Package 'STITCH'

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Type Package

Title STITCH - Sequencing To Imputation Through Constructing Haplotypes

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Description STITCH performs imputation of individuals sequenced to low coverage in a read aware fashion without a reference panel.

Installation To install, first install dependencies, then run the install.packages command, pointing to the downloaded tarball (STITCH.tar.gz)

Getting started A minimum run requires the following options to be set: the chromosome being run (chr); a path to a file with a set of bi-allelic SNP sites (posfile); a choice of K, the number of internally modelled haplotypes (K); a path to an output directory (outputdir); a path to a temporary directory, ideally on fast disks or a RAM disk (tempdir); a list of bam files (bamlist); and the number of generations since founding (nGen), which can be approximated from a choice of K for wild populations from 4 * Ne / K. Additional useful options relate to what region to impute (regionStart, regionEnd, buffer), whether to use validation data to benchmark imputation (genfile), the number of cores to use (nCores), whether imputation is run on a server or cluster (environment), the number of EM iterations (niterations), whether to run in diploid or pseudoHaploid mode (method), and if run in pseudoHaploid mode, what iteration to switch from pseudoHaploid to diploid (switchModelIteration).

Depends R (>= 3.6.0), parallel, rrbgen (>= 0.0.4)

Imports Rcpp, data.table

Suggests testthat

Remotes github::rwdavies/rrbgen/rrbgen

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```
LinkingTo Rcpp, RcppArmadillo
```

RoxygenNote 7.3.2

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BugReports https://github.com/rwdavies/STITCH/issues

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        extract_hd_to_cube
        Extract ancestral haplotype dosage to RData cube
```

Description

Extract ancestral haplotype dosage to RData cube

Usage

```
extract_hd_to_cube(
   vcf_file,
   ref,
   bcftools = "bcftools",
   gatk_jar =
      "/data/smew1/rdavies/stitch_richard_paper/bin/gatk/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1c3ef/GenomeAnalysisTK-3.8-1-0-gf15c1
```

Arguments

```
vcf_file
                  path to VCF
ref
                  path to reference fasta (required by GATK)
                  path to veftools (or just "beftools" if in path)
bcftools
                  path to GATK jar
gatk_jar
samples
                  vector of sample names (or NULL for all)
                  pos matrix, a matrix with at least two columns where the first two columns
pos
                  are chrom and 1-based physical position, respectively. specifies which SNPs to
                  extract (or NULL for all)
field
                  What to get from the VCF. Default HD for haplotype dosages
```

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Value

A cube with dimensions of SNPs x samples x ancestral haplotypes

Author(s)

Robert Davies

make_STITCH_cli

Make STITCH command line interface

Description

Make STITCH command line interface

Usage

```
make_STITCH_cli(
  function_file,
  cli_output_file,
  integer_vectors = c("shuffleHaplotypeIterations", "splitReadIterations",
        "refillIterations", "reference_shuffleHaplotypeIterations"),
  character_vectors = c("reference_populations"),
  other_logical_params = NULL,
  other_integer_params = NULL,
  other_double_params = NULL,
  other_character_params = NULL,
  function_name = "STITCH",
  library_name = "STITCH"
)
```

