Application of a reaction-diffusion equation to the spread of the invasive ascidian *Botrylloides* violaceus

Ruby Krasnow 2024-04-29

Abstract

Modeling is a critical tool for minimizing the harmful effects of invasive species on natural ecosystems. Despite the recent popularity of correlative environmental niche models, mechanistic reaction-diffusion models remain powerful tools for predicting and controlling the spread of invasive species. The colonial ascidian Botrylloides violaceus has rapidly spread across the Gulf of Maine, outcompeting and smothering native species and presenting a major nuisance for shellfish aquaculture. Here, we apply a relationship between the rate of spread of an invasive species and select biological parameters (namely, the population growth rate and diffusion coefficient) derived from the generalized reaction-diffusion equation. We validated the predicted rate of spread for B. violaceus against rates calculated from species occurrence data from the citizen science platform iNaturalist. The model predicted a mean speed of \sim 49 km/year, which was extremely close to the observed mean of \sim 40 km/year. Although simple reaction-diffusion models have limited biological realism, the accuracy of our results suggest that estimating spreading speed based on empirical data on local diffusion potentials and population growth rates could be an underutilized approach in invasion ecology.

Introduction

Reaction-diffusion models are a type of partial differential equation (PDE) that have long been used by ecologists to analyze the migration and population dynamics of invasive species (Cosner, 2008). Recent work has led to innumerable modifications on the original formulations by Fisher (1937) and Skellam (1951). Beyond the fundamental processes of growth and spread, extensions of reaction-diffusion models can be applied to age- and size-structured populations

(Neubert & Caswell, 2000), and incorporate environmental heterogeneity, interspecies interactions (competition, predation, mutualism, etc.), Allee effects, invasion control efforts, and long-distance dispersal. For an introduction to the use of PDEs to represent dispersal in a population ecology context, the reader is directed to Holmes et al. (1994) and the seminal papers referenced therein. For an accessible review of recent advancements in modeling invasion using reaction-diffusion equations and integro-difference equations, see Hastings et al. (2005); for a comprehensive treatment, see the textbooks by Shigesada and Kawasaki (1997) or Lewis et al. (2016).

Here, we use a simple two-dimensional reaction-diffusion model to represent the spread of the invasive ascidian *Botrylloides violaceus* in the Gulf of Maine. Colonial ascidians (or tunicates) are compound organisms formed by a mass of small zooids organized within a common gelatinous matrix (Wagstaff, 2017). These colonies often form visible crusts on rocks, vessel hulls, and other artificial floating structures and submerged objects (Wagstaff, 2017). Colonial ascidians also commonly grow on macroalgae and bivalves such as mussels and oysters. Ascidian growth on cultivation equipment and the cultivated organisms themselves can have many harmful effects, including smothering the bivalves, reducing local water circulation, consuming nutrients and dissolved oxygen, and causing structural damage to cultivation equipment (Aldred & Clare, 2014; Cahill et al., 2012; Fitridge et al., 2012). The detrimental impacts of ascidians on farm production and efficiency present significant economic costs for the North American aquaculture industry (Fitridge et al., 2012).

In New England and Atlantic Canada, aquaculturists have had little success controlling the spread or impact of invasive ascidians, particularly species like *B. violaceus* that have few natural predators and can outcompete other fouling organisms (Bullard et al., 2013; M. Carman et al., 2016; Carver et al., 2006; Gittenberger & Moons, 2011). *B. violaceus* is the most common fouling ascidian at shellfish aquaculture sites along the East coast of North America and the most widely distributed nonindigenous ascidian in British Columbia. (M. R. Carman et al., 2010; Simkanin et al., 2013).

B. violaceus has a complex life cycle, including a cyclical growth pattern dependent on both sexual and asexual reproduction. The growth and reproduction of colonial tunicates can be highly variable based on a number of environmental factors, including food availability, temperature, salinity, sediment load, and water pollution (Carver et al., 2006; Nelson, 2014). The microscale mechanisms behind tunicate dispersion are therefore very complicated, and a fully-resolved spatiotemporal model of B. violaceus spread would need also to incorporate local hydrodynamic conditions, human-mediated transport, competition, and predation.

However, simple reaction-diffusion models can accurately capture population spread without replying on many assumptions and often unknown information about organism-level behavior. The value of these models is derived from their tremendous utility in ecological applications, despite abstracting many biological details (Andow et al., 1990). By eliminating unnecessary complexity and noise, simple reaction-diffusion models present a tangible method of connecting the mathematical theory of partial differential equations to observed data on invasive species, with implications for real-world ecological and economic problems.

