



2. Graphics

**Ken Rice
Thomas Lumley**

Universities of Washington and Auckland

Seattle, July 2014

Important pre-takeoff announcement:

We are assuming you know;

- ... that graphics are useful! (and may be worth \leq 1000 words)
- How to make some simple plots e.g. making a scatterplot with `plot()`, adding to existing plots using `points()`, `lines()`, `text()`, and `legend()`
- That these functions can take many three letter arguments; `lwd`, `lty`, `pch` and many others, which can be looked up via `?par`
- That, ultimately, we want PDFs, JPEGs and other output formats – not just a window in an R session

Plotting large & high-dimensional data

'Simple' plots involve two-dimensional data, which we measure on the x and y axes.

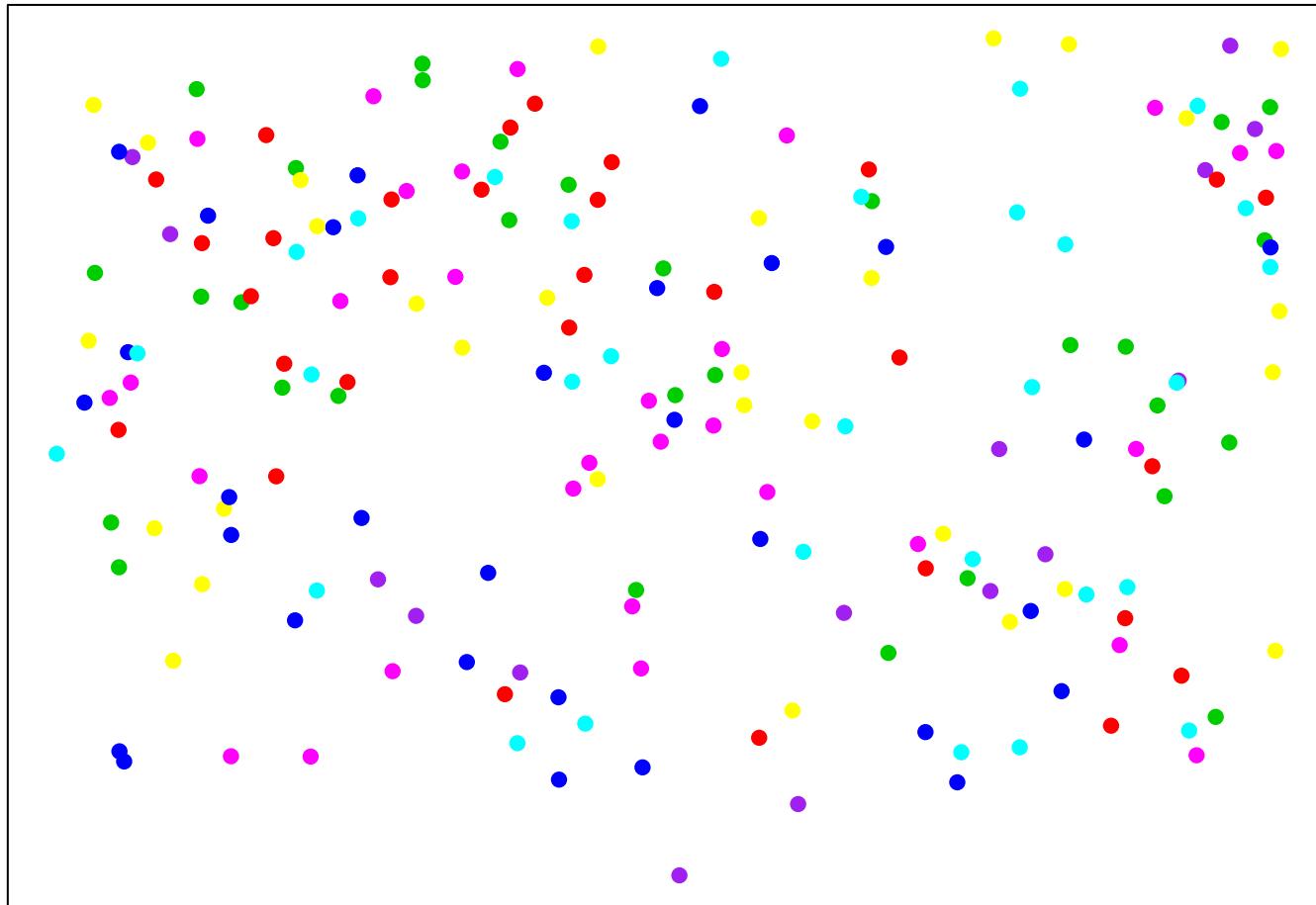
For higher-dimensions, some traditional approaches are;

- Different colors for e.g. men, women (`col`)
- Different-shaped symbols (`pch`), or different sizes (`cex`)

For ≤ 100 's of data points, **modest** use of these is fine. But your eye is not good at concentrating e.g. just on the purple points, in a fully Technicolor plot;

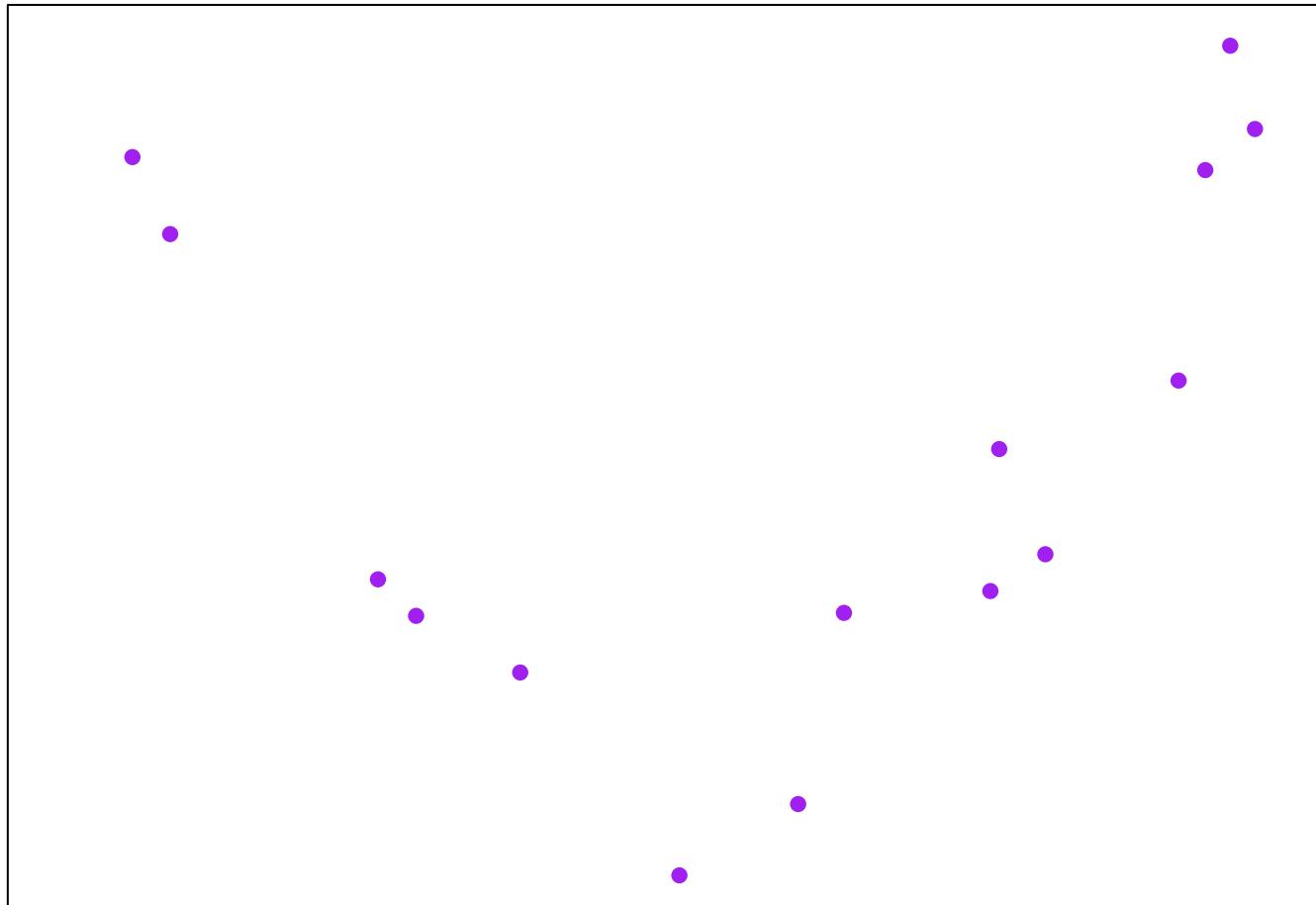
Plotting large & high-dimensional data

Some of these points are not like the others...



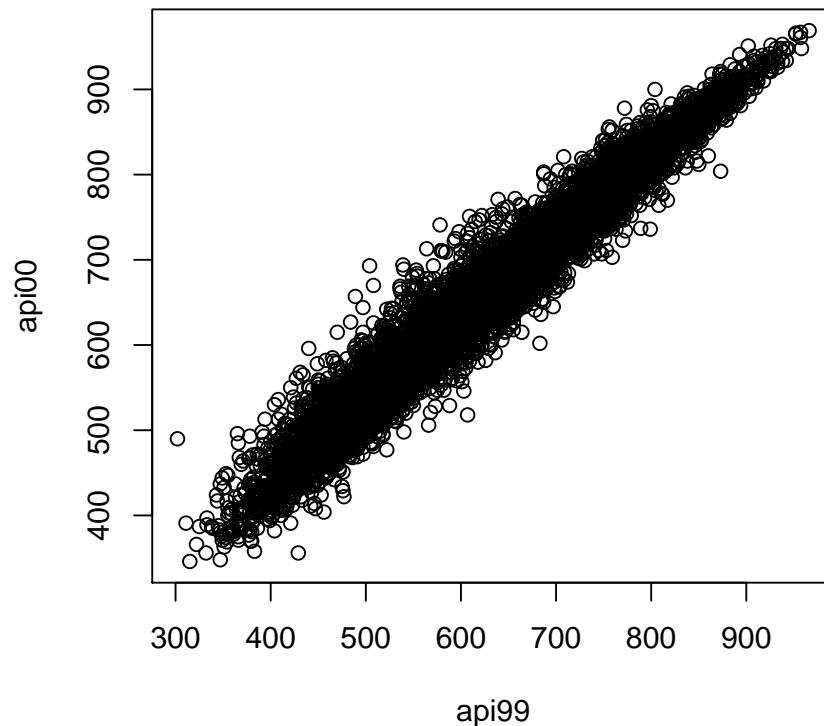
Plotting large & high-dimensional data

Some of these points are not like the others...