Arguments

STITCH

Sequencing To Imputation Through Constructing Haplotypes

Description

Sequencing To Imputation Through Constructing Haplotypes

Usage

```
STITCH(
  chr,
 nGen,
 posfile,
 Κ,
  S = 1,
 outputdir,
 nStarts,
  tempdir = NA,
 bamlist = "",
  cramlist = ""
  sampleNames_file = "",
  reference = ""
 genfile = "",
 method = "diploid",
 output_format = "bgvcf",
 B_bit_prob = 16,
  outputInputInVCFFormat = FALSE,
  downsampleToCov = 50,
  downsampleFraction = 1,
  readAware = TRUE,
  chrStart = NA,
  chrEnd = NA,
  regionStart = NA,
  regionEnd = NA,
 buffer = NA,
 maxDifferenceBetweenReads = 1000,
 maxEmissionMatrixDifference = 1e+10,
  alphaMatThreshold = 1e-04,
  emissionThreshold = 1e-04,
  iSizeUpperLimit = as.integer(600),
  bqFilter = as.integer(17),
  niterations = 40,
  shuffleHaplotypeIterations = c(4, 8, 12, 16),
  splitReadIterations = 25,
  nCores = 1,
  expRate = 0.5,
 maxRate = 100,
```

```
minRate = 0.1,
  Jmax = 1000,
  regenerateInput = TRUE,
  originalRegionName = NA,
  keepInterimFiles = FALSE,
  keepTempDir = FALSE,
  outputHaplotypeProbabilities = FALSE,
  switchModelIteration = NA,
  generateInputOnly = FALSE,
  restartIterations = NA,
  refillIterations = c(6, 10, 14, 18),
  downsampleSamples = 1,
  downsampleSamplesKeepList = NA,
  subsetSNPsfile = NA,
  useSoftClippedBases = FALSE,
  outputBlockSize = 1000,
  outputSNPBlockSize = 10000,
  inputBundleBlockSize = NA,
  genetic_map_file = "",
  reference_haplotype_file = "",
  reference_legend_file = "",
  reference_sample_file = "",
  reference_populations = NA,
  reference_phred = 20,
  reference_iterations = 40,
  reference_shuffleHaplotypeIterations = c(4, 8, 12, 16),
  output_filename = NULL,
  initial_min_hapProb = 0.2,
  initial_max_hapProb = 0.8,
  regenerateInputWithDefaultValues = FALSE,
  plotHapSumDuringIterations = FALSE,
  plot_shuffle_haplotype_attempts = FALSE,
  plotAfterImputation = TRUE,
  plotReferenceAlleleCount = TRUE,
  save_sampleReadsInfo = FALSE,
  gridWindowSize = NA,
  shuffle_bin_nSNPs = NULL,
  shuffle_bin_radius = 5000,
  keepSampleReadsInRAM = FALSE,
  useTempdirWhileWriting = FALSE,
  output_haplotype_dosages = FALSE,
  use_bx_tag = TRUE,
  bxTagUpperLimit = 50000,
  do_phasing = FALSE
)
```

Arguments

chr

What chromosome to run. Should match BAM headers

nGen Number of generations since founding or mixing. Note that the algorithm is

relatively robust to this. Use nGen = 4 * Ne / K if unsure

posfile Where to find file with positions to run. File is tab seperated with no header,

one row per SNP, with col 1 = chromosome, col 2 = physical position (sorted from smallest to largest), col 3 = reference base, col 4 = alternate base. Bases

are capitalized. Example first row: 1<tab>1000<tab>A<tab>G<tab>

K How many founder / mosaic haplotypes to use

S How many sets of founder / mosaic haplotypes to use

outputdir What output directory to use

tempdir What directory to use as temporary directory. If set to NA, use default R tempdir.

If possible, use ramdisk, like /dev/shm/

bamlist Path to file with bam file locations. File is one row per entry, path to bam files.

Bam index files should exist in same directory as for each bam, suffixed either

.bam.bai or .bai

cramlist Path to file with cram file locations. File is one row per entry, path to cram files.

cram files are converted to bam files on the fly for parsing into STITCH

sampleNames_file

Optional, if not specified, sampleNames are taken from the SM tag in the header of the BAM / CRAM file. This argument is the path to file with sampleNames for samples. It is used directly to name samples in the order they appear in the

bamlist / cramlist

reference Path to reference fasta used for making cram files. Only required if cramlist is

defined

genfile Path to gen file with high coverage results. Empty for no genfile. File has a

header row with a name for each sample, matching what is found in the bam file. Each subject is then a tab seperated column, with 0 = hom ref, 1 = het, 2 = hom alt and NA indicating missing genotype, with rows corresponding to rows of the posfile. Note therefore this file has one more row than posfile which has

no header

method How to run imputation - either diploid, pseudoHaploid, or diploid-inbred. Please

see main README for more information. All methods assume diploid samples. diploid is the most accurate but slowest, while pseudoHaploid may be advantageous for large sample sizes and K. diploid-inbred assumes all samples are inbred and invokes an internal haploid mathematical model but outputs diploid

genotypes and probabilities

output_format one of bgvcf (i.e. bgziped VCF) or bgen (Layout = 2, CompressedSNPBlocks =