Model Development & Theory

Here, we follow the derivation method of Lewis et al. (2016) to justify our use of the following equation to represent the rate of spread of *B. violaceus*:

$$V = \sqrt{4rD} \tag{1}$$

where V > 0 is the velocity (distance/time) of the advancing front of the invasive population and D and r are positive constants representing the diffusion coefficient and maximum per capita growth rate, respectively.

Since B. violaceus has overlapping generations and can be considered to be continuously reproducing, it is appropriate to represent the invasion dynamics using a continuous-time PDE rather than a discrete integrodifference equation. We represent the population density of B. violaceus at time t and spatial coordinate x as u(t,x).

Assume that the population initially occupies a finite section of the New England coastline of length L and that the initial population distribution is compact.

$$u(0,x) = \begin{cases} \phi(x) > 0 & 0 < x < L \\ 0 & \text{otherwise} \end{cases}$$
 (2)

The reaction-diffusion model describing the population dynamics takes the form

$$\frac{\partial u}{\partial t} = D \frac{\partial u^2}{\partial x^2} + F(u) \tag{3}$$

Determining the correct form of the population growth term in (3) is challenging. Colonial ascidians generally display indeterminate growth, meaning that colony area tends to increase exponentially with time (Jackson, 1977). If we assumed simple linear population growth, our PDE would take the form

$$\frac{\partial U}{\partial t} = D \frac{\partial U^2}{\partial x^2} + rU \tag{4}$$

This is the simplest version of the general reaction-diffusion model, for a homogeneous, unstructured population growing exponentially and dispersing in a uniform, one-dimensional environment (Andow et al., 1990; Skellam, 1951). However, it is unlikely that colony-scale dynamics would apply over much larger spatial scales. Leaving F(u) as an unspecified function allows for the possibility of logistic growth, as in the Fisher-KPP equation:

$$\frac{\partial u}{\partial t} = D \frac{\partial u^2}{\partial x^2} + ru \left(1 - \frac{u}{K} \right) \tag{5}$$

We can generalize our section of the coastline of length L to an n-dimensional domain Ω , with boundary $\partial\Omega$. If the function u(t,x) and $\partial\Omega$ are smooth and $\phi(x)$ is continuous on the $\Omega\cup\partial\Omega$, the problem presented by (2) and (3) will be well posed in the sense that it will have a unique solution u(x,t) on $\Omega\times(0,T]$ for some T>0, with u(x,t) depending continuously on $\phi(x)$ (Cosner, 2008). Further discussion of the existence, uniqueness, and smoothness of solutions to reaction-diffusion equations can be found in Cosner (2008), along with a review of the relevant maximum principle and comparison theorem and a discussion of the stability and propagation of solutions.

The use of nonlinear reaction diffusion models in a biological context is well-established (see Cherniha & Davydovych, 2017). However, note that for biologically reasonable choices of F(u), the growth function will be bounded by the linear function, such that $F(u) \leq ru$. Thus,

$$D\frac{\partial u^2}{\partial x^2} + F(u) \le D\frac{\partial U^2}{\partial x^2} + rU \tag{6}$$

Now, note that for compact initial conditions like (2), there exist positive constants G and σ such that we bound the initial distribution from above by a Gaussian distribution:

$$U(x,0) = \frac{G}{\sqrt{4\pi\sigma^2}} \exp\left(-\frac{x^2}{4\sigma^2}\right) \tag{7}$$

such that
$$u(0,x) \le U(0,x)$$
 for all x (8)

From (6) and (8), we use the comparison theorem for PDEs to find that the solution to the linear equation in (6) provides an upper bound to the solution of the potentially nonlinear equation (3).

A detailed derivation of the solution of (4) can be found in Lewis et al. (2016). In brief, we utilize the change of variables $U(t,x) = W(t,x)e^{rt}$, reducing (4) to the standard diffusion equation for W. We then apply Fourier and inverse Fourier transforms to the new PDE and the initial condition (7) to find that

$$W(t,x) = \frac{G}{\sqrt{4\pi(\sigma^2 + Dt)}} \exp\left(-\frac{x^2}{4(\sigma^2 + Dt)}\right)$$

Converting back to our original variable, we conclude that the solution to (4) with the initial condition (7) is a normal distribution:

$$U(t,x) = \frac{G}{\sqrt{4\pi(\sigma^2 + Dt)}} \exp\left(-\frac{x^2}{4(\sigma^2 + Dt)} + rt\right)$$
(9)

Now, we can quantify the spatial spread of the population by considering how quickly a level set $U(x,t) = U_c$ moves in space, where U_c could be considered a threshold of detection. In other words, below U_c it is so difficult to find the invasive organisms that for practical purposes they are not present. By rearranging (9), we can get the following expression

$$\left(\frac{x}{t}\right)^2 = 4rD + \frac{4r\sigma^2}{t} - \frac{4(\sigma^2 + Dt)}{t^2} \ln\left(\frac{U_c\sqrt{4\pi(\sigma^2 + Dt)}}{G}\right)$$

As t becomes large, this formula simplifies to $\frac{|x|}{t} \to \sqrt{4rD}$. Thus, the asymptotic rate of spread is $V = \sqrt{4rD}$, as we initially sought to show.