Plotting large & high-dimensional data

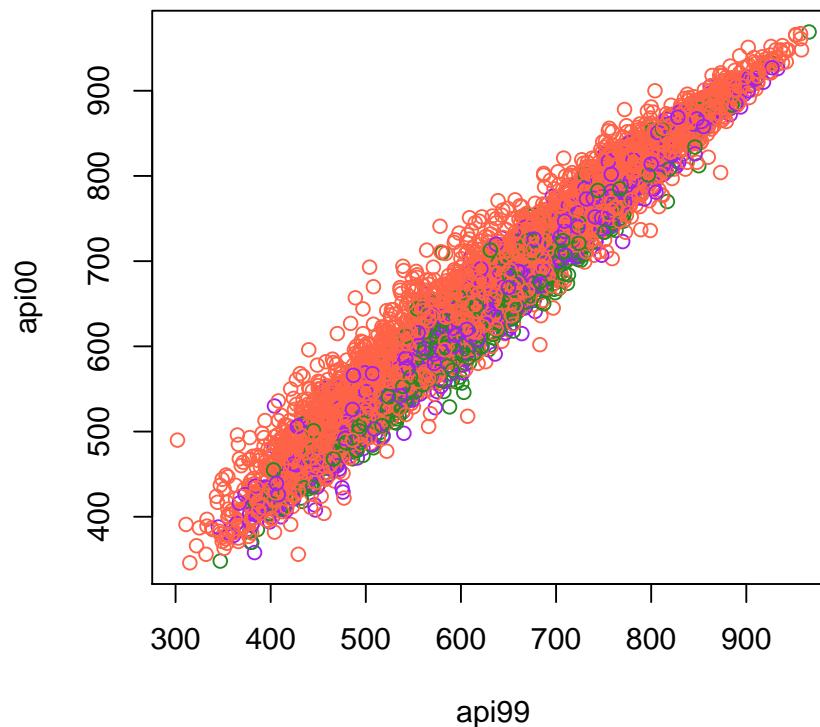
For large(ish) data, ‘overlap’ is a fundamental problem...



(California Academic Performance Index on 6194 schools)

Plotting large & high-dimensional data

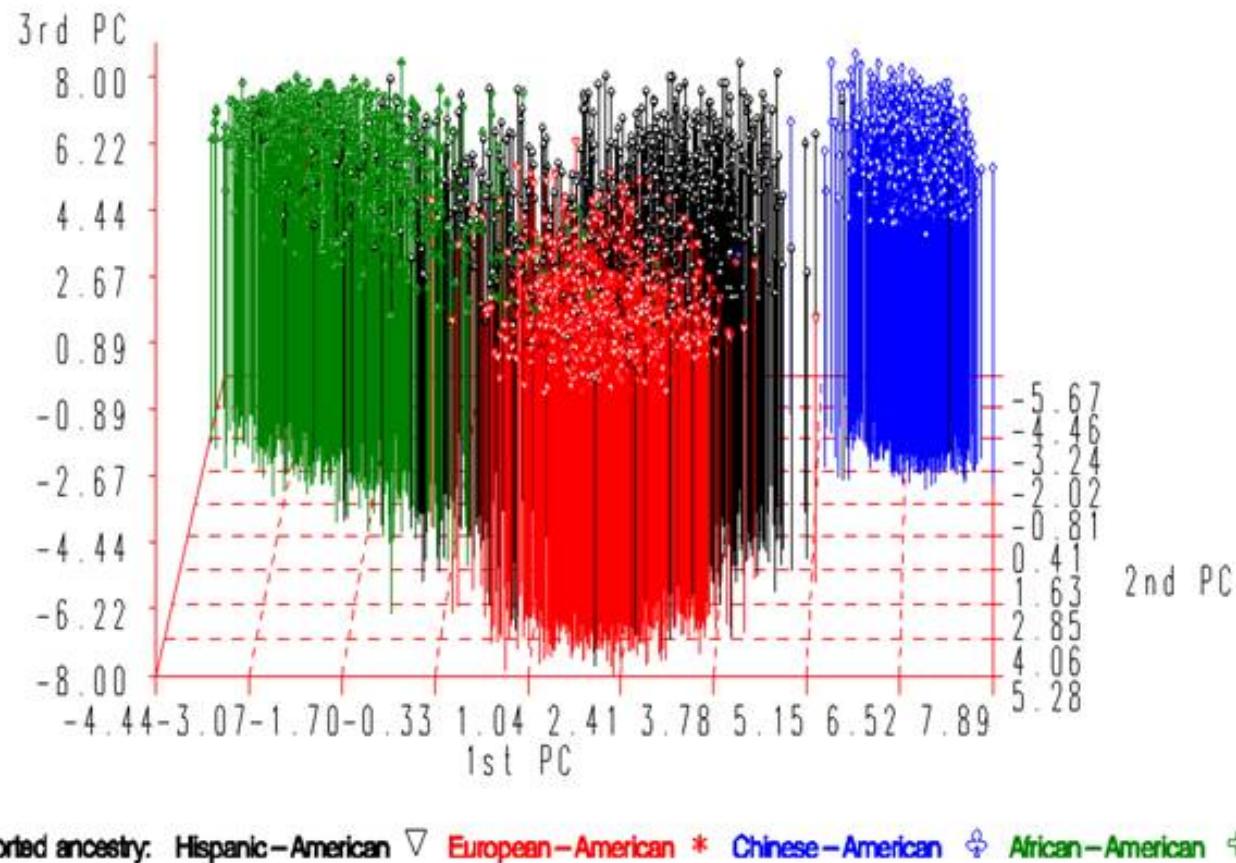
... which remains, when we color-code.



Colors denote Elementary, Middle & High Schools

Plotting large & high-dimensional data

With three dimensions + color-codes, this can happen:



(R does have `persp()`, for occasional use)

Conditioning plots

A typical goal for measuring Z is to see whether the $Y - X$ relationship changes at different values of Z . For example, we might want to see if a Blood Pressure/genotype association varies by Body Mass Index (weight/height²)

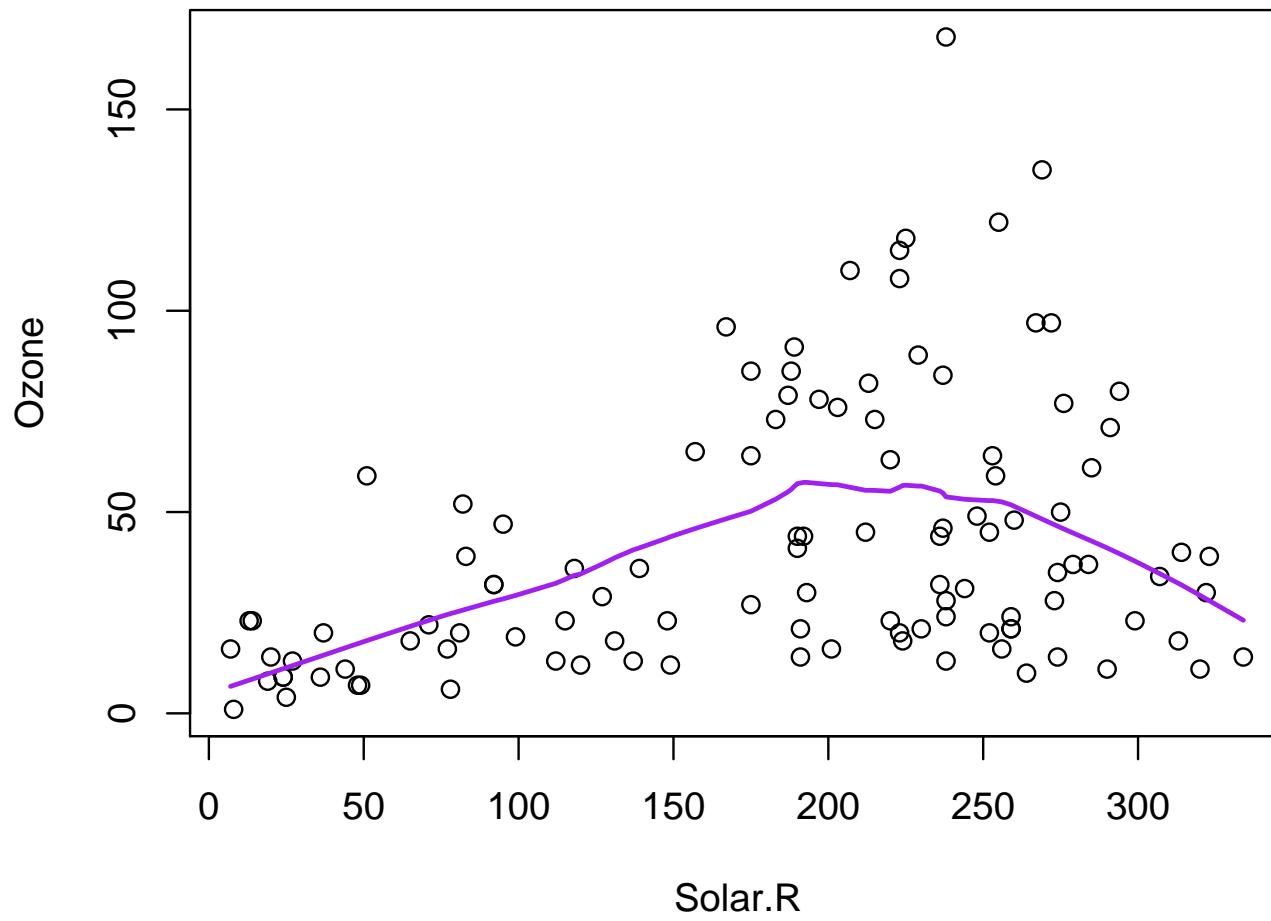
In this case, it's useful to show plots of Y against X conditioned on the value of Z , i.e. Y versus X for all data with Z in a small range. This is known as a **conditioning plot**, and can be produced with `coplot()`.