1)

B_bit_prob when using bgen, how many bits to use to store each double. Optiosn are 8, 16,

24 or 32

outputInputInVCFFormat

Whether to output the input in vcf format

downsampleToCov

What coverage to downsample individual sites to. This ensures no floating point

errors at sites with really high coverage

downsampleFraction

Downsample BAMs by choosing a fraction of reads to retain. Must be value

0<downsampleFraction<1

readAware Whether to run the algorithm is read aware mode. If false, then reads are split

into new reads, one per SNP per read

chrStart When loading from BAM, some start position, before SNPs occur. Default NA

will infer this from either regionStart, regionEnd and buffer, or posfile

chrEnd When loading from BAM, some end position, after SNPs occur. Default NA

will infer this from either regionStart, regionEnd and buffer, or posfile

regionStart When running imputation, where to start from. The 1-based position x is kept if

regionStart <= x <= regionEnd

regionEnd When running imputation, where to stop.

buffer Buffer of region to perform imputation over. So imputation is run form regionStart-

buffer to regionEnd+buffer, and reported for regionStart to regionEnd, including

the bases of regionStart and regionEnd

maxDifferenceBetweenReads

How much of a difference to allow the reads to make in the forward backward probability calculation. For example, if P(read | state 1)=1 and P(read | state 2)=1e-6, re-scale so that their ratio is this value. This helps prevent any individual read as having too much of an influence on state changes, helping prevent

against influence by false positive SNPs

maxEmissionMatrixDifference

Similar to maxDifferenceBetweenReads, specifies ratio of how much larger the most probable state can be than the least probable state, but across all reads rather than for a single read. This helps to limit overflow in C++ calculations

alphaMatThreshold

Minimum (maximum is 1 minus this) state switching into probabilities

emissionThreshold

Emission probability bounds. emissionThreshold < P(alt read | state k) < (1-

emissionThreshold)

iSizeUpperLimit

Do not use reads with an insert size of more than this value

bqFilter Minimum BQ for a SNP in a read. Also, the algorithm uses bq<=mq, so if

mapping quality is less than this, the read isnt used

niterations Number of EM iterations.

shuffleHaplotypeIterations

Iterations on which to perform heuristic attempt to shuffle founder haplotypes

for better fit. To disable set to NA.

splitReadIterations

Iterations to try and split reads which may span recombination breakpoints for a

better fit

nCores How many cores to use

expRate Expected recombination rate in cM/Mb

maxRate Maximum recomb rate cM/Mb

minRate Minimum recomb rate cM/Mb

Jmax Maximum number of SNPs on a read

regenerateInput

Whether to regenerate input files. If this is FALSE, please using the same region-Start, regionEnd, buffer and posfile as you used to generate the input. Setting any of those to different values can cause the previous input data to be improperly interpreted. Please also see originalRegionName and regenerateInputWithDe-

faultValues

original Region Name

If regenerateInput is FALSE (i.e. using existing data), this is the name of the original region name (chr.regionStart.regionEnd). This is necessary to load past variables

keepInterimFiles

Whether to keep interim parameter estimates

keepTempDir Whether to keep files in temporary directory

switchModelIteration

Whether to switch from pseudoHaploid to diploid and at what iteration (NA for no switching)

generateInputOnly

Whether to just generate input data then quit

restartIterations

In pseudoHaploid method, which iterations to look for collapsed haplotype prnobabilities to resolve

refillIterations

When to try and refill some of the less frequently used haplotypes

downsampleSamples

What fraction of samples to retain. Useful for checking effect of N on imputation. Not meant for general use

downsampleSamplesKeepList

When downsampling samples, specify a numeric list of samples to keep

subsetSNPsfile If input data has already been made for a region, then subset down to a new set of SNPs, as given by this file. Not meant for general use

useSoftClippedBases

Whether to use (TRUE) or not use (FALSE) bases in soft clipped portions of reads

outputBlockSize

How many samples to write out to disk at the same time when making temporary VCFs that are later pasted together at the end to make the final VCF. Smaller means lower RAM footprint, larger means faster write.

