Indexing the (compacted) colored de Bruijn graph



NOTE: This lecture is being recorded

Scaling up fast reference-based indices

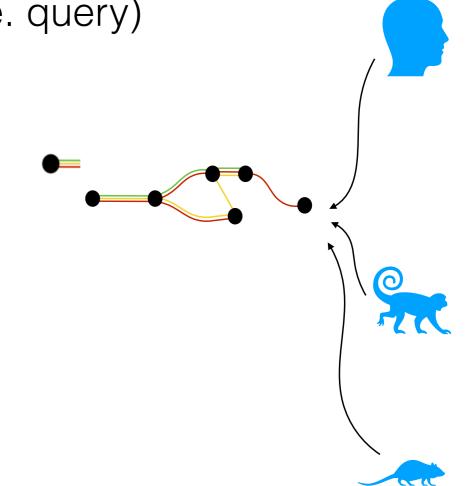
Motivation: Indices used in "ultra-fast" mapping approaches are typically very memory hungry. This is **OK** for transcriptome mapping, but **not scalable** to genomic, metagenomic, pangenomic or population mapping.

Goal: Develop an index with practical memory requirements that maintains the desirable performance (i.e. query) characteristics of the "ultra-fast" indices.

Compacted colored de Bruijn graph (ccdBG)

Built over 1 or more genomes / sequence collections

Index makes use of minimum perfect hashing succinct bit vector representations and (optionally) a new sampling scheme



Pufferfish: An efficient index for the ccdBG

A space and time-efficient index for the compacted colored de Bruijn graph

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Appeared at ISMB 2018

- The past decade has largely been dominated by SA/BWT/FM-indexbased approaches to reference sequence indexing (e.g. Bowtie, BWA, BWA-MEM, Bowtie2, STAR, etc.)
- There has been a renaissance of sorts for hash-based indexing (deBGA, Brownie, kallisto, mashmap, minimap & minimap2, etc.)
- Pufferfish goes the hashing-based route; with a twist.
- Not considering generalized path indices on general seq (e.g. GCSA2 (VG), HISAT2). Interesting, but a different problem.

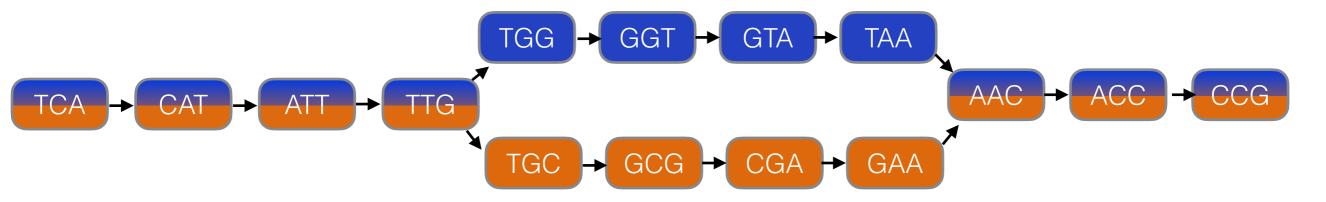
https://github.com/COMBINE-lab/pufferfish

Recall the "colored" de Bruijn Graph

Nodes are k-mers (here k=3)

Edges exist between nodes that overlap by k-1 (in the input)*

Colors encode "origin" of k-mers (e.g., references where they exist)



compacted colored de Bruijn graph



Example from: https://algolab.files.wordpress.com/2016/10/chikhi-milan-18nov.pdf

There are multiple related (but distinct) definitions of the dBG in practice. We adopt the **edge-explicit** version.

The compacted colored dBG as a sequence index

• **Key idea**: represent a collection of sequences using the colored de Bruijn graph (dBG) (lqbal '12).

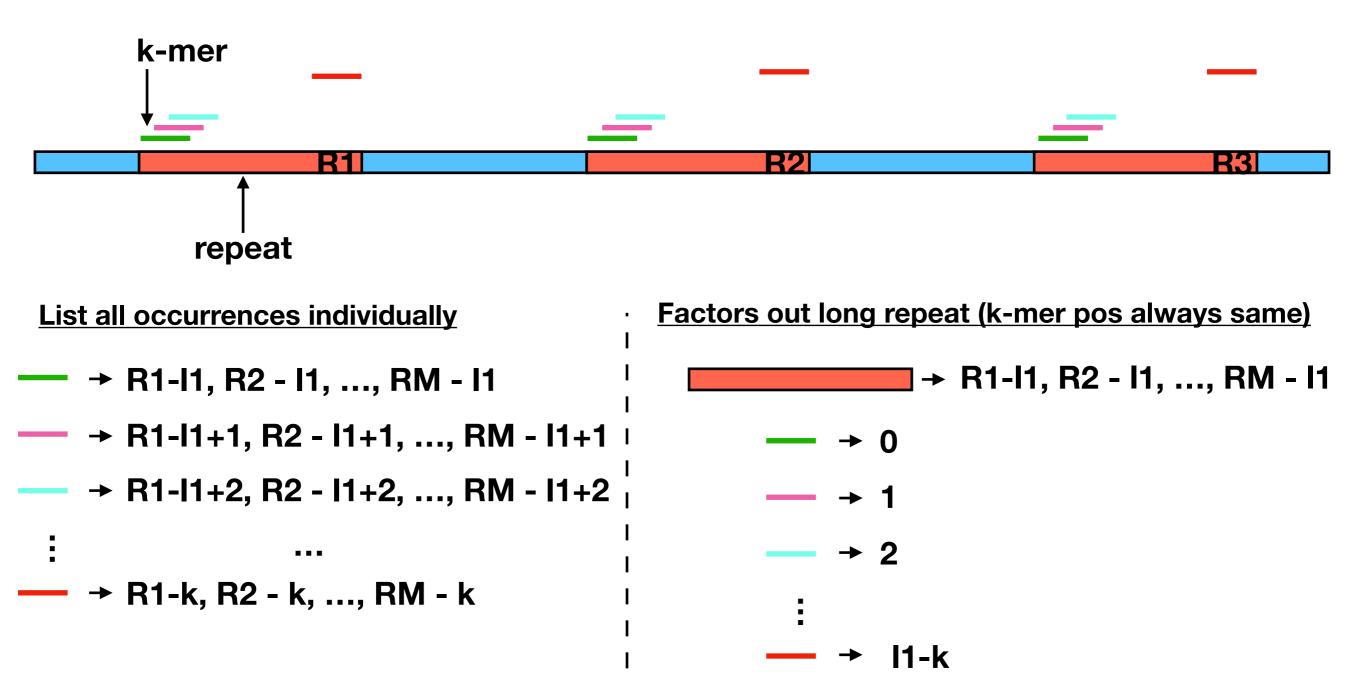
Each color is an input reference (e.g. genome or transcript).