Conditioning plots

Ozone is a **secondary pollutant**, it is produced from organic compounds and atmospheric oxygen in reactions catalyzed by nitrogen oxides and powered by sunlight.

However, looking at ozone concentrations in NY in summer (Y) we see a non-monotone relationship with sunlight (X)

Conditioning plots

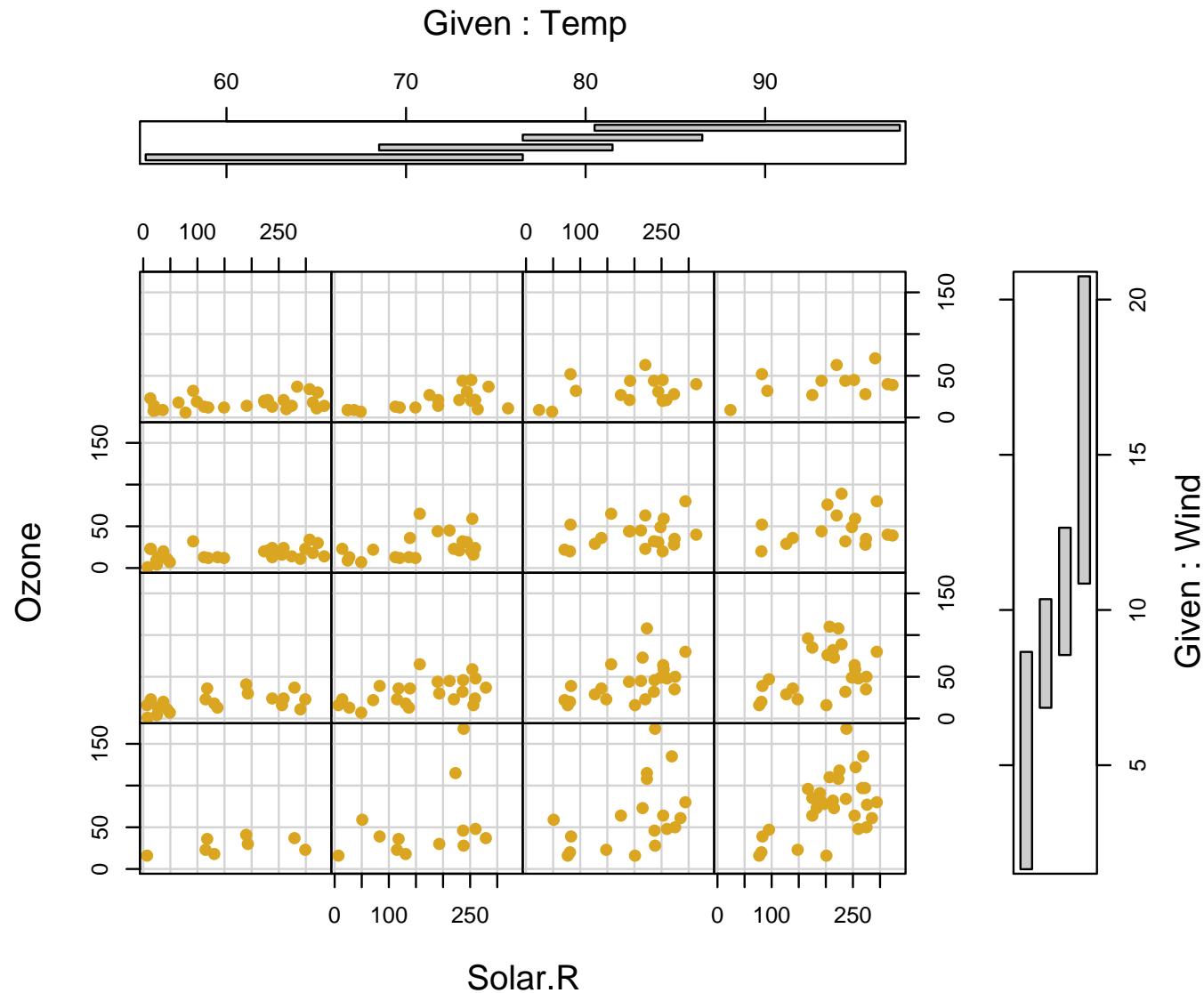


Conditioning plots

Here we draw a scatterplot of `Ozone` vs `Solar.R` for various subranges of `Temp` and `Wind`. For more examples like this, see the commands in the `lattice` package.

```
data(airquality)
coplot(Ozone ~ Solar.R | Temp * Wind, number = c(4, 4),
       data = airquality,
       pch = 21, col = "goldenrod", bg = "goldenrod")
```

Conditioning plots

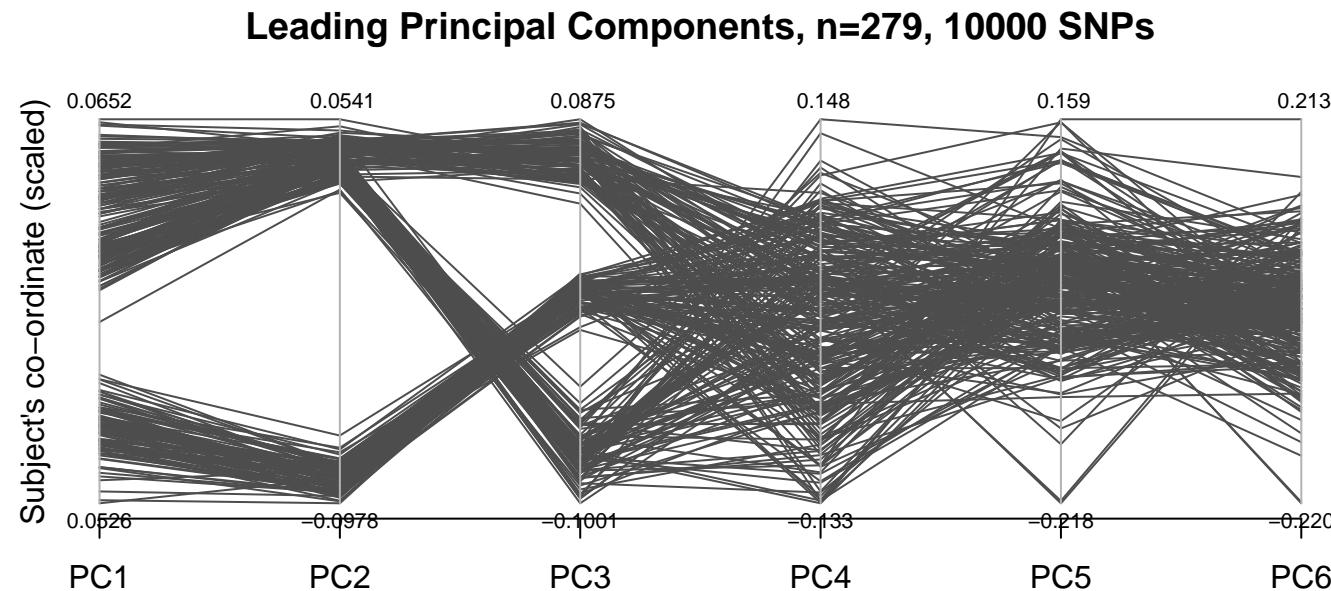


Conditioning plots

- A 4-D relationship is illustrated; the Ozone/sunlight relationship changes in strength depending on both the Temperature and Wind
- The vertical bar | is statistician-speak for ‘conditioning on’ (nb this is different to use of |’s meaning as Boolean ‘OR’)
- The horizontal/vertical ‘shingles’ tell you which data appear in which plot. The overlap can be set to zero, if preferred
- `coplot()`’s default layout is a bit odd; try setting `rows`, `columns` to different values
- For more plotting commands that support conditioning, see `library(help="lattice")`

Parallel Coordinate Plots

For even higher-dimensional data, scatterplots can not provide adequate summaries. For data where the dimensions can be ordered, the **parallel co-ordinates plot** is useful;

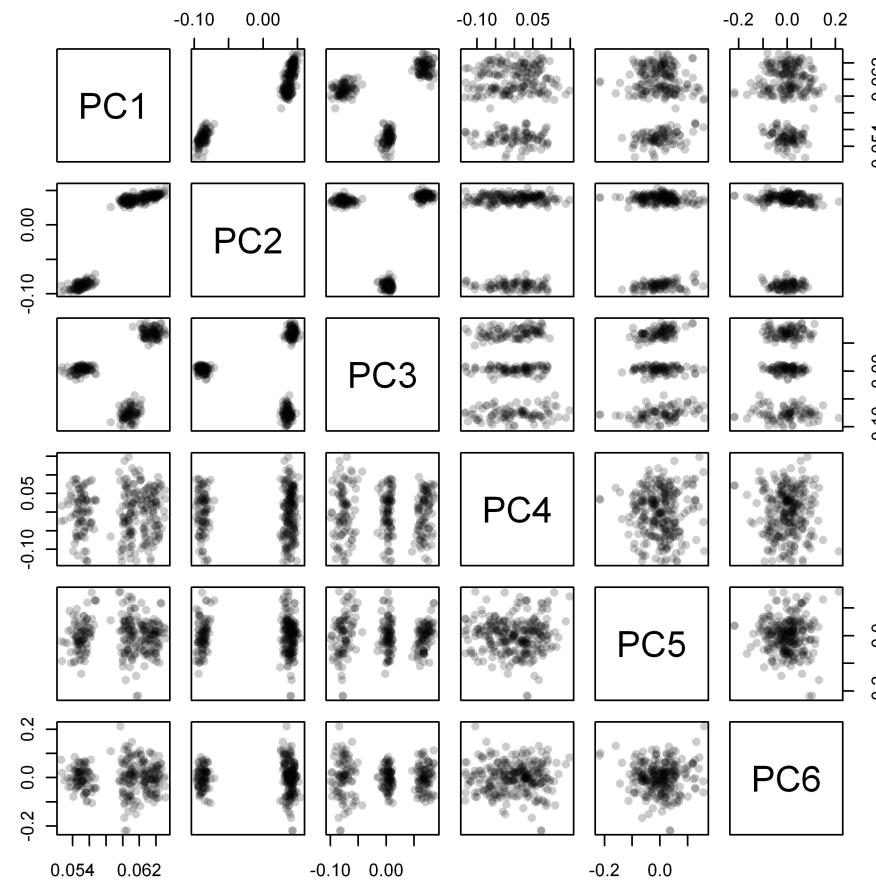


Parallel Coordinate Plots

- Each multi-dimensional data point (i.e. each person) is represented by a line – not a point
- `parcoord()` in the MASS package is one simple implementation – writing your own version is not a big job
- Coloring the lines also helps (example later)
- Scaling of axes, and their vertical positions are arbitrary
- Doing ‘Principal Components Analysis’ is just choosing axes for your data so that their variance is maximized on axis 1, then axis 2, ...