This relationship between V, r, and D holds true for nonlinear forms of F(u), can be extended to higher dimensions, and does not need to rely on the idea of a threshold of detection—in fact, true advancing fronts with asymptotic speeds given by (1) form (Holmes et al., 1994; Okubo & Levin, 2001; Skellam, 1951). For invasion of a two-dimensional homogeneous habitat described by the reaction-diffusion equation

$$\frac{\partial u(x,y)}{\partial t} = D \left[\frac{\partial u^2}{\partial x^2} + \frac{\partial u^2}{\partial y^2} \right] + F(u), \tag{10}$$

the radius of the species range should increase linearly with time, with the slope of the increase given by $\sqrt{4rD}$.

This theory has been tested for numerous ecological systems and often agrees well with the empirical relationship between measured spread rates, per capita growth rates, and diffusion coefficients. Microscale observations can be used to parameterize (1) and predict the spreading rate for a given species at a geographic scale. Such spreading-speed analyses have been conducted for a number of terrestrial animal species, including the muskrat, cereal leaf beetle, European starling, small cabbage white butterfly, and grey squirrel (Andow et al., 1990; Grosholz, 1996; Holmes, 1993).

Spreading speed theory has also been applied to numerous aquatic species, including the brown trout, green crab, hermit crab, periwinkle, barnacle, mussels, and molluscs (Gharouni et al., 2015; Grosholz, 1996; MacDonald et al., 2022; Marculis & Lui, 2016). Possibly the only application of this method to a tunicate was performed by Grozholz (1996) for $B.\ leachi$, but his estimates of r=1.8 and D=1800 yielded a predicted rate of spread an order of magnitude larger than observed rates. In addition to providing insight on the spread of a specific invasive species of commercial concern, comparing the accuracy of our estimation for $B.\ violaceus$ to the previous results for $B.\ leachi$ could allow for broader conclusions to be drawn regarding the applicability of this approach for colonial tunicates in general.

Application

Predicting invasion speed

The diffusion coefficient, D, was calculated using the formula

$$D = \frac{2M_D(t)^2}{\pi t}$$

where $M_D(t)$ is the mean displacement of tracked organisms t time units after they begin moving (Andow et al., 1990; Othmer et al., 1988).

We estimated M_D using information on the dispersal potential of post-larval Botrylloides sp. colonies collected by Worcester (1994) (Table 1). B. violaceus can disperse naturally by rafting on eelgrass and other floating vegetation or debris (M. Carman & Grunden, 2010; Carver et al., 2006; Worcester, 1994). The dispersal distance/speed of rafting colonies was used rather than swimming larvae to better reflect the likely mechanisms by which B. violaceus has spread along the coastline.

Table 1: Calculating the diffusion coefficient

Parameter	Value
Distance traveled per colony per day (km)	0.225
Probability that a given colony will raft	0.667
Probability that a given rafting colony will reach a suitable habitat	0.5
Mean displacement (km/year)	27.38
$D (km^2/year)$	477.1

Eight estimates of population growth rate (r) were taken from Grey (2011), ranging from 0.78-1.89. Each r value was combined with our estimate of D to find a predicted invasion speed, V, in km/year.

Model validation

We validated our invasion speed estimates against the rate of boundary displacement for the region in which *B. violaceus* has been observed. We retrieved species occurrence data from the Global Biodiversity Information Facility (GBIF, https://www.gbif.org/) using the package 'spocc' (Owens et al., 2024). GBIF is an international database of biological data that has been used in over 10,000 peer-reviewed scientific articles and incorporates data from a variety of sources, primarily the citizen science platform iNaturalist.

Although citizen science data can contain spatial and temporal bias (Di Cecco et al., 2021), it is a valuable and widely-used tool for monitoring the spread of invasive species (Encarnação

et al., 2021). Furthermore, iNaturalist observations must meet certain quality standards to be included in GBIF (https://www.inaturalist.org/pages/help#GBIFdata), and tunicates like *B. violaceus* have a relatively low risk of misidentification by citizen volunteers (Lawson et al., 2022).