 Use the compacted colored dBG as an index for reference-based sequence search.

 Redundant sequences (repeats) are implicitly collapsed. Why is this potentially much better than a naive hash?

The compacted colored dBG as a sequence index

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The compacted colored dBG as a sequence index

 Redundant sequences (repeats) are implicitly collapsed. Why is this potentially much better than a naive hash?

Still, the biggest problem for these schemes, in practice, is memory usage

The main culprit is the hash table itself

The cdBG removes redundancy by providing an extra level of indirection

Recall: Minimum Perfect Hashing

Minimum Perfect Hash Function (MPHF)

$$\mathcal{K} \subseteq \mathcal{U}, \ f: \mathcal{K} \to \mathbb{N}^+$$

if
$$x \in \mathcal{K}$$
 then $f(x) \in [1, |\mathcal{K}|]$

if
$$x \in \mathcal{U} \setminus \mathcal{K}$$
 then $f(x) \in [1, |\mathcal{U}|]$ (Like "false positives")

f is a complete, injective function from $\mathcal{K} \to [1, |\mathcal{K}|]$

Best methods achieve ~2.1 bits/key regardless of key size

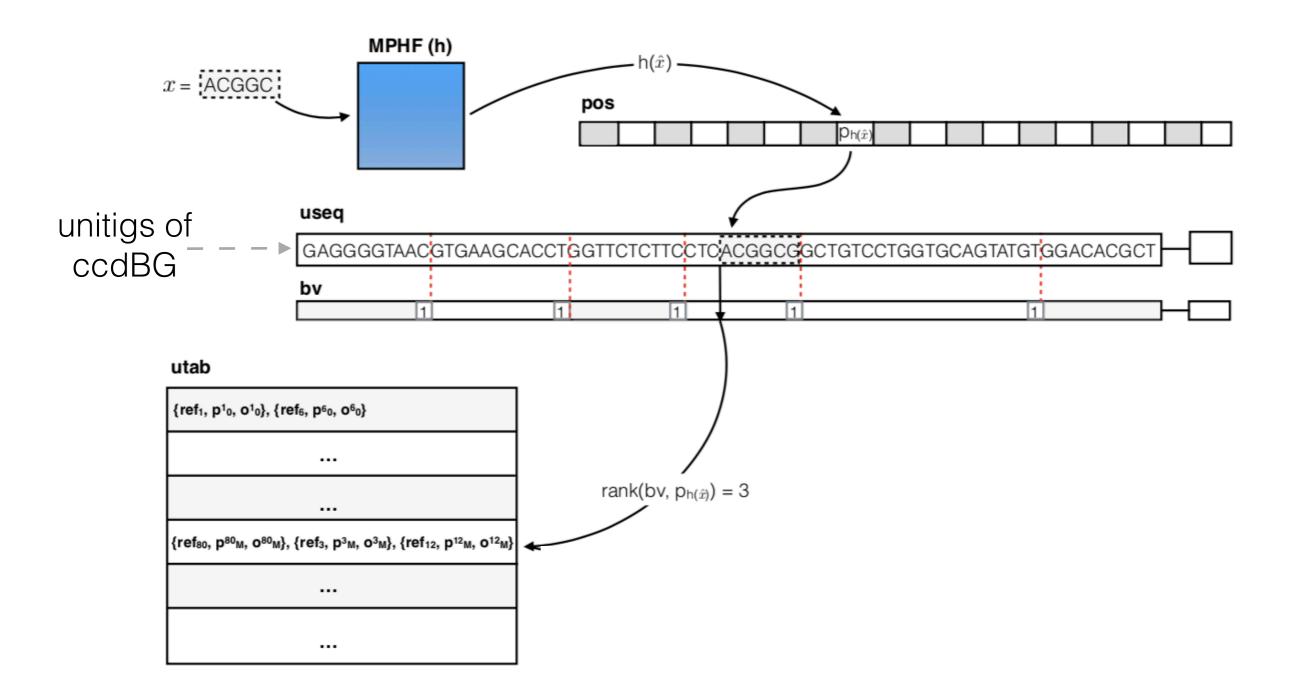
Use BBHash:)

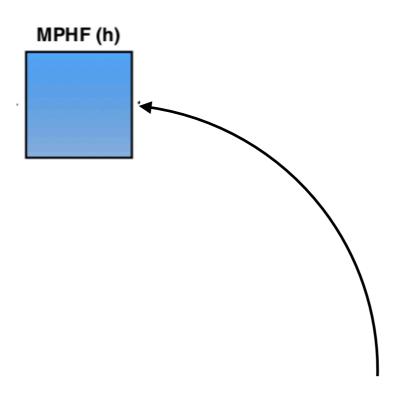
Fast and scalable minimal perfect hashing for massive key sets

Antoine Limasset¹, Guillaume Rizk¹, Rayan Chikhi², and Pierre Peterlongo¹

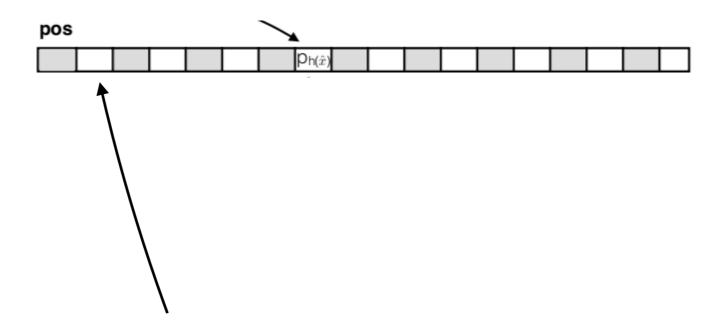
2 CNRS, CRIStAL, Université de Lille, Inria Lille - Nord Europe, France

¹ IRISA Inria Rennes Bretagne Atlantique, GenScale team, Campus de Beaulieu 35042 Rennes, France