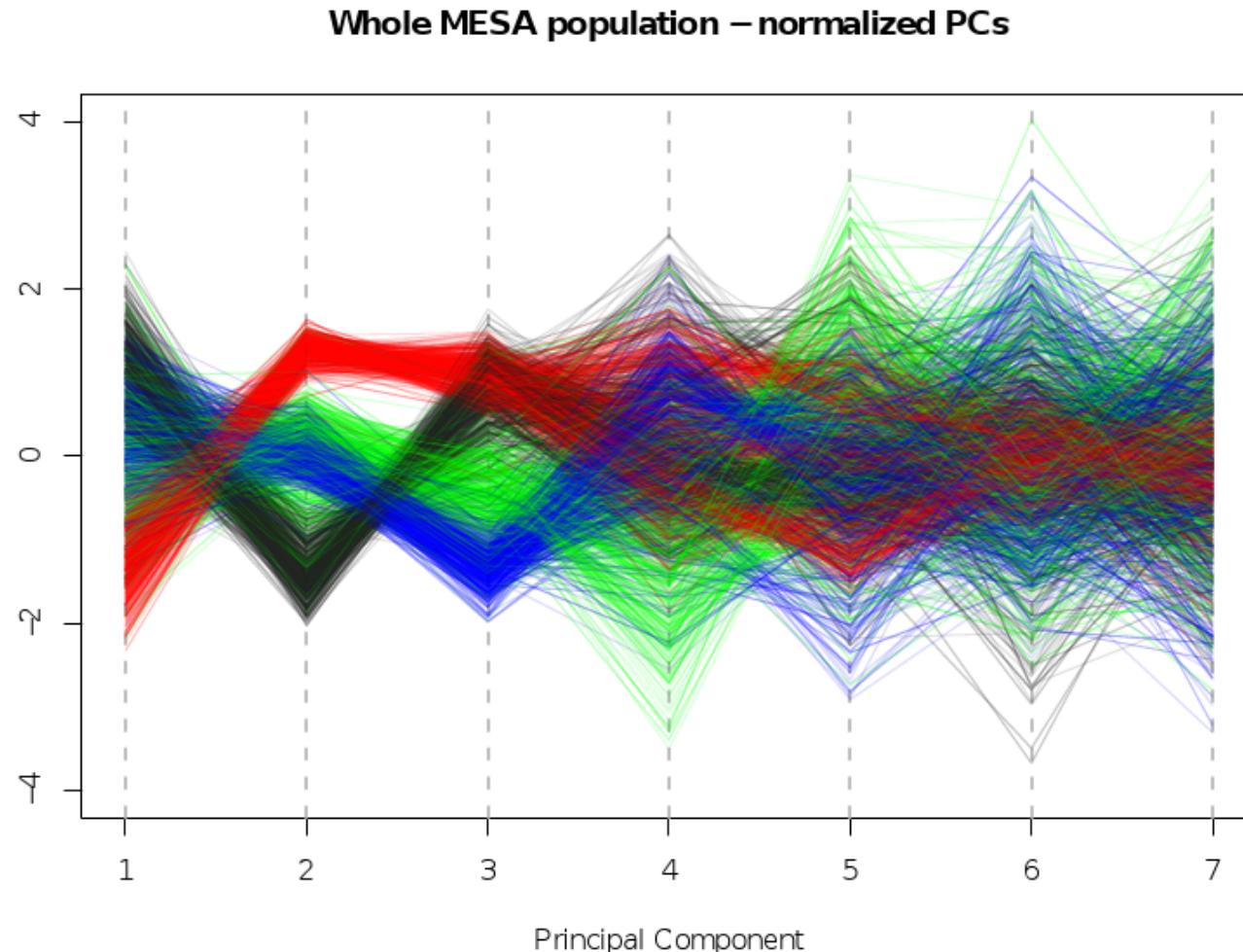
Parallel Coordinate Plots

A `pairs()` plot of the same thing; (nasty!)



Parallel Coordinate Plots

The pin cushion data++ : colors indicate self-report ancestry



Transparency

The colors in the last examples were **transparent**. As well as specifying e.g. `col=2` or `col="red"`, you can also specify

```
col="#FF000033"
```

– coded as RRGGBB in hexadecimal, with transparency 33 (also hexadecimal). This is a ‘pale’ red – $33/\text{FF} \approx 20\%$.

Get from color names to RGB with `col2rgb()`, and from base 10 to base 16 using `format(as.hexmode(11), width=2)`

Transparency

An example; (also shows other graphics commands)

```
curve(0.8*dnorm(x), 0, 6, col="blue", ylab="density", xlab="z")
curve(0.2*dnorm(x,3,2), 0, 6, col="red", add=T)

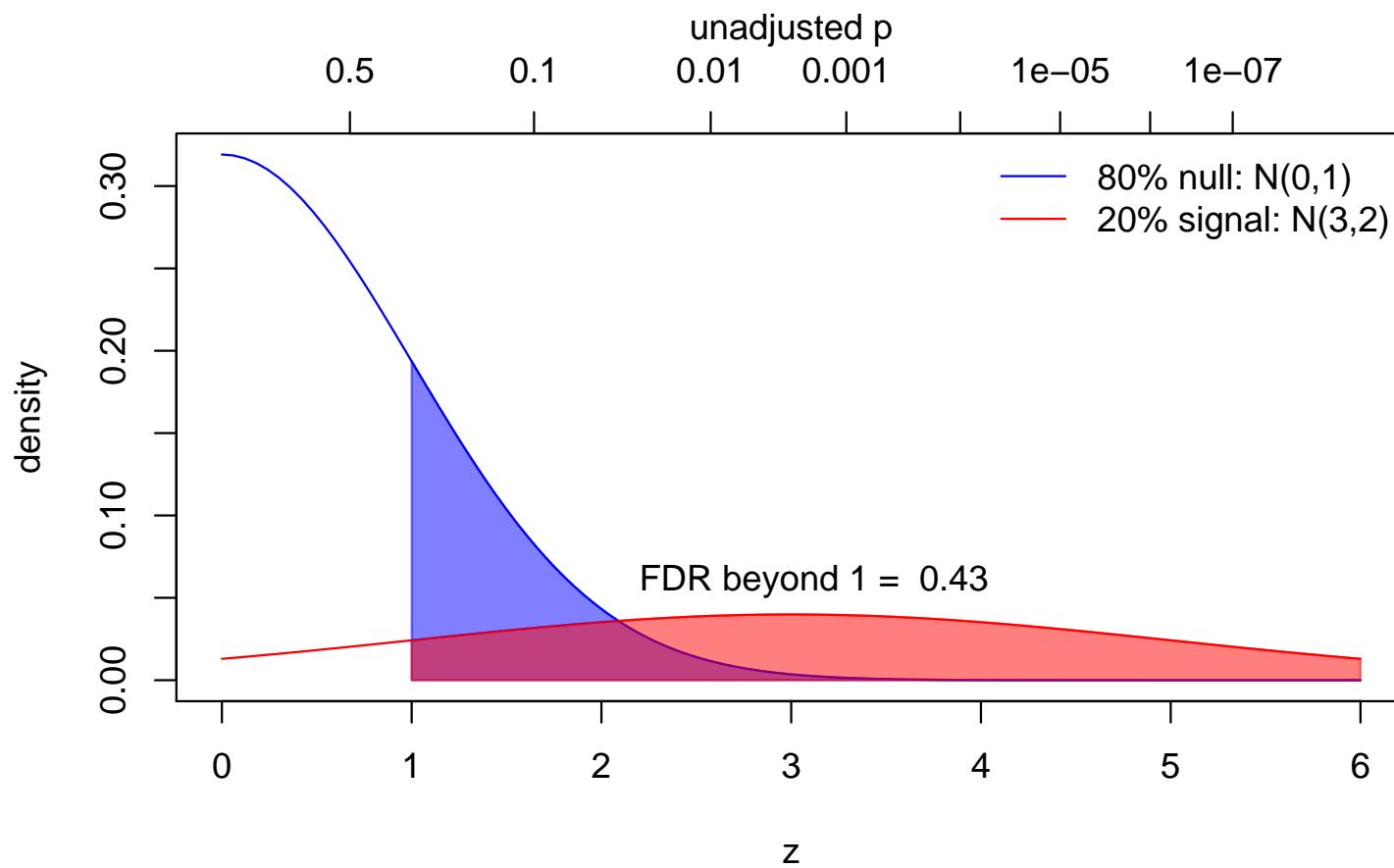
xvals <- seq(1, 6, l=101)
polygon(
c(xvals,6,1), c(0.8*dnorm(xvals), 0,0),
density=NA, col="#0000FF80" ) # transparent blue
polygon(
c(xvals,6,1), c(0.2*dnorm(xvals,3,2), 0,0),
density=NA, col="#FF000080" ) # transparent red

legend("topright", bty="n", lty=1, col=c("blue","red"),
c("80% null: N(0,1)", "20% signal: N(3,2)"))
axis(3, at=qnorm(c(0.25, 0.5*10^(-1:-7)), lower=F), c(0.5, 10^(-1:-7)) )
mtext(side=3, line=2, "unadjusted p")

text(2.2, 0.07, adj=c(0,1), paste("FDR beyond 1 = ",
round(0.8*pnorm(1,lower=F)/(0.8*pnorm(1,lower=F) + 0.2*pnorm(1,3,2,lower=F)),3)))
```

