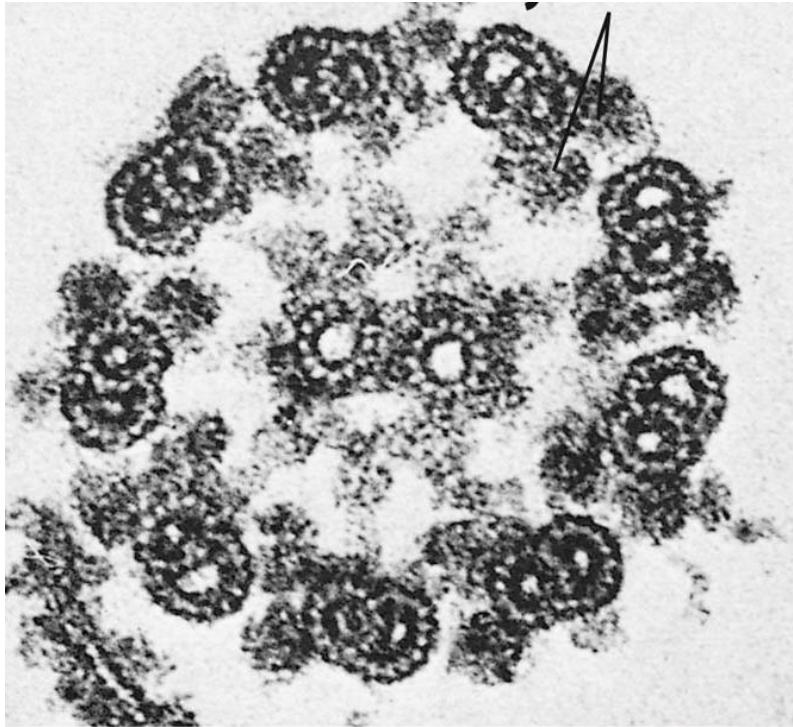
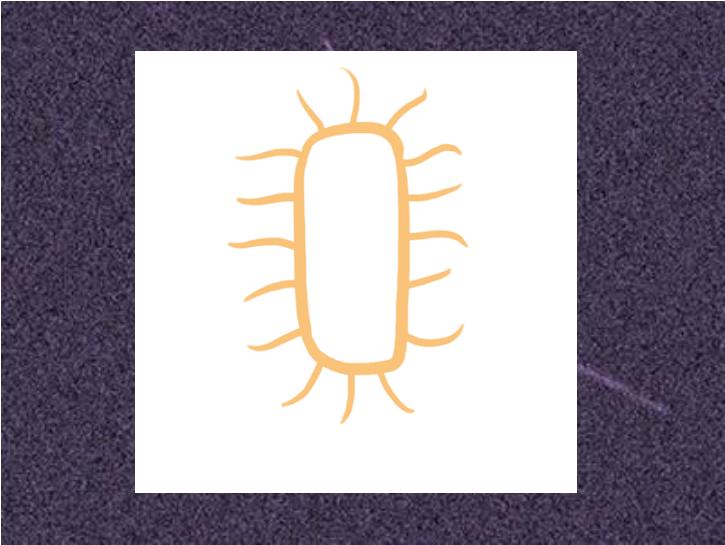


# biol303- cytoskeleton 2

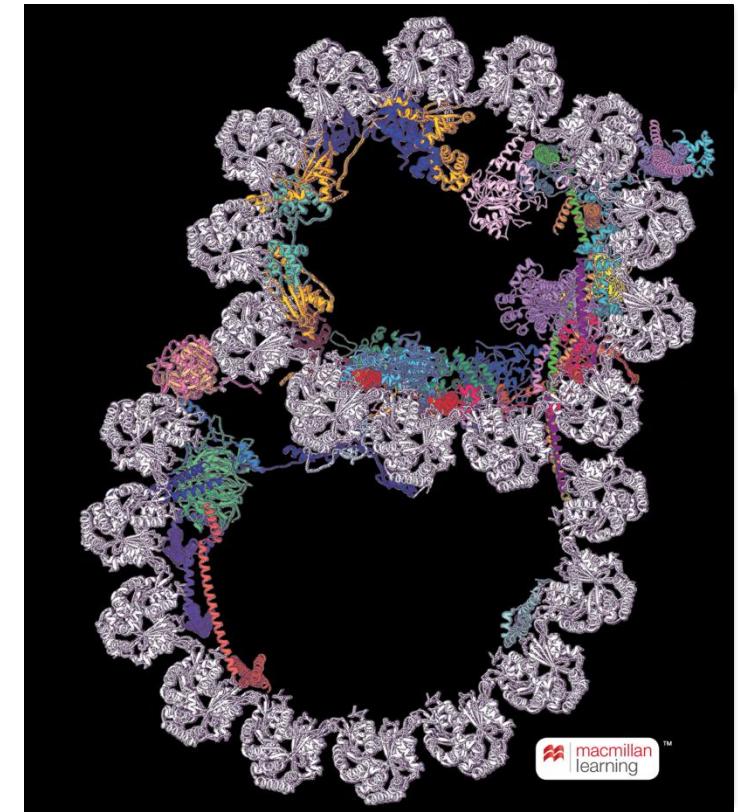
## microtubules, cilia and flagella



now playing: whip it devo



30a Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)



Lodish Ch 18

# overview

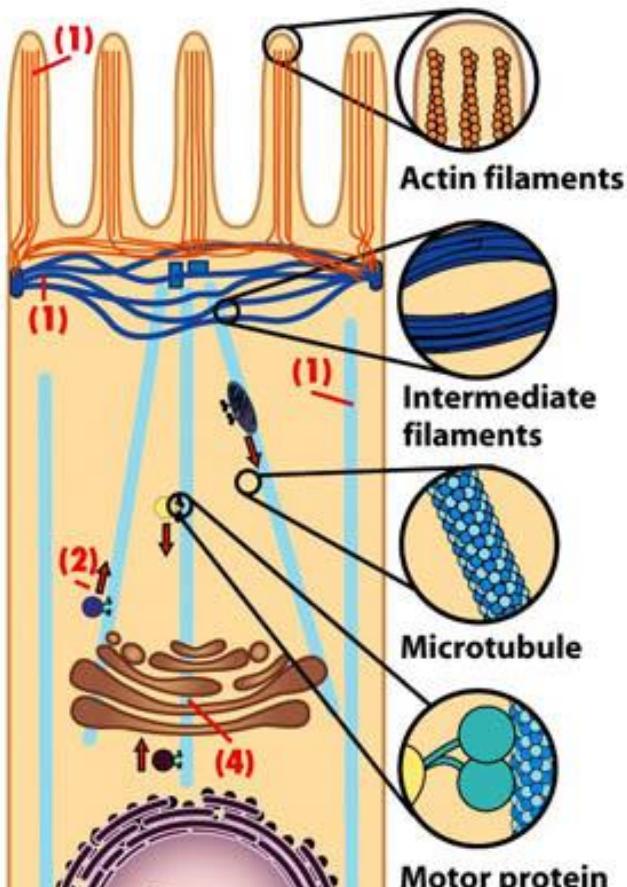
## Key to Cytoskeletal Functions

**(1) Structure and Support**

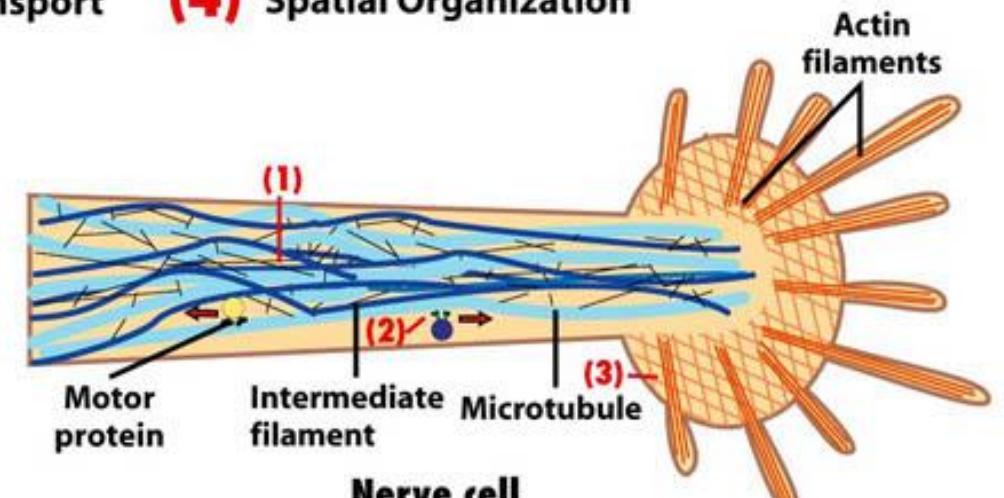
**(2) Intracellular Transport**

**(3) Contractility and Motility**

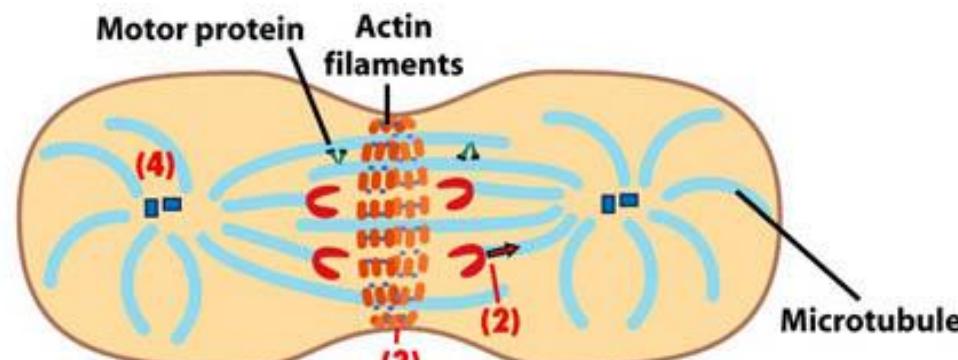
**(4) Spatial Organization**



**(a) Epithelial cell**



**(b)**

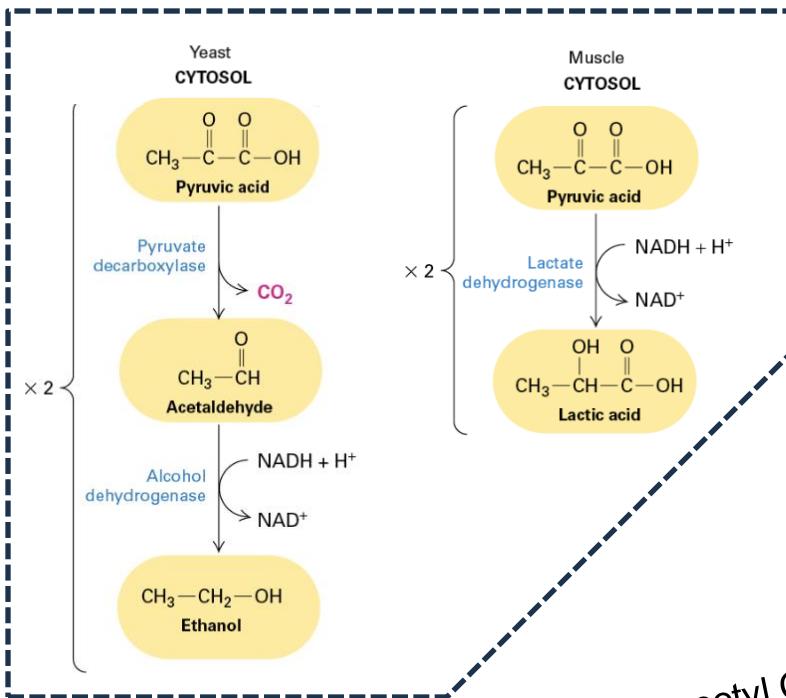
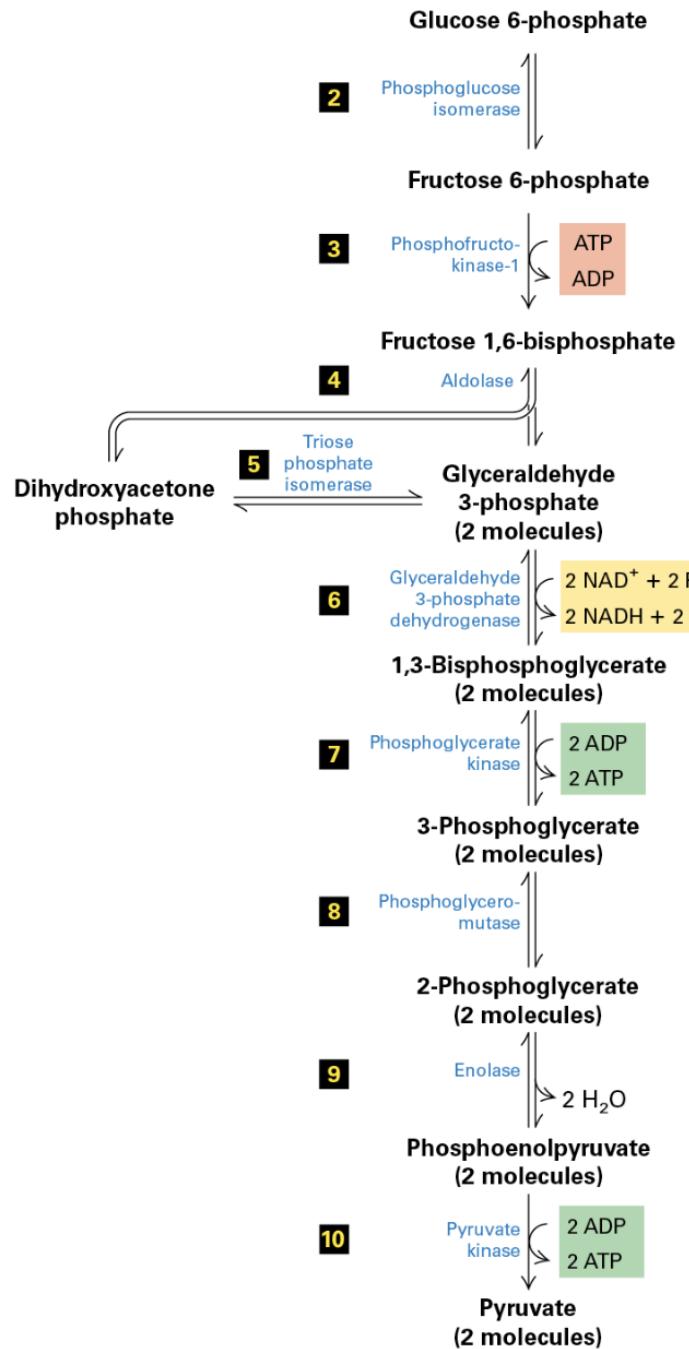


**(c)**

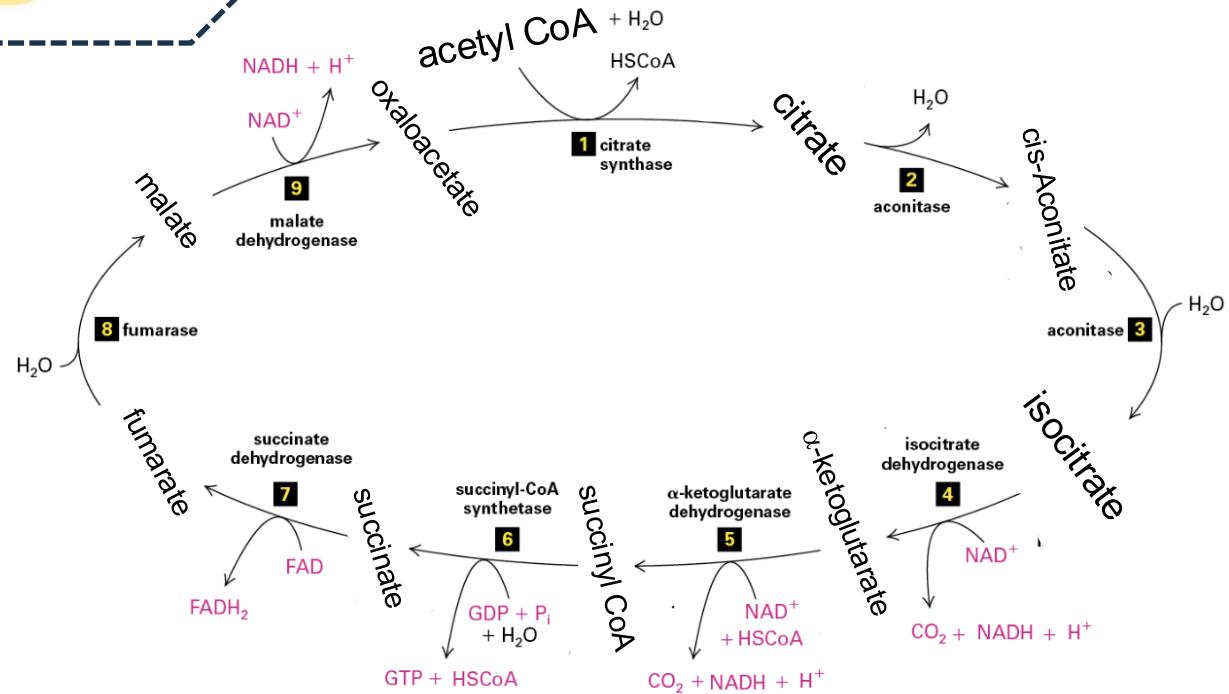
# administrative

- error on Achieve assignment and related things...
- There is discussion homework this week (week 7) and Achieve homework due Friday
- project instructions and grading rubric is posted- more details to follow as you need them – next step is a figure summary
- No Friday office hours with msg- instead: Thurs Oct 16, 4:30-5:30pm this week only
- Exam 2 is in one week- some old exam questions are posted for practice; use your resources (discussion board, SI-PASS, study groups, tutoring, office hours) and know your vocabulary. There will be a reference sheet for substrates/products/enzymes (but not structures) for glycolysis, gluconeogenesis, fermentation, and the Krebs cycle

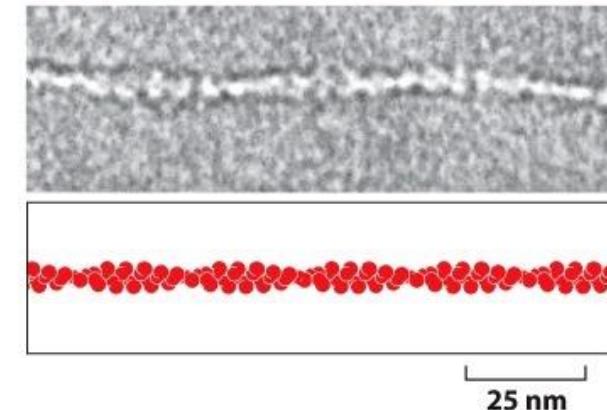
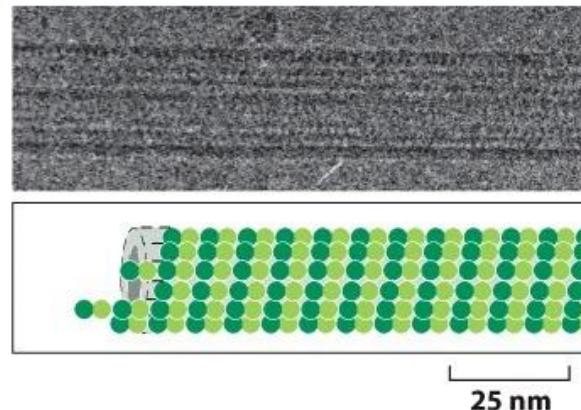
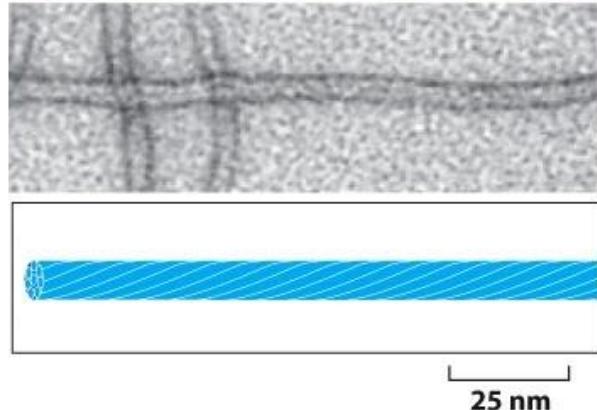
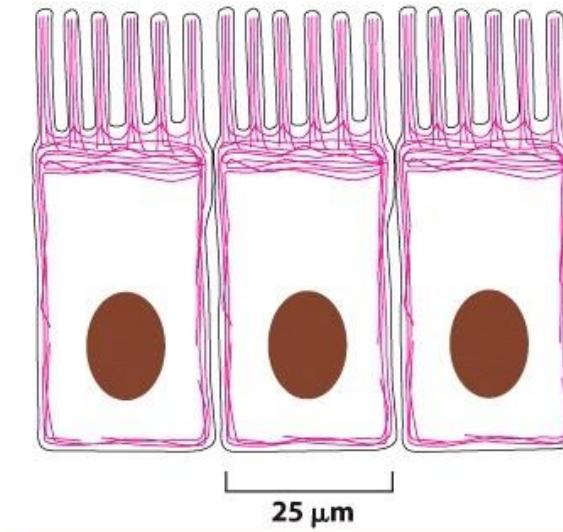
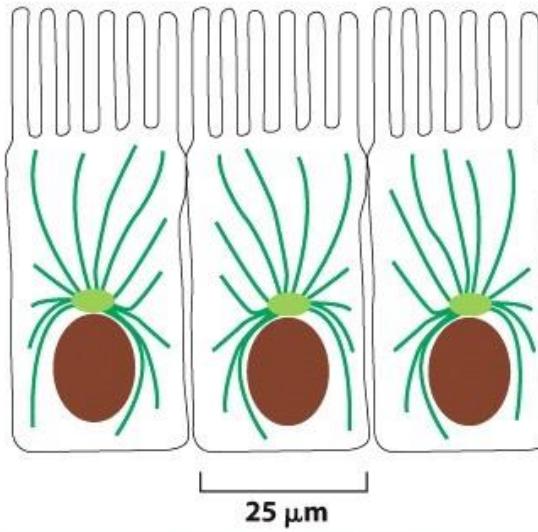
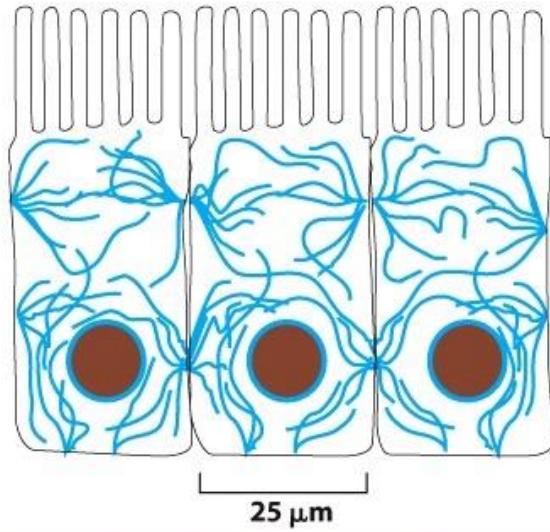
# Metabolic reactions reference sheet



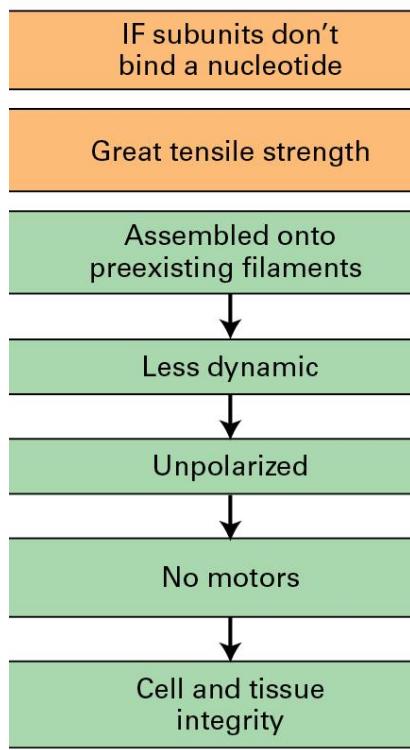
(make sure you know which pathway here is which!)



