'] = df51['drug_classes'].astype(str)

# Check if conversion fixed the issue
print("\nAfter conversion, first few rows of 'drug_classes':")
print(df51['drug_classes'].head(10))
```

After conversion, first few rows of 'drug classes':

```
Streptomycin
0 Streptomycin
1 Streptomycin
2 Amoxicillin-Clavulanic Acid
3 Amoxicillin-Clavulanic Acid
4 Amoxicillin-Clavulanic Acid
5 Amoxicillin-Clavulanic Acid
6 Amoxicillin-Clavulanic Acid
7 Tetracycline
8 Tetracycline
9 Streptomycin
Name: drug classes, dtype: object
```

```
In [23]: # Strip whitespace from the 'drug classes' column
df51['drug classes'] = df51['drug classes'].str.strip()

# Check the unique values again
unique_drug_classes = df51['drug classes'].unique()
print("\nUnique values in 'drug classes' after stripping whitespace:")
print(unique_drug_classes)
```

Unique values in 'drug classes' after stripping whitespace:

```
['Streptomycin' 'Amoxicillin-Clavulanic Acid' 'Tetracycline' 'Ampicillin'
'Sulfamethoxazole-Sulfisoxazole' 'Chloramphenicol' 'Ciprofloxacin'
'Nalidixic Acid' 'Trimethoprim-Sulfamethoxazole' 'Gentamicin'
'Ceftriaxone']
```

```
In [25]: # Verify if expected drug classes are present
expected_drug_classes = [
    'Streptomycin', 'Amoxicillin-Clavulanic Acid', 'Tetracycline', 'Ampicillin',
    'Sulfamethoxazole-Sulfisoxazole', 'Chloramphenicol', 'Ciprofloxacin',
    'Nalidixic Acid', 'Trimethoprim-Sulfamethoxazole', 'Gentamicin', 'Ceftriaxone'
]

# Display a message if any expected drug class is missing
missing_classes = [drug for drug in expected_drug_classes if drug not in unique_drug_classes]
if missing_classes:
    print("\nMissing drug classes:")
    print(missing_classes)
else:
    print("\nAll expected drug classes are present.")
```

All expected drug classes are present.

```
In [37]: # Initialize a dictionary to store unique column names for each drug class
drug_class_columns = {drug_class: [] for drug_class in df51['drug classes'].unique()}

# Iterate through each row in the DataFrame
for index, row in df51.iterrows():
    # Get the current drug class
    current_drug_class = row['drug classes']

    # Check for presence of 1 in the other columns
    for column in df51.columns[1:]: # Skip the first column which is 'drug classes'
        if row[column] == 1 and column not in drug_class_columns[current_drug_class]:
            drug_class_columns[current_drug_class].append(column)