The region of interest for the GBIF query was arbitrarily defined to be the rectangular region bounded by the coordinates (72°W, 41°N, 59°W, 50°N). We only included observations labeled as "Human Observation" in order to exclude preserved specimens located in museum collections. Although *B. violaceus* may have been introduced to the east coast of the United States as early as the 1970s (Bock et al., 2011; Dijkstra et al., 2007), the earliest record in the GBIF database that did not have any issues or flags is from 2005.

For each year (t) since the year of the initial observation ($t_0 = 2005$), we calculated V as

$$V = \frac{\max|x_i - x_0|}{t - t_0}$$

where x_0 was the coordinate location of the 2005 observation and x_i is a single observation within the vector x of locations where B. violaceus had been observed over the period t_0 to t. While there are various methods of quantifying macroscale patterns of species' range expansions, similar boundary displacement approaches are common in invasion ecology (Clark et al., 2018; Tobin et al., 2007). We discarded the very low speed estimated for 2006 (1.1 km/year) as a clear outlier, but all other data was retained.

All calculations were performed in R version 4.3.2 (R Core Team, 2023). R packages central to data analysis and visualization included 'spocc', 'sf', 'leaflet', and the tidyverse collection (Cheng et al., 2023; Owens et al., 2024; Pebesma & Bivand, 2023; Wickham et al., 2019).

Results

Predicted invasion speeds using (1) varied from 38.5 to 60.0 km/year, corresponding to the lowest (r = 0.78) and highest (r = 1.89) population growth rates assumed for *B. violaceus* (Table 2).

Table 2: Summary statistics for observed and predicted *B. violaceus* invasion speed (km/year).

	Observed	Predicted
Min.	27.3	38.5
1st Qu.	32.0	42.5
Median	40.4	48.5
Mean	40.6	49.4
3rd Qu.	45.5	57.1
Max.	64.0	60.0

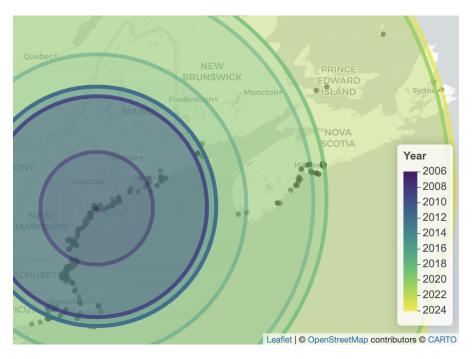


Figure 1: A map of the spread of *B. violaceus* in the Gulf of Maine. Black dots represent confirmed observations of *B. violaceus* from the GBIF database. Colored circles (centered at the location of the first observation) illustrate the expanding range.

There was significant overlap between the range of predicted speeds and the distribution of "true" invasion speeds, estimated by dividing the distance between the introduction point and furthest observation of $B.\ violaceus$ in each year by the time elapsed since introduction (Figure 2). The mean observed and predicted invasion speeds differed by fewer than $9 \, \mathrm{km/year}$, a remarkably small disparity (Table 2).

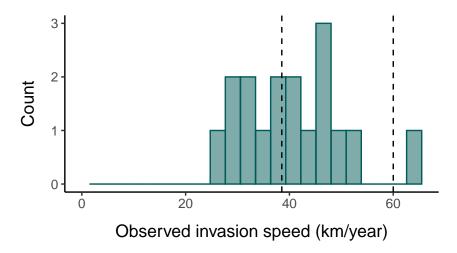


Figure 2: Histogram of observed invasion speeds for *B. violaceus* in the Gulf of Maine, calculated from GBIF species occurrence data from 2006-2024. Dashed lines represent the minimum and maximum predicted invasion speeds when varying population growth rates were used as input values.

Discussion

This model attempted to predict the macro-scale dispersal of an invasive tunicate using a simple reaction-diffusion model. Given the low biological realism of our simple diffusion model, the very strong agreement between observed and predicted invasion speeds is quite remarkable. Most studies that have applied this method of predicting invasion speed characterized their results as successful if estimates were within the same order of magnitude as observed spreading speeds. For example, Grosholz (1996) predicted the spreading speed of the tunicate $B.\ leachi$ (a member of the same genus as to be $B.\ violaceus$) to be 114 km/year, which was much faster than the observed rate of 16 km/year.

In contrast, our predicted speed of ~ 49 km/year was extremely close to the mean of ~ 40 km/year derived from observations of B. violaceus made in the Gulf of Maine over the past two decades (Table 2). Both the observed and predicted invasion speeds were generally in the range of 40-60 km/year (Fig 2). While such accurate prediction is unlikely to be replicable with other data sets, it does suggest that estimating spreading speed from empirical data on local diffusion potential and experimentally-derived population growth rates could be an underutilized approach in invasion ecology.