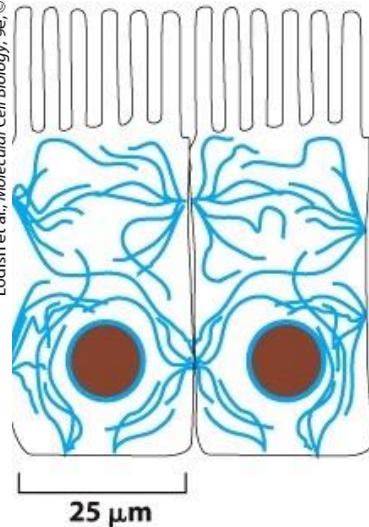
# three types of cytoskeletal filaments...



## Intermediate Filaments



Part (b) Courtesy of Keith Burridge. Part (c) Courtesy of William J. Brown, Cornell University. Part (d) Courtesy of Elaine Fuchs.



## Microtubules

$\alpha\beta$ -Tubulin binds GTP

Rigid and not easily bent

Regulated assembly from a small number of locations

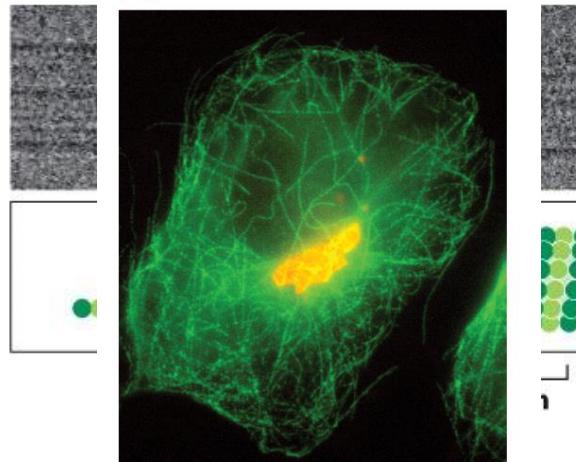
Highly dynamic

Polarized

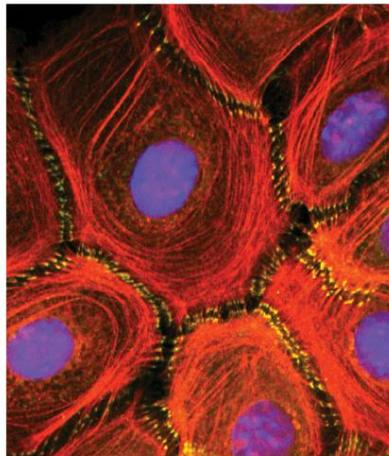
Tracks for kinesins and dyneins

Organization and long-range transport of organelles

(c)



(d)



## Microfilaments

Actin binds ATP

Form rigid gels, networks, and linear bundles

Regulated assembly from a large number of locations

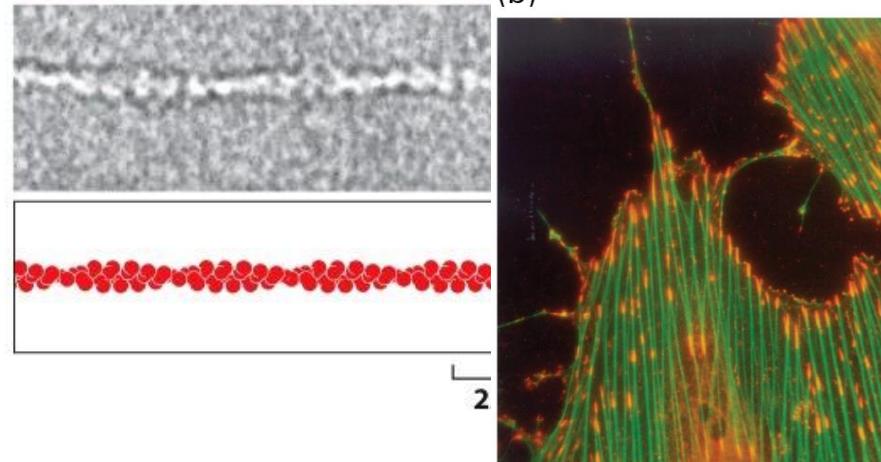
Highly dynamic

Polarized

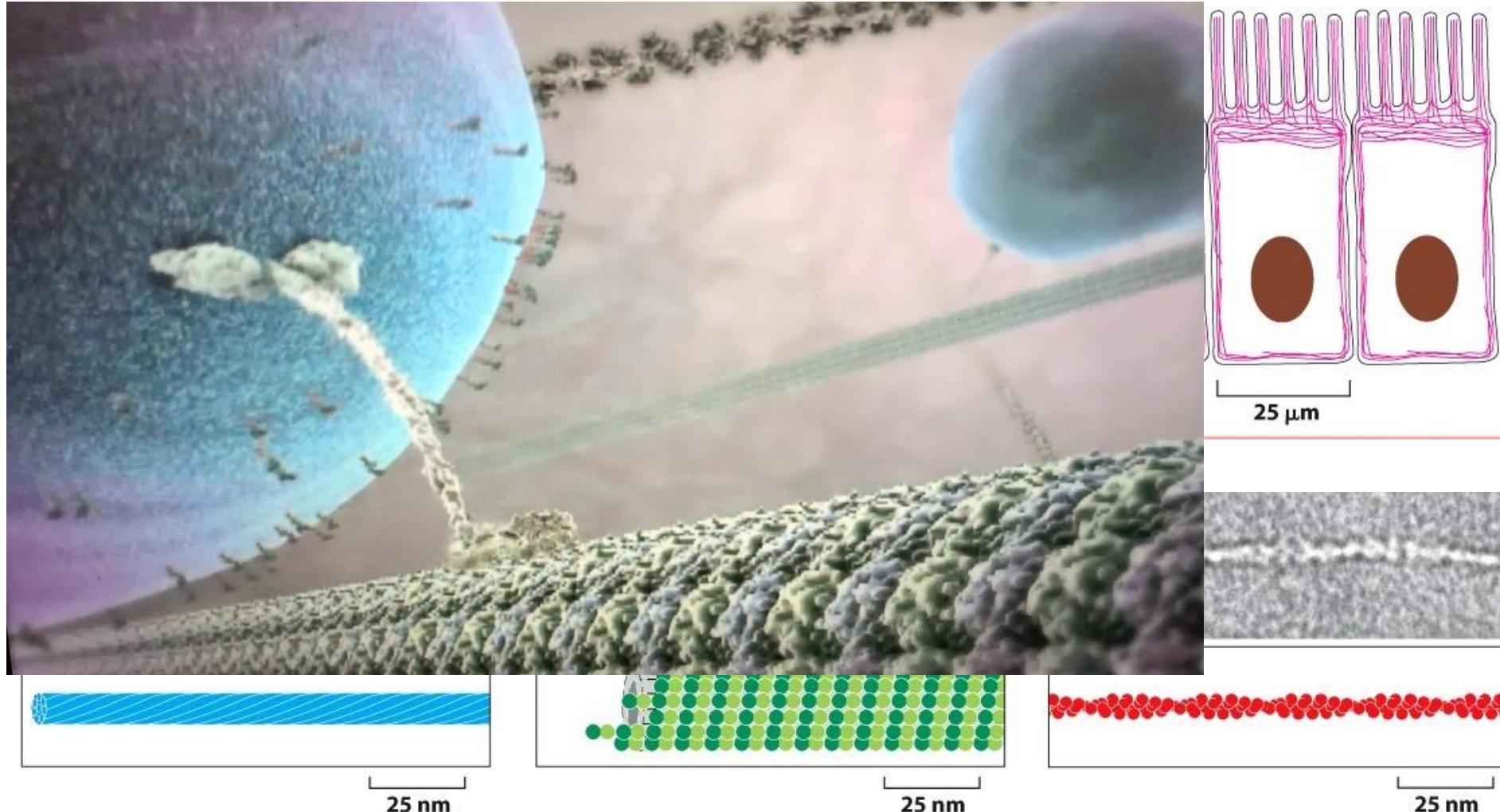
Tracks for myosins

Contractile machinery and network at the cell cortex

(b)



# three types of cytoskeletal filaments... and...



# three major motor proteins

motor proteins use **energy** to move directionally along cytoskeletal polymers

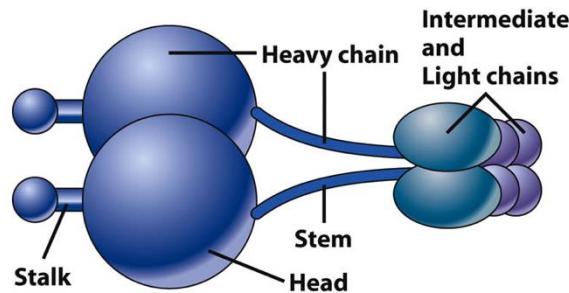


Figure 9-17a Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

**Dynein** (moves along Tub to - end)

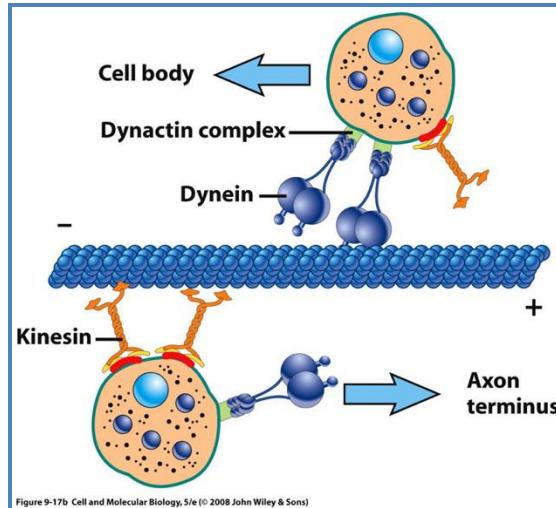


Figure 9-17b Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

**Myosin**  
(moves along Actin)

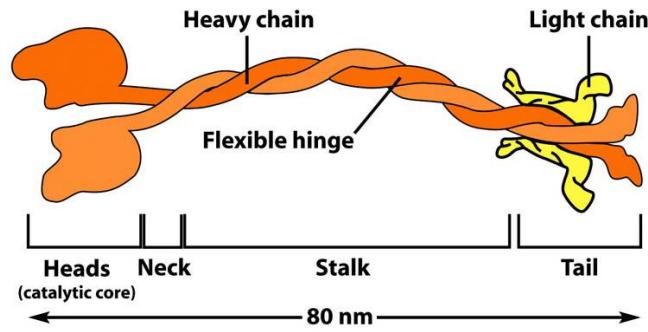
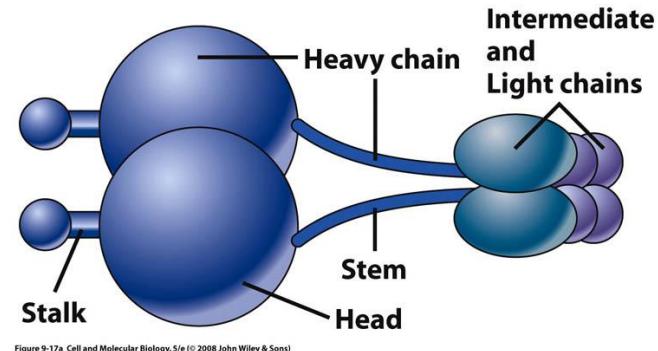
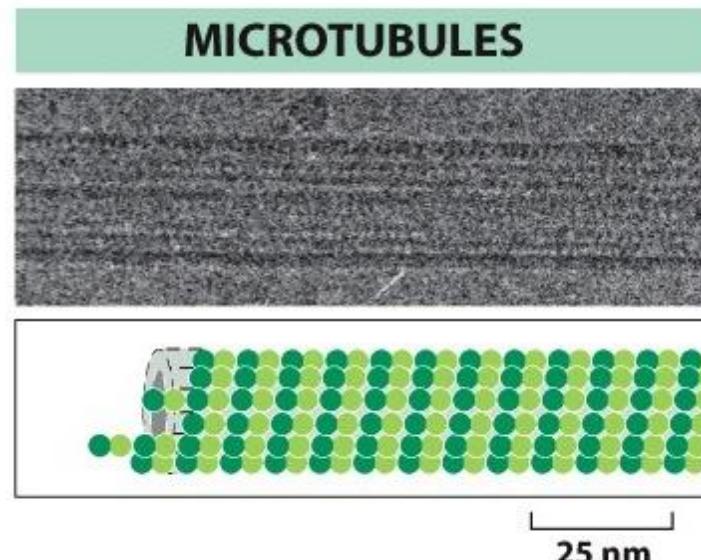
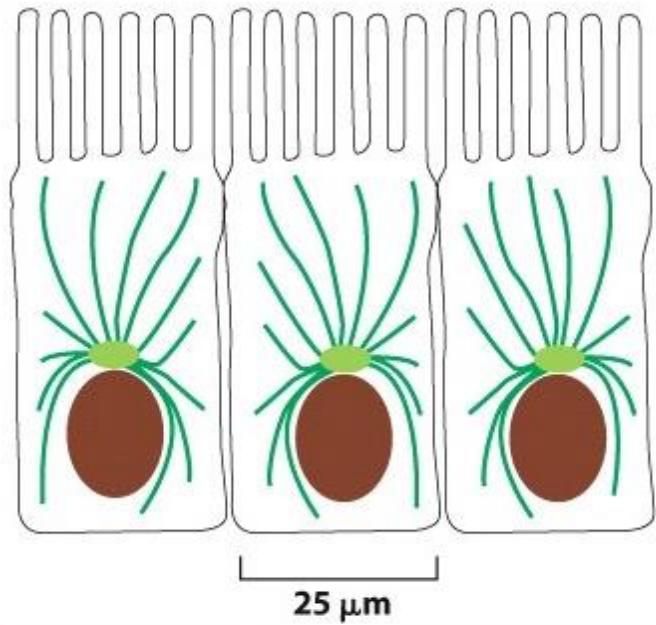


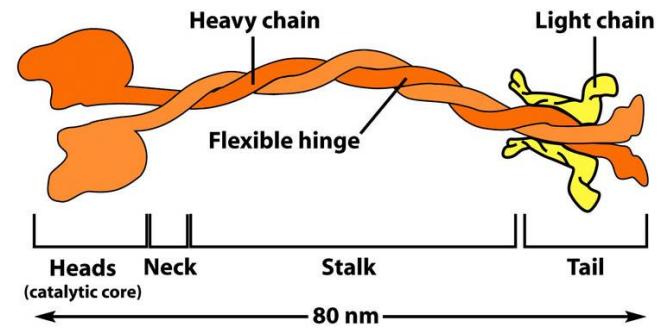
Figure 9-15a Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

**Kinesin**  
(moves along Tub to + end)

# today: one filament, two motors



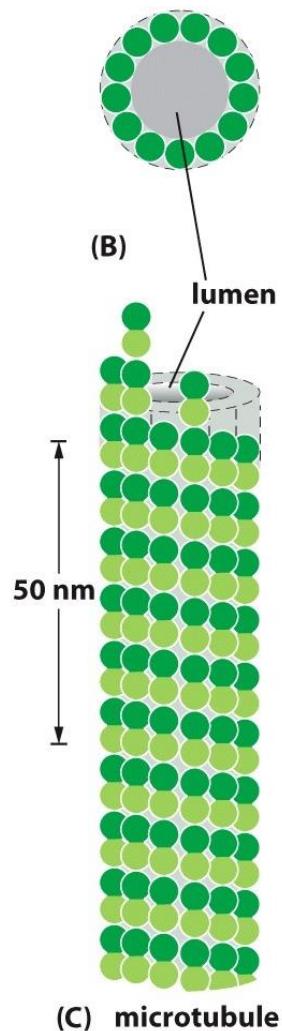
Dynein (moves along Tub to - end)



Kinesin  
(moves along  
Tub to + end)

# microtubule learning objectives

by the end of this lecture you should be able to:



- describe the key functions of microtubules
- describe how microtubules assemble and breakdown
- know which molecules are required for microtubule assembly and stability and what drugs disrupt these
- explain “dynamic instability” and explain advantages of polymer structure
- compare and contrast the motor proteins associated with microtubules and their functions
- describe the organization of the axoneme structure and explain how microtubules and dynein contribute to flagellar and ciliary movement
- explain the importance of cilia in human biology

# overview: **microtubules** contribute to structural support, organization, motility within the cell

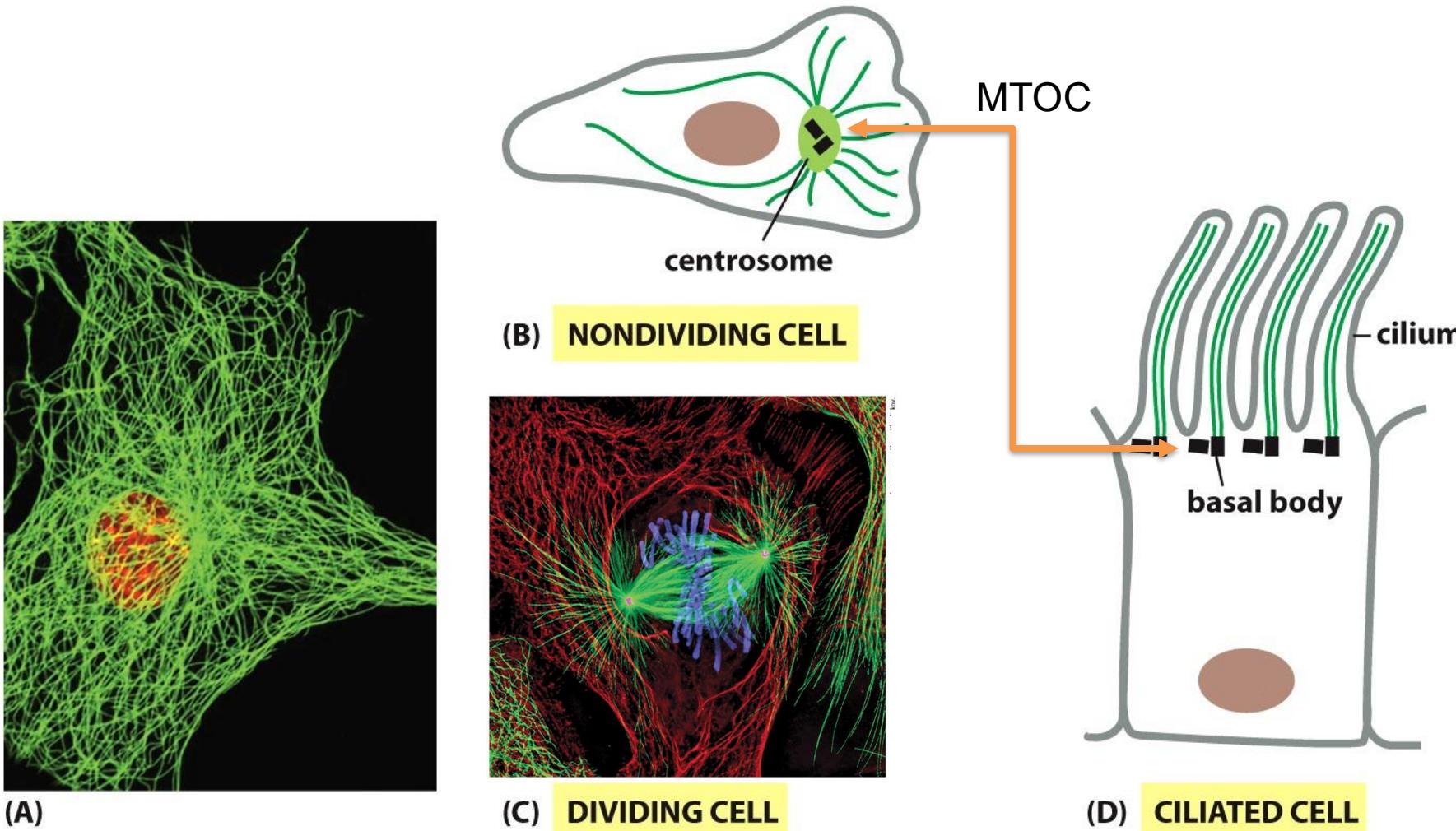


Figure 17-10 Essential Cell Biology, 4th ed. (© Garland Science 2014)

+

# microtubules are polymers

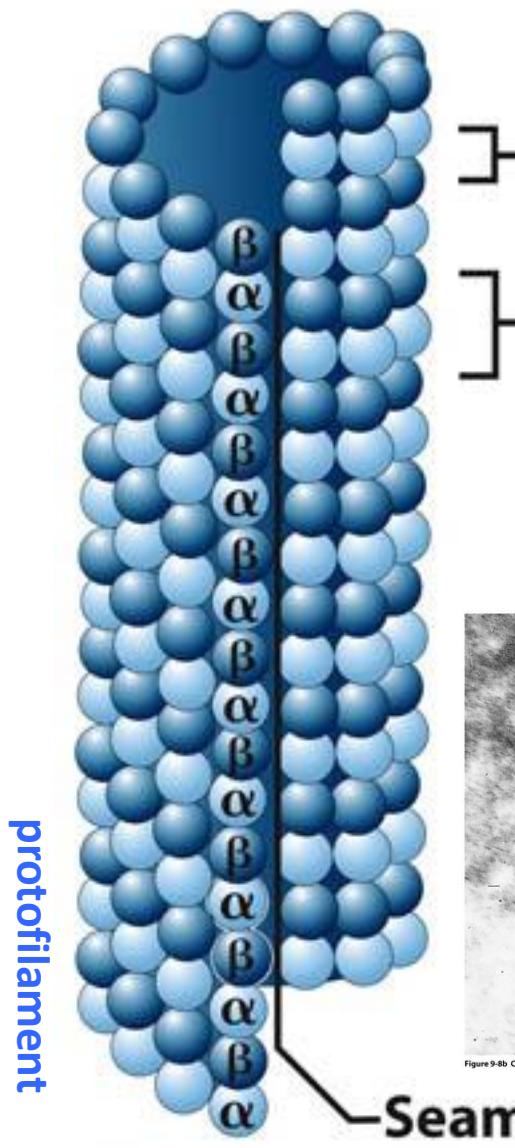
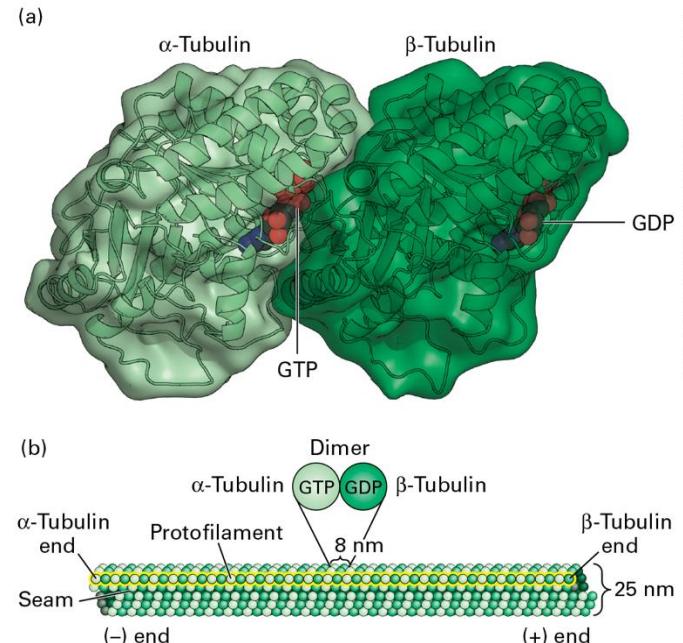


