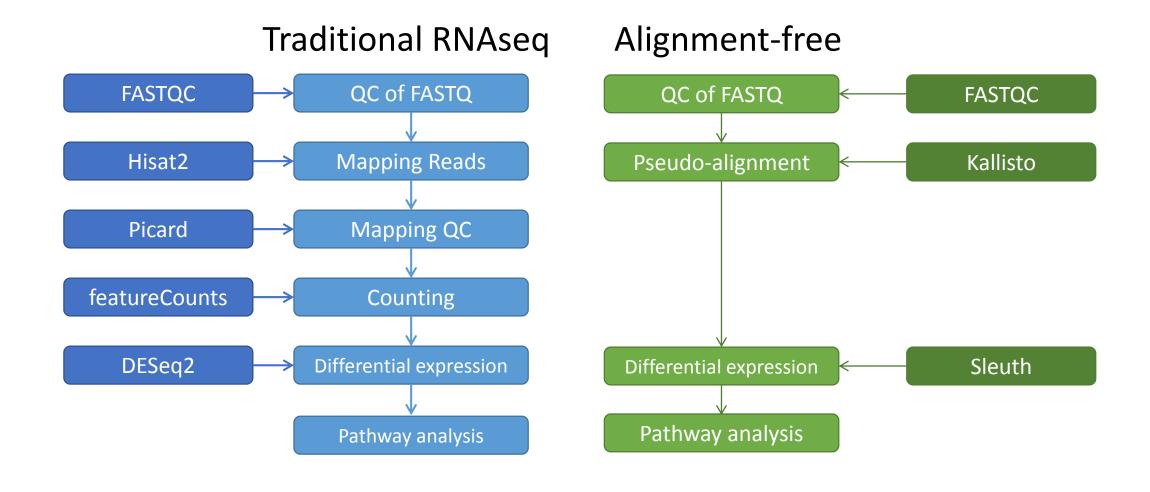
# Alignment-free RNAseq

Oxford Biomedical Data Science Training Programme



### Alignment-free Quantification

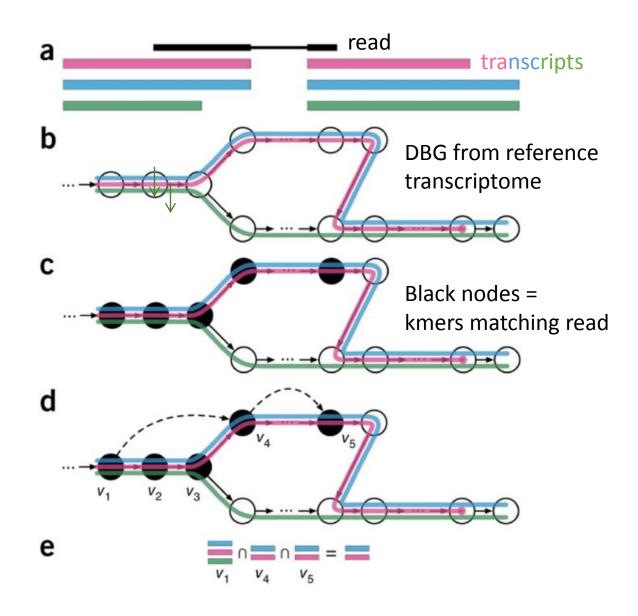
- Also known as Pseudoalignment
- Work directly from Fastq
- Specify which transcript the read is from NOT where in sequence it is!
- No base level information
- Transcript level quantification
- Output is a quantification table for each sample -> quant.sf
  - Rows = transcripts
  - Quantification Values = Transcripts Per Million
- Very fast can run on laptop
- Better accuracy then traditional method
- Can only quantify known transcripts
- Does not produce a BAM file no visualisation

#### Alignment-free Quantification Tools

NATURE BIOTECHNOLOGY | RESEARCH | BRIEF COMMUNICATIONS 日本語要約 Sailfish enables alignment-free isoform quantification from RNA-seq reads using lightweight algorithms Rob Patro, Stephen M Mount & Carl Kingsford Affiliations | Contributions | Corresponding author NATURE BIOTECHNOLOGY | RESEARCH | BRIEF COMMUNICATIONS 日本語要約 Nature Biotechnology 32, 462-464 (2014) | doi:10.1038/nbt.2862 Near-optimal probabilistic RNA-seq quantification Nicolas L Bray, Harold Pimentel, Páll Melsted & Lior Pachter Affiliations | Contributions | Corresponding author Nature Biotechnology **34**, 525–527 (2016) | doi:10.1038/nbt.3519 NATURE METHODS | BRIEF COMMUNICATION http://robpatro.com/blog/?p=248 Salmon provides fast and bias-aware quantification of transcript expression Not-quite alignments: Salmon, **Kallisto and Efficient** Rob Patro, Geet Duggal, Michael I Love, Rafael A Irizarry & Carl Kingsford **Quantification of RNA-Seq** Affiliations | Contributions | Corresponding authors data Nature Methods 14, 417-419 (2017) | doi:10.1038/nmeth.4197

#### For each read find compatible transcripts

- Build a coloured De-Bruijn graph from all transcripts
  - Each circle/node is a Kmer
  - Each kmer is associated with set of transcripts
    - 'kmer compatibility class'
- Decompose each read into kmers
  - Compare with graph
- Take the intersection of compatibility classes the read matches
  - V1 = match 3 transcripts
  - V4 = match 2 transcripts
  - Intersection = 2 transcripts
- Fast because it doesn't not look up every kmer
  - skips to the node in the next equivalence class



#### Bootstrapping

- 1) Sample reads randomly with replacement
- 2) Estimate transcript abundances
- 3) Repeat 100 times

#### Output:

- Get a confidence interval for abundance of each transcript Purpose:
- Identify transcripts with good quality quantification estimates

ONLY POSSIBLE BECAUSE IT'S SO FAST

#### Gene Annotations

- The quality of the annotations you provide will effect the quality of the output
- What geneset are you using?
  - Ensembl
    - Manually curated
    - Wide range of species
  - Refseq
    - Better for small RNAs
    - Longer first exons

RESEARCH OPEN ACCESS

Isoform prefiltering improves performance of count-based methods for analysis of differential transcript usage

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† Contributed equally

Genome Biology 2016 17:12 | DOI: 10.1186/s13059-015-0862-3 | © Soneson et al. 2016 Received: 11 October 2015 | Accepted: 29 December 2015 | Published: 26 January 2016

## Differential Expression Analysis

- Output = TPM counts table
  - need to convert to use with tools requiring raw counts (DESeq2/edgeR)
  - R txiimport package
- Sleuth is a differential expression program built to interface directly with pseudoalignments
- Transcript level differential expression is hard
  - recommend quantifying at gene level

## Keeping up to date

- Lior Pactor https://liorpachter.wordpress.com
- Rob Patro http://robpatro.com/blog/
- RNASeq blog http://www.rna-seqblog.com/
- Other people to look out for/keep an eye on:
  - Mike Love, Charlotte Soneson, Wolfgang Huber, Simon Anders, Jeff Leek
- For a book that explains the RNA-Seq workflow and considerations from library prep up to differential expression very well see:

RNA-seq Data Analysis: A Practical Approach

Eija Korpelainen, Jarno Tuimala, Panu Somervuo, Mikael Huss, Garry Wong