Transparency

Here's the output;



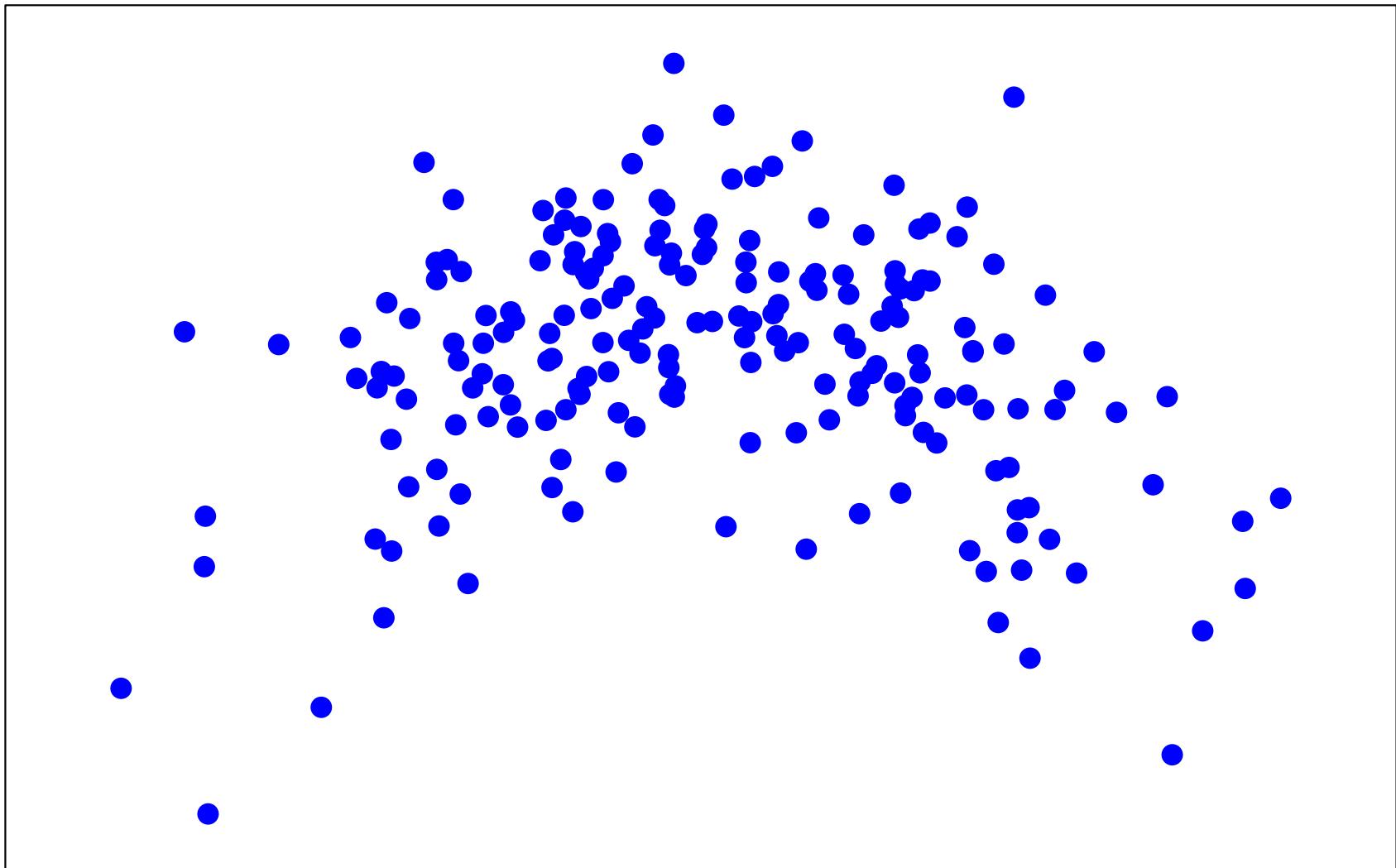
Hexagonal binning

Using transparent plotting symbols is a quick-and-dirty way to adapt scatterplots for use with large datasets.

A better method is ‘hexagonal binning’; this is a 2D analog of a histogram – where you would count the number of data in one area, and then draw a bar with height proportional to that count.

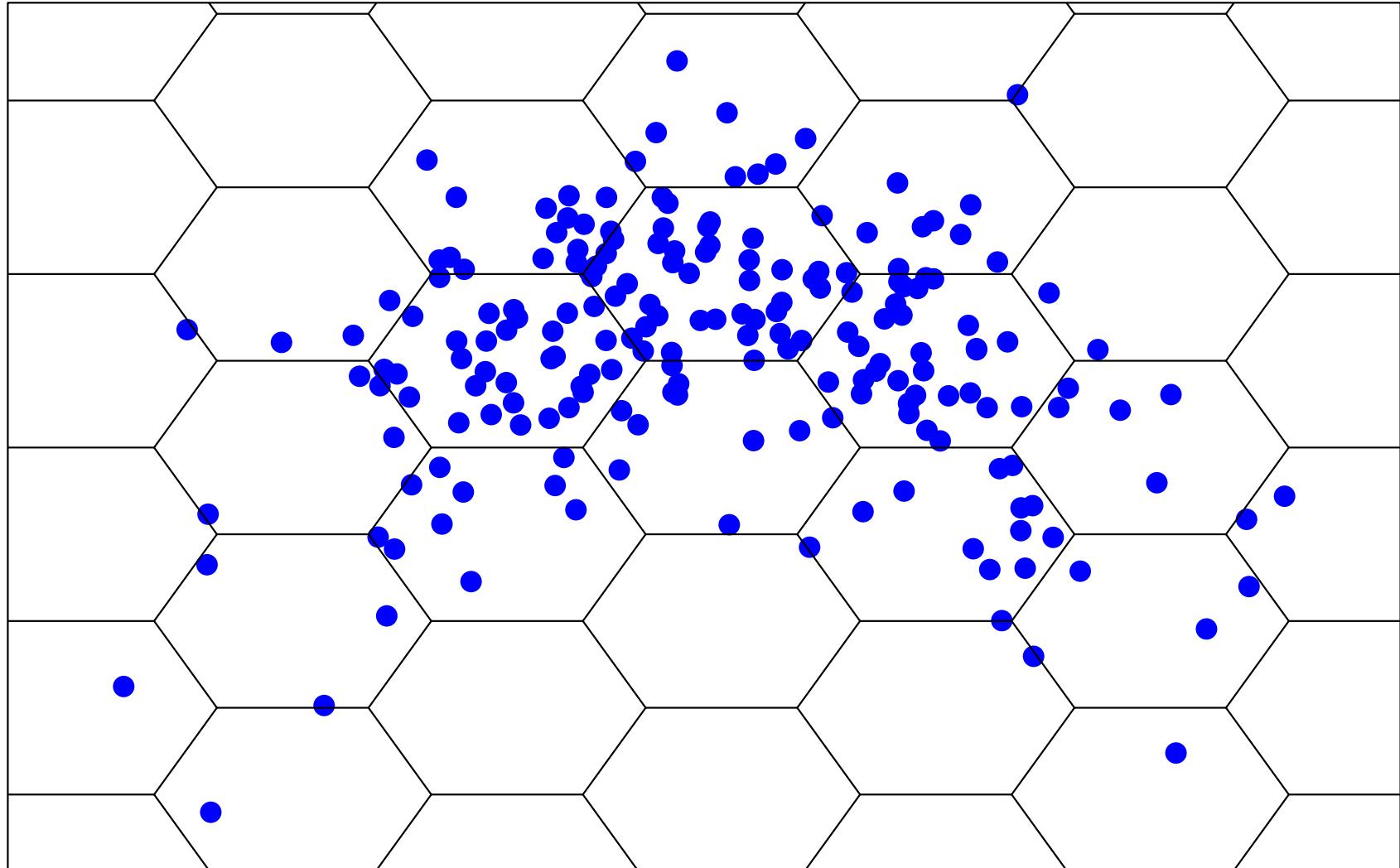
Hexagonal binning

Binning in two dimensions;