Figure 9-8d Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

- polarized +/-, hollow tube \_\_\_ nm diameter
- made up of  $\alpha$  and  $\beta$  subunits (heterodimers) that can be added and released (13 protofilaments)
- stabilized by associated proteins



Lodish et al., Molecular Cell Biology, 9/e, © 2021 W.H. Freeman and Company

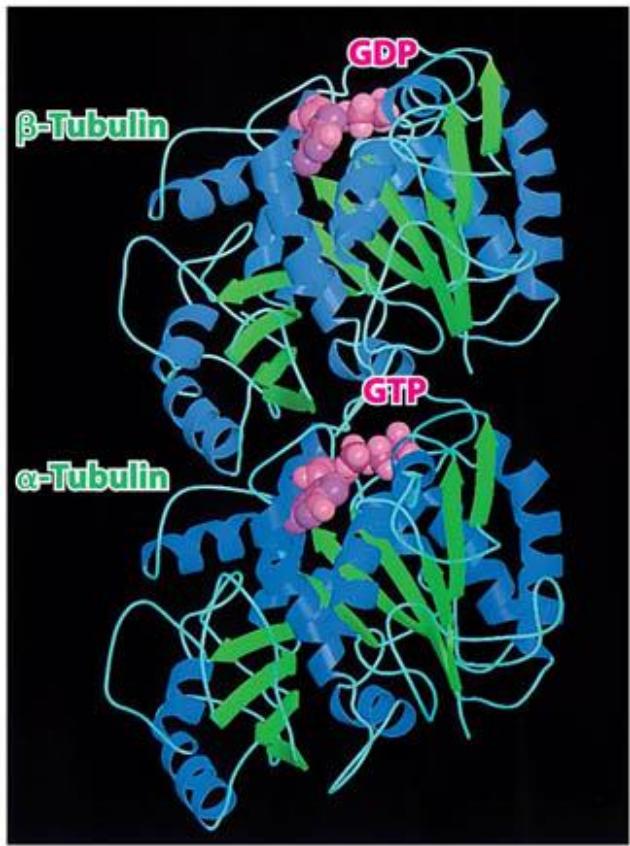
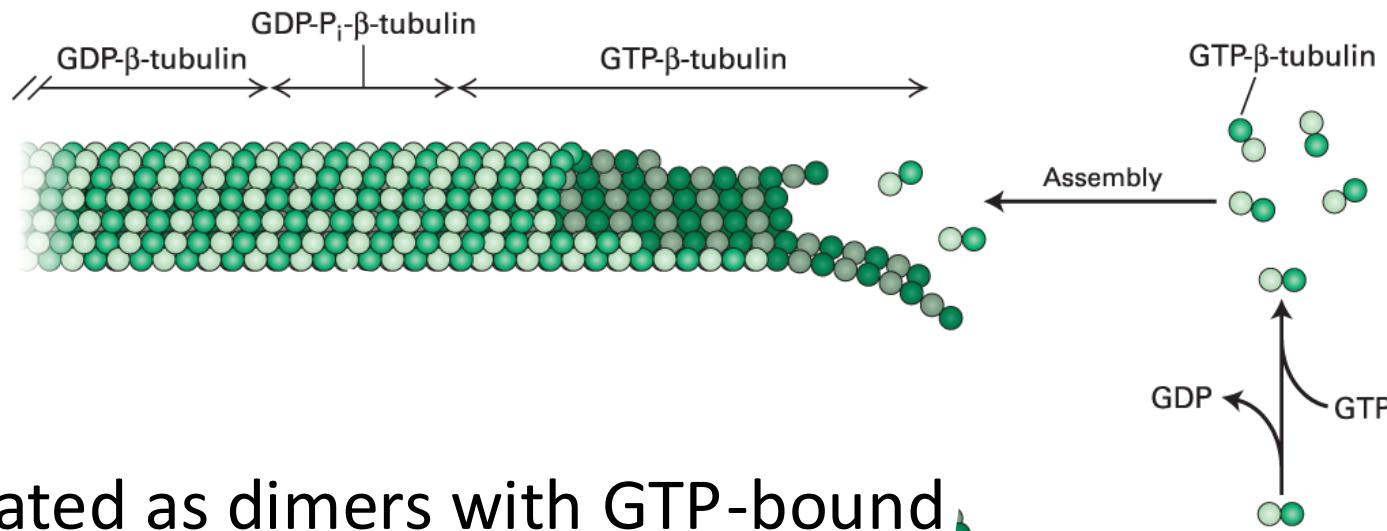
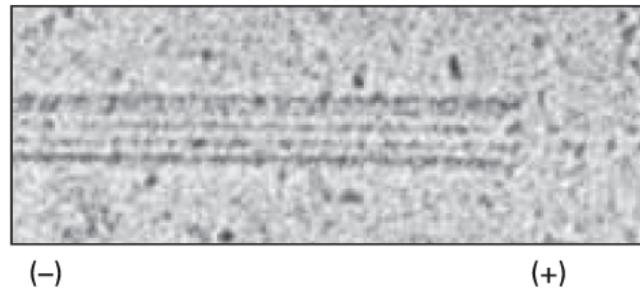
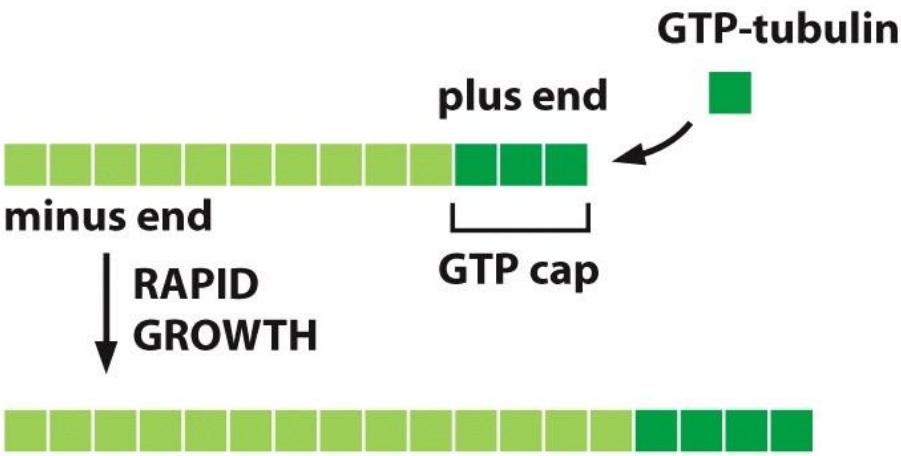


Figure 9-8c Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# microtubules can lengthen...

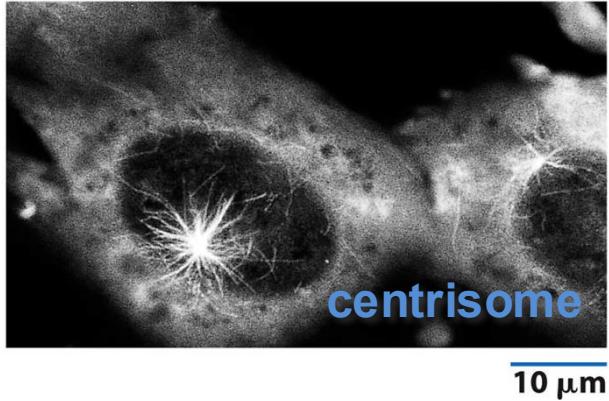


- subunits are rapidly incorporated as dimers with GTP-bound  $\beta$ -tubulin to the  $\beta$ -subunit, then GTP is hydrolyzed (expensive!)

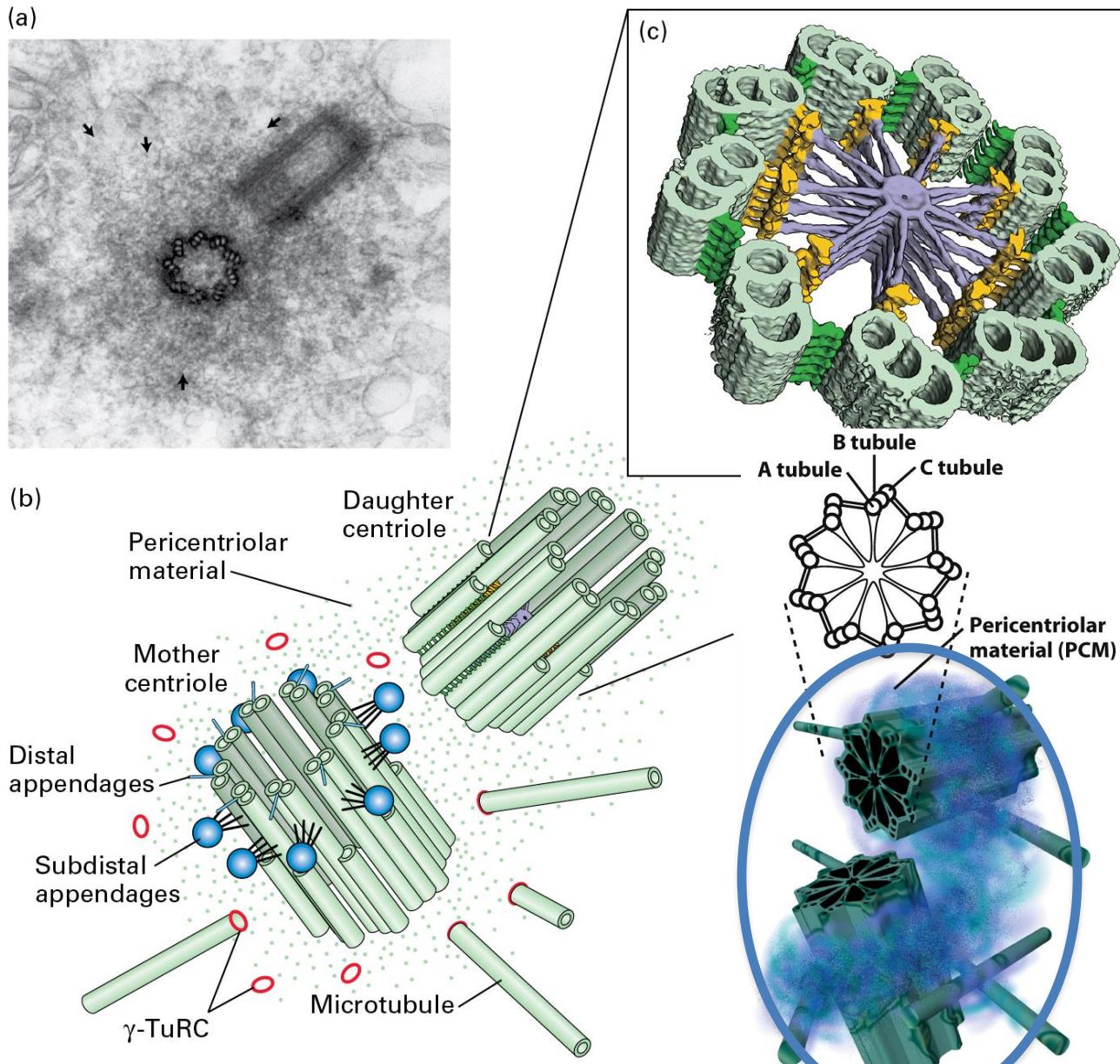
- GDP on  $\beta$  is exchanged in the cytosol for GTP

soluble pool  
of dimers in  
cytosol

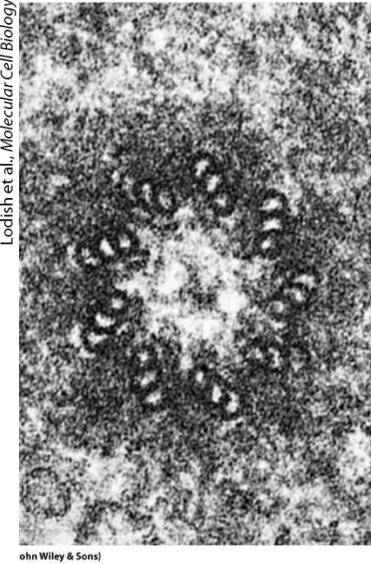
# MTOCs govern MT assembly



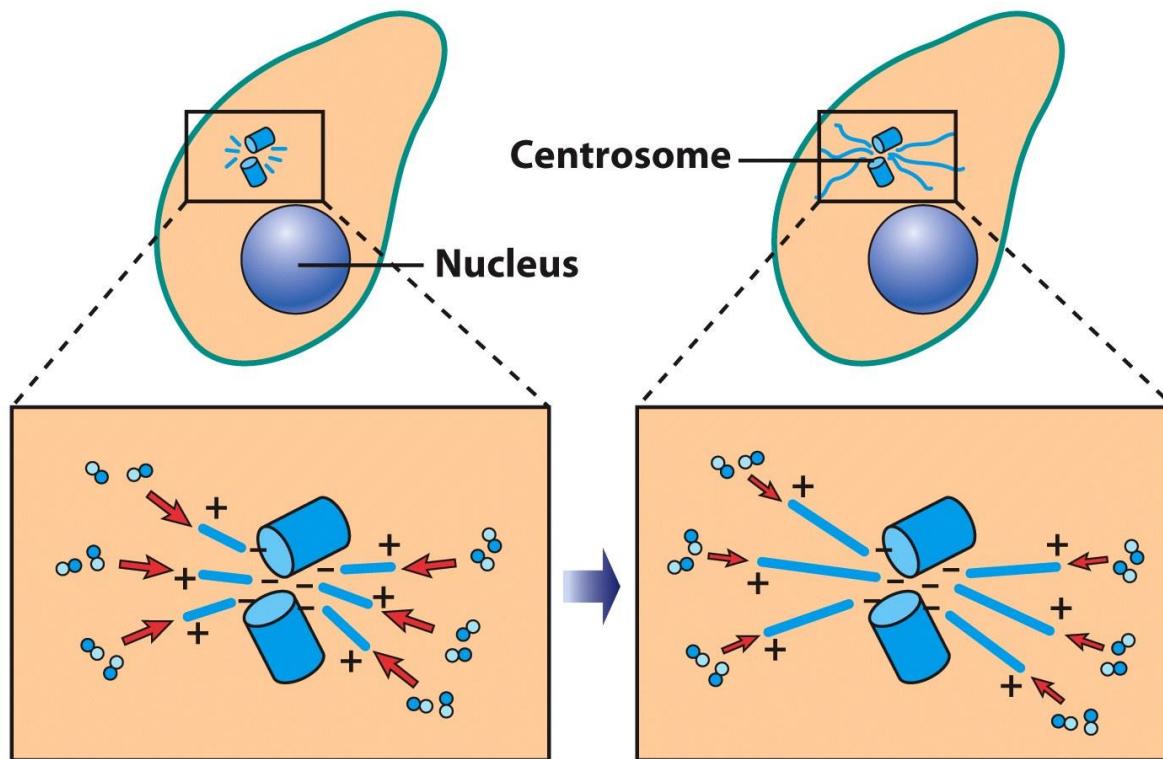
- MTOC - microtubule-organizing centers
- control #, polarity, time, location, of nucleation
- **centrosome and basal body** in most animal cells (nucleus in plants)
- centrosome is made up of 2 **centrioles** and **PCM** (pericentriolar material)



Part (d) Courtesy of Ryoko Kuriyama. © 2021 W.H. Freeman and Company



# microtubule assembly



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nucleation of MTs is slow  
elongation is fast

minus end is associated with centrosome;  
dimers are added to (and lost from)  
the PLUS end

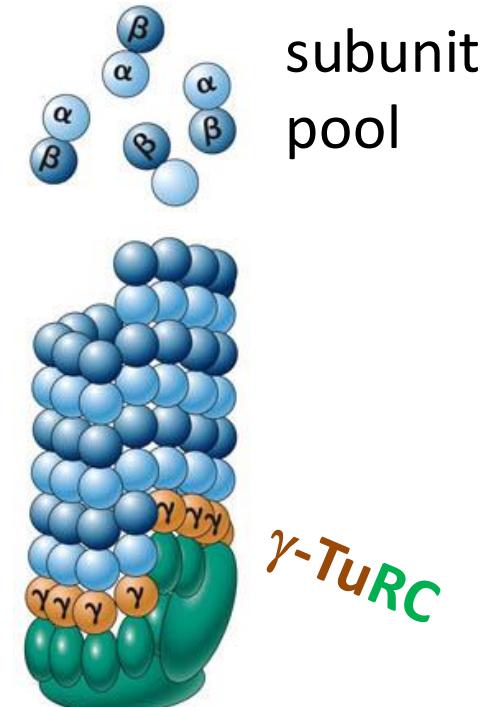


Figure 9-20c Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

**$\gamma$ -Tubulin** sits in a  
ring in PCM and dimers  
bind to it to form  
microtubule

# microtubules are very dynamic – grow & shrink

- subunits are rapidly incorporated as dimers with GTP-bound to the  $\beta$ -subunit, then GTP is hydrolyzed (expensive!)
- if all GTP is hydrolyzed, the microtubule can rapidly depolymerize
- GDP on  $\beta$  is exchanged in the cytosol for GTP

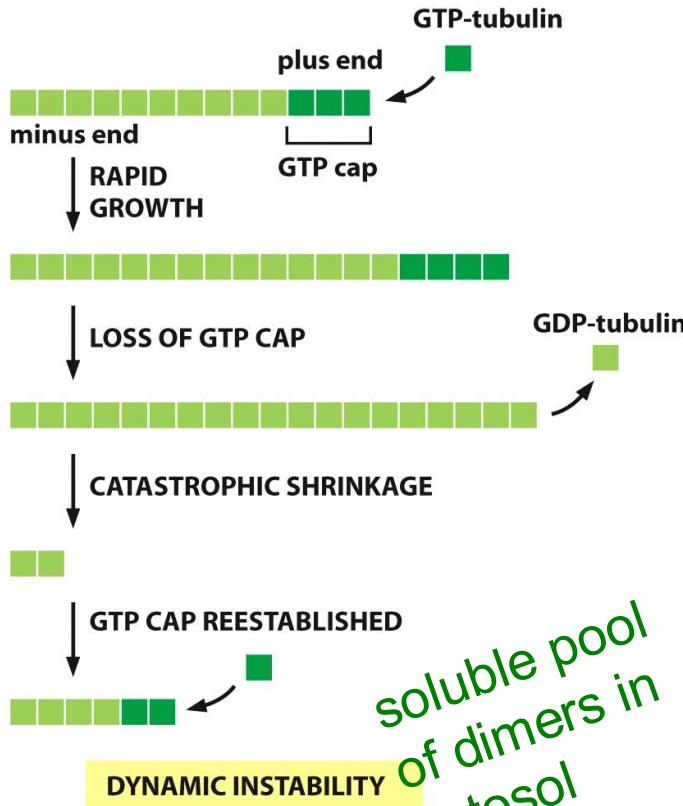
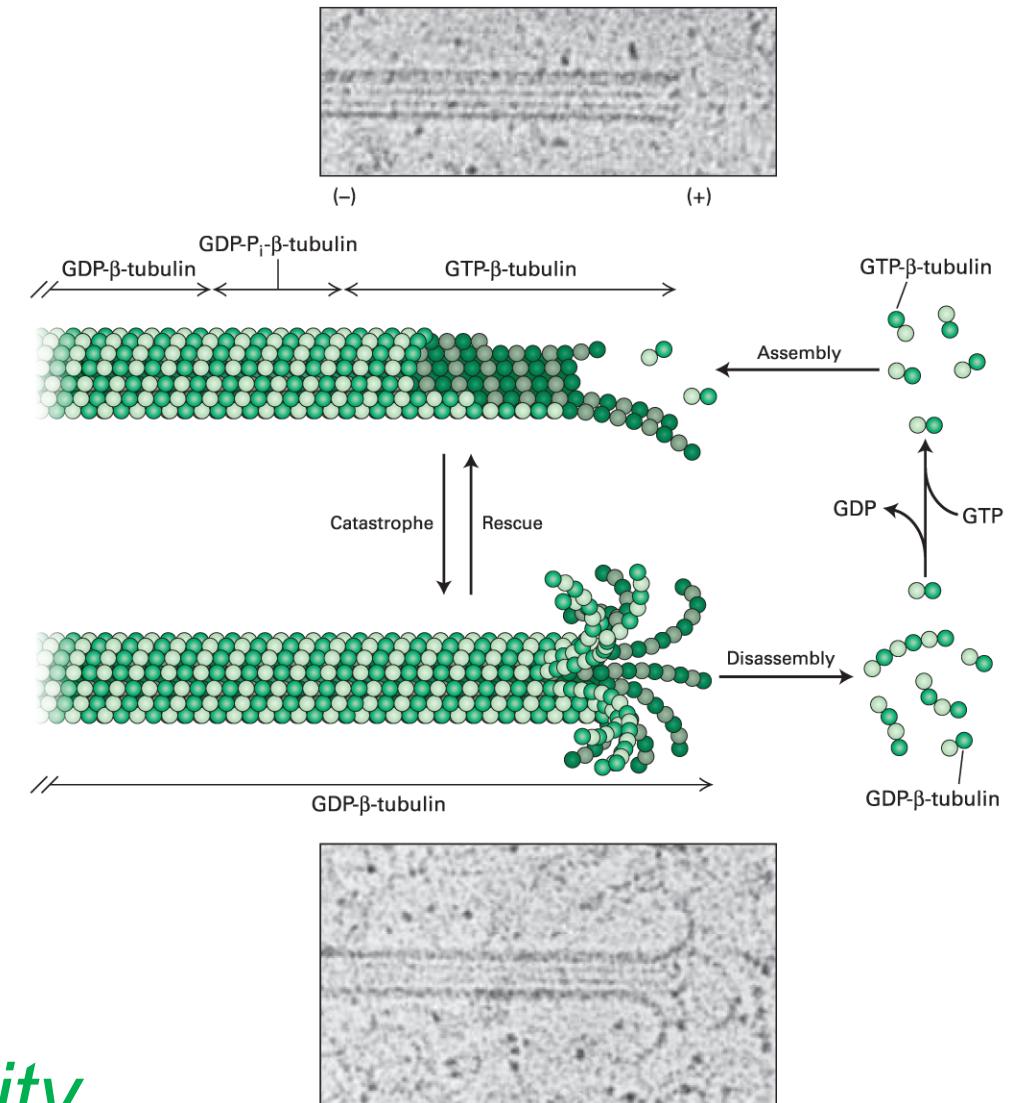