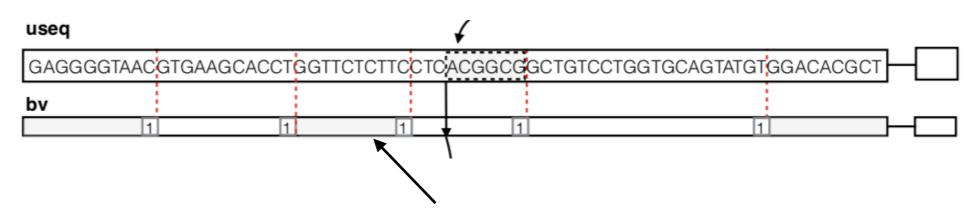




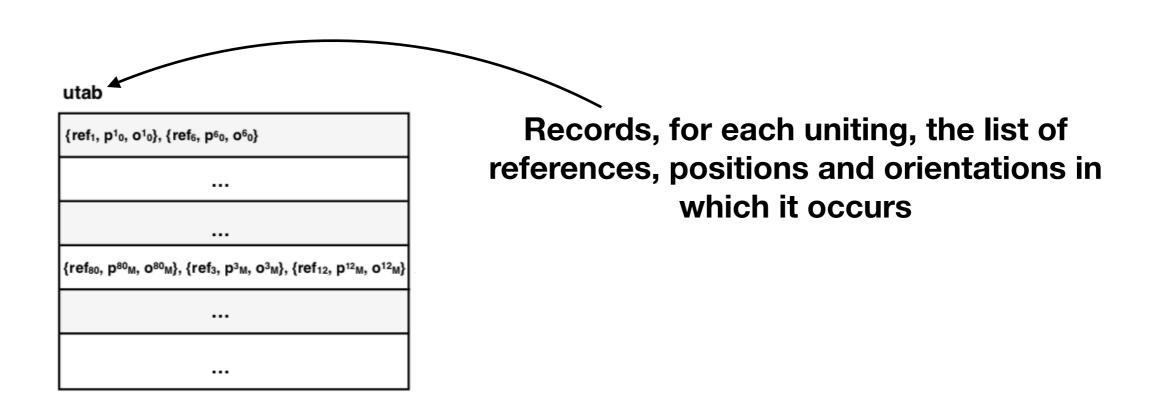
Maps each valid k-mer to some number in [0,N)

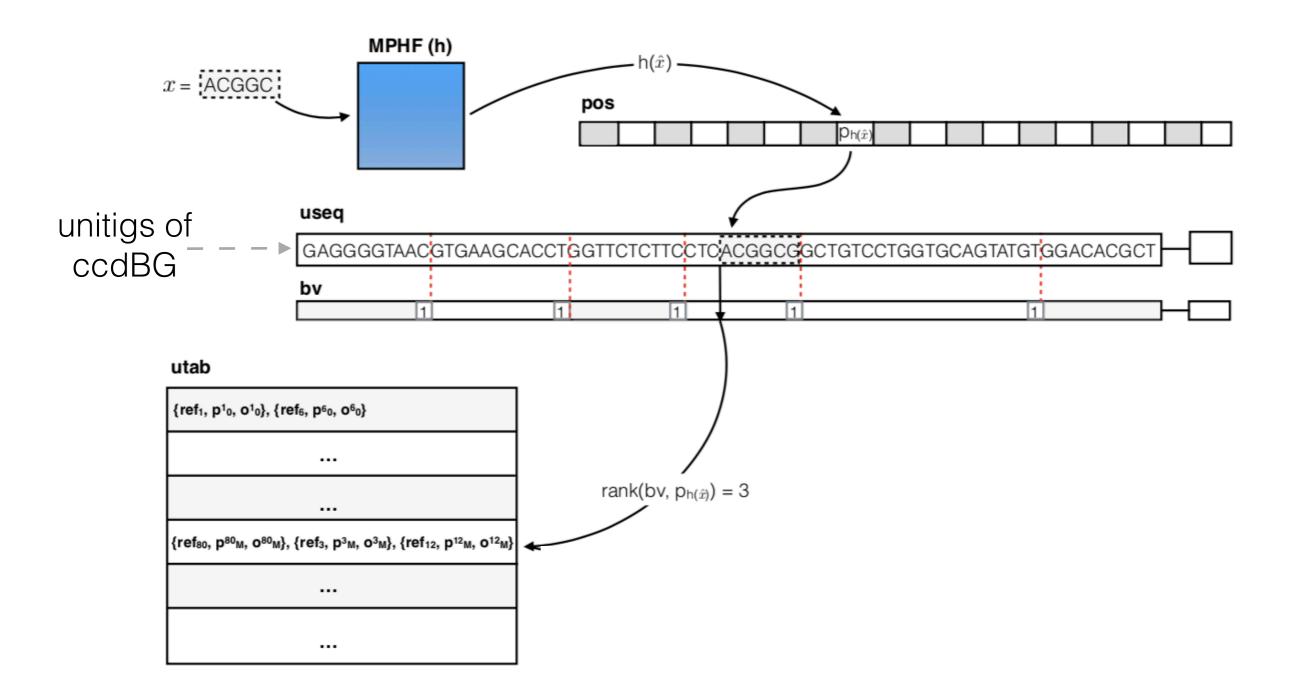


At index h(x), this table contains the position, in the list of unitigs, of this k-mer