# Display the results
for drug_class, columns in drug_class_columns.items():
    print(f"{drug_class}: {columns}")
```

Streptomycin: ['aph(6)-Ic', 'aph(6)-Id', 'blaCMY', 'blaCMY-2', 'tet', 'tet(B)', 'tet(A)', 'aadA2', 'sul1', 'aadA1', 'sul3', 'blaCARB-2', 'floR', 'tet(G)', 'tet(M)', 'aadA7', 'blaTEM', 'blaTEM-1', 'parC\_S80I', 'gyrA\_D87G', 'gyrA\_S83F', 'sul2', 'dfrA1', 'blaOXA-1', 'blaCTX-M', 'blaCTX-M-2', 'qnrB1', 'qnrB19', 'aadA12', 'aadA15', 'aadA22', 'dfrA14', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'gyrA\_S83Y', 'dfrA12', 'blaCTX-M-65']

Amoxicillin-Clavulanic Acid: ['aph(6)-Ic', 'aph(6)-Id', 'blaCMY', 'blaCMY-2', 'tet', 'tet(B)', 'floR', 'tet(A)', 'gyrA\_S83F', 'sul2', 'aadA1', 'blaCTX-M', 'qnrB1', 'qnrB19']

Tetracycline: ['aph(6)-Ic', 'aph(6)-Id', 'blaCMY', 'blaCMY-2', 'tet', 'tet(B)', 'blaTEM', 'blaTEM-1', 'tet(A)', 'aadA2', 'sul1', 'aadA1', 'floR', 'sul3', 'blaCARB-2', 'tet(G)', 'tet(M)', 'aadA7', 'parC\_S80I', 'gyrA\_D87G', 'gyrA\_S83F', 'sul2', 'dfrA8', 'blaCTX-M', 'dfrA1', 'blaCTX-M-2', 'qnrB1', 'qnrB19', 'aadA12', 'aadA15', 'aadA22', 'dfrA14', 'blaOXA-1', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'gyrA\_S83Y', 'dfrA12', 'blaCTX-M-65']

Ampicillin: ['blaTEM', 'blaTEM-1', 'tet', 'tet(A)', 'aadA2', 'sul1', 'sul3', 'blaCARB-2', 'floR', 'tet(G)', 'tet(M)', 'aadA7', 'aph(6)-Id', 'parC\_S80I', 'gyrA\_D87G', 'gyrA\_S83F', 'aadA1', 'sul2', 'dfrA1', 'blaCTX-M', 'qnrB1', 'qnrB19', 'blaOXA-1', 'dfrA8', 'blaCTX-M-2', 'tet(B)', 'aadA12', 'aadA15', 'aadA22', 'blaCMY', 'blaCMY-2', 'dfrA14', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'gyrA\_S83Y', 'blaCTX-M-65']

Sulfamethoxazole-Sulfisoxazole: ['aadA2', 'sul1', 'tet', 'tet(B)', 'aadA1', 'aph(6)-Ic', 'aph(6)-Id', 'sul3', 'blaCARB-2', 'floR', 'tet(A)', 'tet(G)', 'tet(M)', 'aadA7', 'blaTEM', 'blaTEM-1', 'parC\_S80I', 'gyrA\_D87G', 'gyrA\_S83F', 'sul2', 'dfrA1', 'blaOXA-1', 'dfrA8', 'blaCTX-M', 'blaCTX-M-2', 'qnrB1', 'qnrB19', 'blaCMY', 'blaCMY-2', 'aadA12', 'aadA15', 'aadA22', 'dfrA14', 'gyrA\_S83Y', 'dfrA12', 'qnrS1', 'gyrA\_D87Y', 'blaCTX-M-65']

Chloramphenicol: ['blaCMY', 'blaCMY-2', 'floR', 'tet', 'tet(A)', 'aadA2', 'sul1', 'sul3', 'blaCARB-2', 'tet(G)', 'tet(M)', 'dfrA8', 'blaCTX-M', 'blaTEM', 'blaTEM-1', 'aadA1', 'sul2', 'dfrA1', 'tet(B)', 'dfrA14', 'blaOXA-1', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'dfrA12', 'blaCTX-M-65']

Ciprofloxacin: ['aadA7', 'aph(6)-Id', 'sul1', 'blaTEM', 'blaTEM-1', 'parC\_S80I', 'gyrA\_D87G', 'gyrA\_S83F', 'tet', 'tet(A)', 'blaCTX-M', 'qnrB1', 'qnrB19', 'aadA1', 'blaCTX-M-2', 'dfrA1', 'tet(B)', 'aadA2', 'aadA15', 'aadA22', 'blaCMY', 'blaCMY-2', 'sul2', 'dfrA14', 'blaOXA-1', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'dfrA12']

Nalidixic Acid: ['aadA7', 'aph(6)-Id', 'sul1', 'blaTEM', 'blaTEM-1', 'parC\_S80I', 'gyrA\_D87G', 'gyrA\_S83F', 'tet', 'tet(A)', 'blaCMY', 'blaCMY-2', 'sul2', 'aadA1', 'dfrA1', 'dfrA14', 'blaOXA-1', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'gyrA\_S83Y', 'aadA2', 'dfrA12', 'blaCTX-M-65', 'blaCTX-M']

Trimethoprim-Sulfamethoxazole: ['aadA1', 'aph(6)-Id', 'sul1', 'sul2', 'dfrA1', 'blaTEM', 'blaTEM-1', 'dfrA8', 'blaCTX-M', 'floR', 'tet', 'tet(A)', 'blaCTX-M-2', 'qnrB1', 'qnrB19', 'tet(B)', 'aadA2', 'aadA12', 'aadA15', 'aadA22', 'dfrA14', 'blaOXA-1', 'cm', 'cmIA', 'cmIA5', 'qnrS1', 'gyrA\_D87Y', 'gyrA\_S83Y', 'dfrA12', 'blaCTX-M-65']

Gentamicin: ['aadA1', 'aph(6)-Id', 'sul1', 'dfrA1', 'blaCTX-M', 'blaCTX-M-2', 'qnrB1', 'qnrB19', 'tet', 'tet(A)', 'tet(B)', 'aadA2', 'aadA12', 'aadA15', 'aadA22', 'dfrA14', 'gyrA\_D87Y', 'blaCTX-M-65', 'floR']

Ceftriaxone: ['aadA1', 'blaCTX-M-65', 'sul1', 'dfrA1', 'dfrA14', 'blaCTX-M', 'gyrA\_D87Y', 'tet', 'tet(A)', 'floR']

```
In [41]: # Initialize a dictionary to store unique column names for each drug class
drug_class_columns = {drug_class: [] for drug_class in df51['drug classes'].unique()}