Figure 17-31b Essential Cell Biology, 4th ed. (© Garland Science 2014)

*dynamic instability*



# microtubule assembly & disassembly

- assembly occurs at microtubule-organizing centers (MTOCs)
- requires energy from **GTP hydrolysis** (beta) to lengthen (expensive)
- *dynamic instability* at the plus end = polymerize and depolymerize

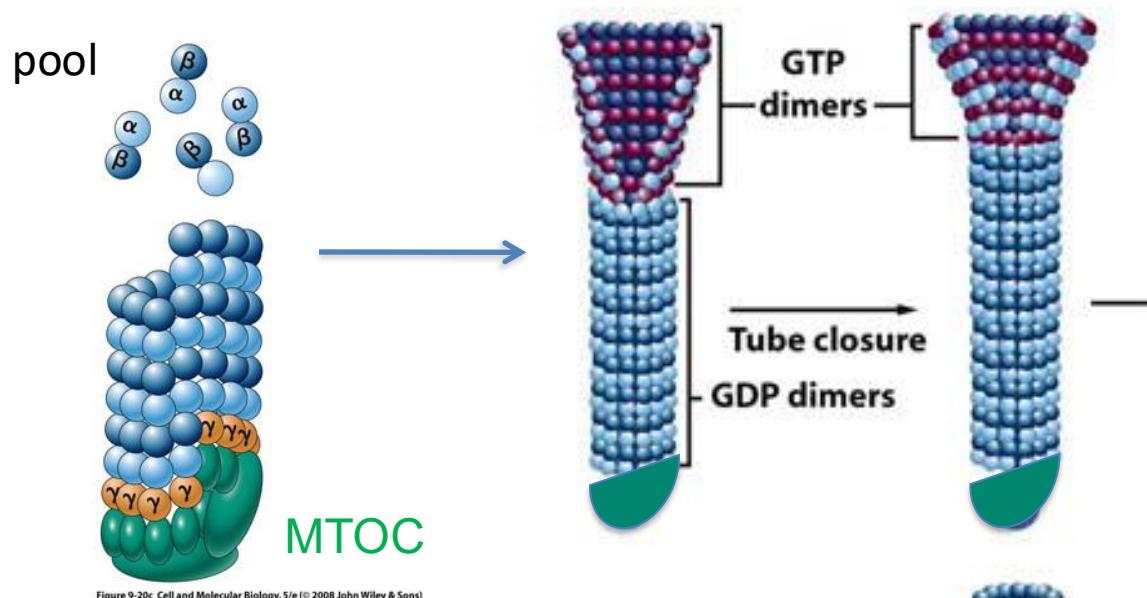


Figure 9-20c Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

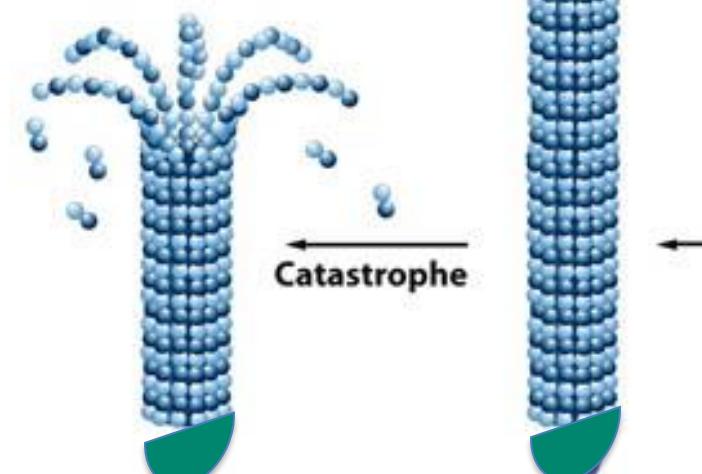
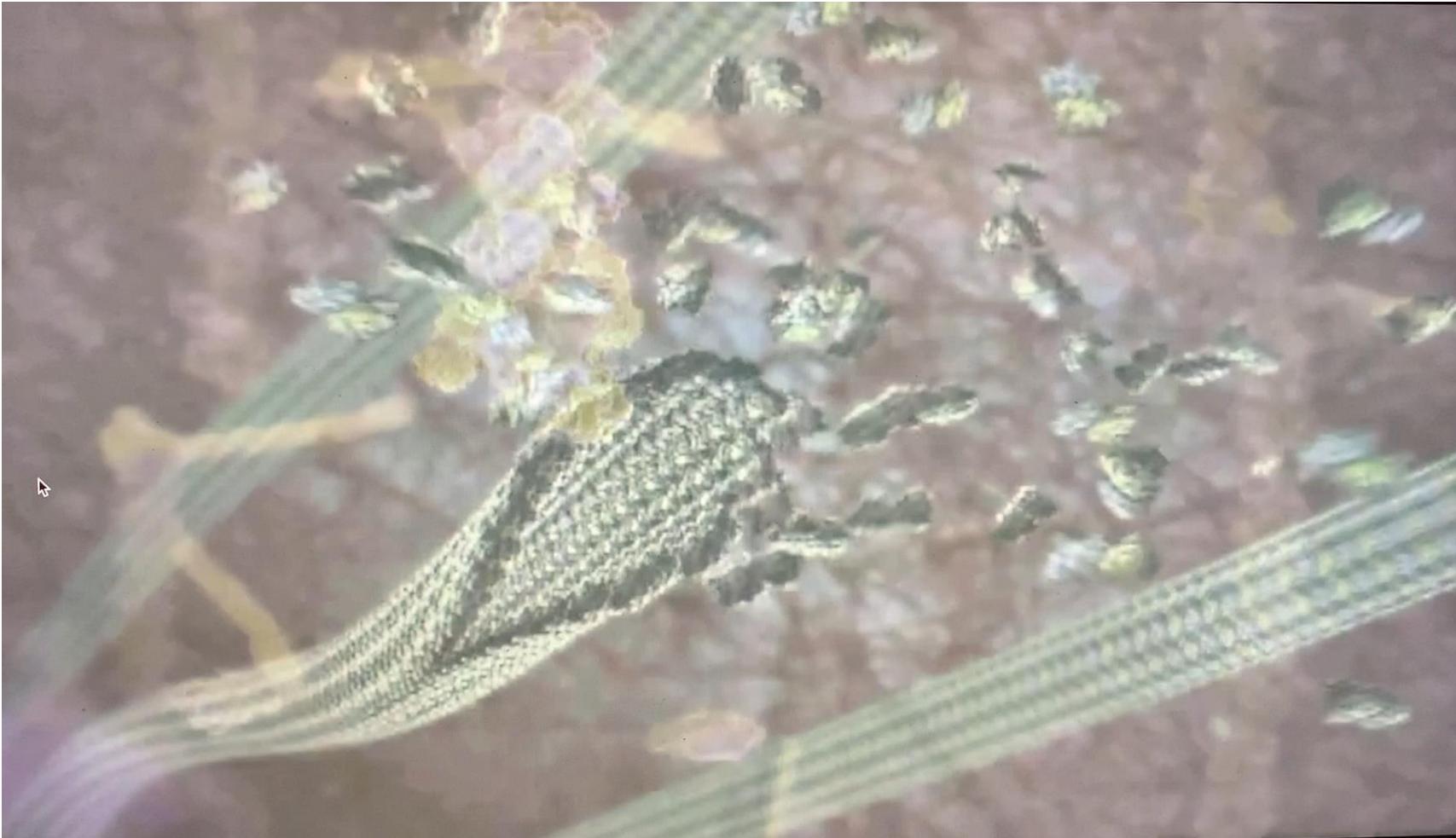


Figure 9-25 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# microtubule assembly & disassembly



<https://www.youtube.com/watch?v=wJyUtn0O5Y>

# some treatments change microtubule dynamics

treatments that disassemble/destabilize microtubules (shift steady state):  
cold, high Ca<sup>++</sup>, and

## drugs:

**colchicine** – binds dimer, adds to tubule, blocks further addition

**nocodazole, vinblastine** - suppress addition to tubule, may detach from MTOC

**taxol** - inhibits disassembly and thereby assembly.

THESE are ANTI-CANCER agents because they interfere with cell division

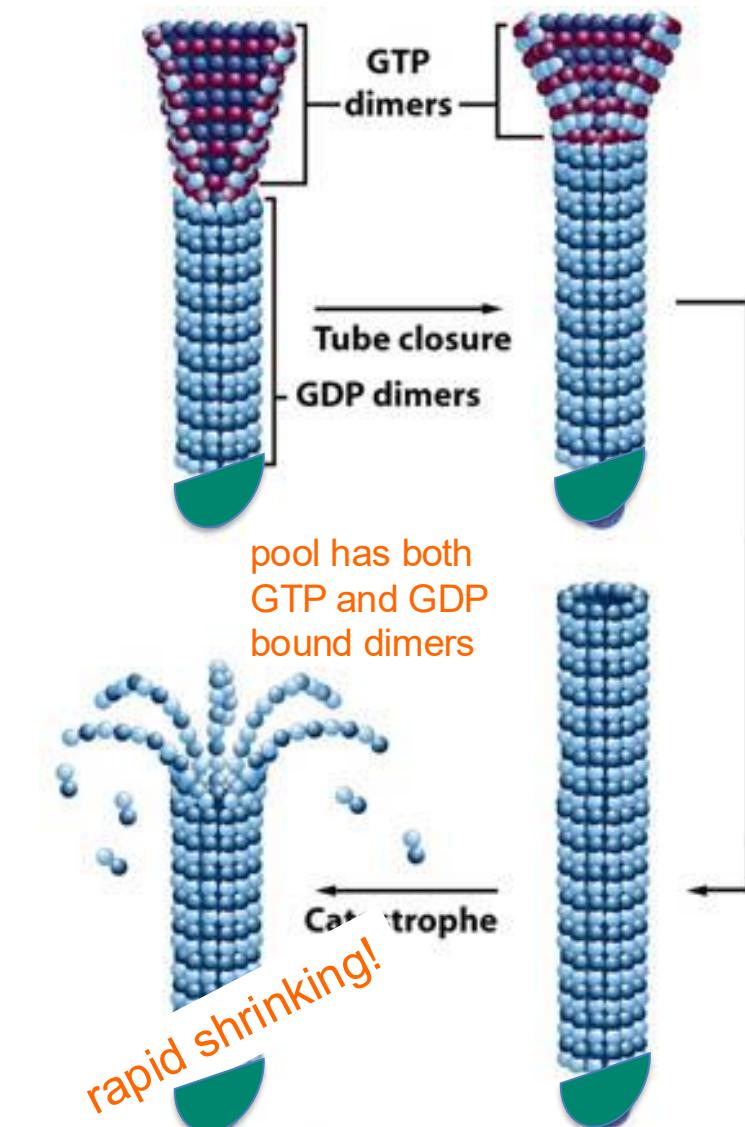
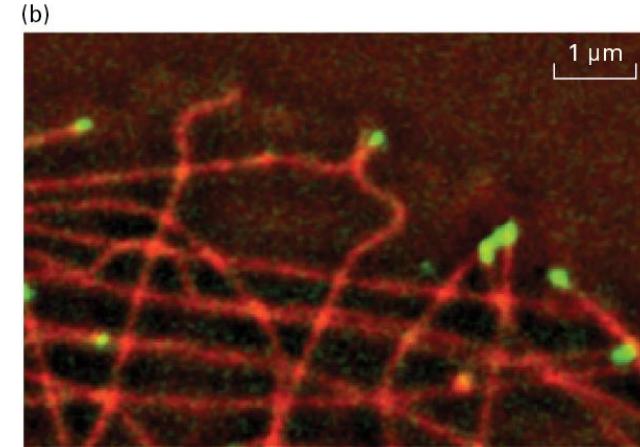
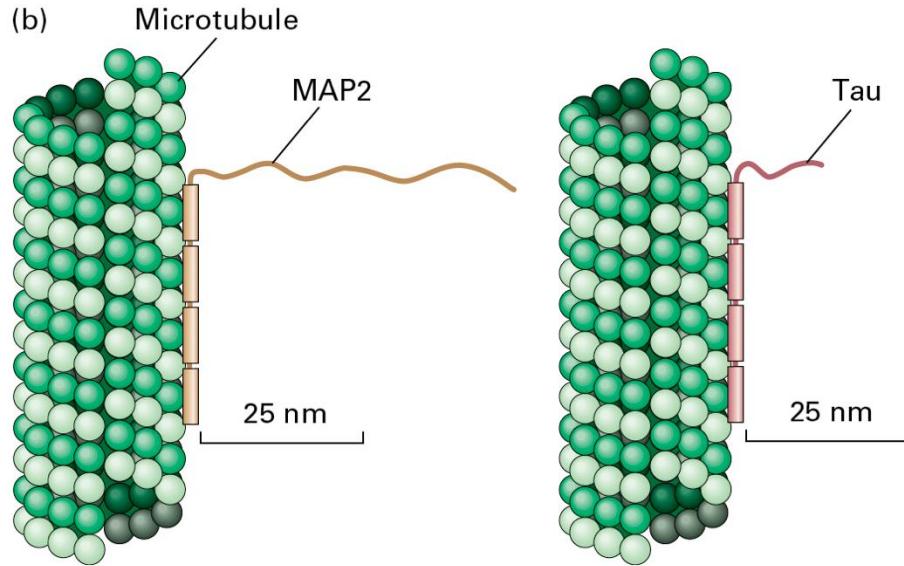
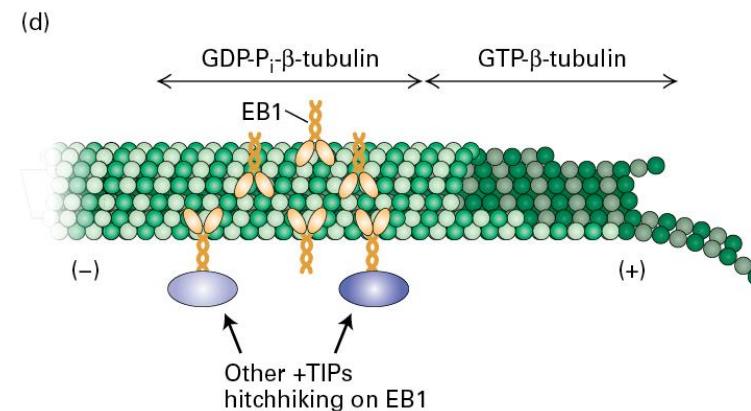


Figure 9-25 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# dynamic microtubule structures are regulated in cells by Microtubule Associated Proteins (MAPs)



■ EB1 or EB3  
■ Tubulin



- MAPs can stabilize and organize microtubules (Microtubule Associated Proteins , eg *tau*, *MAP2*)
- EB1 binds near the end of microtubules
  - + end tracking proteins
  - As can post-translational modifications (acetylation), dependent on the context



## a microtubule experiment – [poll 1](#)

- you attempt to make tubulin polymers *in vitro*
- you add cell extracts to large amounts of  $\alpha$ - and  $\beta$ -Tubulin, ATP, GTP, GDP, and adjust the temperature to 37.

Poll 1: after you get polymers, which of the following will be depleted from the test tube?

- a) ATP
- b) ADP
- c) GTP
- d) GDP

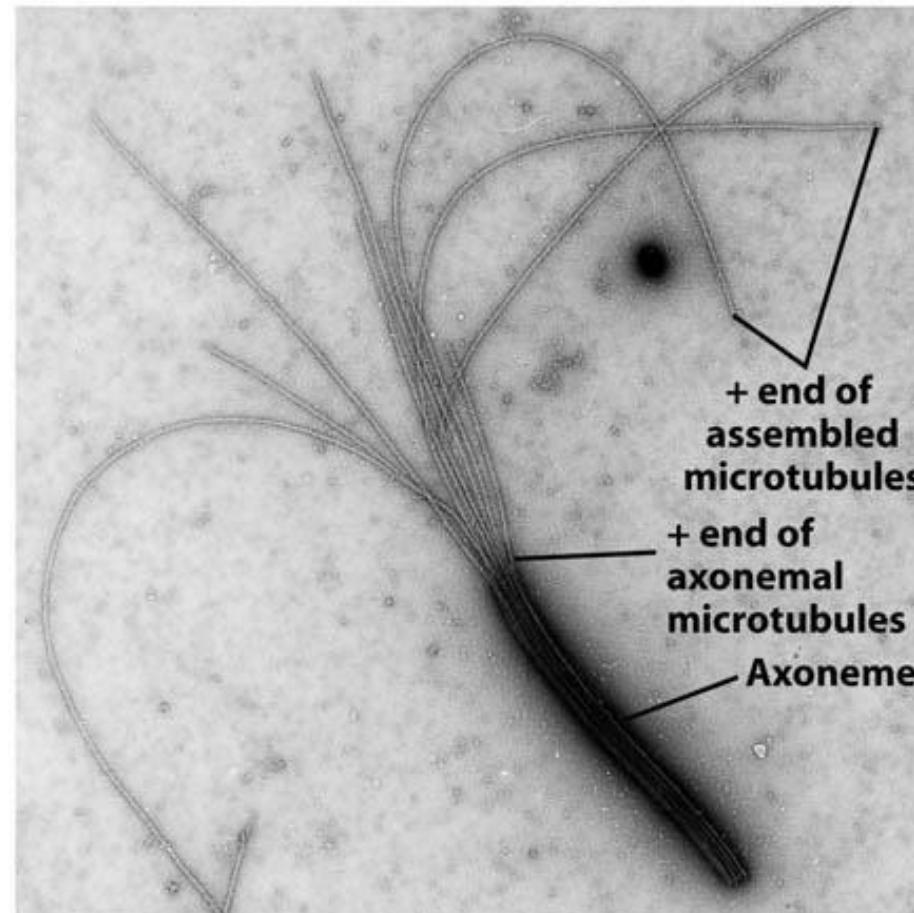


Figure 9-24 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

## an MT experiment – poll 2

- you attempt to make tubulin polymers *in vitro*
- you add cell extracts to large amounts of  $\alpha$ - and  $\beta$ -Tubulin, ATP, GTP, GDP, and adjust the temperature to 37.
- you have made tubulin polymers *in vitro*, but they are *transient*, which is not great for taking pictures.  
**which drug might help you maintain the microtubules?**

- a) colchicine
- b) phalloidin
- c) nocodazole
- d) taxol

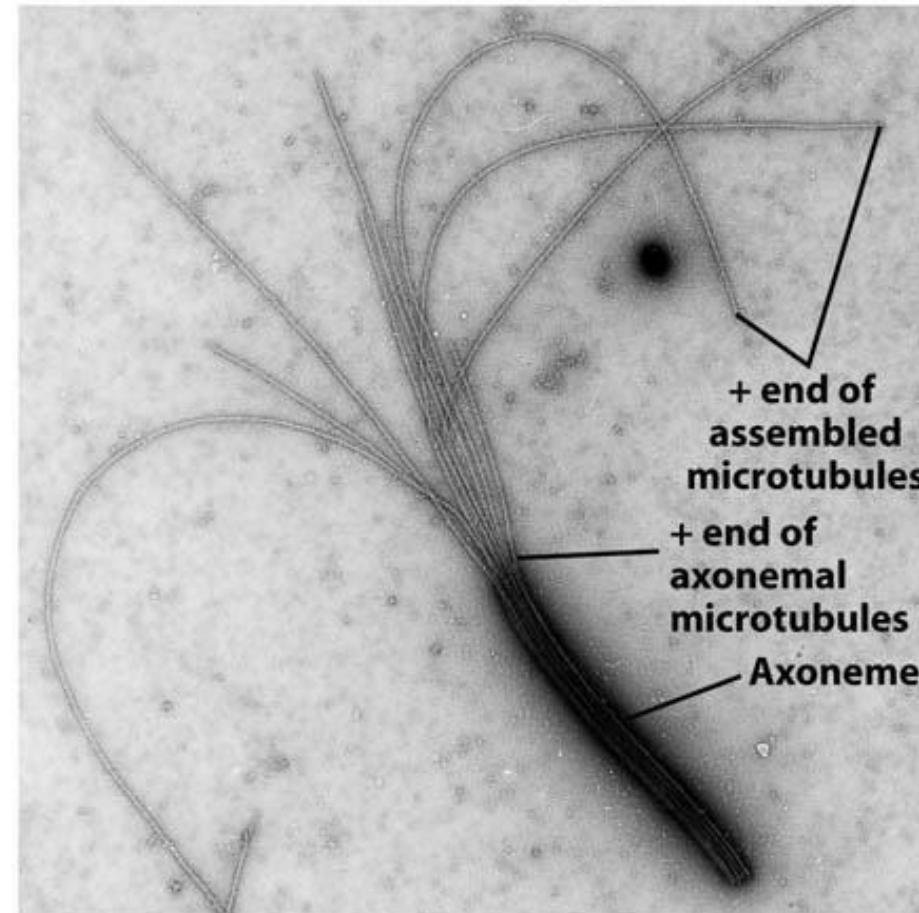


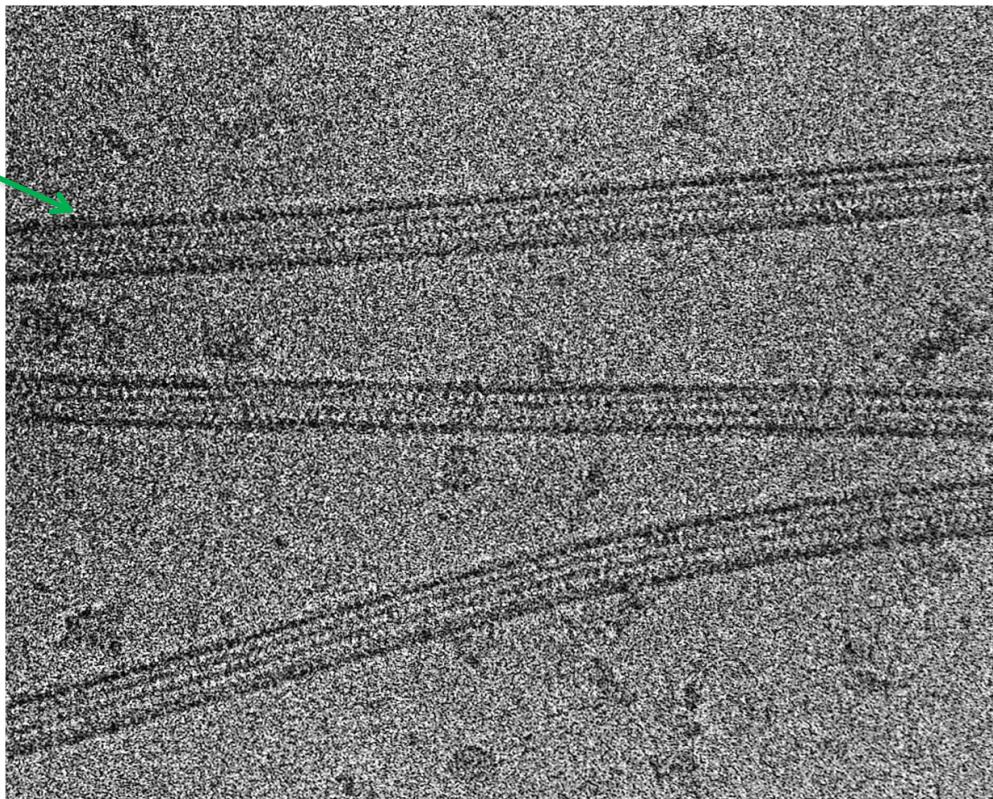
Figure 9-24 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# About your *in vitro* microtubules...

after much work (EM), you realize your polymers are funny-looking because the tubes are too small in diameter ☹

Poll 3: which of these additions would help?