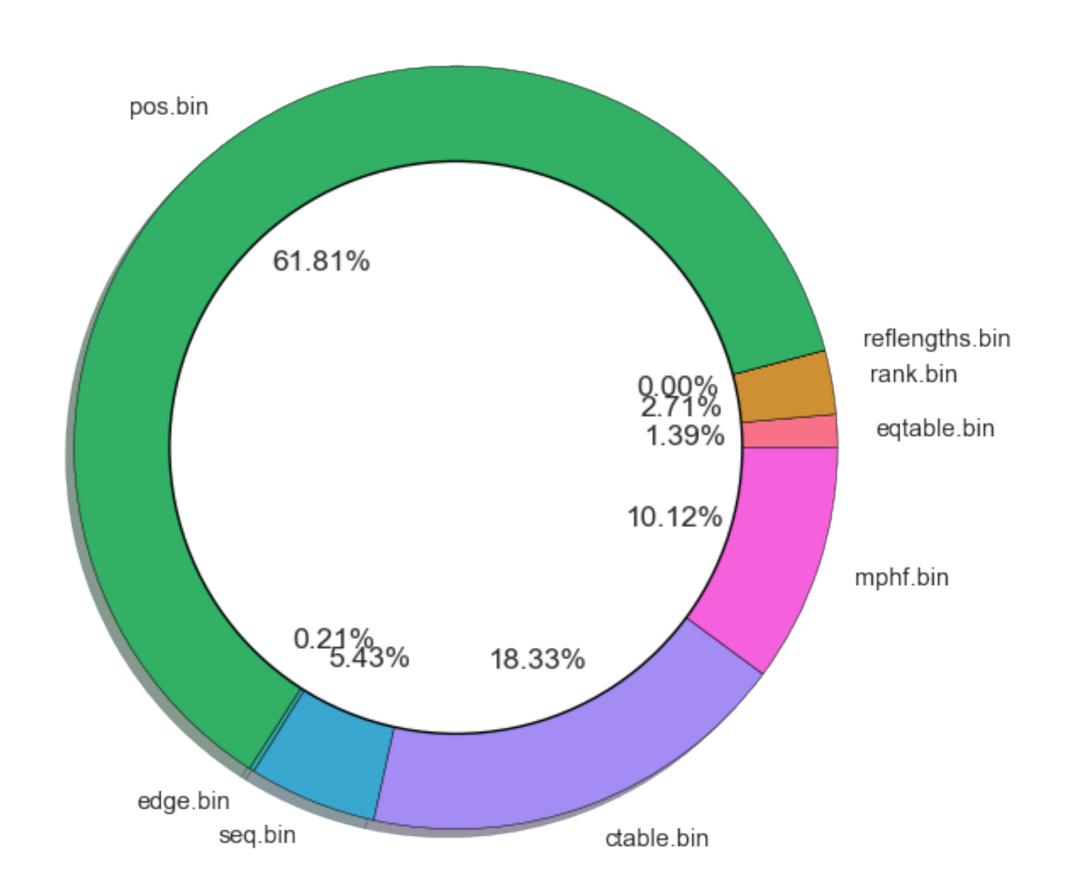


- useq contains the uniting sequences concatenated together
- bv is a boundary vector that records a 1
 at the end of each uniting, and a 0
 elsewhere

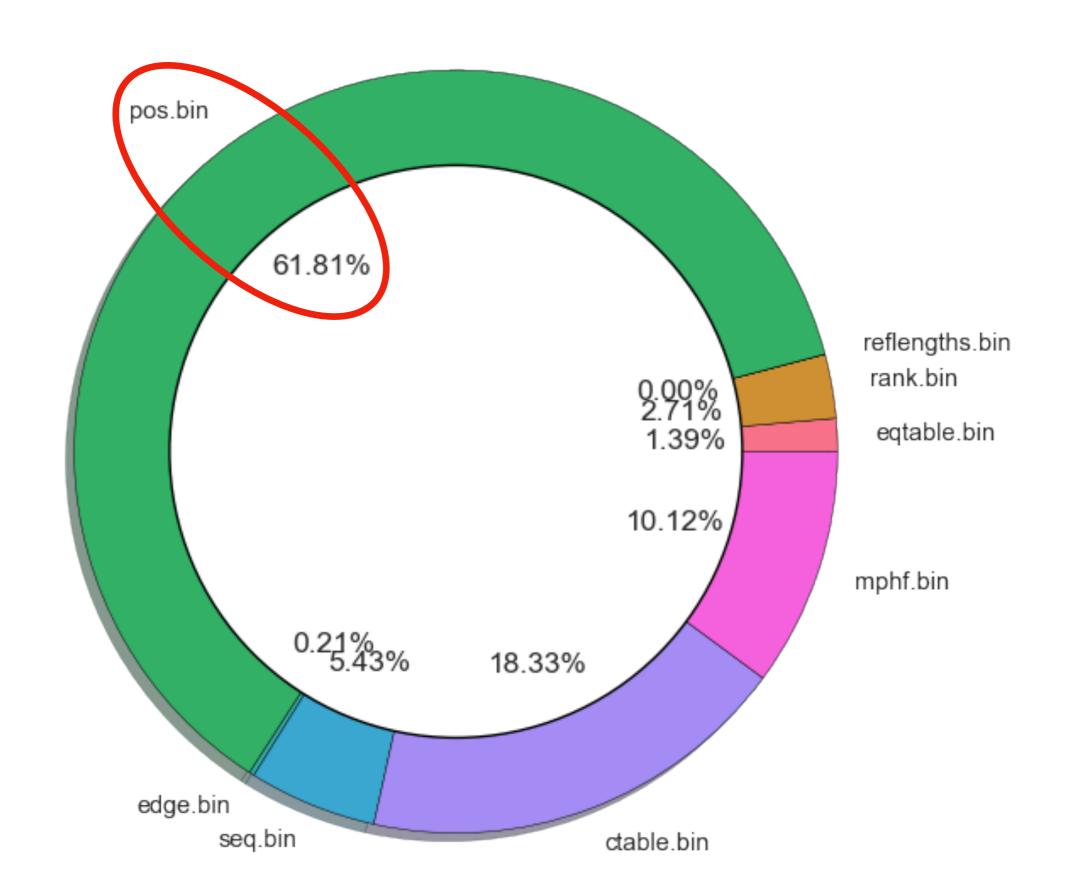




Who's the culprit?

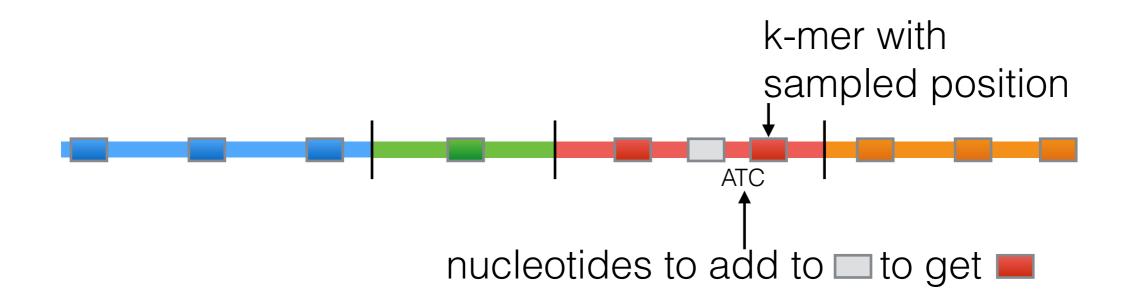


Who's the culprit?