Multiple avenues are apparent for future work modeling the spread of *B. violaceus* using diffusion-type equations. First, there exist many other possible formulations for the term representing population growth. The Fisher equation, which assumes logistic rather than exponential growth, has been well-studied from a mathematical perspective and has seen

widespread application across ecological contexts (Berestycki et al., 2009; e.g., Holmes et al., 1994). Other diffusion models incorporate Allee effects, an ecological phenomenon characterized by decreased individual fitness (lower per capita growth rates) at small population sizes. Accounting for Allee effects in continuous reaction-diffusion models or analogous discrete integrodifference models can alter predictions about the potential success or failure of an invasion, rates of spread, and plausible ranges of an invasive species (Keitt et al., 2001; Kot et al., 1996; Liu & Kot, 2019).

The model from which (1) was derived assumes a uniform environment and isotropic dispersal. In reality, invasive species are more likely to disperse into more favorable habitats, so assuming that the rate of spread does not vary based on the direction of movement is not realistic. For example, the growth and reproduction of $B.\ violaceus$ generally increase with warmer temperatures, so in a relatively cold climate, southward dispersal may be more likely than northward dispersal.

Human-mediated introduction via hull fouling on recreational or commercial watercraft and aquaculture activities are also major pathways by which invasive tunicates are introduced to new locations (Carver et al., 2006). Although *B. violaceus* has a relatively low attachment strength, the small drag coefficient associated with their low profile means that its displacement velocity is above the top speeds of sailboats, barges, and other slower-moving vessels (Clarke Murray et al., 2012). Clarke Murray et al. (2011) found that *B. violaceus* was attached to the hulls of 9.8% of recreational boats at nine distinct marinas in British Columbia.

Future work could utilize discrete integrodifference equations with leptokurtic dispersion kernels to account for transport via hull fouling and other potential vectors of long-distance dispersion. The potential for long-distance dispersal events via natural or human-mediated pathways often prompts the use of integrodifference equations for diffusion models of invasive populations (Kot & Neubert, 2008; Lewis et al., 2016; Lutscher, 2019; Neubert & Caswell, 2000). These models assume that growth and dispersal occur in distinct and asynchronous stages. The ability of *B. violaceus* to utilize multiple forms of reproduction and the fact that both larvae and post-larvae can contribute to the colonization of new habitat potentially violates the assumption of temporally separated growth and dispersion. However, stratified models that separate spatially continuous short-distance dispersal from discrete long-distance dispersal can be valuable tools to reconstruct realistic invasion dynamics (e.g., Gilioli et al., 2013) and their potential for modeling the spread of *B. violaceus* merits further investigation.

Recent work coupling deterministic models with a stochastic component has led to major improvements in the biological realism of PDE-based invasion models. These stochastic models can account for both process and observation error, incorporating the randomness inherent in natural systems. For example, Roques et al. (2011) used a "statistical-reaction-diffusion approach" to analyze the expansion of the pine processionary moth based on moth nest presence/absence data. Using a "mechanistic-statistical" diffusion model assuming logistical growth and imperfect species observation data, Louvrier et al. (2020) were able to forecast the expanding distribution of the grey wolf in France. Applying similar methods to data on B. violaceus would be valuable for developing useful predictions of potential future invasions.

Finally, competition-diffusion models that represent the population growth and spread of two competing species in the same habitat admit similar wavefront solutions to those described above, and may result in periodic spatial patterns driven by the dynamics of the interspecific interactions (Carrère, 2018; Holmes et al., 1994). It is often interesting to investigate the stability and long-term behavior of a system of competition-diffusion models for different parameter values, including varying the propagation speeds of the two organisms or the differential impacts of the competitive interactions on each species (Carrère, 2018; Ducrot et al., 2014). Experimental evidence suggests that *B. violaceus* colonies can outcompete *Botryllus schlosseri*, a different invasive tunicate that has been equally or more problematic for the New England aquaculture industry (Dijkstra et al., 2007; Gittenberger & Moons, 2011). However, these studies have been limited to small-scale experiments conducted on a single substrate like a PVC panel (Wagstaff, 2017). Our single-species reaction diffusion model could be extended to a system of competition-diffusion equations to examine the potential impacts of competition between *B. violaceus* and *B. schlosseri* on the spread of these two species on a broader scale.