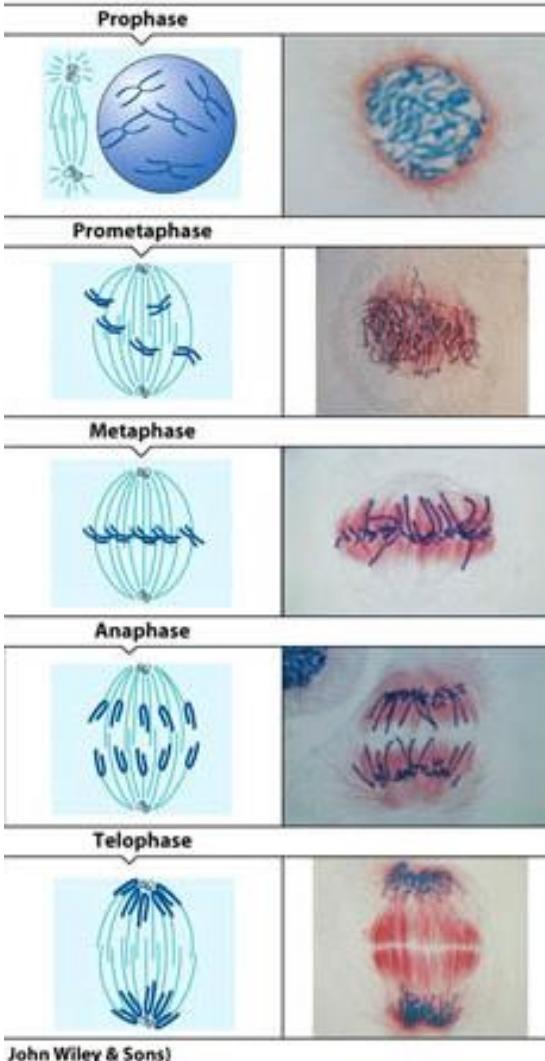
- 1) more GTP
- 2) more  $\beta$ -subunits
- 3) *in vitro*-made centrioles
- 4) centrosomes =  $\gamma$ -Tubulin ring complex



Courtesy of R. H. Wade, Institut de Biologie Structurale, Grenoble, France.

# why all the dynamics? one example:

prophase



prometaphase

metaphase

anaphase

telophase

photos  
are  
of  
a plant  
cell

## stages of mitosis (M phase)

# mitosis example: prometaphase- movement of chromosomes to spindle equator requires:

## tubulin dynamics

and

## motor proteins

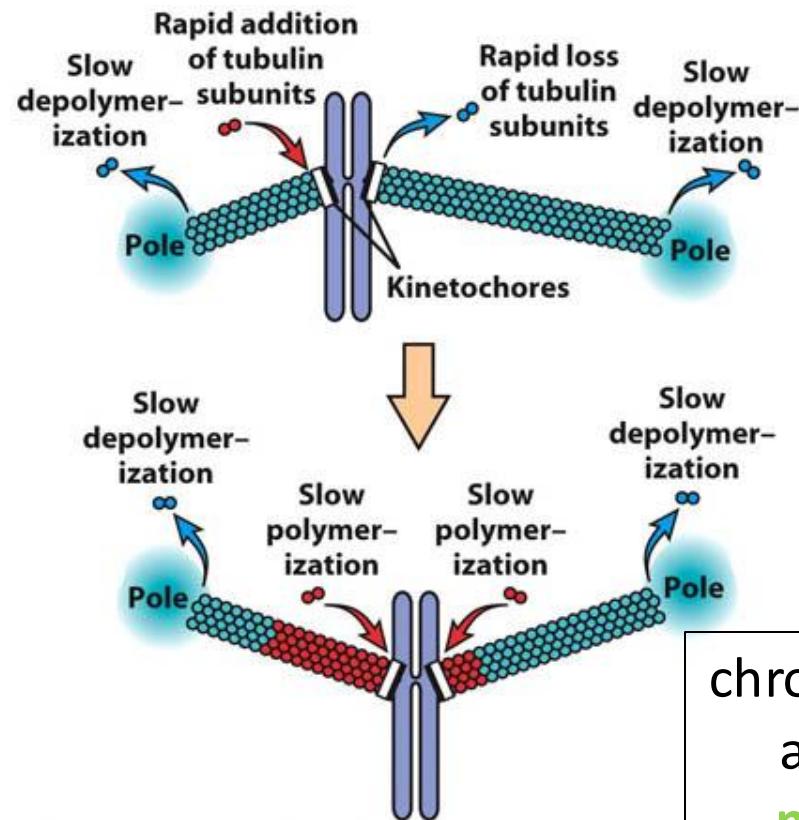


Figure 14-22 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

chromosomes align at the  
metaphase plate

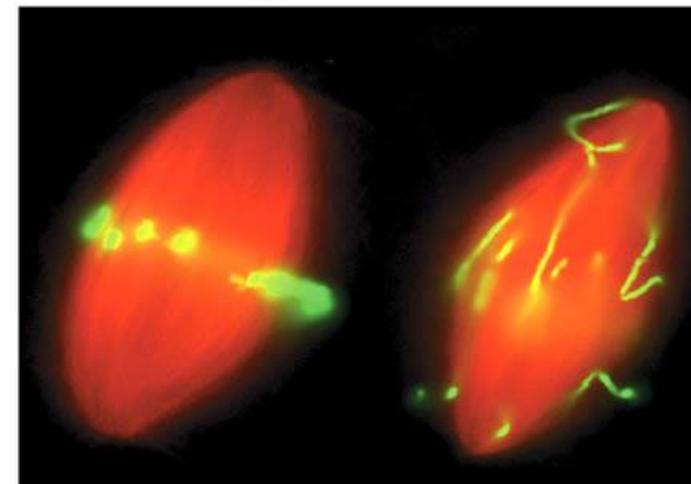
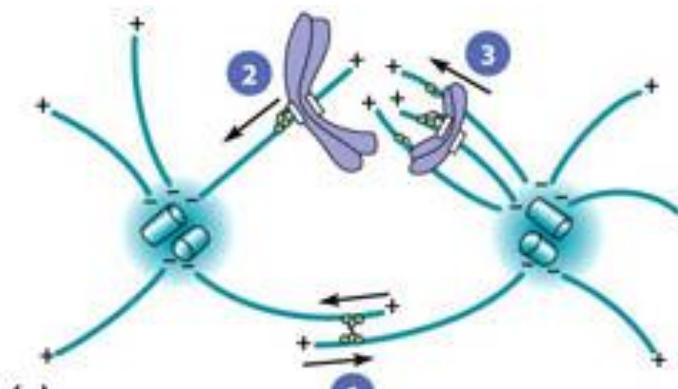
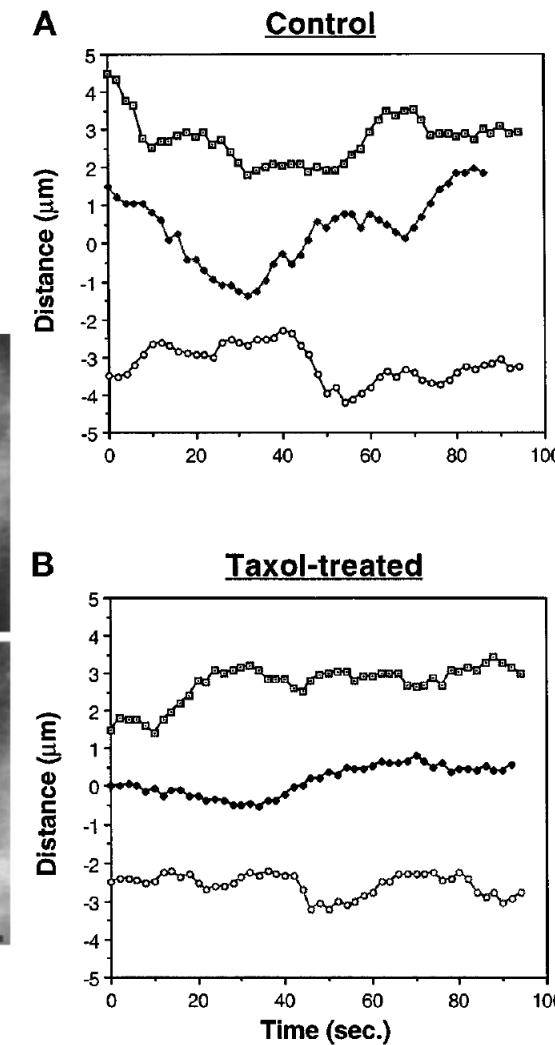
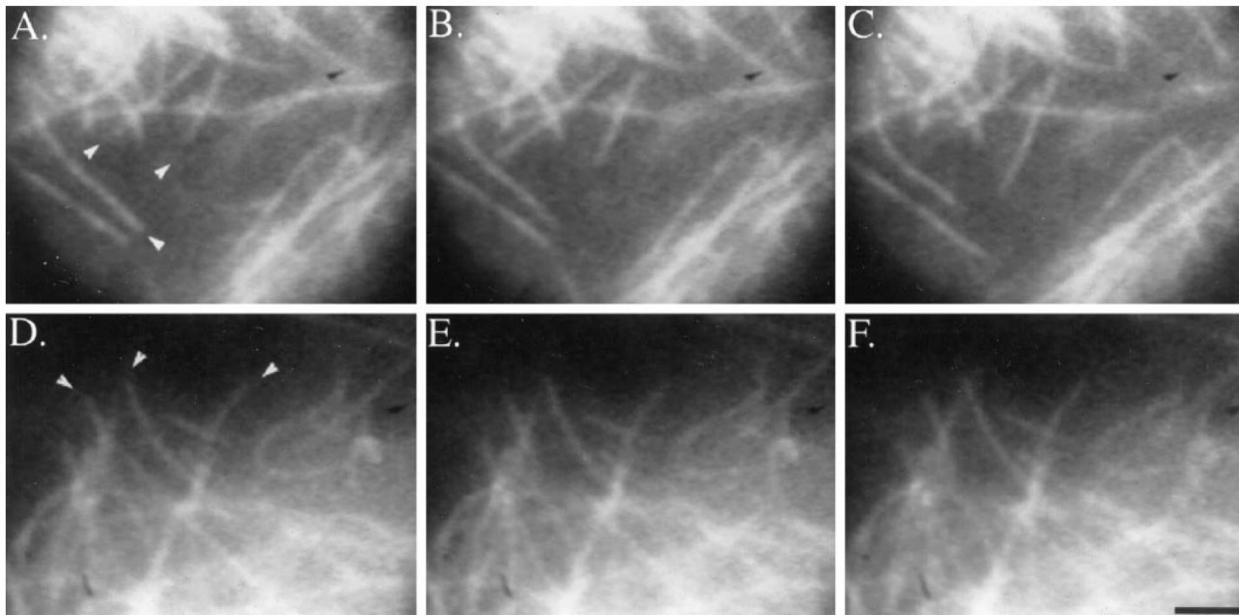


Figure 14-21 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# Taxol-treated cells have less dynamic microtubules



- Caov-3 cells (ovarian adenocarcinoma)  
Control (top), or with 30nM taxol (bottom), scale bar 2 $\mu\text{m}$
- Data from primary paper: Yvon, Wadsworth, and Jordan (1999) “Taxol Suppresses Dynamics of Individual Microtubules in Living Human Tumor Cells.” *Molecular Biology of the Cell*. Vol 10, April, 947-959.



# microtubule polarity is important and consistent within cell types

microtubules stabilize the location of organelles such as ER and Golgi.

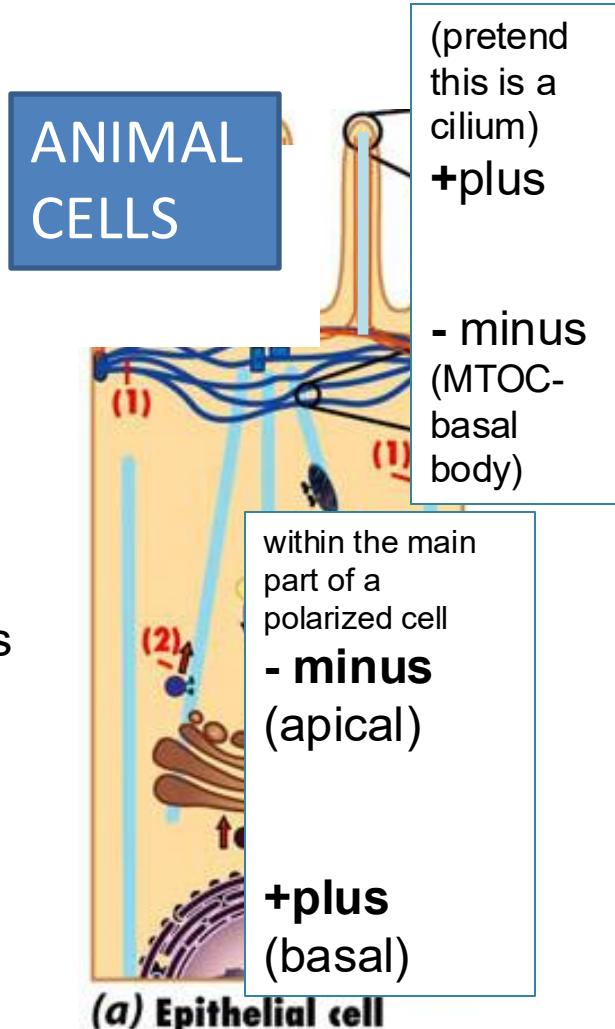
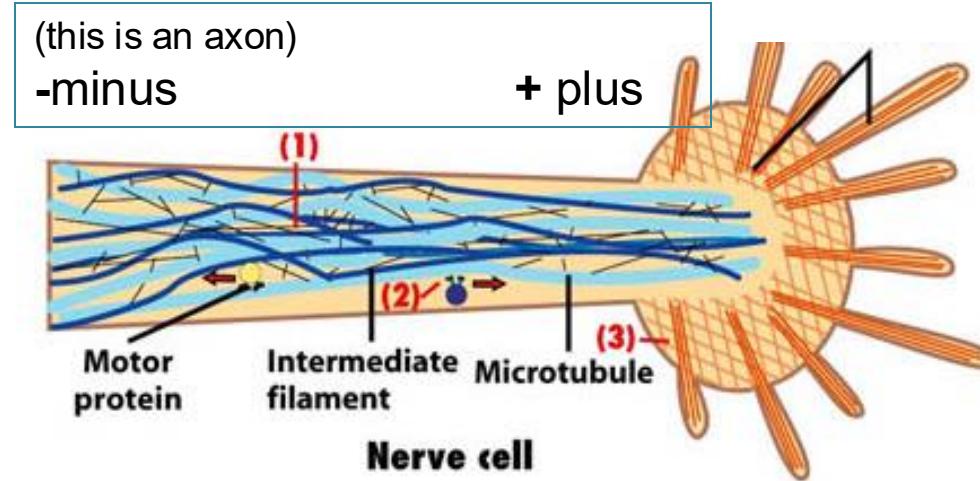
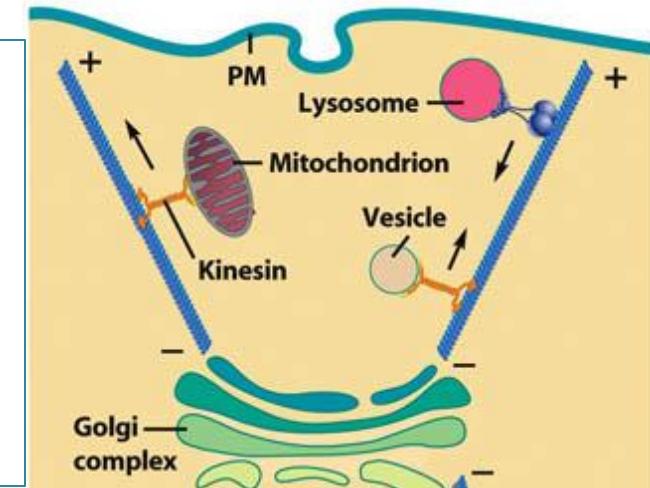


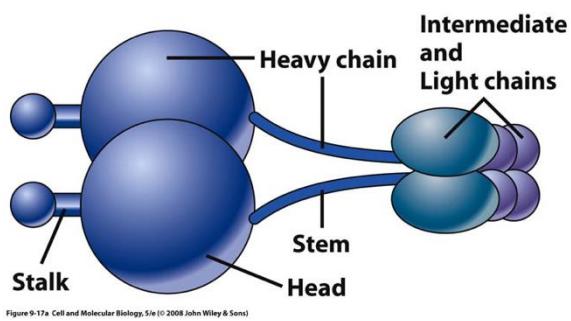
Figure 9-1 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)



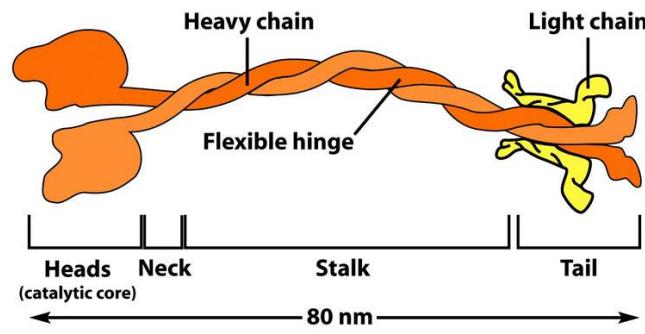
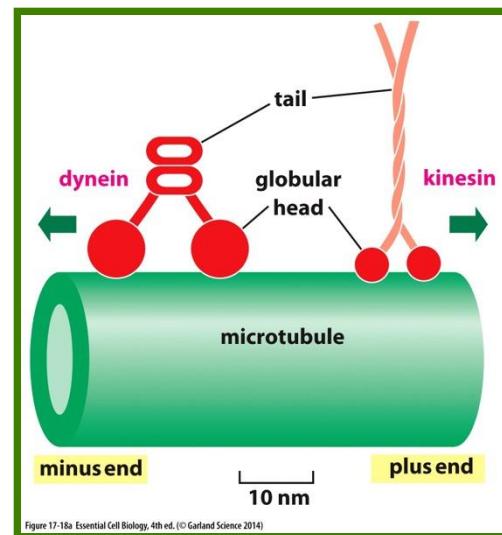
non polarized cell  
+ plus  
- minus (at MTOC)



# two motor protein families use energy to move in opposite directions along microtubules



**Dynein**  
(moves along  
Tub to - end)



**Kinesin**  
(moves along  
Tub to + end)

# Dynein is a minus-end directed microtubular motor

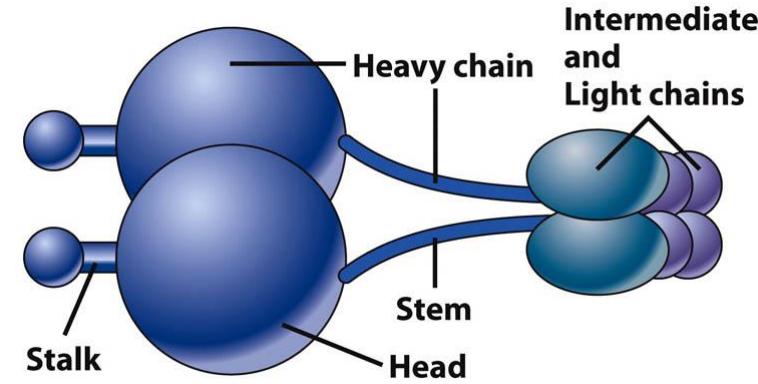
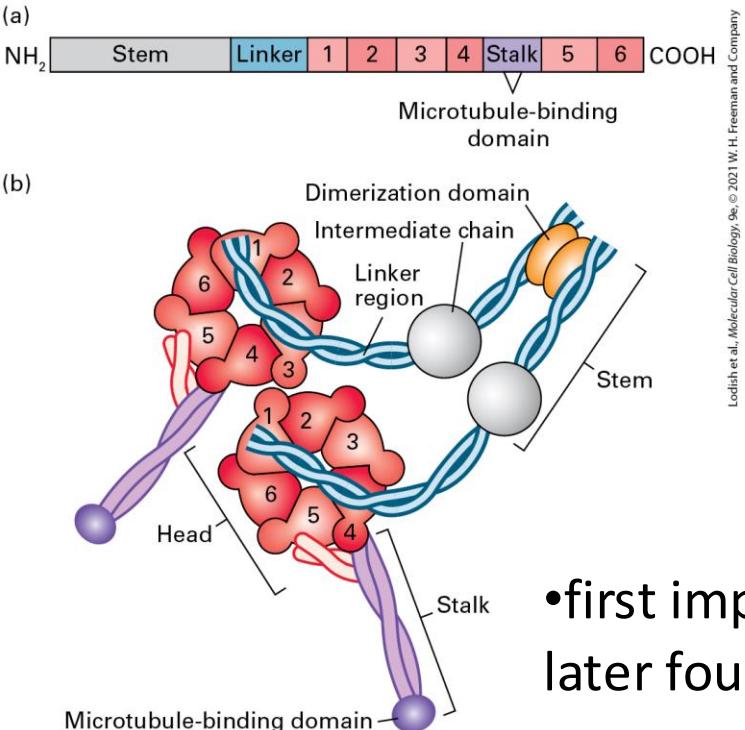


Figure 9-17a Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

- first implicated in cilia and flagella movement; later found in all animal cells (cytoplasmic)
- stalk binds MTs; head generates force; requires **ATP**
- binds cargo via *dynactin* protein
- moves organelles, vesicles, particles; positions Golgi, centrosomes, mitotic spindle (required for cell division)