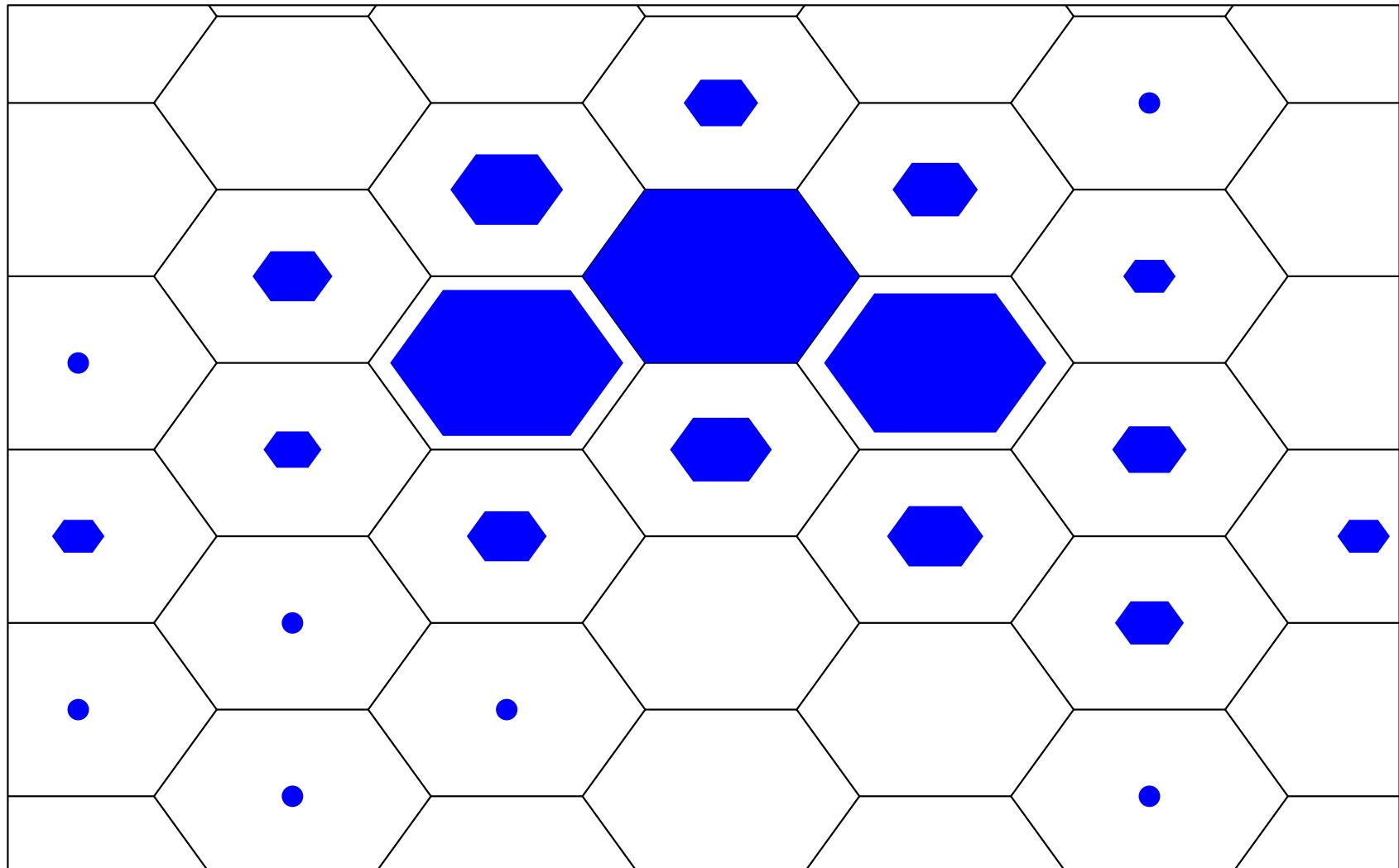
Hexagonal binning

Binning in two dimensions;



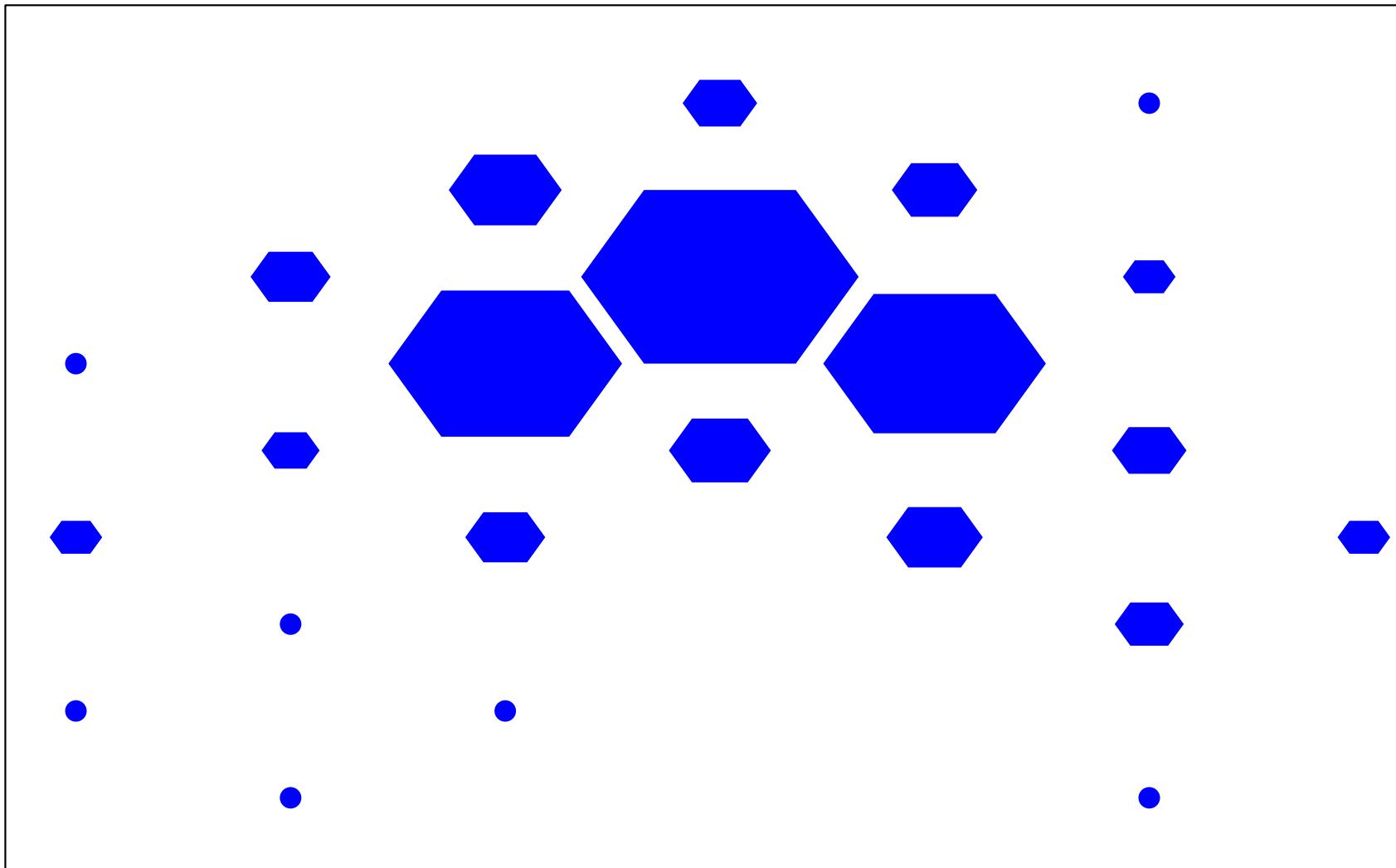
Hexagonal binning

Binning in two dimensions;



Hexagonal binning

Binning in two dimensions;



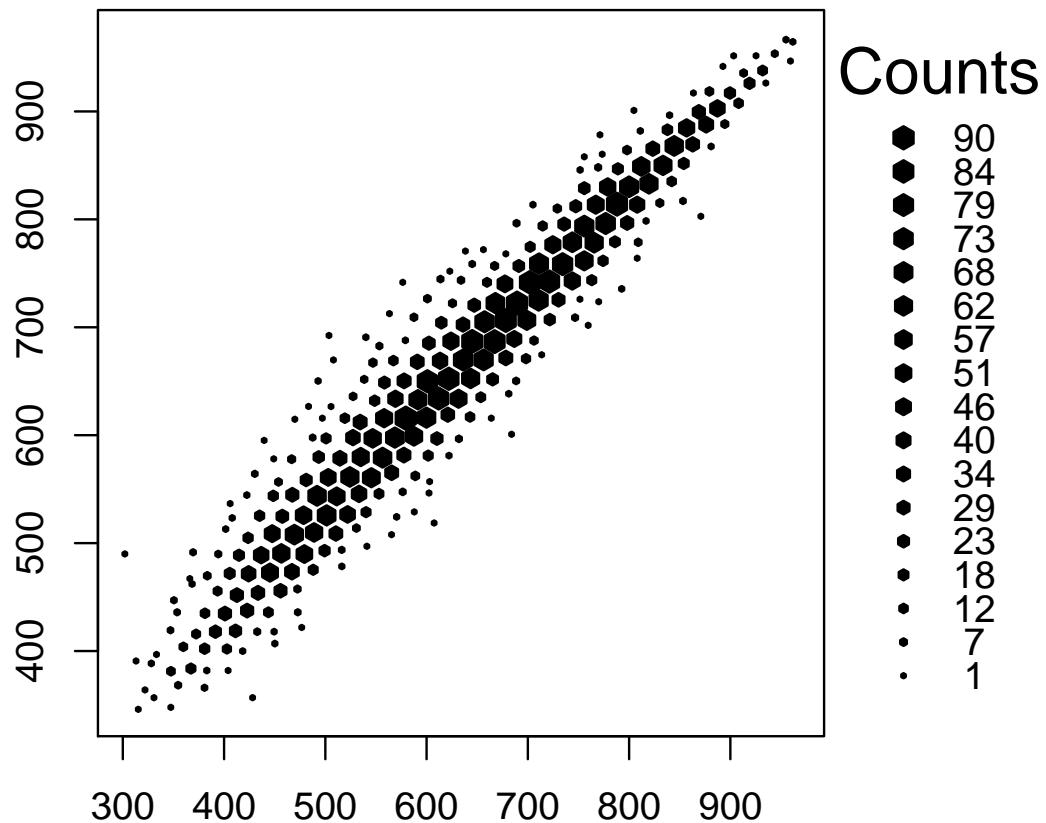
Hexagonal binning

The `hexbin()` package does all the bin construction, and counting. It has a plot method for its `hexbin` objects;

```
install.packages(c("hexbin", "survey"))
library("hexbin")
library("survey")# for apipop data frame

with(apipop, plot(hexbin(api99,api00), style="centroids"))
```

Hexagonal binning



Hexagonal binning

Hexbin is used when you don't *really* care about the exact location of every single point

- Singleton points are plotted 'as usual'; you do (perhaps) care about them
- hexbin centers the 'ink' at the cell data's 'center of gravity'
- `style="centroids"` gives the center-of-gravity version; the default `style` is `colorscale` – usually grayscale. See `?gplot.hexagons` for more options

