Bradley's Notes on Doing Random Stuff with PDB Structures

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1 Creating Maps and Kinemages for a Given PDB

This is a rather easy thing to do. We will use phenix.fetch_pdb, phenix.maps and phenix.kinemage.

- 1. Fetch the PDB files (including SFs) from the PDB and create an SF mtz.
 - \$ phenix.fetch_pdb --mtz 10RN
- 2. Create kinemage.
 - \$ phenix.kinemage 10RN.pdb
- 3. Calculate $2mF_o$ - DF_c and mF_o - DF_c maps as well as map coefficients (needed for mmtbx.flipbase).
 - \$ phenix.maps map.map_type=2mFo-DFc map.map_type=mFo-DFc
 map_coefficients.map_type=2mFo-DFc 10RN.pdb 10RN.mtz

1.1 bash script

Here it is in a bash script.

Save this as get_pdb.sh. To run: \$ bash get_pdb.sh 10RN

2 DNA Stuff

2.1 Flip A Purine 180°

There is a tool within cctbx that can do this and is called *mmtbx.flipbase*. It works by flipping the given purine by 180° about the glycosidic bond and then doing 3 cycles of real-space refinement to bring the purine into the optimal place in the density. This means that you actually need density for this tool to work reasonably – no wishful flipping. As of today (11/8/2016) mmtbx.flipbase cannot handle ligands that require cif restraint files in refinement. This is super annoying because mmtbx.flipbase only refines the given residues and you should only be flippind canonical DNA bases (if you can and want to fix this issue everyone would appreciate it). The following steps are how to get arround this issue. If you don't have ligands that require cif restraints then you can run mmtbx.flipbase on the unmodified PDB (Step 3).

- 1. Copy the original file.
 - \$ cp 10RN.pdb 10RNNOHETS.pdb
- 2. Delete the HETATM records for the ligands (waters and metals ar OK to keep).
- 3. Run mmtbx.flipbase. It will create 1ORNNOHETS_flipbase.pdb. \$ mmtbx.flipbase 10RNNOHETS.pdb 10RN_map_coeffs.mtz chain=B res_num=1
- 4. Copy the original file.
 - \$ cp 10RN.pdb 10RN_flip.pdb
- 5. Replace the ATOM records for the purine of interest in 1ORN_flip.pdb with the ones in 1ORNNOHETS_flipbase.pdb.