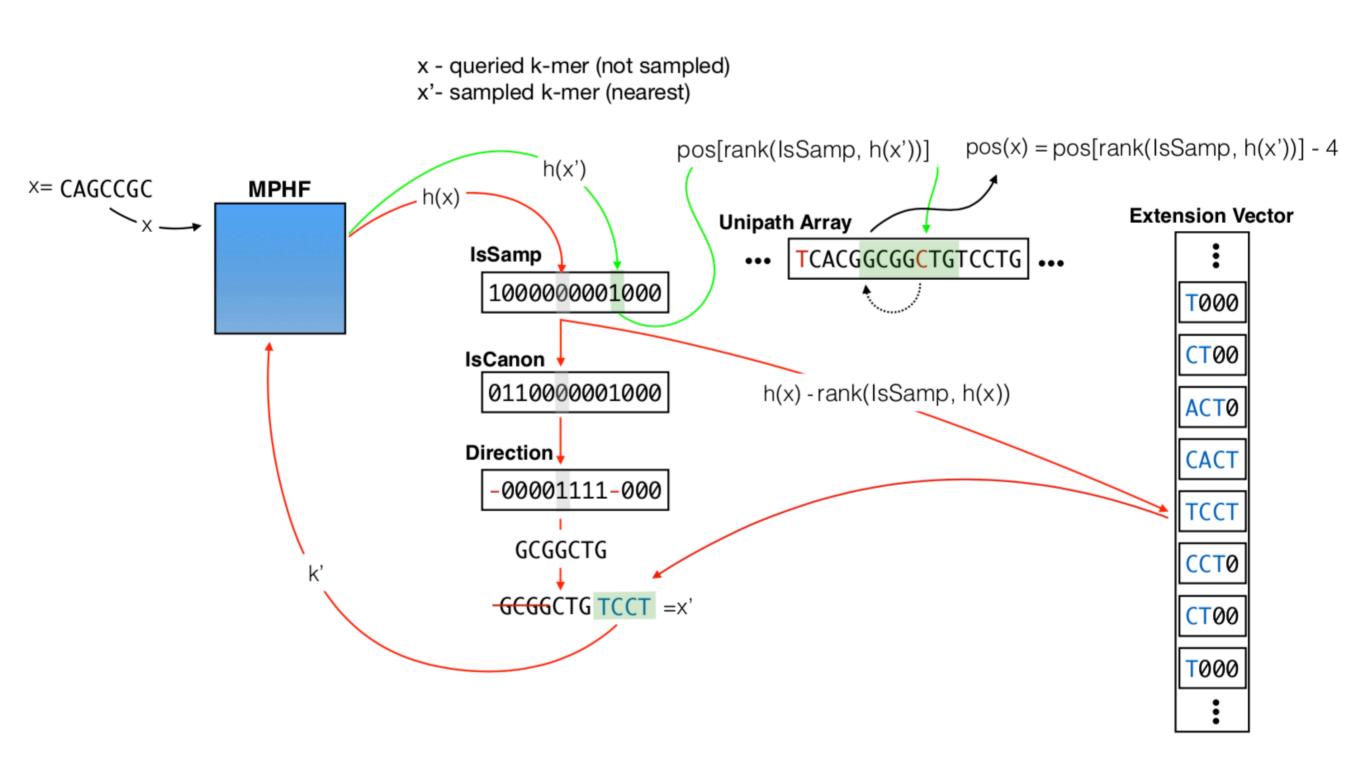
The sparse Pufferfish index

In large indices, the position table dominates index size



Intuition: Successors and predecessors in unipaths are *globally unique*, instead of storing position information for all k-mers, store positions only at sampled "landmarks" and say how to navigate to these landmarks (similar to bi-directional sampling in the FM-index).

The sparse Pufferfish index (in detail)

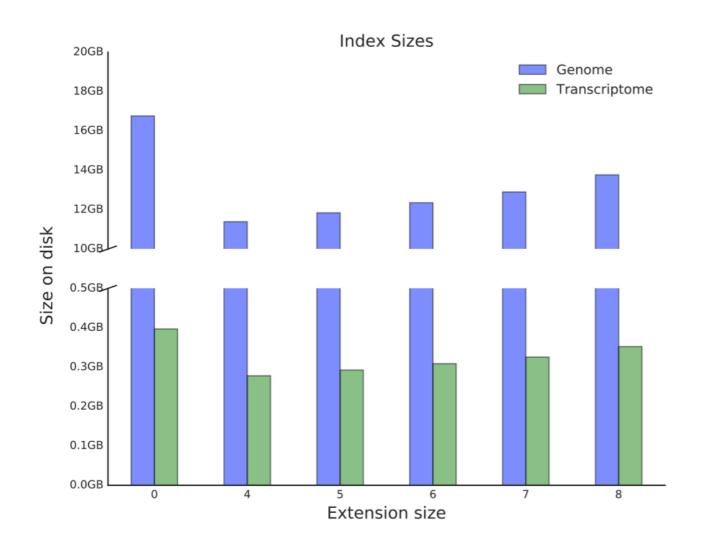


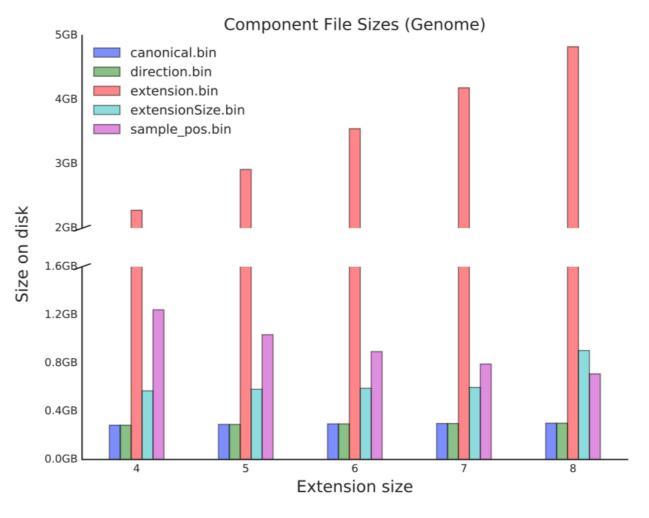
What sampling factor is right?