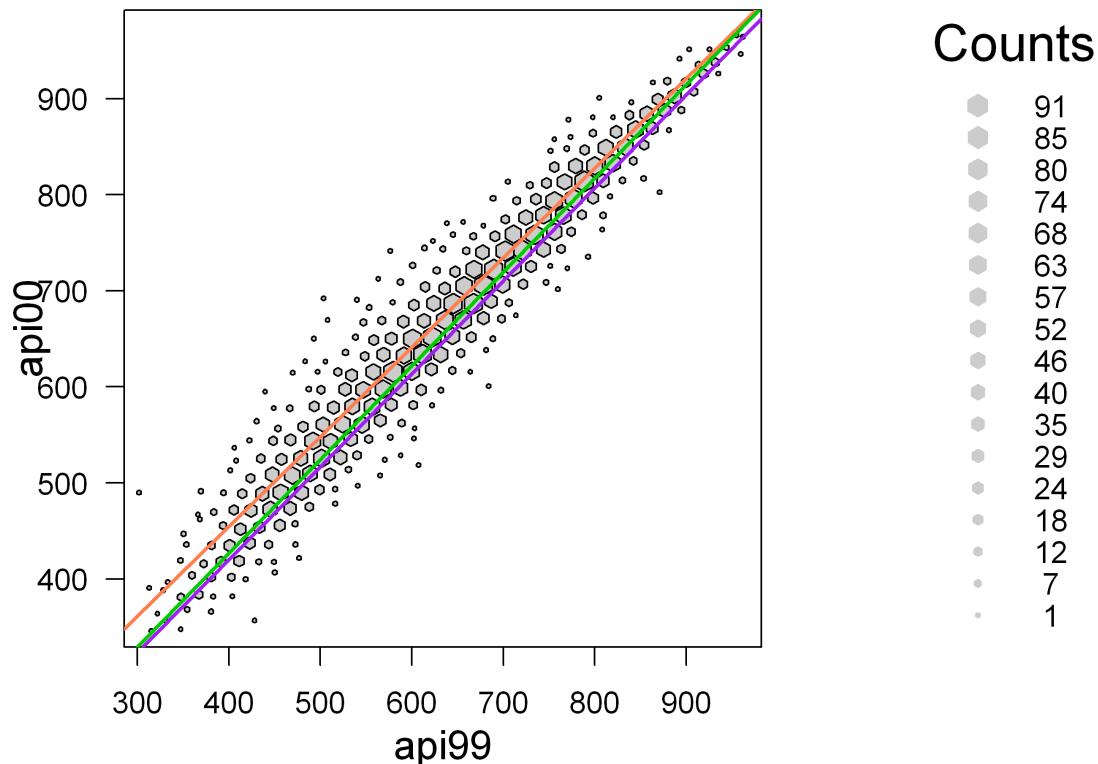
Hexagonal binning

For keen people: the `hexbin` package doesn't use the standard R graphics plotting devices; instead, it operates through the `Grid` system (in the `grid` package) which defines rectangular regions on a graphics device; these `viewport` regions can have a number of coordinate systems. To add lines to a hexbin plot, the options are;

- Use `hexVP.abline()` to add these directly
- Move everything into 'standard' graphics – not `Grid` graphics (see `?Grid`). The `Grid` system lets you alter graphics *after* plotting them
- Write your own plot method for `hexbin` objects, with standard R graphics commands
- Make do with `hexBinning()` in the `fMultivar` package

Hexagonal binning

An example; color-coded lines of best fit, by school type;



```
lm.e <- coef(lm(api00~api99, data=apiipop, subset=stype=="E"))
lm.m <- coef(lm(api00~api99, data=apiipop, subset=stype=="M"))
lm.h <- coef(lm(api00~api99, data=apiipop, subset=stype=="H"))

hexVP.abline(vp1$plot.vp, lm.e[1], lm.e[2], col="coral")
```

File formats

Ultimately, we want to output the graph in an appropriate file format. (Cut-and-paste is possible, but not recommended)

R knows more about font sizes and spacing than most users – so first design the graph at the size it will end up, eg:

```
## on Windows  
windows(height=4,width=6)  
## on Unix  
x11(height=4,width=6)
```

... and, when that's done, write a version to a file

File formats

For example, for a 6×4 PDF file;

```
pdf("myprettypic.pdf", height=4, width=6) # inches  
... plotting commands here ...  
dev.off()                      # close the file
```

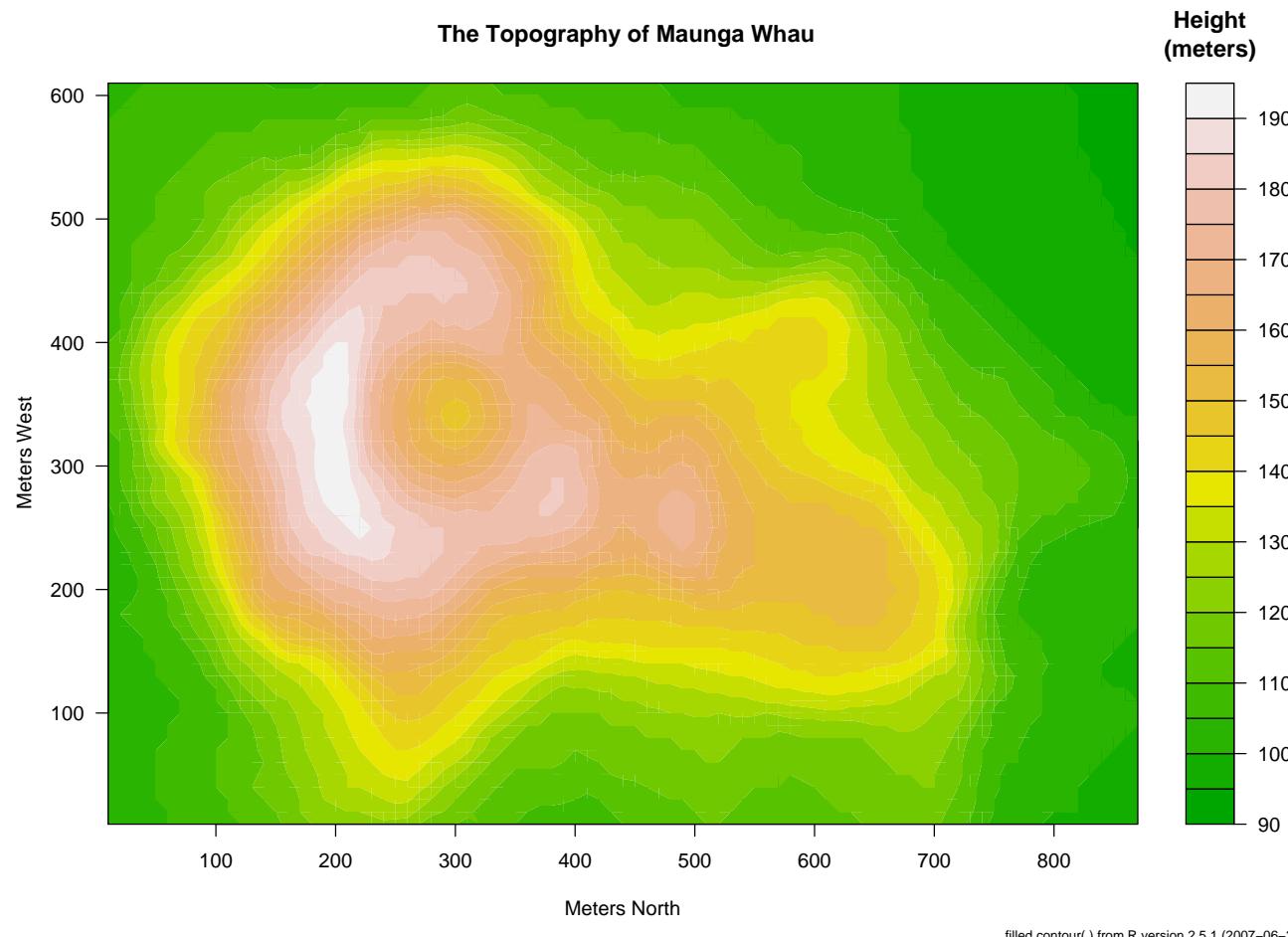
Some other formats: (see `?Devices` for a full list)

- `jpeg("mypic.jpg", w=6*288, h=4*288, res=288)` – lossy
 - `png("mypic.png", w=6*288, h=4*288, res=288)` – lossless
- point size of text can also be manipulated, which can be useful when making posters

PowerPoint, or Word, or L^AT_EX can all rescale graphs. But when the graph gets smaller, so do the axis labels...

