

Nanocarriers for targeted drug delivery

Benefits and challenges of nanotechnology for medicine

Russell Maguire

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Durham University

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Nanocarriers

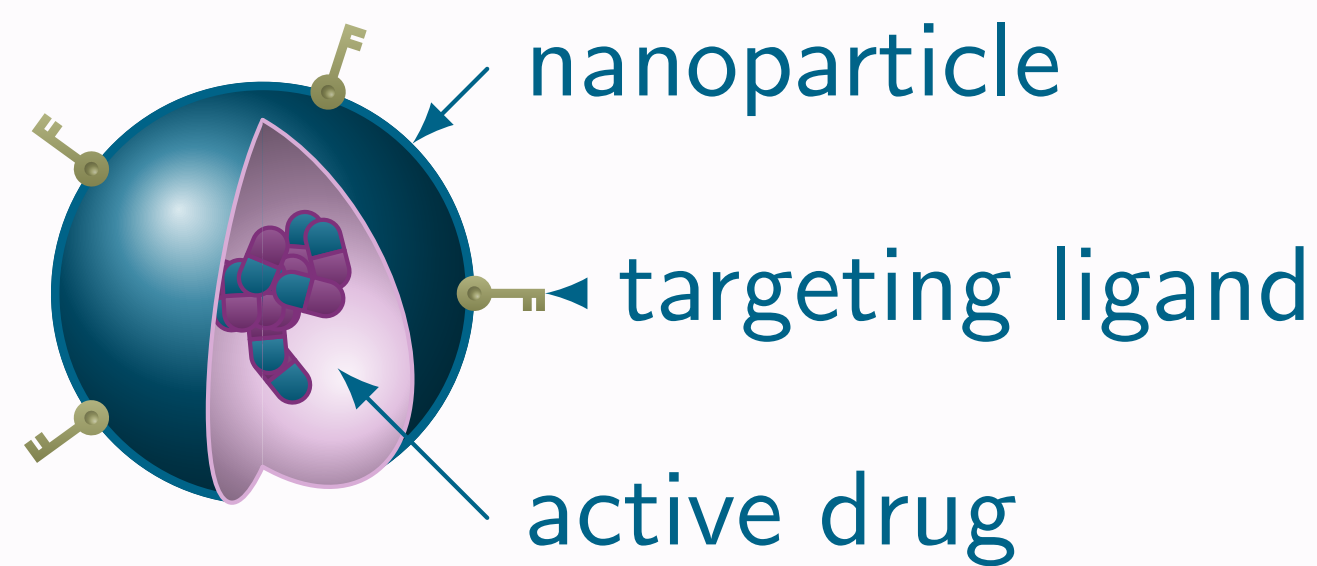


Figure 1: General structure of a *nanocarrier*, including the optional *active targeting ligands*.

A nanocarrier is a **biocompatible nanoparticle** encapsulating a drug, with size of the order 1 nm to 100 nm.

Nanocarriers provide a range of improvements over free drugs, including **reduced toxicity** and **immunogenic response**; **longer circulation time**; and **improved solubility**.

This is achieved using encapsulants with a hydrophilic surface; typically phospholipids or polymers which form stable **single layer micelles** and **bilayer liposomes** sus-

pended in water and blood. **Drug conjugates** are distinct in that the active drug is covalently bonded to a protein, polymer or antibody.

Polyethylene glycol (PEG) is a hydrophilic polymer which has been shown to mask immunogens such as the phospholipids used to build liposomes [1], resulting in a new class of nanoparticles—**immunoliposomes**.

However employing nanocarriers for drug delivery raises additional challenges: new **toxic side-effects**; **biodegradability**; the **cost** and **complexity** of formulation.