References

- Aldred, N., & Clare, A. S. (2014). Mini-review: Impact and dynamics of surface fouling by solitary and compound ascidians. *Biofouling*, 30(3), 259270. https://doi.org/10.1080/08927014.2013.866653
- Andow, D. A., Kareiva, P. M., Levin, S. A., & Okubo, A. (1990). Spread of invading organisms. Landscape Ecology, 4(2), 177–188. https://doi.org/10.1007/BF00132860
- Berestycki, H., Nadin, G., Perthame, B., & Ryzhik, L. (2009). The non-local Fisher–KPP equation: travelling waves and steady states. *Nonlinearity*, 22(12), 2813. https://doi.org/10.1088/0951-7715/22/12/002
- Bock, D. G., Zhan, A., Lejeusne, C., MacISAAC, H. J., & Cristescu, M. E. (2011). Looking at both sides of the invasion: patterns of colonization in the violet tunicate Botrylloides violaceus. *Molecular Ecology*, 20(3), 503–516. https://doi.org/10.1111/j.1365-294X.2010.04971.x
- Bullard, S. G., Davis, C. V., & Shumway, S. E. (2013). Seasonal patterns of ascidian settlement at an aquaculture facility in the damariscotta river, maine. *Journal of Shellfish Research*, 32(2), 255–264. https://doi.org/10.2983/035.032.0202
- Cahill, P., Heasman, K., Jeffs, A., Kuhajek, J., & Mountfort, D. (2012). Preventing ascidian fouling in aquaculture: Screening selected allelochemicals for anti-metamorphic properties in ascidian larvae. *Biofouling*, 28(1), 3949. https://doi.org/10.1080/08927014.2011.648624
- Carman, M. R., Morris, J. A., Karney, R. C., & Grunden, D. W. (2010). An initial assessment of native and invasive tunicates in shellfish aquaculture of the North American east coast. Journal of Applied Ichthyology, 26, 8–11. https://doi.org/10.1111/j.1439-0426.2010.01495.
- Carman, M., & Grunden, D. (2010). First occurrence of the invasive tunicate didemnum vexillum in eelgrass habitat. Aquatic Invasions, 5(1), 23–29. https://doi.org/10.3391/ai.

2010.5.1.4

- Carman, M., Lindell, S., Green-Beach, E., & Starczak, V. (2016). Treatments to eradicate invasive tunicate fouling from blue mussel seed and aquaculture socks. *Management of Biological Invasions*, 7(1), 101–110. https://doi.org/10.3391/mbi.2016.7.1.12
- Carrère, C. (2018). Spreading speeds for a two-species competition-diffusion system. *Journal of Differential Equations*, 264(3), 2133–2156. https://doi.org/10.1016/j.jde.2017.10.017
- Carver, C. E., Mallet, A. L., & Vercamaer, B. (2006). Biological synopsis of the colonial tunicates, Botryllus schlosseri and Botrylloides violaceus (p. 42). publications.gc.ca/pub?id=9.562383&sl=0
- Cheng, J., Schloerke, B., Karambelkar, B., & Xie, Y. (2023). Leaflet: Create interactive web maps with the JavaScript 'leaflet' library. https://CRAN.R-project.org/package=leaflet
- Cherniha, R., & Davydovych, V. (2017). Nonlinear Reaction-Diffusion Systems (Vol. 2196). Springer International Publishing. https://doi.org/10.1007/978-3-319-65467-6
- Clark, T. L., Iannone, B. V. I., & Fei, S. (2018). Metrics for macroscale invasion and dispersal patterns. *Journal of Plant Ecology*, 11(1), 64–72. https://doi.org/10.1093/jpe/rtw136
- Clarke Murray, C., Pakhomov, E. A., & Therriault, T. W. (2011). Recreational boating: a large unregulated vector transporting marine invasive species. *Diversity and Distributions*, 17(6), 1161–1172. https://doi.org/10.1111/j.1472-4642.2011.00798.x
- Clarke Murray, C., Therriault, T. W., & Martone, P. T. (2012). Adapted for invasion? Comparing attachment, drag and dislodgment of native and nonindigenous hull fouling species. Biological Invasions, 14(8), 1651–1663. https://doi.org/10.1007/s10530-012-0178-0
- Cosner, C. (2008). Reaction–Diffusion Equations and Ecological Modeling (A. Friedman, Ed.; pp. 77–115). Springer. https://doi.org/10.1007/978-3-540-74331-6_3
- Di Cecco, G. J., Barve, V., Belitz, M. W., Stucky, B. J., Guralnick, R. P., & Hurlbert, A. H. (2021). Observing the observers: How participants contribute data to iNaturalist and implications for biodiversity science. *BioScience*, 71(11), 1179–1188. https://doi.org/10.1093/biosci/biab093
- Dijkstra, J., Harris, L. G., & Westerman, E. (2007). Distribution and long-term temporal patterns of four invasive colonial ascidians in the Gulf of Maine. *Journal of Experimental Marine Biology and Ecology*, 342(1), 61–68. https://doi.org/10.1016/j.jembe.2006.10.015
- Ducrot, A., Giletti, T., & Matano, H. (2014). Existence and convergence to a propagating terrace in one-dimensional reaction-diffusion equations. *Transactions of the American Mathematical Society*, 366(10), 5541–5566. https://doi.org/10.1090/S0002-9947-2014-06105-9
- Encarnação, J., Teodósio, M. A., & Morais, P. (2021). Citizen science and biological invasions: A review. Frontiers in Environmental Science, 8. https://doi.org/10.3389/fenvs.2020. 602980
- Fisher, R. A. (1937). THE WAVE OF ADVANCE OF ADVANTAGEOUS GENES. Annals of Eugenics, 7(4), 355-369. https://doi.org/10.1111/j.1469-1809.1937.tb02153.x
- Fitridge, I., Dempster, T., Guenther, J., & Nys, R. de. (2012). The impact and control of biofouling in marine aquaculture: A review. *Biofouling*, 28(7), 649–669. https://doi.org/10.1080/08927014.2012.700478
- Gharouni, A., Barbeau, M., Locke, A., Wang, L., & Watmough, J. (2015). Sensitivity of invasion speed to dispersal and demography: an application of spreading speed theory to