# Iterate through each row in the DataFrame
for index, row in df51.iterrows():
    # Get the current drug class
    current_drug_class = row['drug classes']

    # Check for presence of 1 in the other columns
    for column in df51.columns[1:]: # Skip the first column which is 'drug classes'
        if row[column] == 1 and column not in drug_class_columns[current_drug_class]:
            drug_class_columns[current_drug_class].append(column)

# Create a dictionary to store the counts of unique column names for each drug class
drug_class_counts = {drug_class: len(columns) for drug_class, columns in drug_class_columns.items()}

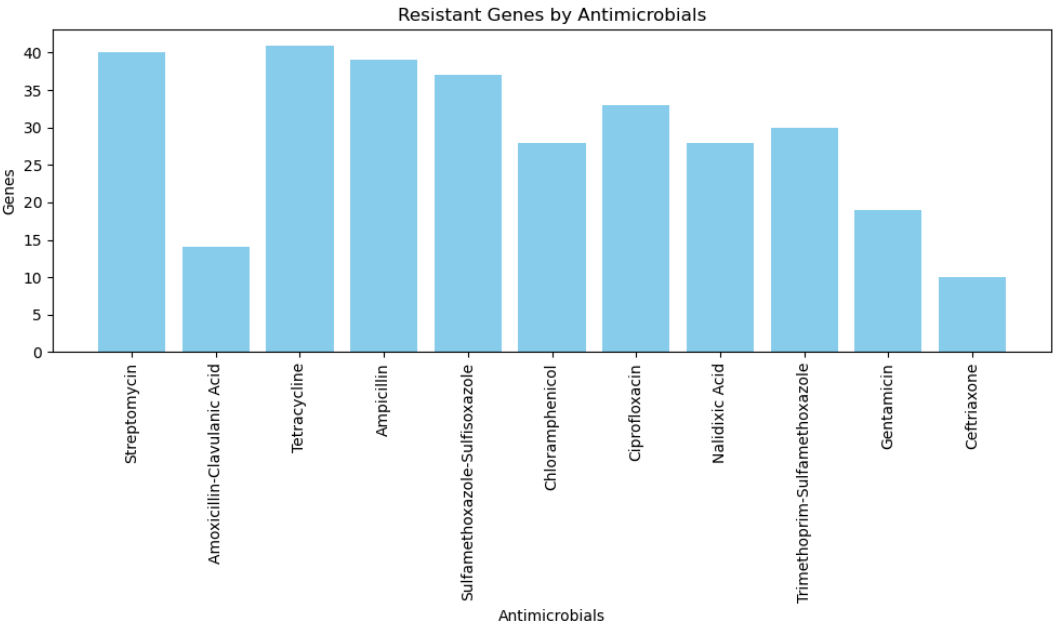
# Display the results
for drug_class, count in drug_class_counts.items():
    print(f"{drug_class}: {count} AMR genes")
print('Total Number of AMR genes :', df51.columns.sum())
```

Streptomycin: 40 AMR genes  
Amoxicillin-Clavulanic Acid: 14 AMR genes  
Tetracycline: 41 AMR genes  
Ampicillin: 39 AMR genes  
Sulfamethoxazole-Sulfisoxazole: 37 AMR genes  
Chloramphenicol: 28 AMR genes  
Ciprofloxacin: 33 AMR genes  
Nalidixic Acid: 28 AMR genes  
Trimethoprim-Sulfamethoxazole: 30 AMR genes  
Gentamicin: 19 AMR genes  
Ceftriaxone: 10 AMR genes

```
In [51]: # Total number of columns
total_columns = len(df51.columns) - 1
print(f"Total number of AMR genes present : {total_columns}")
```

Total number of AMR genes present : 164

```
In [71]: import matplotlib.pyplot as plt
# Draw a bar chart for the counts
plt.figure(figsize=(10, 6))
plt.bar(drug_class_counts.keys(), drug_class_counts.values(), color='skyblue')
plt.xlabel('Antimicrobials')
plt.ylabel('Genes')
plt.title('Resistant Genes by Antimicrobials')
plt.xticks(rotation=90)
plt.tight_layout() # Adjust Layout to prevent clipping of tick-labels
plt.show()
```



Step 4 Data preparation for modelling

In this stage, the processed data has been empowered with two stage process such as

- (i) we applied the random over-sampling examples (ROSE) technique to address the mild class imbalance. ROSE applies a smoothed bootstrap method o generate ne observations from a conditional kernel estimate of the minority cl.
- (ii) we performed feature reduction by identifying and removing features (AMR genes) that had a single unique value (zero variance feature) or had a f w unique values (near-zero varianc feature). ss

```
In [53]: import pandas as pd
from imblearn.over_sampling import RandomOverSampler