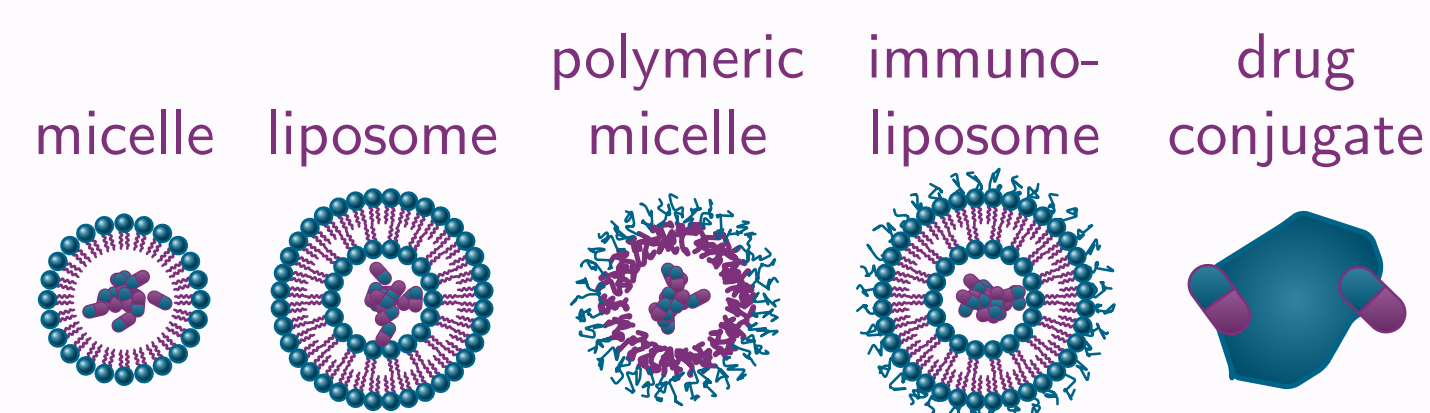


Figure 2: Structure of common types of organic nanocarriers
■ hydrophobic/lipophilic, ■ hydrophilic

Targeted drug delivery

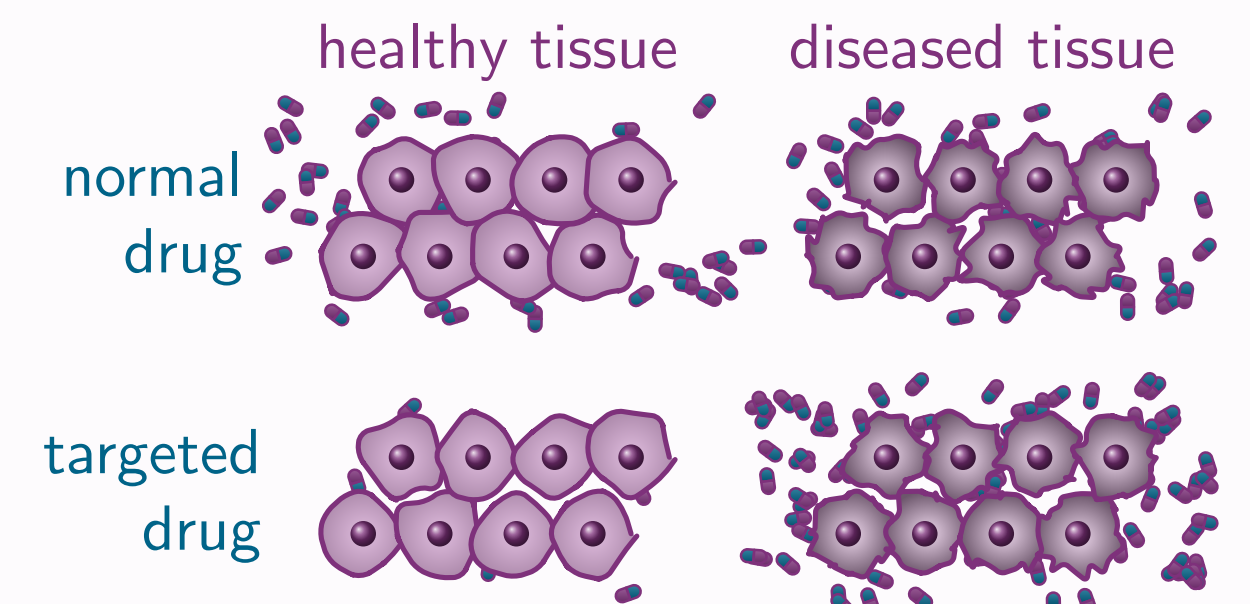


Figure 3: Effect of drug distribution in healthy and diseased tissue when using drugs targeted to a specific diseased tissue

The goal of targeted drug delivery is to **maximise therapeutic benefit** and **minimise side-effects** by increasing the concentration ratio of the active drug in the diseased tissue compared to healthy tissue.

The textbook example of targeted drug delivery is chemotherapy for tumour treatment.