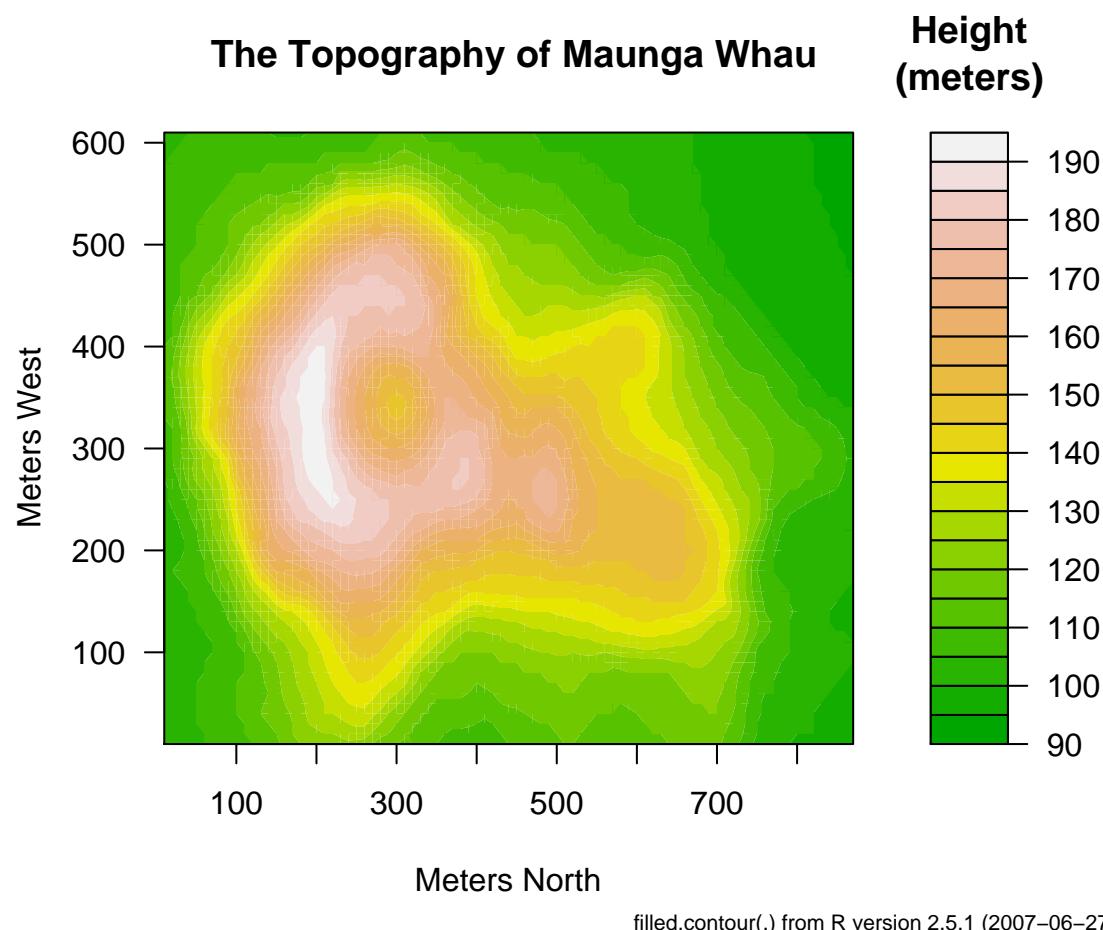
File formats

Created at full-page size (11×8.5 inches)



File formats

Created at 6×5 inches



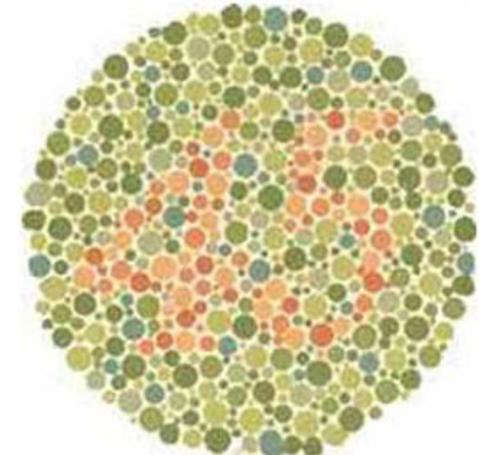
Color schemes

Color choice is best left to experts, or people with taste.

<http://www.colorbrewer.org> has color schemes designed for the National Cancer Atlas, also in package `RColorBrewer`

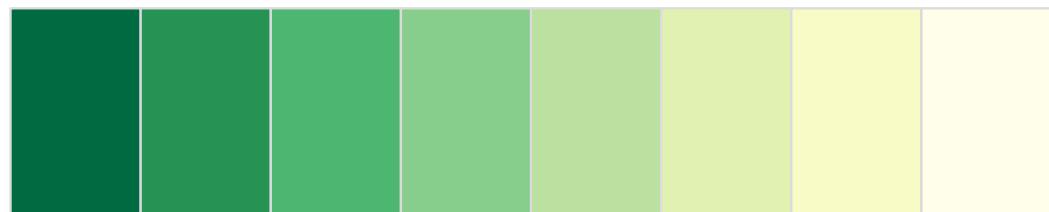
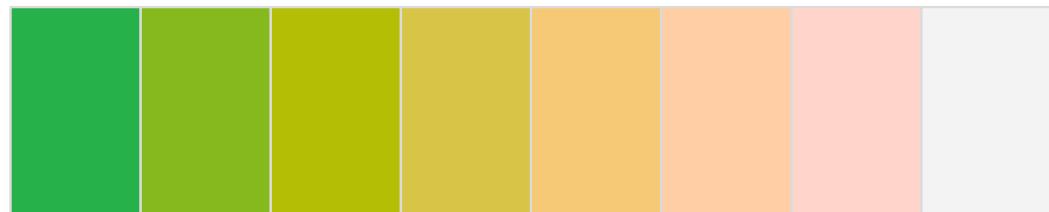
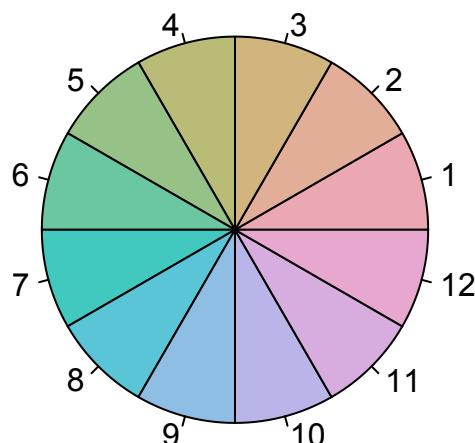
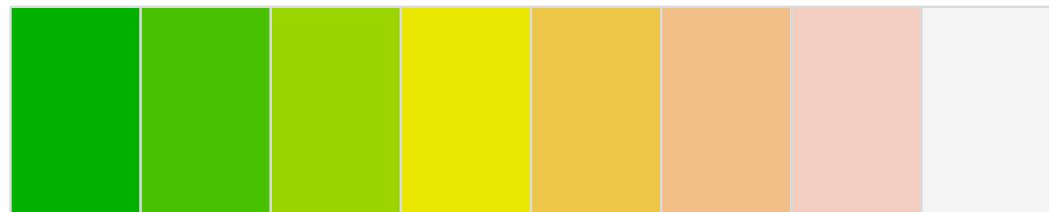
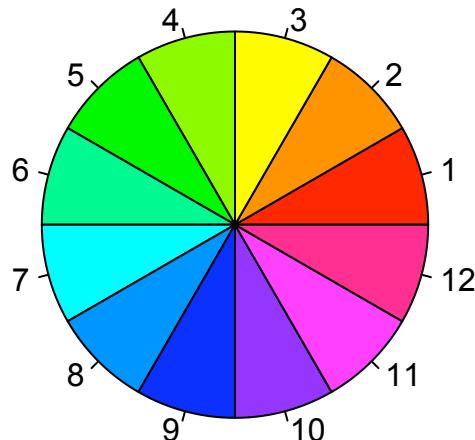
`colorspace` package has color schemes based on straight lines in a perceptually-based color space (rather than RGB).

`dichromat` package attempts to show the impact of red:green color blindness on your R color schemes.



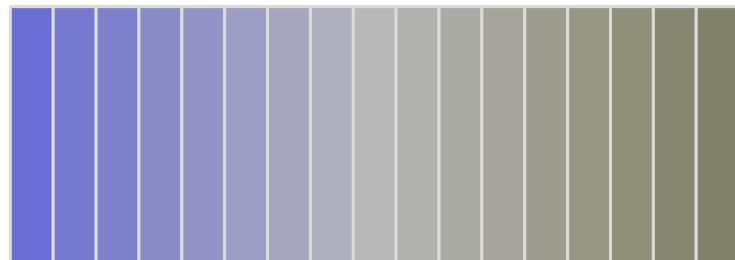
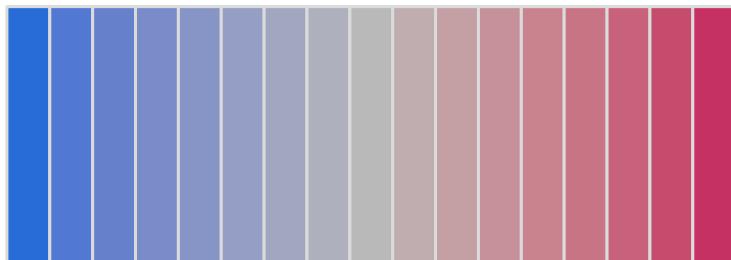
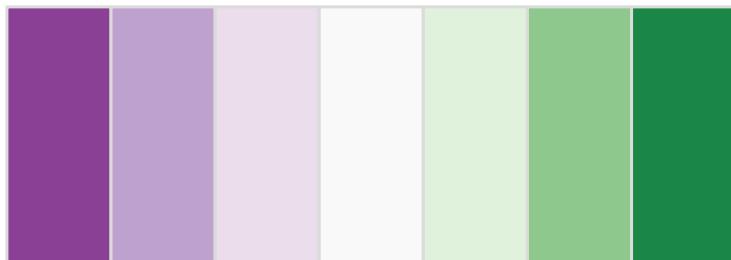
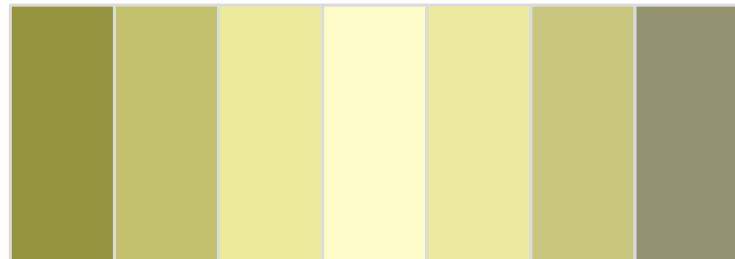
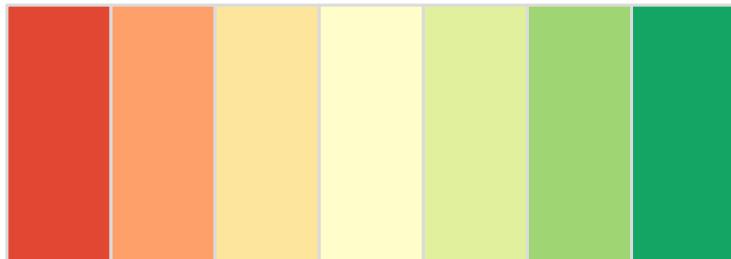
[Code for examples is in file `colorpalettes.R` on course website]

Color choice



(nb B&W printed copies of this slide may not be helpful!)

Color blindness



(nb B&W printed copies of this slide may not be helpful!)