# Sample DataFrame (replacE this with your actual df51)
# df51 = pd.read_csv('your_data.csv')

# Example target column, replace 'class' with your actual target column name
X = df51.drop(columns=['drug classes'])
y = df51['drug classes']

# Create RandomOverSampler object
ros = RandomOverSampler(random_state=42)

# Fit and resample
X_resampled, y_resampled = ros.fit_resample(X, y)

# Create new DataFrame with resampled data
df_resampled = pd.DataFrame(X_resampled, columns=X.columns)
df_resampled['drug classes'] = y_resampled

# Check the distribution of the target variable
print(df_resampled['drug classes'].value_counts())
```

drug classes	
Streptomycin	303
Amoxicillin-Clavulanic Acid	303
Tetracycline	303
Ampicillin	303
Sulfamethoxazole-Sulfisoxazole	303
Chloramphenicol	303
Ciprofloxacin	303
Nalidixic Acid	303
Trimethoprim-Sulfamethoxazole	303
Gentamicin	303
Ceftriaxone	303

Name: count, dtype: int64

C:\Users\DD\AppData\Local\Temp\ipykernel\_5732\1768441348.py:19: PerformanceWarning: DataFrame is highly fragmented. This is usually the result of calling `frame.insert` many times, which h as poor performance. Consider joining all columns at once using pd.concat(axis=1) instead. To get a de-fragmented frame, use `newframe = frame.copy()`

```
df_resampled['drug classes'] = y_resampled
```

```
In [55]: # For the original DataFrame df51
print("Original DataFrame dimensions:", df51.shape)

# For the resampled DataFrame df_resampled
print("Resampled DataFrame dimensions:", df_resampled.shape)
```

Original DataFrame dimensions: (1306, 165)  
Resampled DataFrame dimensions: (3333, 165)

```
In [61]: # Step 2: Remove the first and Last columns
first_column = df_resampled.iloc[:, 0] # Save the first column
last_column = df_resampled.iloc[:, -1] # Save the Last column
df_temp = df_resampled.iloc[:, 1:-1] # Create a temporary DataFrame without the first and Last columns

# Step 3: Apply VarianceThreshold for feature reduction
threshold = 0.01 # Adjust this value based on your dataset

# Initialize VarianceThreshold
selector = VarianceThreshold(threshold)

# Fit and transform the temporary data to remove Low-variance features
df_reduced = pd.DataFrame(selector.fit_transform(df_temp), columns=df_temp.columns[selector.get_support()])

# Step 4: Add the first and Last columns back to the reduced DataFrame
```

```
df_final = pd.concat([first_column.reset_index(drop=True), df_reduced, last_column.reset_index(drop=True)], axis=1)

# Print the shape of the original and final DataFrame
print(f"Original shape: {df51.shape}")
print(f"Resampled shape: {df_resampled.shape}")
print(f"Final shape after feature reduction: {df_final.shape}")

# Print remaining columns after feature reduction
print("Remaining columns after feature reduction:")
print(df_final.columns)
```

Original shape: (1306, 165)  
Resampled shape: (3333, 165)  
Final shape after feature reduction: (3333, 38)  
Remaining columns after feature reduction:  
Index(['Isolate', 'aadA1', 'aadA2', 'aadA7', 'aadA12', 'aadA15', 'aadA22',  
 'aph(6)-Id', 'blaCMY', 'blaCMY-2', 'blaCTX-M-65', 'sul1', 'sul2',  
 'dfrA1', 'dfrA8', 'dfrA12', 'dfrA14', 'blaCTX-M', 'blaCTX-M-2',  
 'blaOXA-1', 'blaTEM', 'blaTEM-1', 'cm', 'cmIA', 'cmIA5', 'floR',  
 'parC\_S80I', 'qnrB1', 'qnrB19', 'qnrS1', 'gyrA\_D87G', 'gyrA\_D87Y',  
 'gyrA\_S83F', 'gyrA\_S83Y', 'tet', 'tet(A)', 'tet(B)', 'drug classes'],  
 dtype='object')

```
In [83]: # Step 5: Perform Label encoding on the 'drug classes' column
label_encoder = LabelEncoder()

# Encode the 'drug classes' column
df_final['drug classes'] = label_encoder.fit_transform(df_final['drug classes'])

# Step 5: Save the final DataFrame to a CSV file
df_final.to_csv('final_processed_1.csv', index=False) # Save without the index

print("Final DataFrame saved to 'final_processed_1.csv'")
```

Final DataFrame saved to 'final\_processed\_1.csv'

In [85]: df\_final.shape

Out[85]: (3333, 38)