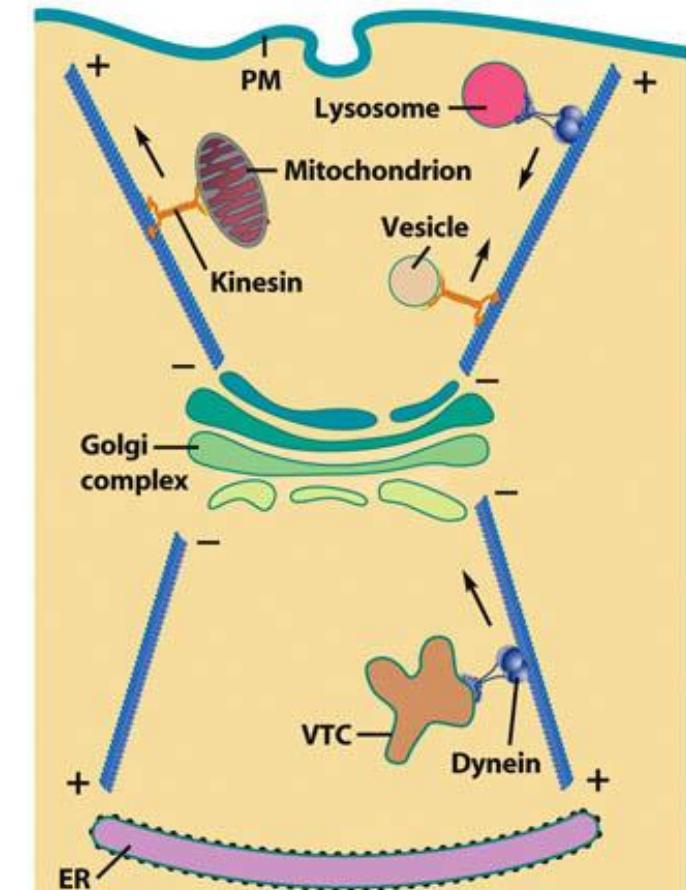
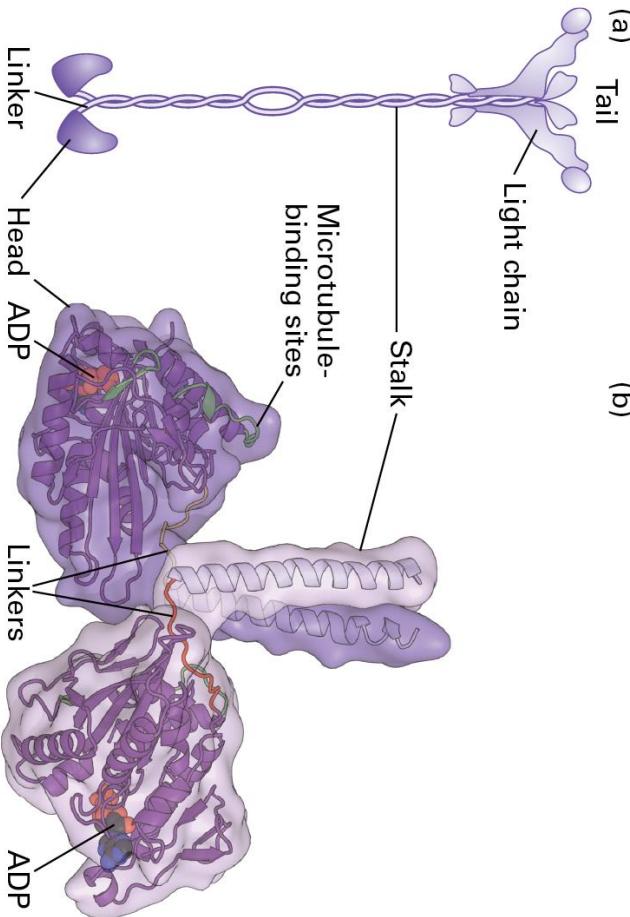


Figure 9-17c Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# Kinesin is a plus-end directed microtubular motor



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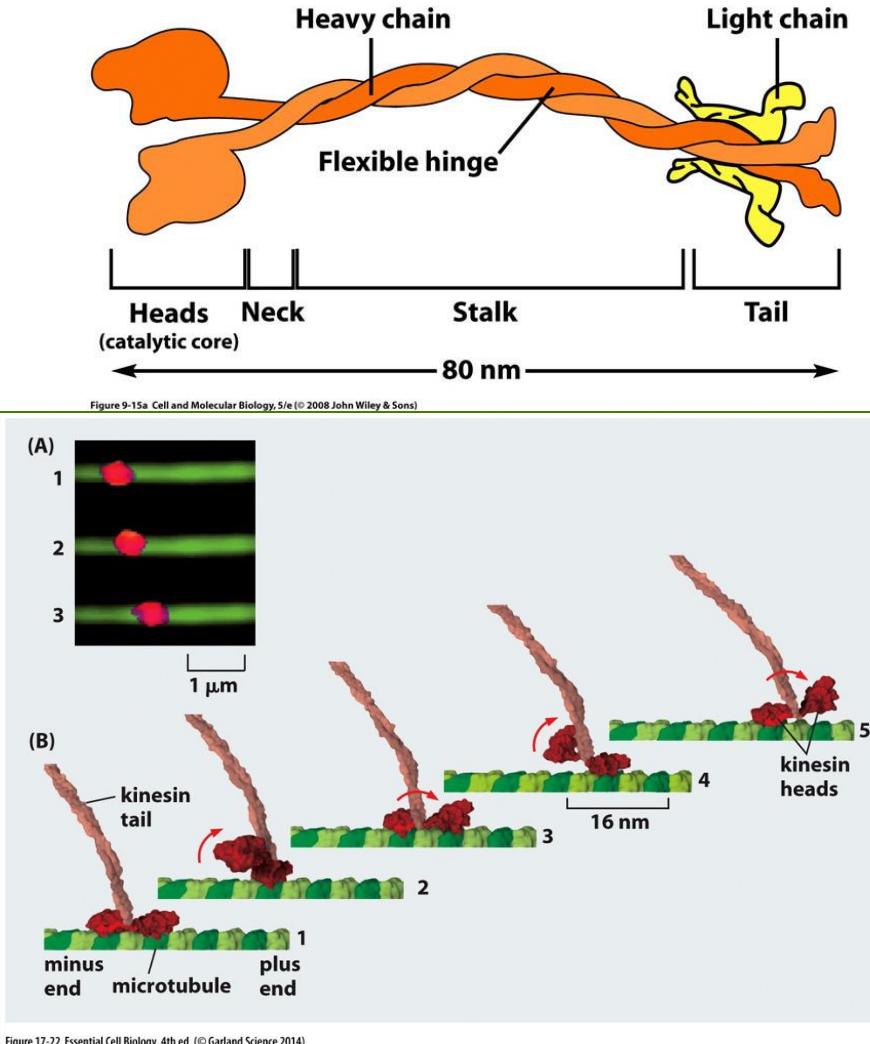


Figure 17-22 Essential Cell Biology, 4th ed. (© Garland Science 2014)

<https://www.youtube.com/watch?v=YAv4g3Pk6k>

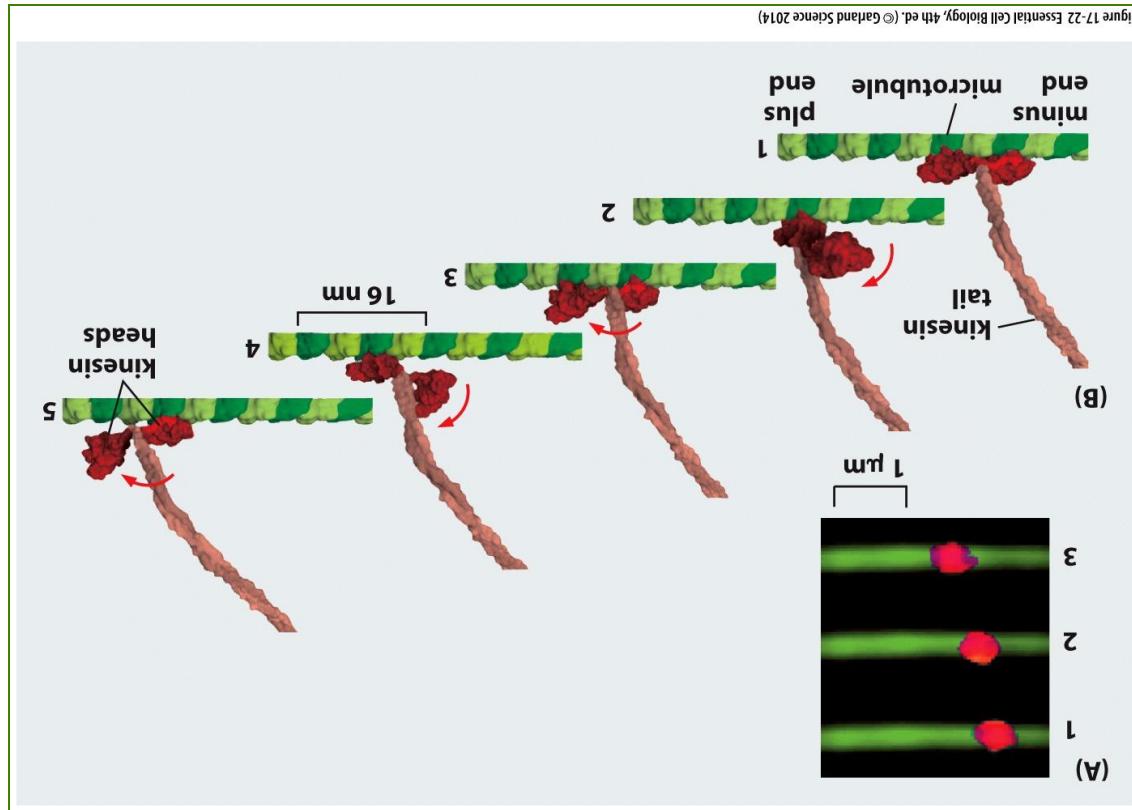
~ 45 different kinesin-like proteins (KLPs) in mammals

- heads “step” along protofilament, 1 tubulin subunit at a time towards + end
- [ATP] dependent
- processive
- tend to move organelles (mitochondria, peroxisomes) and vesicles outward

movie!

# Lightning question: Can Kinesin work upside down?

Yes  
No



# cargo moves along microtubule “tracks”

nerve cell

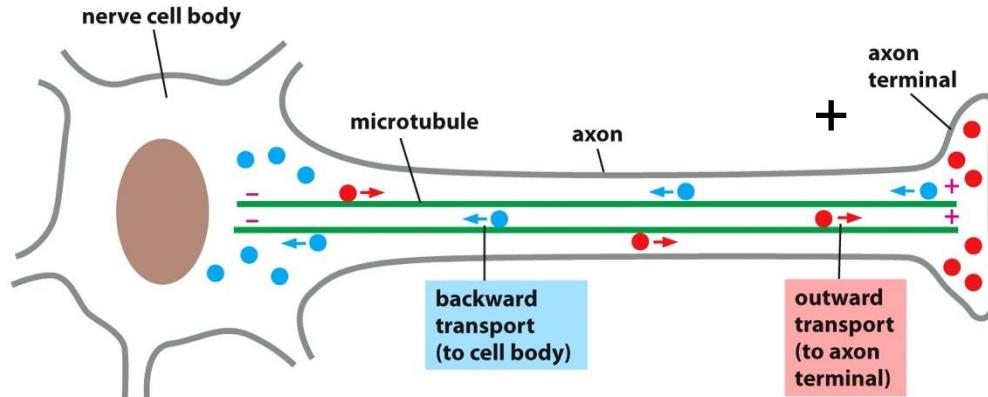


Figure 17-16 Essential Cell Biology, 4th ed. (© Garland Science 2014)

retrograde/anterograde

microtubules provide a path for movement of “cargo” such as vesicles or other protein complexes.

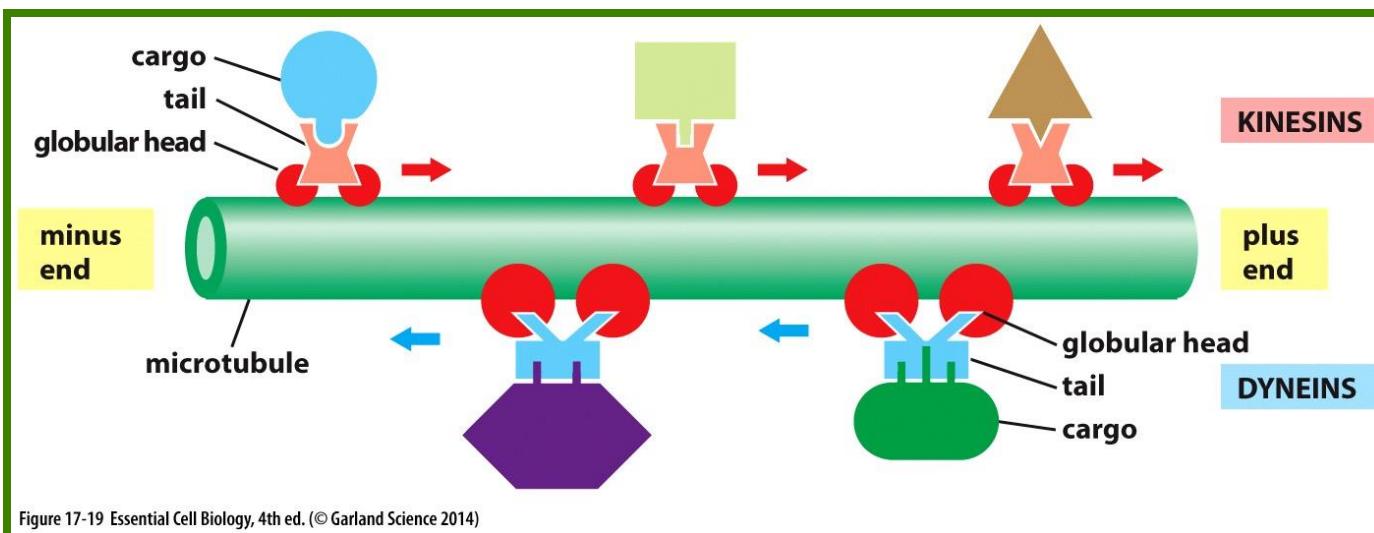


Figure 17-19 Essential Cell Biology, 4th ed. (© Garland Science 2014)

# Poll 4: how fast (and expensive) is Kinesin movement?

one step is about 8nm

let's say the distance from Golgi to the plasma membrane of an epithelial cell is 16 $\mu$ m, and that kinesin can carry cargo there in 16 seconds.

each step expends 1 ATP

How many ATPs does this trip use?

- 1)  $2 \times 10$
- 2)  $2 \times 10^2$
- 3)  $2 \times 10^3$
- 4)  $2 \times 10^4$

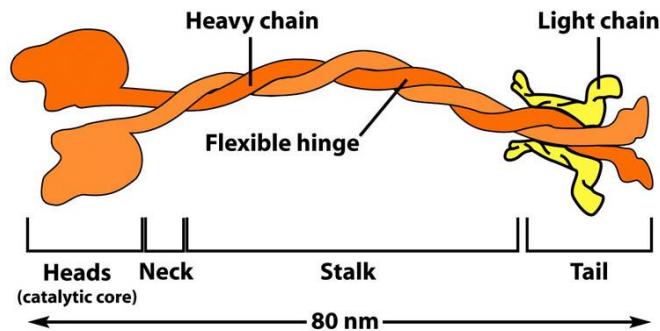
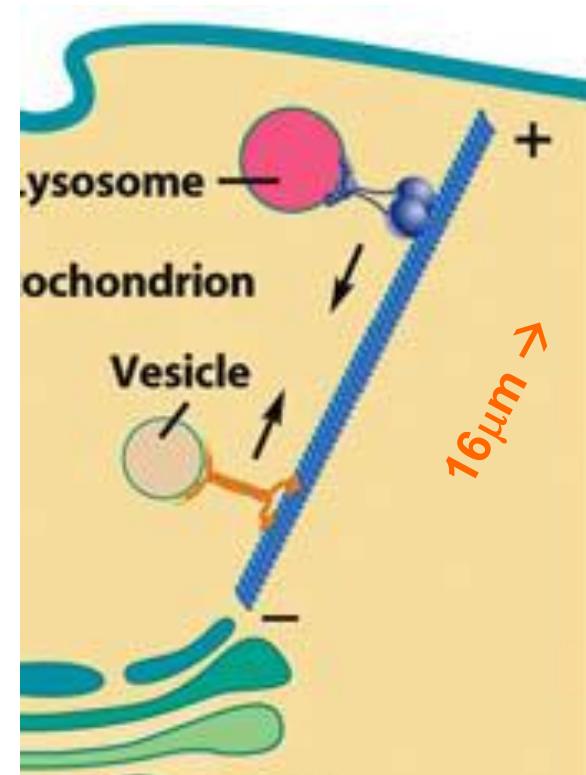
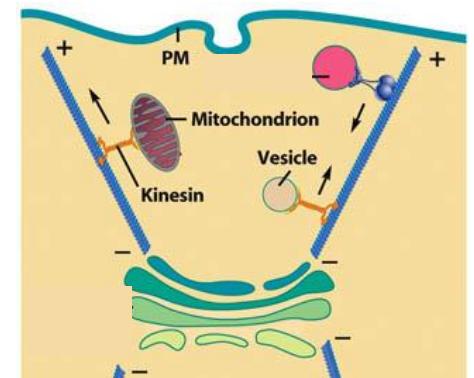
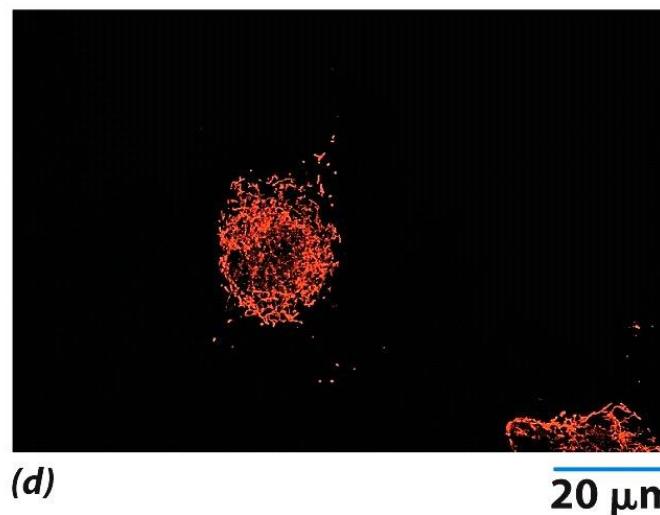
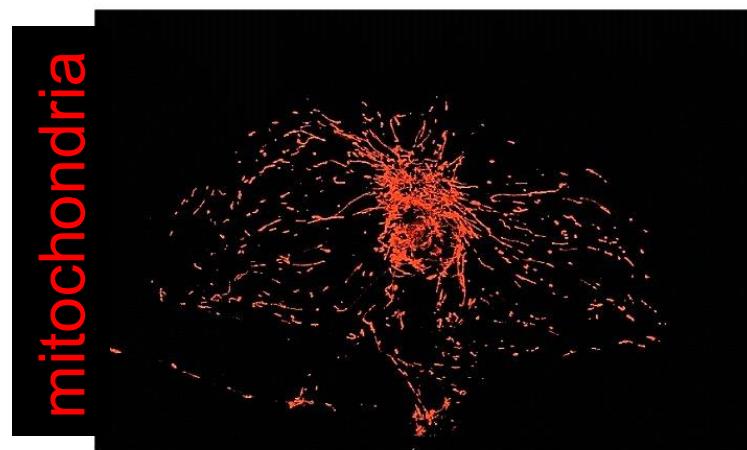
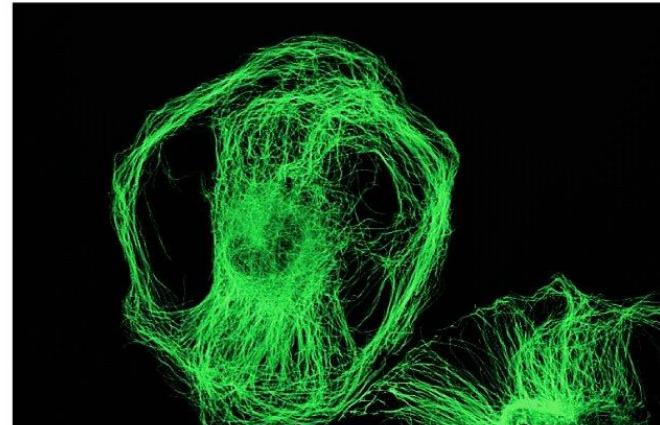
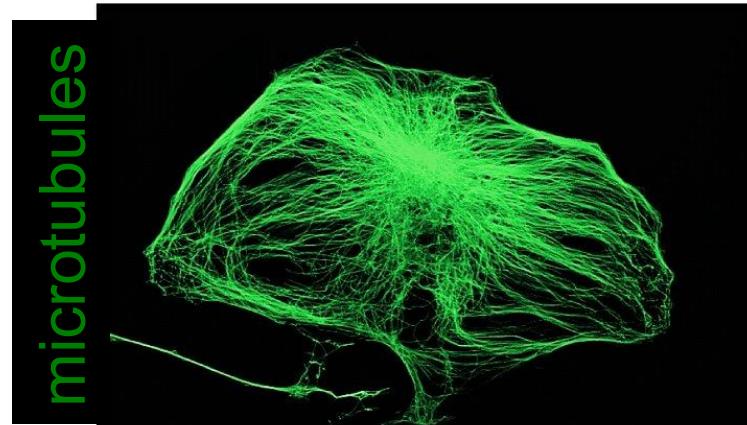


Figure 9-15a Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

# What does this result tell us about microtubules and motors?



From Yosuke Tanaka et al., courtesy of Nobutaka Hirokawa, *Cell* 93:1150, 1998; by permission of Cell Press.

Kinesin KO

# experiment

- you isolated a new motor protein and generated a **GFP-fusion protein**
- yippee! you can see it move when you put it in neuronal cells!
- you treat the cells with **nocodazole** and you see **Green** but no movement
- you conclude your motor associates with:
  - \_\_\_\_\_

in cultured neurons, your GFP fusion motor protein tends to move toward the cell body. is your motor protein a plus-end or minus-end directed motor?

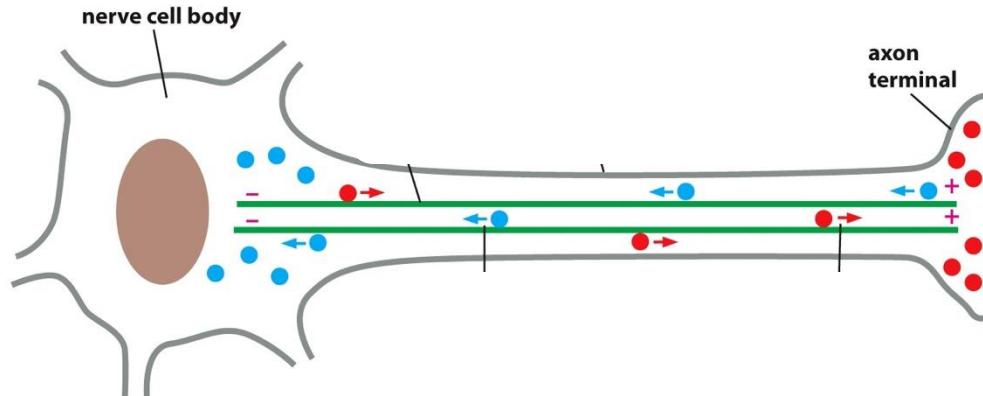


Figure 17-16 Essential Cell Biology, 4th ed. (© Garland Science 2014)

Poll 5

- a) plus
- b) minus
- c) could be either

in cultured neurons, your GFP fusion motor protein tends to move toward the cell body.  
what known kind of motor protein does this resemble?