- the green crab invasion on the northwest Atlantic coast. *Marine Ecology Progress Series*, 541, 135–150. https://doi.org/10.3354/meps11508
- Gilioli, G., Pasquali, S., Tramontini, S., & Riolo, F. (2013). Modelling local and long-distance dispersal of invasive chestnut gall wasp in europe. *Ecological Modelling*, 263, 281–290. https://doi.org/10.1016/j.ecolmodel.2013.05.011
- Gittenberger, A., & Moons, J. (2011). Settlement and possible competition for space between the invasive violet tunicate botrylloides violaceus and the native star tunicate botryllus schlosseri in the netherlands. *Aquatic Invasions*, 6(4), 435–440. https://doi.org/10.3391/ai.2011.6.4.08
- Grey, E. K. (2011). Relative effects of environment and direct species interactions on the population growth rate of an exotic ascidian. *Oecologia*, 166(4), 935–947. https://www.jstor.org/stable/41499896
- Grosholz, E. D. (1996). Contrasting Rates of Spread for Introduced Species in Terrestrial and Marine Systems. *Ecology*, 77(6), 1680–1686. https://doi.org/10.2307/2265773
- Hastings, A., Cuddington, K., Davies, K. F., Dugaw, C. J., Elmendorf, S., Freestone, A., Harrison, S., Holland, M., Lambrinos, J., Malvadkar, U., Melbourne, B. A., Moore, K., Taylor, C., & Thomson, D. (2005). The spatial spread of invasions: new developments in theory and evidence. *Ecology Letters*, 8(1), 91–101. https://doi.org/10.1111/j.1461-0248.2004.00687.x
- Holmes, E. E. (1993). Are diffusion models too simple? A comparison with telegraph models of invasion. *The American Naturalist*, 142(5), 779–795. https://www.jstor.org/stable/2462716
- Holmes, E. E., Lewis, M. A., Banks, J. E., & Veit, R. R. (1994). Partial Differential Equations in Ecology: Spatial Interactions and Population Dynamics. *Ecology*, 75(1), 17–29. https://doi.org/10.2307/1939378
- Jackson, J. B. C. (1977). Competition on Marine Hard Substrata: The Adaptive Significance of Solitary and Colonial Strategies. The American Naturalist, 111(980), 743–767. https://doi.org/10.1086/283203
- Keitt, T. H., Lewis, M. A., & Holt, R. D. (2001). Allee effects, invasion pinning, and species' borders. *The American Naturalist*, 157(2), 203–216. https://doi.org/10.1086/318633
- Kot, M., Lewis, M. A., & Driessche, P. van den. (1996). Dispersal Data and the Spread of Invading Organisms. *Ecology*, 77(7), 2027–2042. https://doi.org/10.2307/2265698
- Kot, M., & Neubert, M. G. (2008). Saddle-Point Approximations, Integrodifference Equations, and Invasions. *Bulletin of Mathematical Biology*, 70(6), 1790–1826. https://doi.org/10.1007/s11538-008-9325-2
- Lawson, K. N., Tracy, B. M., Sharova, M., Muirhead, J. R., & Cawood, A. (2022). Engaging online citizen scientists and the consensus method to monitor the marine biofouling community. Frontiers in Marine Science, 9. https://doi.org/10.3389/fmars.2022.862430
- Lewis, M. A., Petrovskii, S. V., & Potts, J. R. (2016). The Mathematics Behind Biological Invasions (Vol. 44). Springer International Publishing. https://doi.org/10.1007/978-3-319-32043-4
- Liu, B. R., & Kot, M. (2019). Accelerating invasions and the asymptotics of fat-tailed dispersal. Journal of Theoretical Biology, 471, 22–41. https://doi.org/10.1016/j.jtbi.2019.03.016