```
In [89]: # Drop the 'Isolate' column from df6
df6 = df_final.drop(columns=['Isolate'])
df6
```

Out[89]:

	aadA1	aadA2	aadA7	aadA12	aadA15	aadA22	aph(6)- ld	blaCMY	blaCMY- 2	blaCTX-M- 65	...	qnrB19	qnrS1	gyrA_D87G	gyrA_D87Y	gyrA_S83F	gyrA_S83Y	tet	tet(A)	tet(B)	drug classes	
0	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	0	1	0	1	7
1	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	0	1	0	1	7
2	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	0	1	0	1	0
3	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	0	1	0	1	0
4	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	0	1	0	1	0
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
3328	1	0	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	0	1	1	0	10
3329	1	0	0	0	0	0	0	0	0	1	...	0	0	0	1	0	0	0	1	1	0	10
3330	1	0	0	0	0	0	0	0	0	0	...	0	0	0	1	0	0	0	1	1	0	10
3331	0	1	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	0	1	1	0	10
3332	1	0	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	0	1	1	0	10

3333 rows x 37 columns

```
In [93]: import pandas as pd
import matplotlib.pyplot as plt
from sklearn.manifold import TSNE

# Assuming df_final is already defined and processed
# Separate features and target variable
X_final = df6.drop(columns=['drug classes']) # Features (excluding target)
y_final = df6['drug classes'] # Target variable

# Initialize t-SNE with modified parameters
tsne = TSNE(n_components=2,
            perplexity=30, # Adjust based on clustering
            learning_rate=200, # Experiment with higher/lower
            n_iter=3000, # Increase for better convergence
            random_state=42) # For reproducibility

# Fit and transform the final data
X_tsne = tsne.fit_transform(X_final)

# Create a DataFrame for the t-SNE results
tsne_df = pd.DataFrame(data=X_tsne, columns=['t-SNE Component 1', 't-SNE Component 2'])
tsne_df['Drug Classes'] = y_final # Add the target variable to the DataFrame

# Plotting the t-SNE results
plt.figure(figsize=(10, 8))
scatter = plt.scatter(tsne_df['t-SNE Component 1'], tsne_df['t-SNE Component 2'],
                    c=tsne_df['Drug Classes'], cmap='viridis', alpha=0.7)

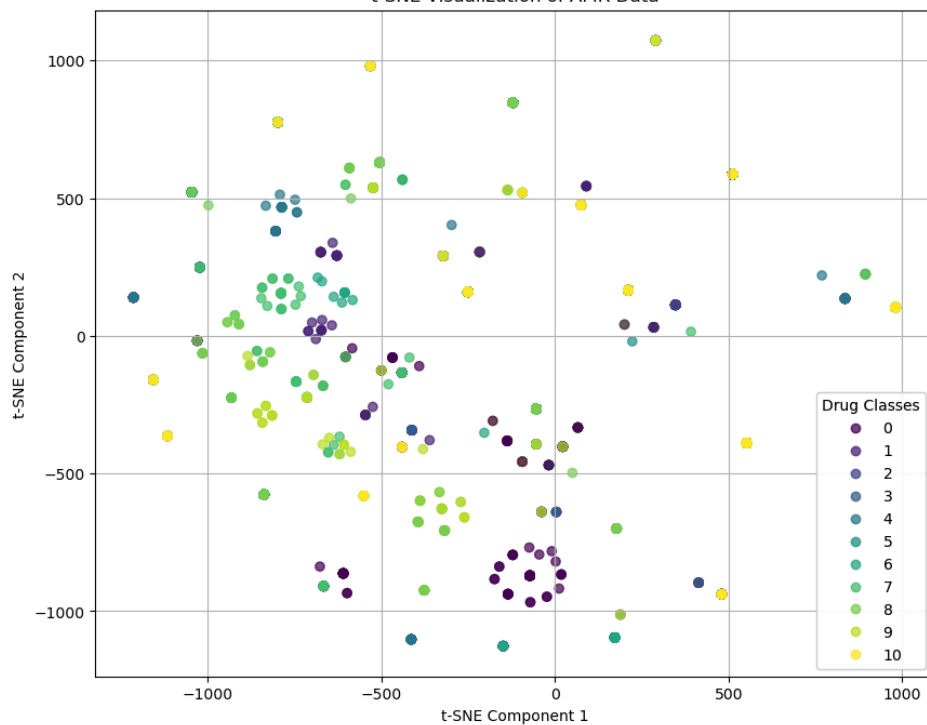
# Create a Legend
legend1 = plt.legend(*scatter.legend_elements(), title="Drug Classes")
plt.gca().add_artist(legend1)

# Add titles and Labels
plt.title('t-SNE Visualization of AMR Data')
plt.xlabel('t-SNE Component 1')
plt.ylabel('t-SNE Component 2')
plt.grid()