Targeted drug delivery can be divided into three categories: first generation **passive targeting**, next generation **stimuli-responsive targeting** and **active targeting**.

Passive targeting

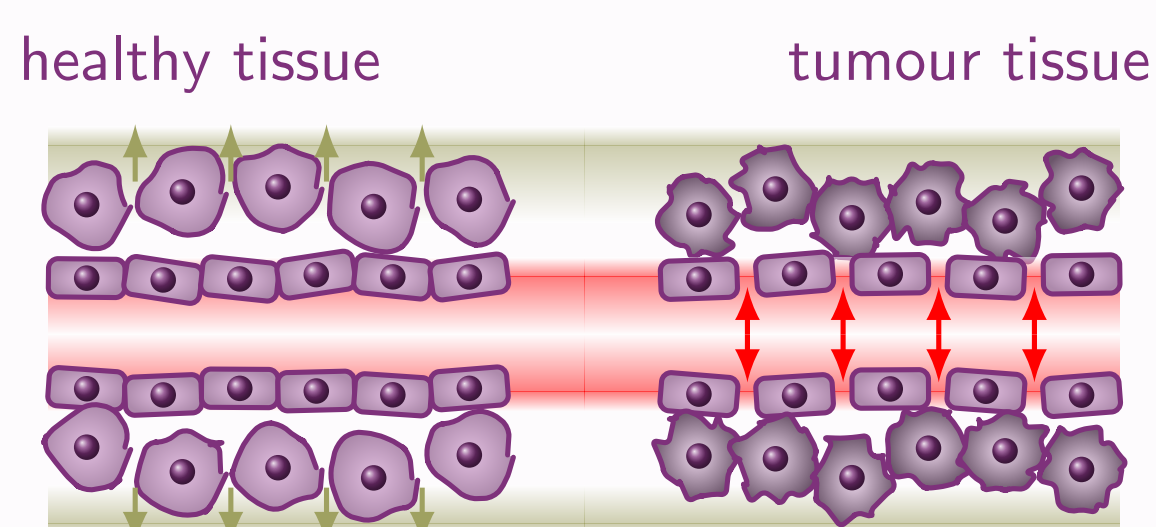


Figure 4: The differences between *lymphatic system drainage* and *blood vessel permeability* in healthy tissue and solid tumours

Passive targeting is dominated by the **enhanced permeability and retention effect (EPR effect)** whereby nanoscale particles can accumulate preferentially in solid tumours, first noticed by Maeda *et al.* [2].

First generation approved nanomedicines [3] rely on the EPR effect.

Permeable blood vessels in solid tumours let nutrients and nanoscale particles easily cross the endothelium. Padera *et al.* discovered rapidly proliferating cancer cells **compress lymph vessels** [4], retaining nanoparticles by preventing the tumour from easily draining.

However, limitations include **poor deep tumour penetration** and **ineffective small tumour targeting**. Jain and Stylianopoulos describe methods for overcoming this barrier [5].

Example

- **Doxin[®]/Caelyx[®]** was the first nanocarrier medicine, approved in 1995 for chemotherapy [6].
- **Doxorubicin** is the active compound, encapsulated in a **pegylated immunoliposome**.
- Benefits include **reduced cardiac side effects** and **increased circulation time** [3].

Stimuli-responsive targeting

Passively, nanocarriers are allowed to break down at the target site releasing the drug, but efficacy can be improved using **stimuli triggered release**.