Tradeoff: Sparser sampling → less space but slower lookup

Fastest: Sampling factor $s > 2 \cdot e + 1$ (Still a range of sizes)

Smallest: Extension size = 1, sampling = s





Index space & K-mer query time

Space of index + query in RAM

Tool	Memory (MB)		
	Human	Human	Bacterial
	Transcriptome	Genome	Genome
BWA	308	4,439	27,535
kallisto	3,336	110,464	232,353
pufferfish dense	454	17,684	41,532
pufferfish sparse	341	$12,\!533$	$30,\!565$

Index space & K-mer query time

Time to look up all fixed-length substrings in an experiment

Tool	Time (h:m:s)		
	Human Transcriptome	Human Genome	Bacterial Genome
BWA	0:17:35	0:50:31	0:14:05
kallisto pufferfish dense	$0:02:01 \\ 0:02:46$	0:19:11 0:10:37	$0:22:25 \\ 0:06:03$
pufferfish sparse		0:22:11	0:08:26
# querie	es: 747,842,900	7,508,576,020	509,143,360

Pufferfish summary (part 1)

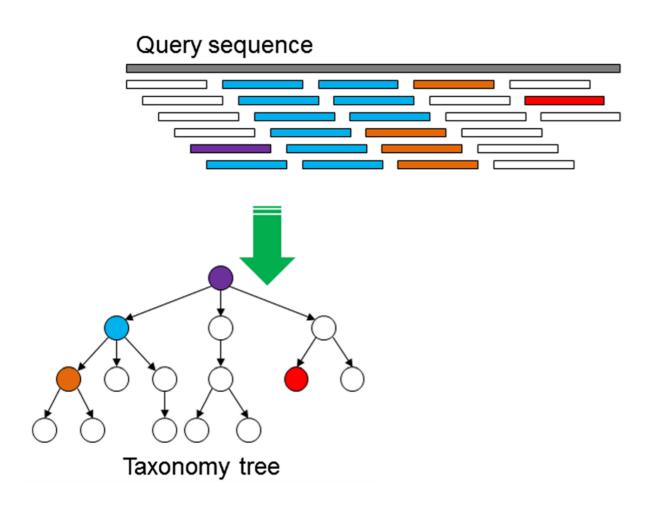
•To keep memory usage reasonable, we have to be quite careful about our hashing-based schemes.

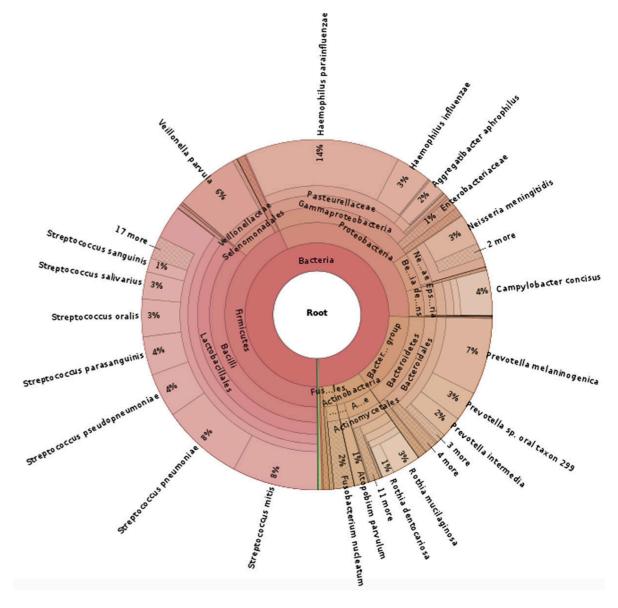
•The dense pufferfish index strikes a good balance between index space and raw query speed.

- •At a constant factor (though not asymptotic) cost, index size is tunable with our sampling scheme.
- •At least for fixed-length patterns, a good hashing approach can be *much faster* than (still asymptotically-optimal) full-text indexes.

An example application of Pufferfish

•Taxonomic read classification — for each read, assign it to the taxon (strain, species, genus) from which we think it derived. Related to, **but distinct from**, taxonomic abundance estimation.





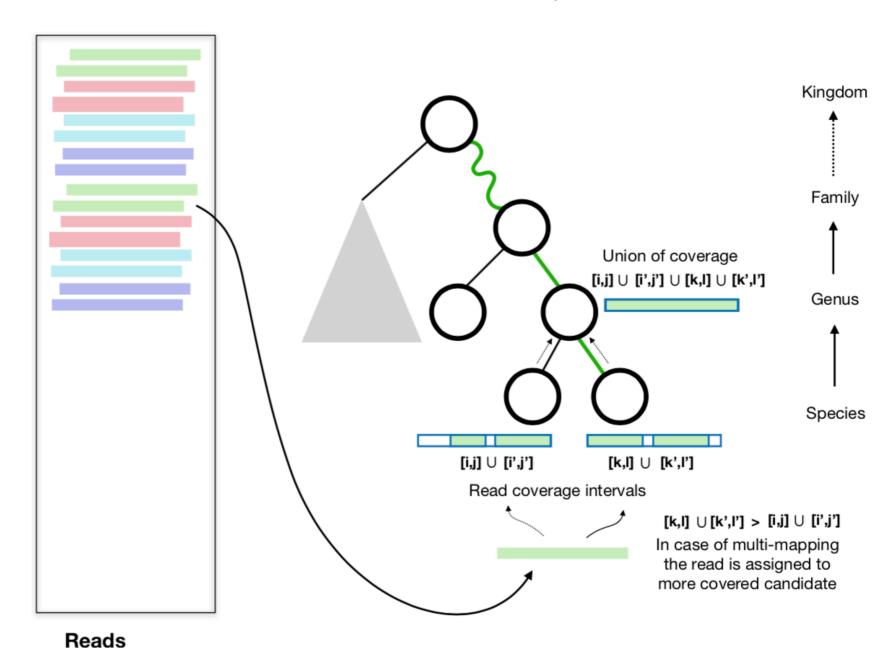
Figures adapted from: Wood, D.E. and Salzberg, S.L., 2014. Kraken: ultrafast metagenomic sequence classification using exact alignments. *Genome biology*, *15*(3), p.R46.

Pufferfish taxonomic assignment

We adopt what is essentially the algorithm of *Kraken**, but replace k-mer counting with lightweight mapping.