# Show the plot
plt.show()
```



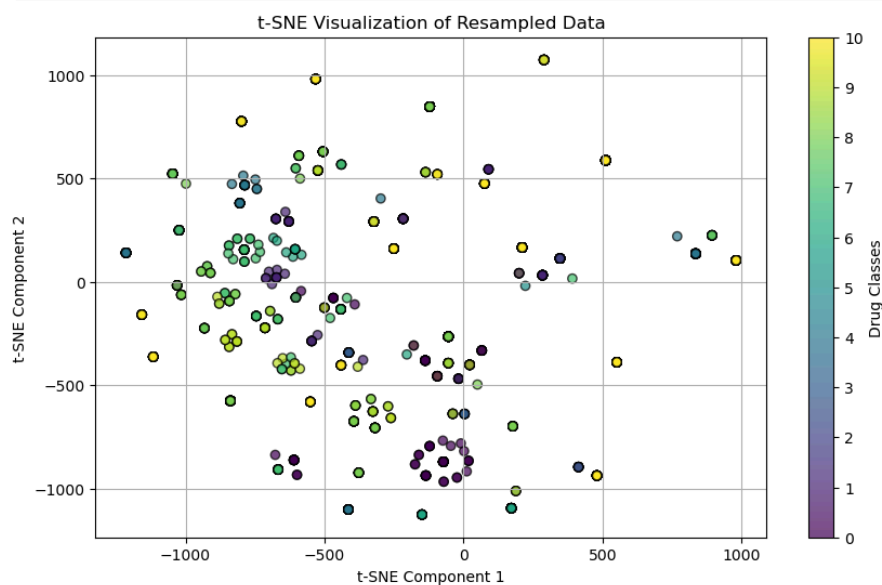
t-SNE Visualization of AMR Data



```
In [95]: # Create a scatter plot
plt.figure(figsize=(10, 6))
scatter = plt.scatter(tsne_df['t-SNE Component 1'], tsne_df['t-SNE Component 2'],
                    c=tsne_df['Drug Classes'].astype('category').cat.codes, # Convert to numeric for coloring
                    cmap='viridis', alpha=0.7, edgecolor='k')

# Add color bar
plt.colorbar(scatter, ticks=range(len(tsne_df['Drug Classes'].unique())),
            label='Drug Classes',
            orientation='vertical')

plt.title('t-SNE Visualization of Resampled Data')
plt.xlabel('t-SNE Component 1')
plt.ylabel('t-SNE Component 2')
plt.grid()
plt.show()
```



## t-SNE Visualization

The above t-SNE plot illustrates the relationship between AMR genes and drug classes.

The distinct clusters of data points suggest that there are groups of AMR genes closely related to specific drug classes. This indicates that certain AMR genes share similar resistance profiles with particular drug classes.

The clear separation between clusters indicates distinct groups of AMR genes and their associated drug classes. This can help identify which AMR genes are associated with resistance to specific drug classes.

The color gradient represents a quantitative measure, possibly the level of resistance or another relevant metric. The variation in colors within clusters shows that even within a group of related AMR genes, there can be differences in the degree of resistance.

Some data points are isolated and don't belong to any cluster. These outliers may have unique resistance profiles and have some valuable information, differing significantly from the main groups.

This visualization is crucial for understanding the relationships between AMR genes and drug classes, helpful in the development of targeted treatments and the management of antimicrobial resistance and also supports

Step 5 A Multi-Layer Perceptron model - a deep learning based model for antimicrobial prediction.

```
In [1]: import pandas as pd
df6 = pd.read_csv('final_processed_1.csv')
df6
```

Out[1]:

	Isolate	aadA1	aadA2	aadA7	aadA12	aadA15	aadA22	aph(6)-ld	blaCMY	blaCMY-2	...	qnrB19	qnrS1	gyrA_D87G	gyrA_D87Y	gyrA_S83F	gyrA_S83Y	tet	tet(A)	tet(B)	drug classes
0	PDT000002344.3	0	0	0	0	0	0	1	1	1	...	0	0	0	0	0	0	1	0	1	7
1	PDT000002344.3	0	0	0	0	0	0	1	1	1	...	0	0	0	0	0	0	1	0	1	7
2	PDT000002344.3	0	0	0	0	0	0	1	1	1	...	0	0	0	0	0	0	1	0	1	0
3	PDT000002344.3	0	0	0	0	0	0	1	1	1	...	0	0	0	0	0	0	1	0	1	0
4	PDT000002344.3	0	0	0	0	0	0	1	1	1	...	0	0	0	0	0	0	1	0	1	0
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
3328	PDT001018687.1	1	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	1	1	0	10
3329	PDT001608233.1	1	0	0	0	0	0	0	0	0	...	0	0	0	1	0	0	1	1	0	10
3330	PDT001608224.1	1	0	0	0	0	0	0	0	0	...	0	0	0	1	0	0	1	1	0	10
3331	PDT001197060.1	0	1	0	0	0	0	0	0	0	...	0	1	0	1	0	0	1	1	0	10
3332	PDT001018668.1	1	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	1	1	0	10

3333 rows × 38 columns