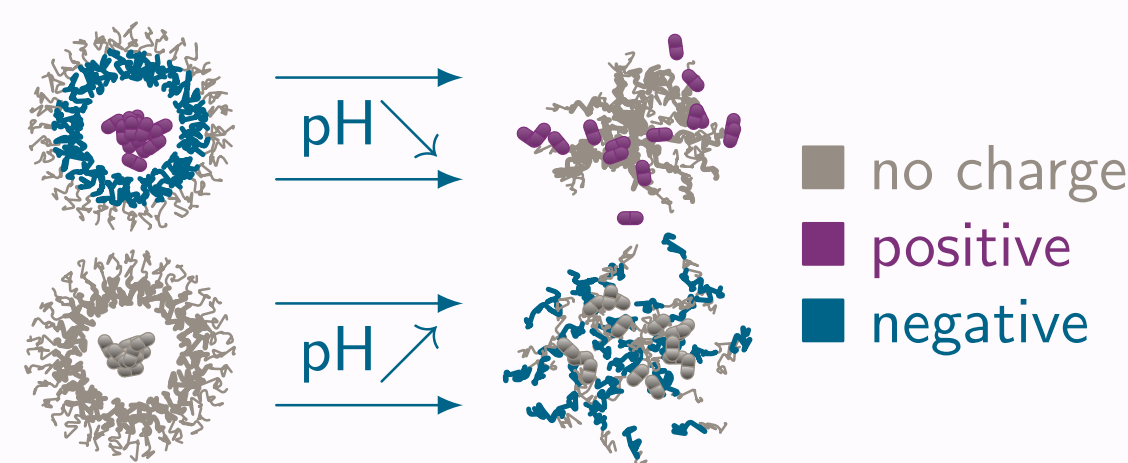


Figure 5: Polymeric micelles with pH-responsive polymers that readily protonate as pH falls, or deprotonate as pH increases, resulting in destructive charge imbalances [7]

Internal stimuli are bioindicators of disease and locality, including **pH**, **temperature**, **redox potential** and **enzymes**.

For example, solid tumours have been observed with a lower pH than healthy tissue [8, 9].

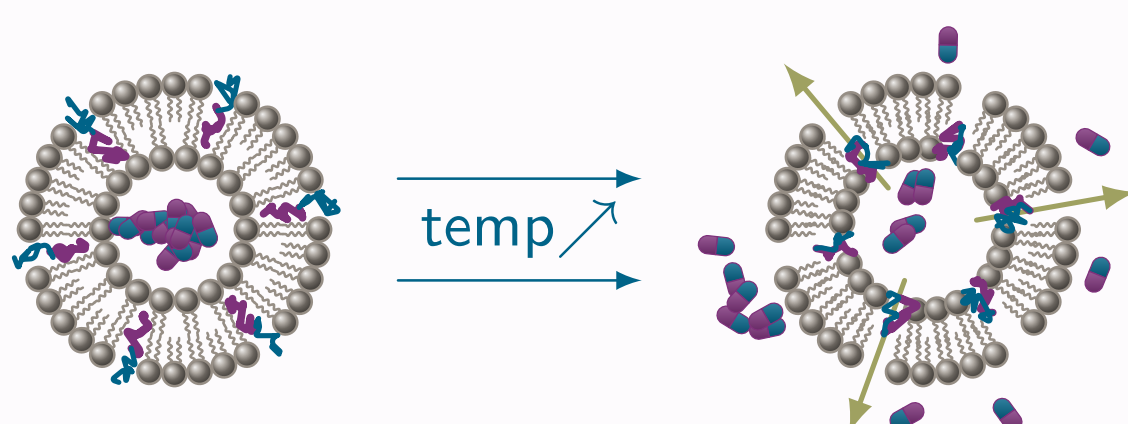


Figure 6: Liposomes modified with thermosensitive polymers which deform at hyperthermic temperatures, disrupting the liposome barrier [10, 11]

External stimuli can be applied **electric** or **magnetic fields**, **ultrasound**, **heat** and **light**.

Hyperthermia can be induced by applying direct heat until the tumour reaches 40 °C to 45 °C [12, 13]. Alternatively iron-oxide containing nanoparticles can be used to convert oscillating magnetic fields into heat [14].