outputSNPBlockSize

How many SNPs to write to disk at one time to reduce RAM usage when making VCFs

inputBundleBlockSize

If NA, disable bundling of input files. If not NA, bundle together input files in sets of <= inputBundleBlockSize together

genetic_map_file

Path to file with genetic map information, a file with 3 white-space delimited entries giving position (1-based), genetic rate map in cM/Mbp, and genetic map in cM

reference_haplotype_file

Path to reference haplotype file in IMPUTE format (file with no header and no rownames, one row per SNP, one column per reference haplotype, space separated, values must be 0 or 1)

reference_legend_file

Path to reference haplotype legend file in IMPUTE format (file with one row per SNP, and a header including position for the physical position in 1 based coordinates, a0 for the reference allele, and a1 for the alternate allele)

reference_sample_file

Path to reference sample file (file with header, one must be POP, corresponding to populations that can be specified using reference_populations)

reference_populations

Vector with character populations to include from reference_sample_file e.g. CHB, CHS

reference_phred

Phred scaled likelihood or an error of reference haplotype. Higher means more confidence in reference haplotype genotypes, lower means less confidence

reference_iterations

When using reference haplotypes, how many iterations to use to train the starting data

reference_shuffleHaplotypeIterations

When using reference haplotypes, how much shuffling to do to lead to better global fit

output_filename

Override the default bgzip-VCF / bgen output name with this given file name. Please note that this does not change the names of inputs or outputs (e.g. RData, plots), so if outputdir is unchanged and if multiple STITCH runs are processing on the same region then they may over-write each others inputs and outputs

initial_min_hapProb

Initial lower bound for probability read comes from haplotype. Double bounded between 0 and 1

initial_max_hapProb

Initial upper bound for probability read comes from haplotype. Double bounded between 0 and 1

regenerateInputWithDefaultValues

If regenerateInput is FALSE and the original input data was made using region-Start, regionEnd and buffer as default values, set this equal to TRUE

plotHapSumDuringIterations

Boolean TRUE/FALSE about whether to make a plot that shows the relative number of individuals using each ancestral haplotype in each iteration

plot_shuffle_haplotype_attempts

Boolean TRUE/FALSE about whether to make a plot that tries to show the selection of ancestral haplotypes to check for shuffling / flipping

plotAfterImputation

Boolean TRUE/FALSE about whether to make plots after imputation has run (can be set to FALSE if this throws errors on systems without x11)

save_sampleReadsInfo

Experimental. Boolean TRUE/FALSE about whether to save additional information about the reads that were extracted

gridWindowSize Whether to work on a grid where reads are binned into windows of this size (1 based, i.e. first bin is bases 1-gridWindowSize). This is particularly appropriate for very low coverage data (e.g. less than 0.2X) and can substantially speed up analyses

shuffle_bin_nSNPs

Parameter that controls how to detect ancestral haplotypes that are shuffled during EM for possible re-setting. If set (not NULL), then break per-SNP (or per-grid) every this many SNPs / grids, and compare each to detect whether haplotypes either 1) are more likely to stay where they are or 2) switch from one haplotype to another. Note that only one of shuffle_bin_nSNPs or shuffle_bin_radius should be non-NULL

shuffle_bin_radius

Parameter that controls how to detect ancestral haplotypes that are shuffled during EM for possible re-setting. If set (not NULL), then recombination rate is calculated around pairs of SNPs in window of twice this value, and those that exceed what should be the maximum (defined by nGen and maxRate) are checked for whether they are shuffled

keepSampleReadsInRAM

Whether to (generally) keep sampleReads in RAM or store them in the temporary directory. STITCH is substantially faster if this is FALSE at the expense of **RAM**

useTempdirWhileWriting

Whether to use temporary directory while writing output file (TRUE), or to keep result in RAM (FALSE). Using temporary directory is slower but uses less RAM output_haplotype_dosages

Whether to output ancestral haplotype dosages, i.e. the expected number of ancestral haplotypes carried by that sample at that locus

Whether to try and use BX tag in same to indicate that reads come from the use_bx_tag same underlying molecule

bxTagUpperLimit

When using BX tag, at what distance between reads to consider reads with the same BX tag to come from different molecules

Whether to try and output phasing (experimental) do_phasing

Value

Results in properly formatted version

Author(s)

Robert Davies

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