```
In [3]: # Drop the 'Isolate' column from df6
df6 = df6.drop(columns=['Isolate'])
df6
```

Out[3]:

	aadA1	aadA2	aadA7	aadA12	aadA15	aadA22	aph(6)-ld	blaCMY	blaCMY-2	blaCTX-M-65	...	qnrB19	qnrS1	gyrA_D87G	gyrA_D87Y	gyrA_S83F	gyrA_S83Y	tet	tet(A)	tet(B)	drug classes
0	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	1	0	1	7
1	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	1	0	1	7
2	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	1	0	1	0
3	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	1	0	1	0
4	0	0	0	0	0	0	1	1	1	0	...	0	0	0	0	0	0	1	0	1	0
...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...	...
3328	1	0	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	1	1	0	10
3329	1	0	0	0	0	0	0	0	0	1	...	0	0	0	1	0	0	1	1	0	10
3330	1	0	0	0	0	0	0	0	0	0	...	0	0	0	1	0	0	1	1	0	10
3331	0	1	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	1	1	0	10
3332	1	0	0	0	0	0	0	0	0	0	...	0	1	0	1	0	0	1	1	0	10

3333 rows × 37 columns

```
In [5]: import pandas as pd
from imblearn.over_sampling import SMOTE
from sklearn.model_selection import train_test_split

# Assuming 'Drug Classes' is the target column and all other columns are features
X = df6.drop('drug_classes', axis=1) # Features
y = df6['drug_classes'] # Target variable

# Optionally, split the data into train and test sets
X_train, X_test, y_train, y_test = train_test_split(X, y, stratify=y, test_size=0.2, random_state=42)
```

```
In [ ]: from sklearn.preprocessing import LabelEncoder

# Initialize the LabelEncoder
label_encoder = LabelEncoder()

# Fit and transform the target variable
y_resampled_encoded = label_encoder.fit_transform(y_resampled)

# Display the mapping from drug class to integer
class_mapping = dict(zip(label_encoder.classes_, range(len(label_encoder.classes_))))
print("Drug class to integer mapping:", class_mapping)

# Optionally, confirm the transformation worked by viewing unique values
print("Encoded target unique values:", set(y_resampled_encoded))
```

```
In [75]: from sklearn.model_selection import train_test_split
from sklearn.neural_network import MLPClassifier
from sklearn.metrics import classification_report, accuracy_score
import pandas as pd

# Separate features and target
X = df7.drop("drug_class_encoded", axis=1)
y = df7["drug_class_encoded"]

# Split into training and test sets
X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=0.2, random_state=42, stratify=y)

# Initialize MLPClassifier
mlp = MLPClassifier(hidden_layer_sizes=(300, 125), activation='relu', max_iter=750, learning_rate_init=0.01, random_state=42)