- Louvrier, J., Papaïx, J., Duchamp, C., & Gimenez, O. (2020). A mechanistic–statistical species distribution model to explain and forecast wolf (*Canis lupus*) colonization in south-eastern france. *Spatial Statistics*, 36, 100428. https://doi.org/10.1016/j.spasta.2020.100428
- Lutscher, F. (2019). *Integrodifference Equations in Spatial Ecology* (Vol. 49). Springer International Publishing. https://doi.org/10.1007/978-3-030-29294-2
- MacDonald, K. J., Leroux, S. J., & Purchase, C. F. (2022). Landscape, demographic, and dispersal parameters influence the spread and establishment dynamics of an aquatic biological invasion. *ResearchSquare*. https://doi.org/10.21203/rs.3.rs-1514165/v1
- Marculis, N. G., & Lui, R. (2016). Modelling the biological invasion of carcinus maenas (the european green crab). *Journal of Biological Dynamics*, 10(1), 140–163. https://doi.org/10.1080/17513758.2015.1115563
- Nelson, J. C. (2014). Species invasion in the marine fouling communities of British Columbia: factors that influence invasion dynamics and how they may affect Botrylloides violaceus [PhD thesis]. https://doi.org/10.14288/1.0167429
- Neubert, M. G., & Caswell, H. (2000). Demography and Dispersal: Calculation and Sensitivity Analysis of Invasion Speed for Structured Populations. *Ecology*, 81(6), 1613–1628. https://doi.org/10.1890/0012-9658(2000)081%5B1613:DADCAS%5D2.0.CO;2
- Okubo, A., & Levin, S. A. (2001). Diffusion and ecological problems: Modern perspectives. Springer New York. https://doi.org/10.1007/978-1-4757-4978-6
- Othmer, H. G., Dunbar, S. R., & Alt, W. (1988). Models of dispersal in biological systems. Journal of Mathematical Biology, 26(3), 263–298. https://doi.org/10.1007/BF00277392
- Owens, H., Barve, V., & Chamberlain, S. (2024). Spoce: Interface to species occurrence data sources. https://CRAN.R-project.org/package=spoce
- Pebesma, E., & Bivand, R. (2023). {Spatial data science: With applications in r}. https://doi.org/10.1201/9780429459016
- R Core Team. (2023). R: A language and environment for statistical computing. R Foundation for Statistical Computing. https://www.R-project.org/
- Roques, L., Soubeyrand, S., & Rousselet, J. (2011). A statistical-reaction-diffusion approach for analyzing expansion processes. *Journal of Theoretical Biology*, 274(1), 43–51. https://doi.org/10.1016/j.jtbi.2011.01.006
- Shigesada, N., & Kawasaki, K. (1997). Biological invasions: Theory and practice. Oxford University Press.
- Simkanin, C., Dower, J. F., Filip, N., Jamieson, G., & Therriault, T. W. (2013). Biotic resistance to the infiltration of natural benthic habitats: Examining the role of predation in the distribution of the invasive ascidian *Botrylloides violaceus*. *Journal of Experimental Marine Biology and Ecology*, 439, 76–83. https://doi.org/10.1016/j.jembe.2012.10.004
- Skellam, J. G. (1951). RANDOM DISPERSAL IN THEORETICAL POPULATIONS. *Biometrika*, 38(1-2), 196–218. https://doi.org/10.1093/biomet/38.1-2.196
- Tobin, P. C., Liebhold, A. M., & Anderson Roberts, E. (2007). Comparison of methods for estimating the spread of a non-indigenous species. *Journal of Biogeography*, 34(2), 305–312. https://doi.org/10.1111/j.1365-2699.2006.01600.x
- Wagstaff, M. (2017). Life history variation of an invasive species botrylloides violaceus (oka, 1927) between novel coastal habitats in the gulf of maine. *Aquatic Invasions*, 12(1), 43–51.

https://doi.org/10.3391/ai.2017.12.1.05

- Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L. D., François, R., Grolemund, G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T. L., Miller, E., Bache, S. M., Müller, K., Ooms, J., Robinson, D., Seidel, D. P., Spinu, V., ... Yutani, H. (2019). Welcome to the {tidyverse}. 4, 1686. https://doi.org/10.21105/joss.01686
- Worcester, S. E. (1994). Adult rafting versus larval swimming: dispersal and recruitment of a botryllid ascidian on eelgrass. *Marine Biology*, 121(2), 309–317. https://doi.org/10.1007/BF00346739