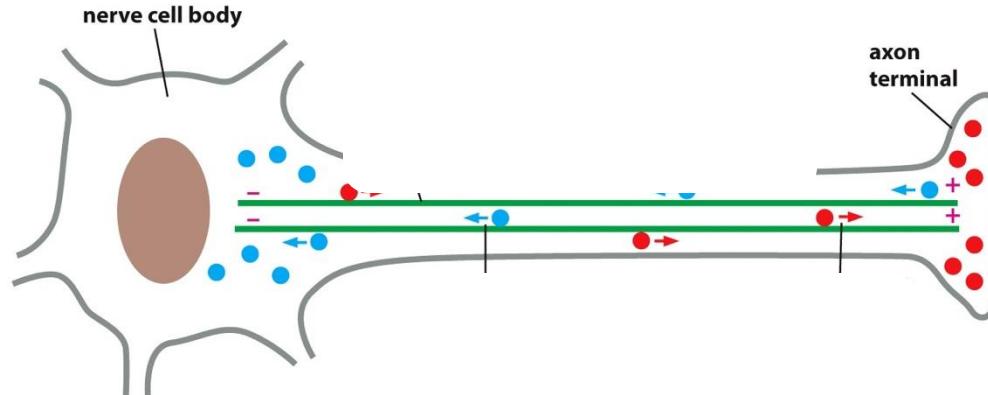


Figure 17-16 Essential Cell Biology, 4th ed. (© Garland Science 2014)

Poll 6

- a) Kinesin
- b) Myosin
- c) Dynein
- d) Keratin

# experiment

- what experiments might you do to determine what kind of cargo associates with your new motor protein? (there are a lot of answers)



# flagella are microtubule-rich structures that beat or wave

- protozoans, algae, sperm use flagella for movement
- not the same molecularly as bacterial flagella
- power stroke is the “push”; recovery stroke swings back into position

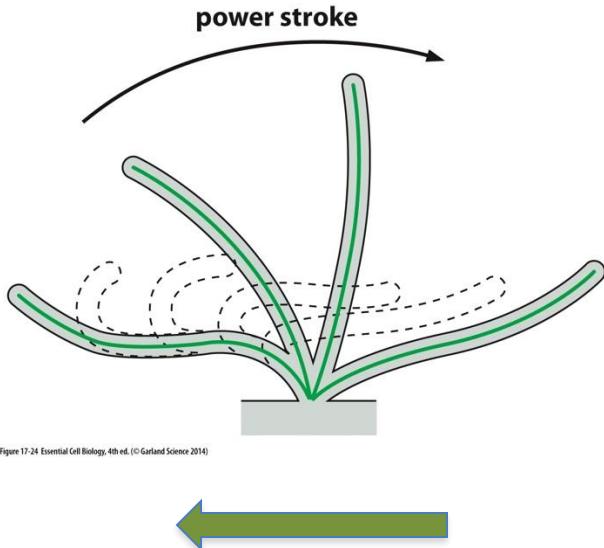


Figure 17-24 Essential Cell Biology, 4th ed. (© Garland Science 2014)

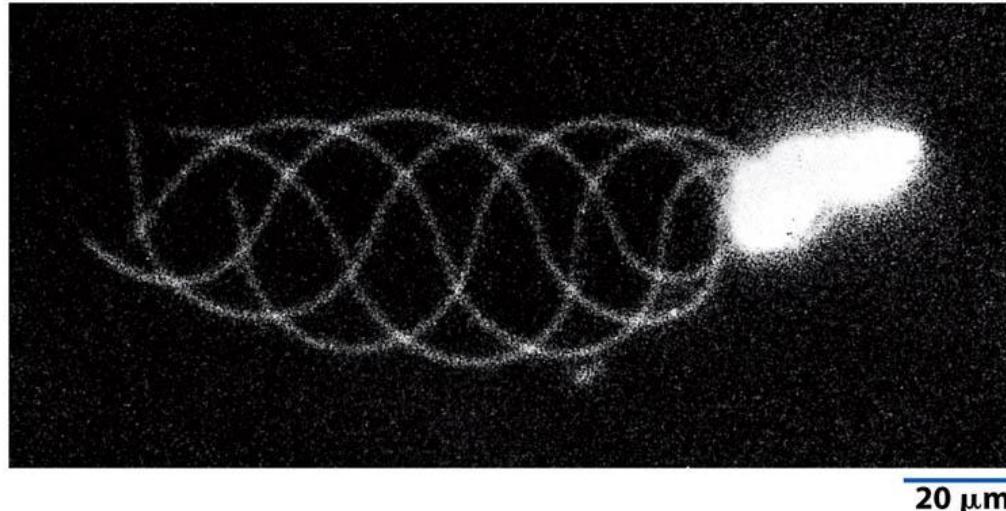
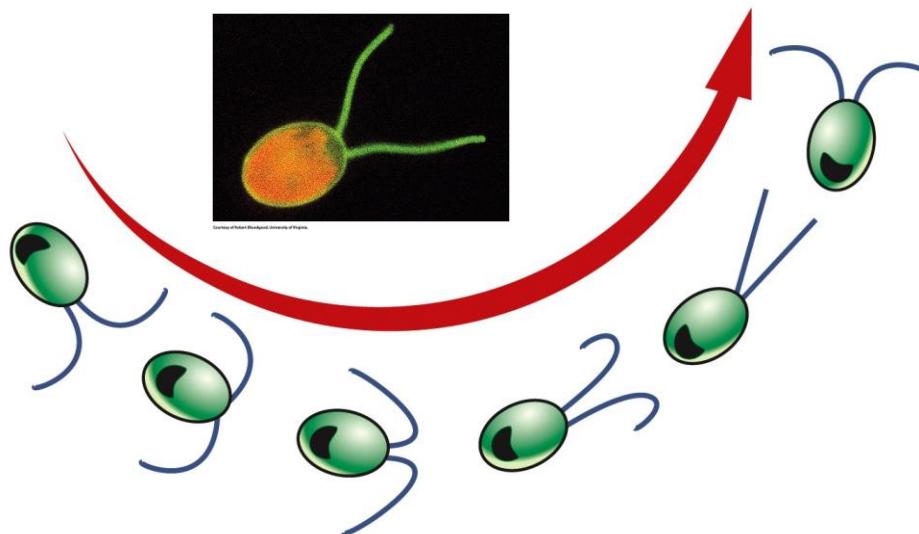
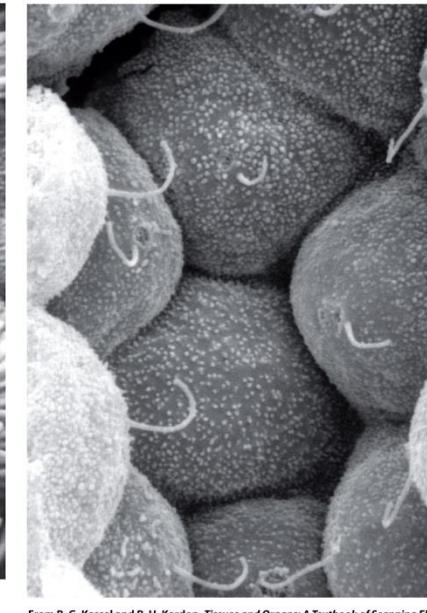
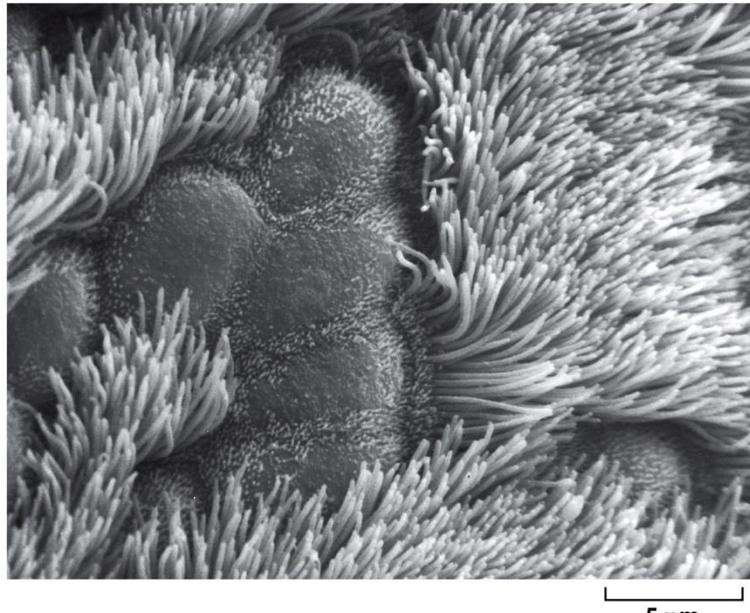


Figure 9-34 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)



# cilia are microtubule rich structures that can wave or be stationary



motile cilia examples

primary cilia

- most cells have one of these structures
- multicellular organisms use cilia to move fluid/particles.  
“primary cilia” are often non-motile, sense the local environment
- defects in cilia can lead to **disease** (Bardet-Biedl Syndrome), polycystic kidney disease, and *situs inversus*

# flagella/cilia – whip it

Classic cilia film:

- <http://www.youtube.com/watch?v=QGAm6hMysTA>

- human sperm:

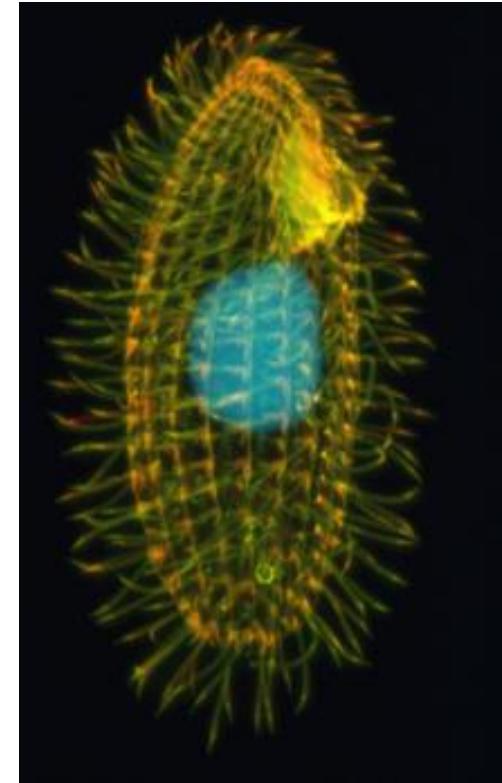
<http://www.youtube.com/watch?v=vvnEsOaKxuw&feature=related>

- soothing cilia on a frog embryo

<http://www.youtube.com/watch?v=-UAgL6GxcS0&feature=related>

airway cilia: (in culture)

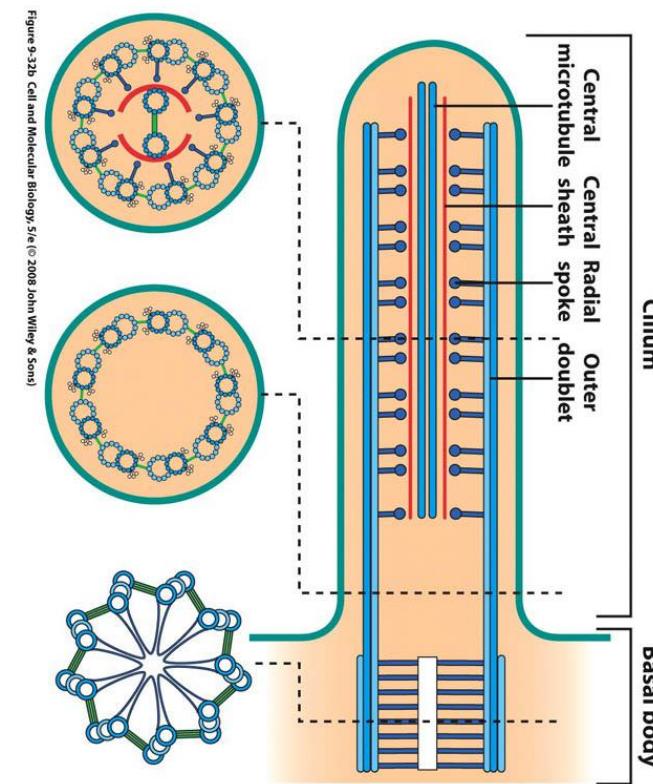
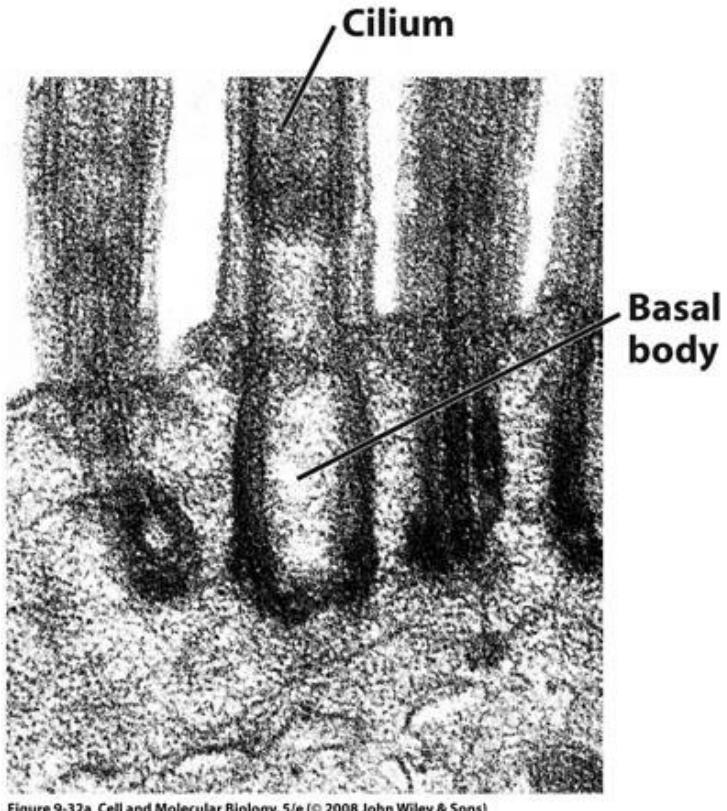
<https://www.youtube.com/watch?v=SDweiaW7fXs>



tetrahymena pic from wikipedia

## HOW DOES THIS WORK?

# the axoneme of cilia/flagella is made up of microtubules and dynein



- the basal body is a microtubule organizing center (MTOC)
- microtubules are organized with the plus end away from the cell body

# motor proteins are also needed to build/maintain the axoneme of cilia/flagella

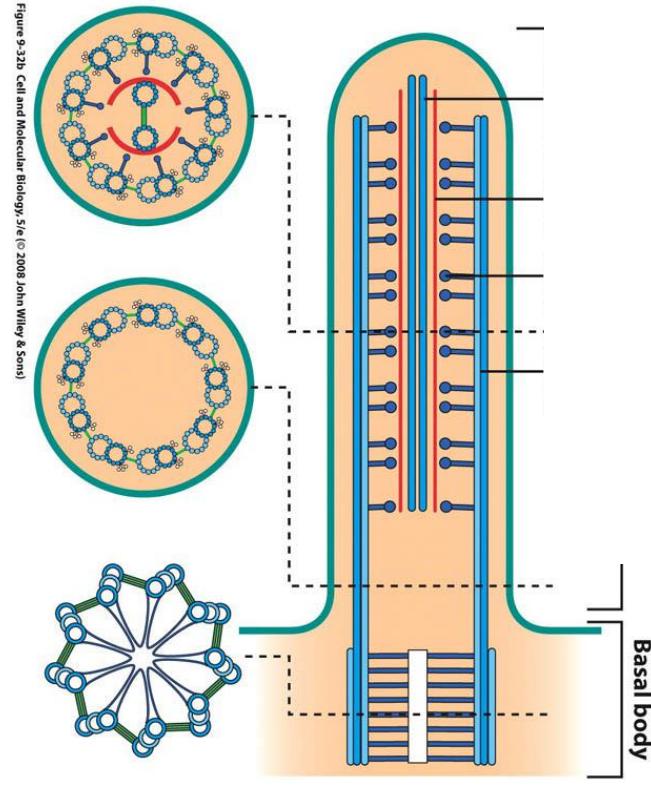
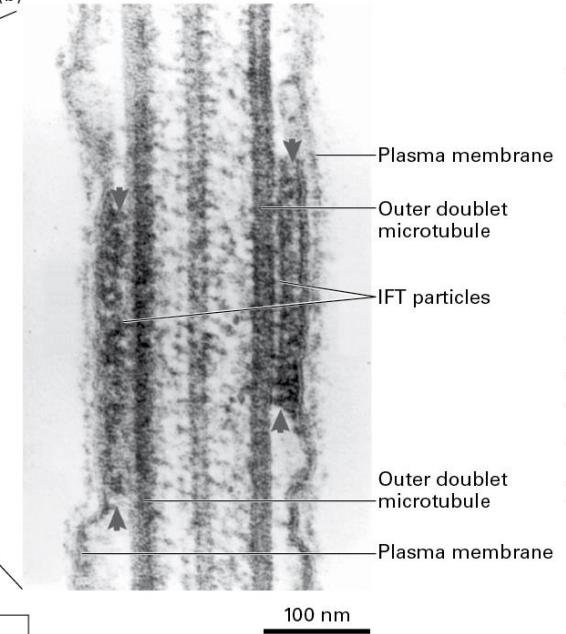
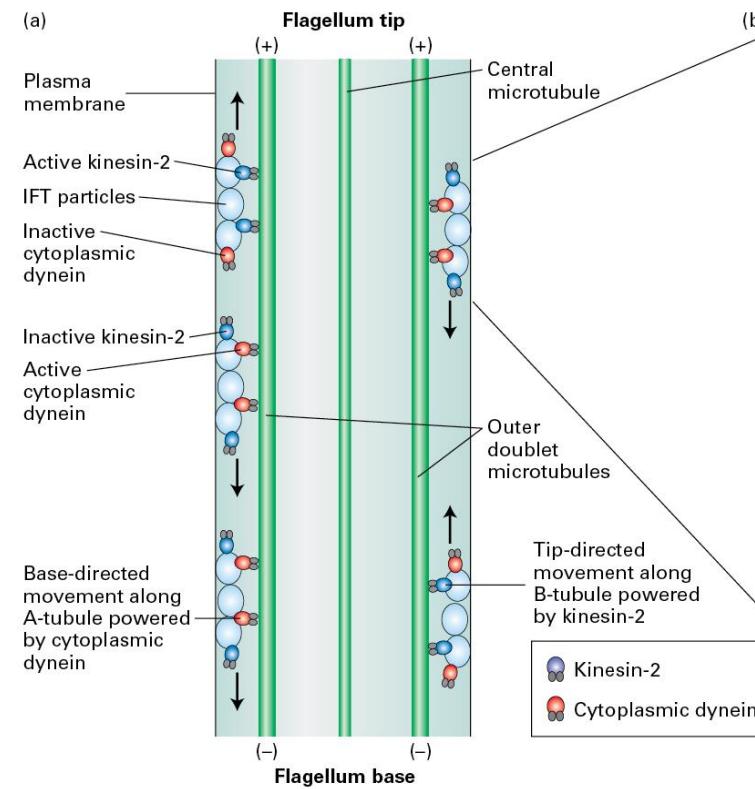


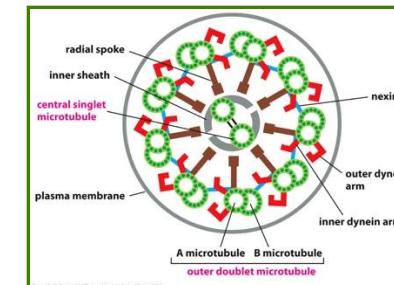
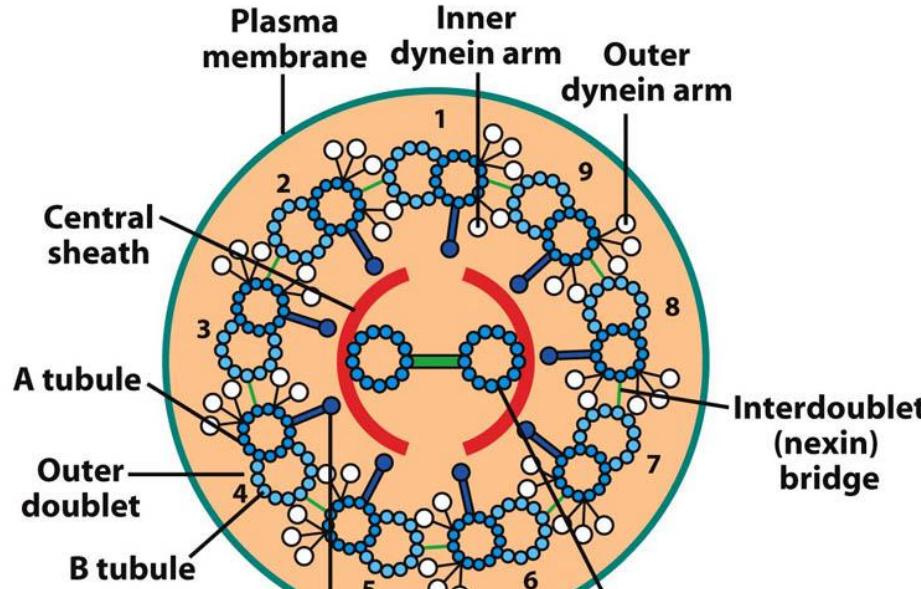
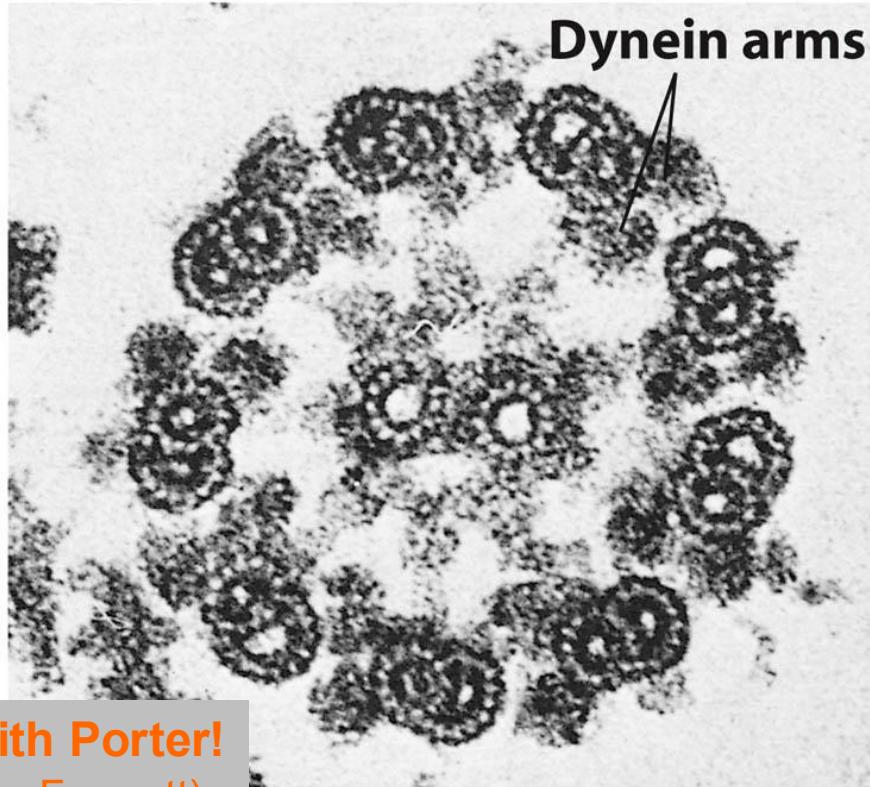
Figure 9-32b Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)