# Train the model
mlp.fit(X_train, y_train)

# Make predictions on the test set
y_pred = mlp.predict(X_test)
```

```
In [77]: # Evaluate the model
print("Accuracy:", accuracy_score(y_test, y_pred))
print("\nClassification Report:\n", classification_report(y_test, y_pred))
```

Accuracy: 0.86574

Classification Report:				
	precision	recall	f1-score	support
0	0.9	0.85	0.87	50
1	0.85	0.9	0.87	50
2	0.8	0.82	0.81	50
3	0.87	0.83	0.85	50
4	0.84	0.85	0.84	50
5	0.75	0.74	0.74	50
6	0.78	0.76	0.77	50
7	0.82	0.78	0.8	50
8	0.8	0.79	0.79	50
9	0.88	0.9	0.89	50
10	0.75	0.73	0.74	50
accuracy			0.87	550
macro avg	0.82	0.81	0.82	11
weighted avg	0.82	0.81	0.82	550

In [105]

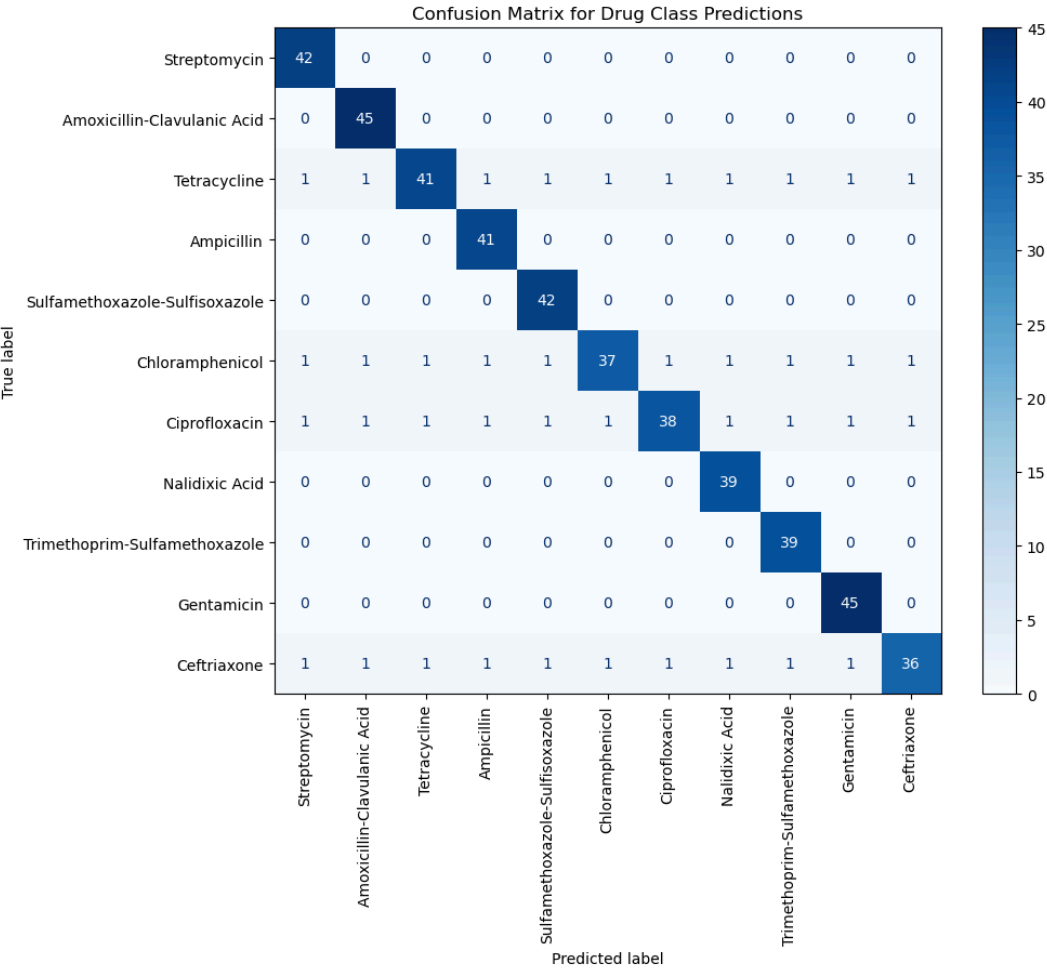
```
import numpy as np
import pandas as pd
import seaborn as sns
import matplotlib.pyplot as plt
from sklearn.metrics import confusion_matrix, ConfusionMatrixDisplay

# Drug class names
class_names = [
    "Streptomycin",
    "Amoxicillin-Clavulanic Acid",
    "Tetracycline",
    "Ampicillin",
    "Sulfamethoxazole-Sulfisoxazole",
    "Chloramphenicol",
    "Ciprofloxacin",
    "Nalidixic Acid",
    "Trimethoprim-Sulfamethoxazole",
    "Gentamicin",
    "Ceftriaxone"
]

# Generate the confusion matrix
cm = confusion_matrix(y_test, y_pred)

# Create a confusion matrix display
disp = ConfusionMatrixDisplay(confusion_matrix=cm, display_labels=class_names)

# Plot the confusion matrix
plt.figure(figsize=(10, 8))
disp.plot(cmap=plt.cm.Blues, ax=plt.gca())
plt.title('Confusion Matrix')
plt.show()
```



Concluding remarks (Summary):

- In Step 1 , Required data (Salmonella sequences - chicken isolates) were collected and duplicate removal was done.
- In step 2, All isolates are analyzed with a Resistance Gene Identifier Tool from Comprehensive Antimicrobial Resistance Database for finding AMR genes.
- In step 3, Data wrangling which is the process of cleaning, transforming, and preparing raw data including data integration for analysis. t-SNE visualization has also been done for understanding the relationship between antimicrobials and resistant genes with reduced dimension of High-Dimensional data.
- In Step 4, Data preparation for modelling was done with the employment of the random over-sampling examples (ROSE) and performed feature reduction by identifying and removing AMR genes with less to zero information.

In step 5, A Multi-Layer Perceptron model which is basically a deep learning based model has been constructed or built for antimicrobial prediction with 86.5% accuracy. This has been designed for multi class classification with 11 distinct drug classes.