Active targeting

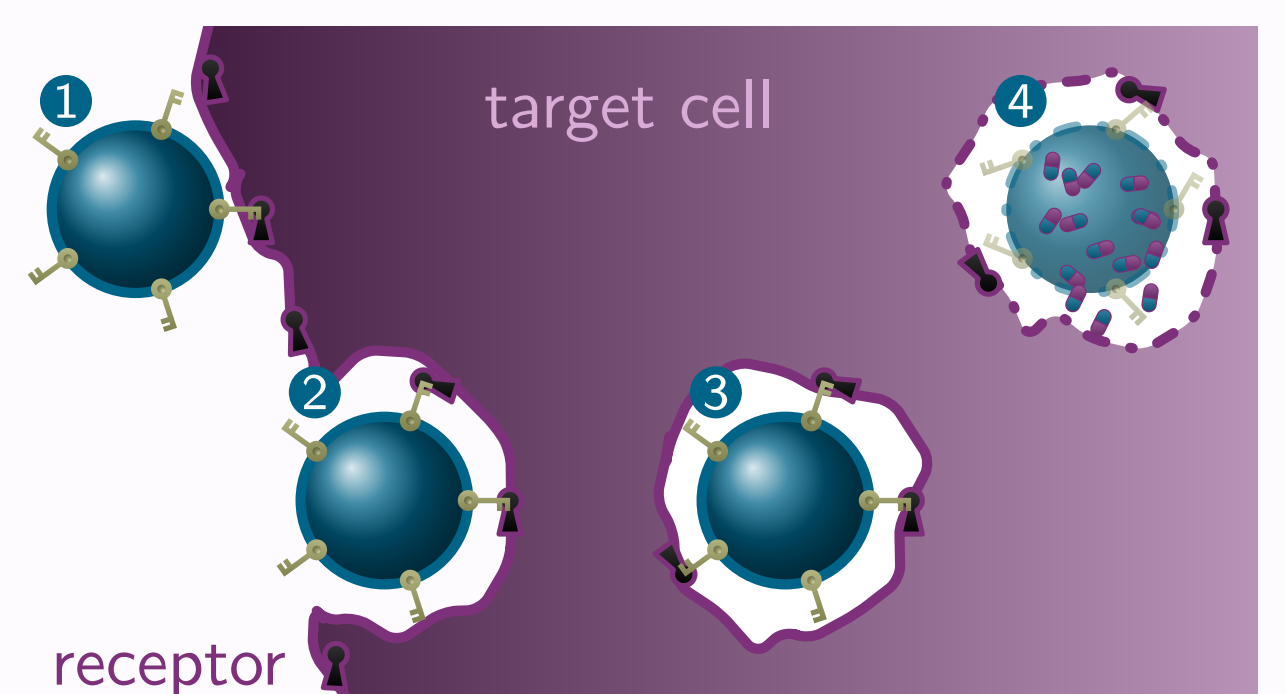


Figure 7: Receptor-mediated endocytosis. In order: **nanocarrier binding** ①, **endocytosis** ②, **endosome transport** ③, and **drug release** ④

Small tumours and blood cancers can be targeted by encouraging cell uptake through **receptor-mediated endocytosis**, where high-affinity **ligands** attached to **nanocarriers** are **targeted to receptors** common in specific cancer cells.

After binding with a receptor, the **cell wall absorbs the nanoparticle** forming an acidic pocket in the cell called an **endosome** [9]. pH-responsive nanocarriers have been designed to release their payload in this environment [7].

Common targeting ligands include **folic acid**, **carbohydrates** and **antibodies**.

To a certain extent, active targeting is still dependent on the **EPR effect** to reach the vicinity of the cancers cells without binding to cells in healthy tissue.

Example

- Variations of **Doxin[®]/Caelyx[®]** with anti-EGFR targeting ligands are in clinical trials [15].
- Targets the epidermal growth factor receptors (EGFR) on rapidly proliferating cancer cells.

1. J. M. Harris, R. B. Chess, *Nature reviews Drug discovery* **2**, 214 (2003).
2. H. Maeda *et al.*, *Journal of controlled release* **65**, 271–284 (2000).
3. A. Wicki *et al.*, *Journal of controlled release* **200**, 138–157 (2015).
4. T. P. Padera *et al.*, *Nature* **427**, 695 (2004).
5. R. K. Jain, T. Stylianopoulos, *Nature reviews Clinical oncology* **7**, 653 (2010).

6. M. Harrison *et al.*, *Journal of clinical oncology* **13**, 914–920 (1995).
7. T. Sun *et al.*, *Angewandte Chemie International Edition* **53**, 12320–12364 (2014).
8. I. F. Tannock, D. Rotin, *Cancer research* **49**, 4373–4384 (1989).
9. L. E. Gerweck, K. Seetharaman, *Cancer research* **56**, 1194–1198 (1996).
10. T. Ta, T. M. Porter, *Journal of controlled release* **169**, 112–125 (2013).

11. K. Kono, *Advanced drug delivery reviews* **53**, 307–319 (2001).
12. S. Ganta *et al.*, *Journal of controlled release* **126**, 187–204 (2008).
13. A. Jhaveri *et al.*, *Journal of controlled release* **190**, 352–370 (2014).
14. F. Scherer *et al.*, *Gene therapy* **9**, 102 (2002).
15. C. Mamot *et al.*, *The lancet oncology* **13**, 1234–1241 (2012).