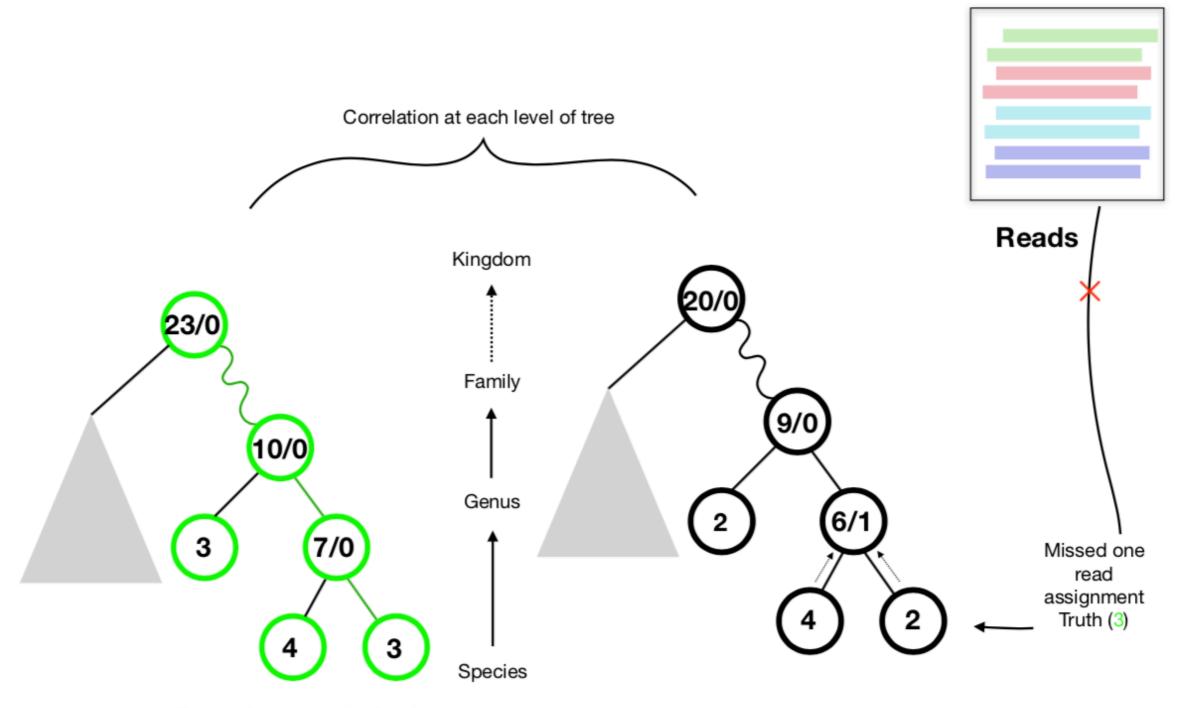
This enforces positional & orientation consistency of matches

- Score all root-to-leaf (RTL) paths
- Assign read to leaf of highest-scoring path
- In case of tie, assign read to LCA of all highest-scoring paths.



^{*}Wood, D.E. and Salzberg, S.L., 2014. Kraken: ultrafast metagenomic sequence classification using exact alignments. Genome biology, 15(3), p.R46.

"Whole taxonomy" accuracy assessment

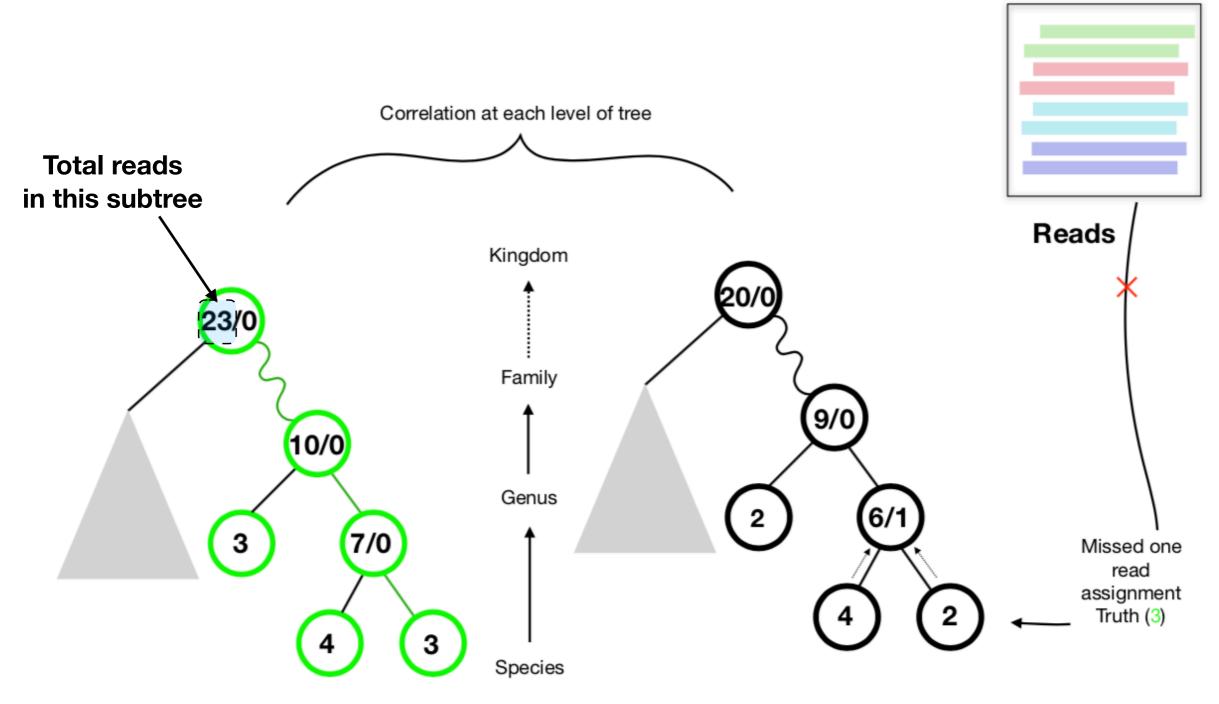


True assignment to leaf nodes

Truth

Pufferfish taxonomic read assignment

"Whole taxonomy" accuracy assessment

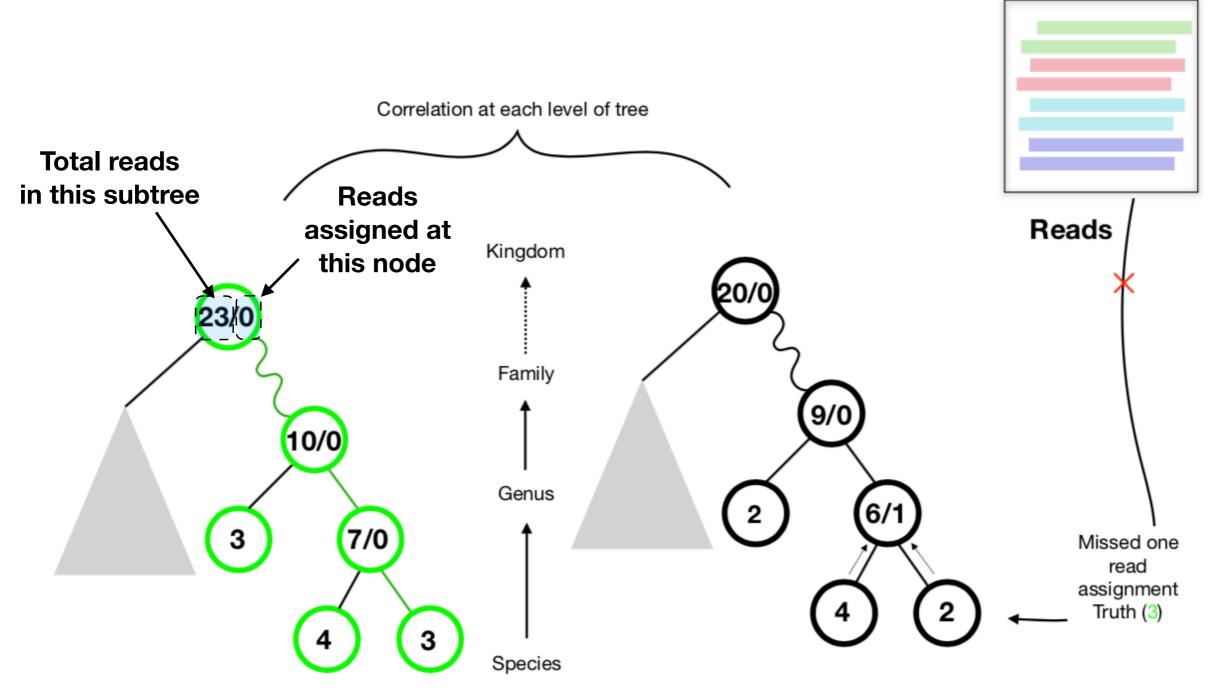


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"Whole taxonomy" accuracy assessment

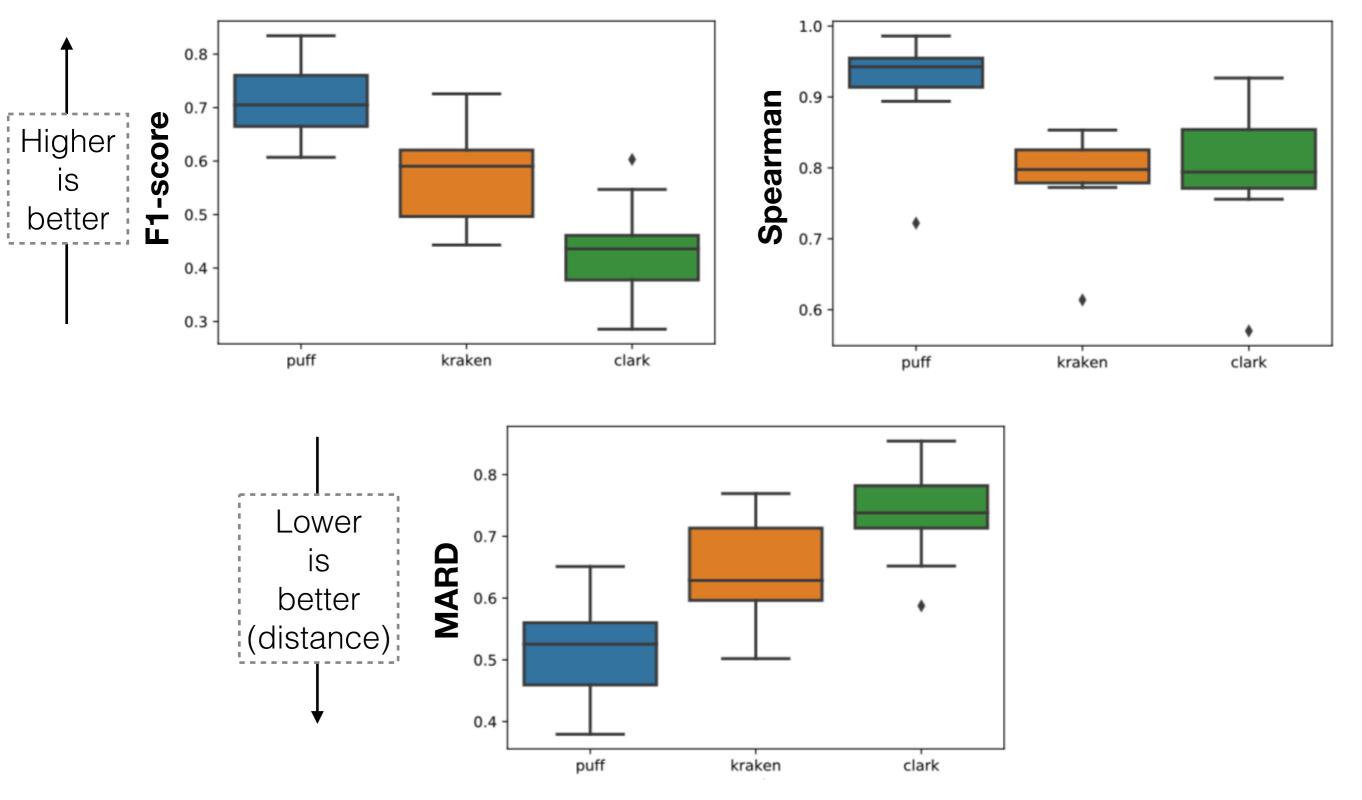


True assignment to leaf nodes

Truth

Pufferfish taxonomic read assignment

Pufferfish taxonomic assignment



Simulated data from : McIntyre, et al. (2017). Comprehensive benchmarking and ensemble approaches for metagenomic classifiers. Genome Biology, 18(1). Simulations: (LC1-8, HC1, HC2)