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Kinesin 2 and **cytoplasmic dynein** transport cargo in and out

# the axoneme of cilia/flagella is made up of microtubules and dynein (cross section)



the **9 + 2** array of microtubules is very well conserved, protists to mammals

# the axoneme of cilia/flagella is made up of microtubules and dynein

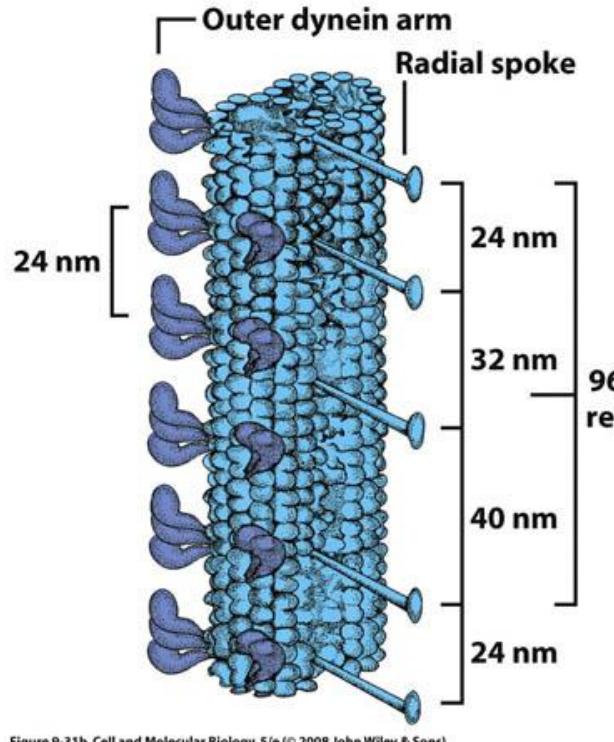


Figure 9-31b Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

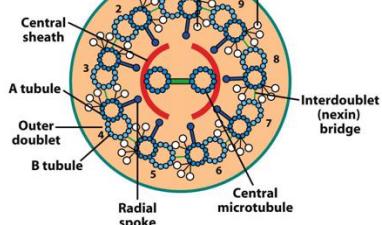
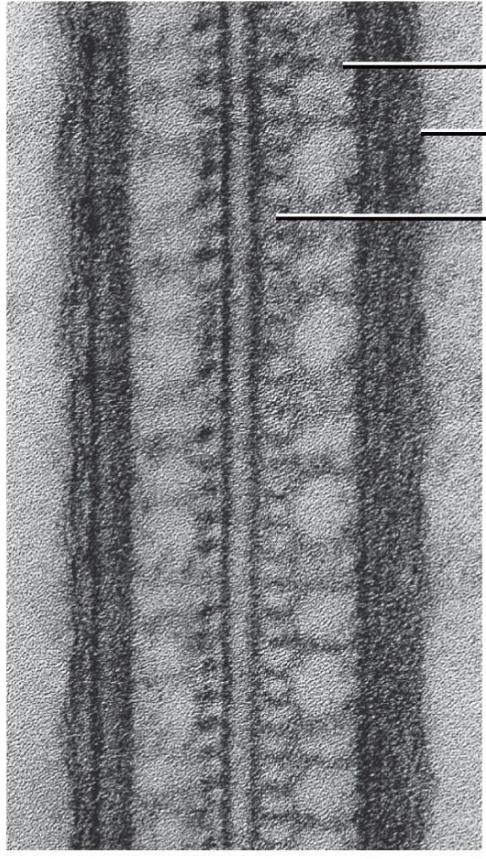
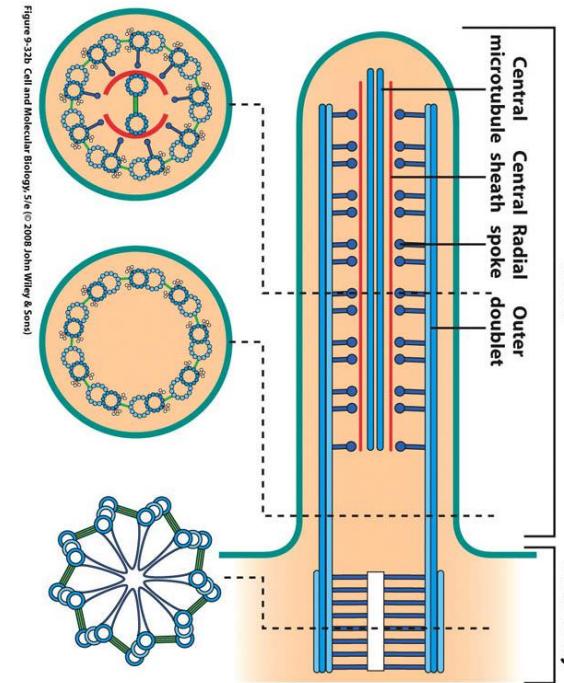


Figure 9-31b Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)



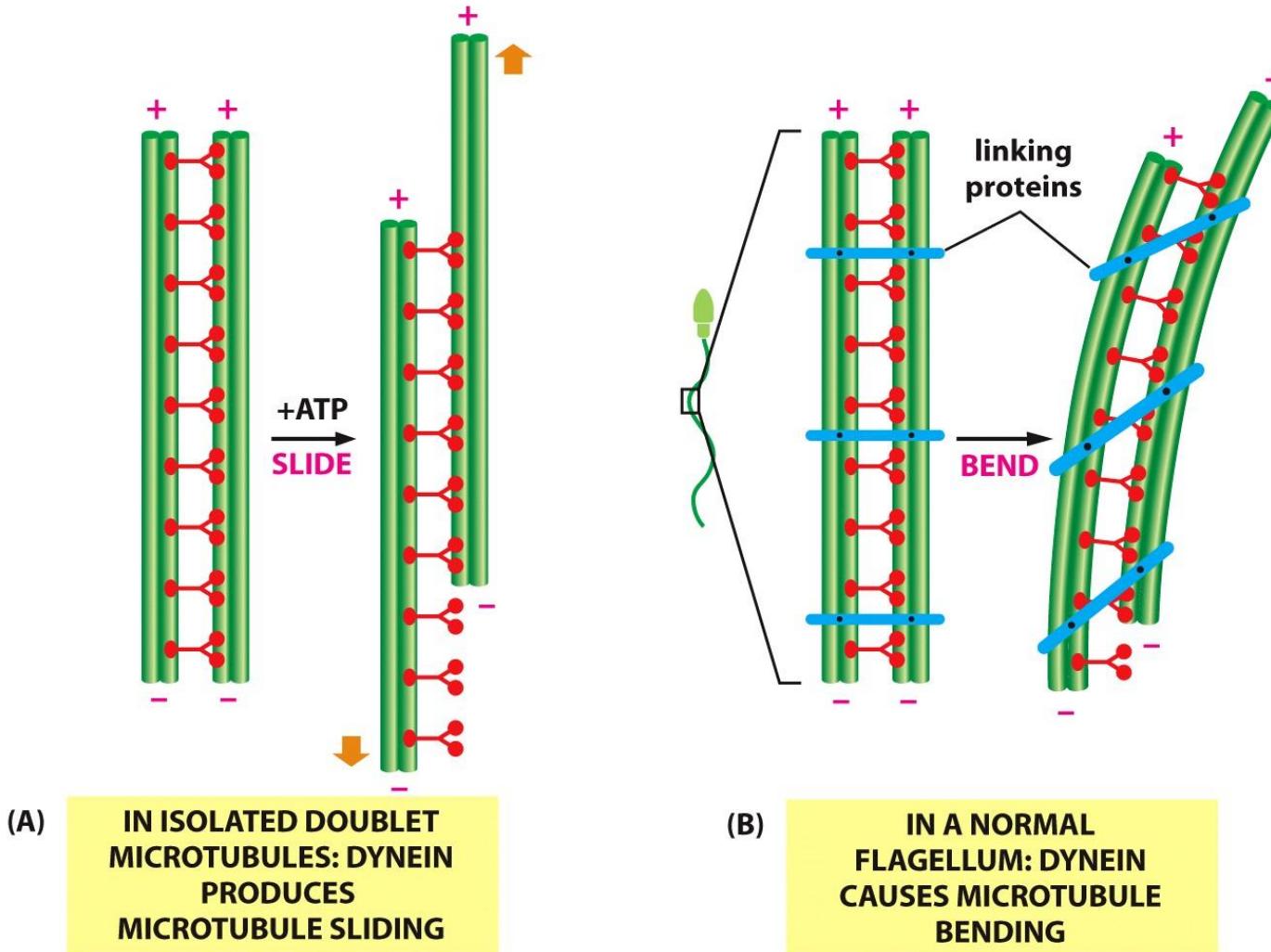
From Fred D. Warner and Peter Satir, J. Cell Biol. 63:41, 1974; by cop



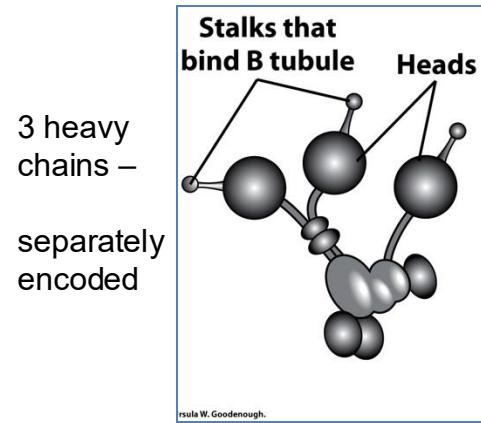
intraflagellar/ciliary transport also occurs - via dynein and kinesin

the 9 + 2 array of microtubules is very well conserved, protists to mammals

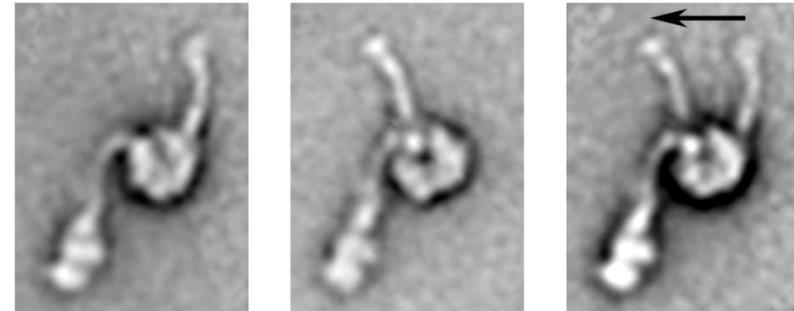
# cilia move by microtubule sliding via ciliary Dynein



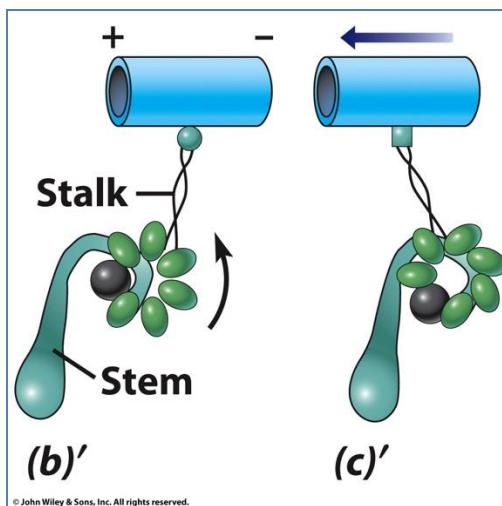
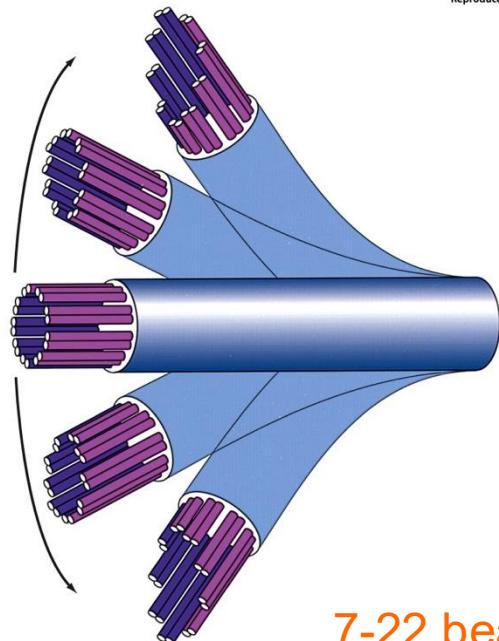
# cilia/flagella move by microtubule sliding via dynein



3 heavy  
chains –  
separately  
encoded



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7-22 beats per second in human cilia

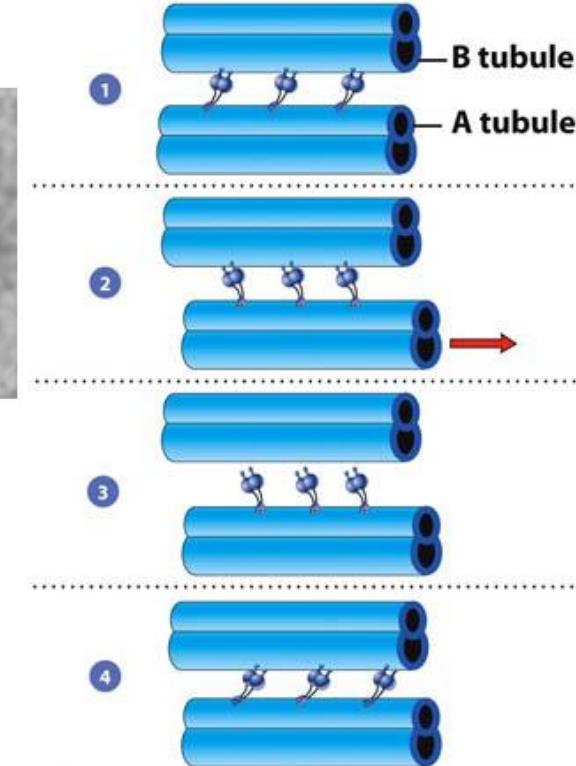
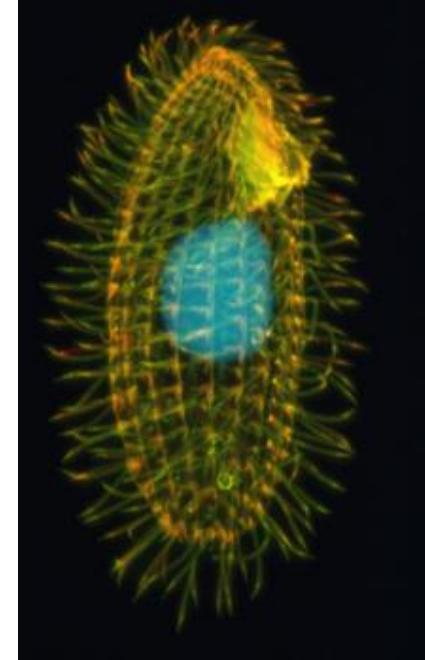
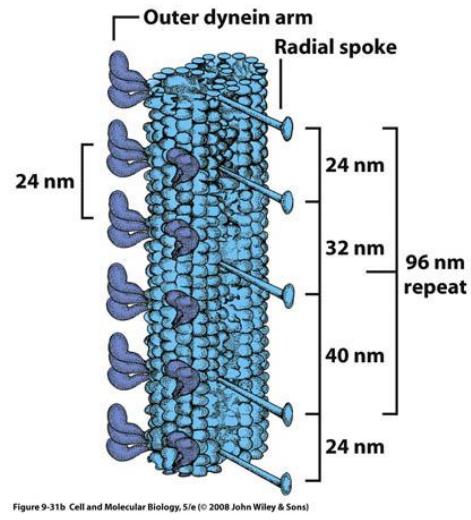
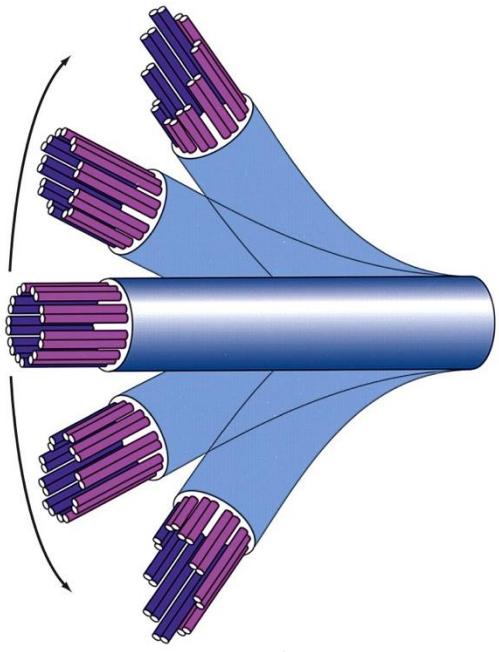


Figure 9-37 Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

stems of dyneins (and intermediate and light chains) are anchored to the A tubule

# more molecular movies...

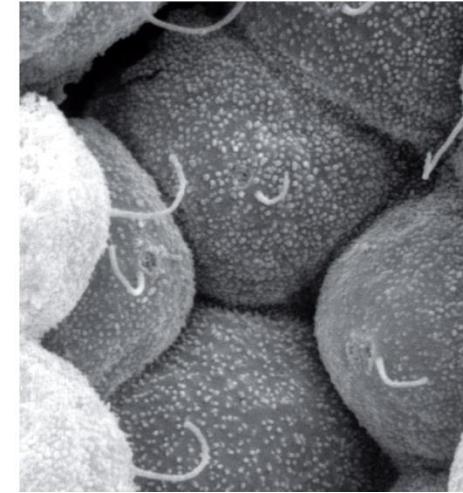
- <https://www.youtube.com/watch?v=9nZYlyFGm50>
- Fan favorite:
- <https://www.youtube.com/watch?v=tMKIPDBRJ1E>



tetrahymena pic from wikipedia

# some diseases are attributable to cilia defects

- may affect motile cilia or primary cilia
- cilia may act as antenna or signal organizing centers
- Bardet-Biedl syndrome – mutations affect cilia/basal bodies, result in polydactyly, situs inversus, congenital defects, retinal degeneration etc...
- left-right asymmetry of organs due to abnormal fluid flow in developing embryo
- Polycystic kidney disease (Ca++ channel) disrupts cell division



# Case study: Kartagener Syndrome

- Primary ciliary dyskinesias (PCD) affects 1 in 30,000 (Kartagener's in a subset)
- Rare, autosomal recessive genetic ciliary disorder (ciliopathy)
- Symptoms: situs inversus, chronic sinusitis, and bronchiectasis  
àear, nose, throat, chest infections, infertility

# Kartagener Syndrome

cilia exist but have defective or no movement

**poll 7.** a mutation that disrupts which of the following proteins is the most likely to result in this outcome:

- a) alpha-tubulin
- b) ciliary dynein
- c) gamma-tubulin
- d) kinesin
- e) ATP synthesizing enzymes

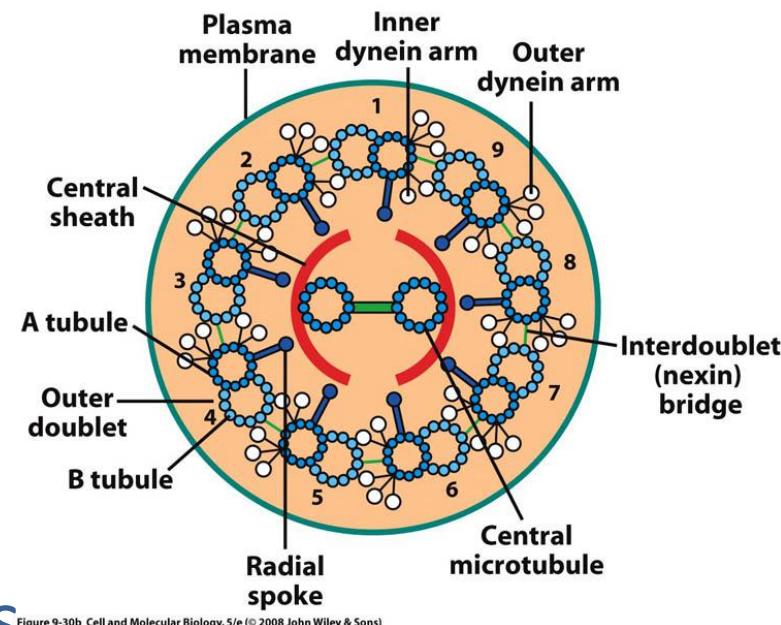


Figure 9-30b Cell and Molecular Biology, 5/e (© 2008 John Wiley & Sons)

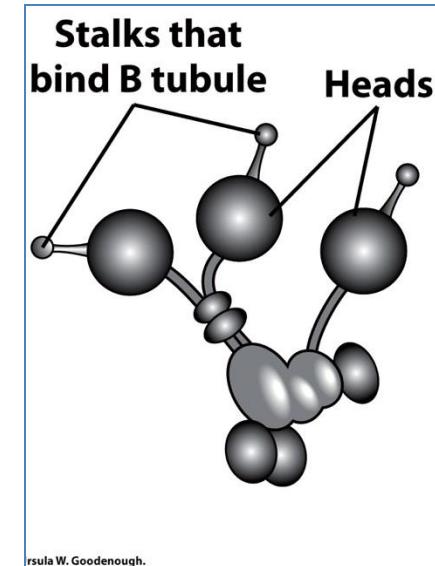
# Kartagener Syndrome

cilia exist but have defective or no movement

you want a mouse model of the disease, so you want to make a genetic mutant. you know that the human patients have only **small changes** in the coding region of the dynein protein, not entire loss, and you know that **cilia from these patients hydrolyze ATP at a similar rate to normal cilia.**

so, which region of the protein(s) might be the best place to make a **small deletion** to mimic the disease?

- a) heavy chain motor (head) domain
- b) heavy chain stem domain
- c) light chain



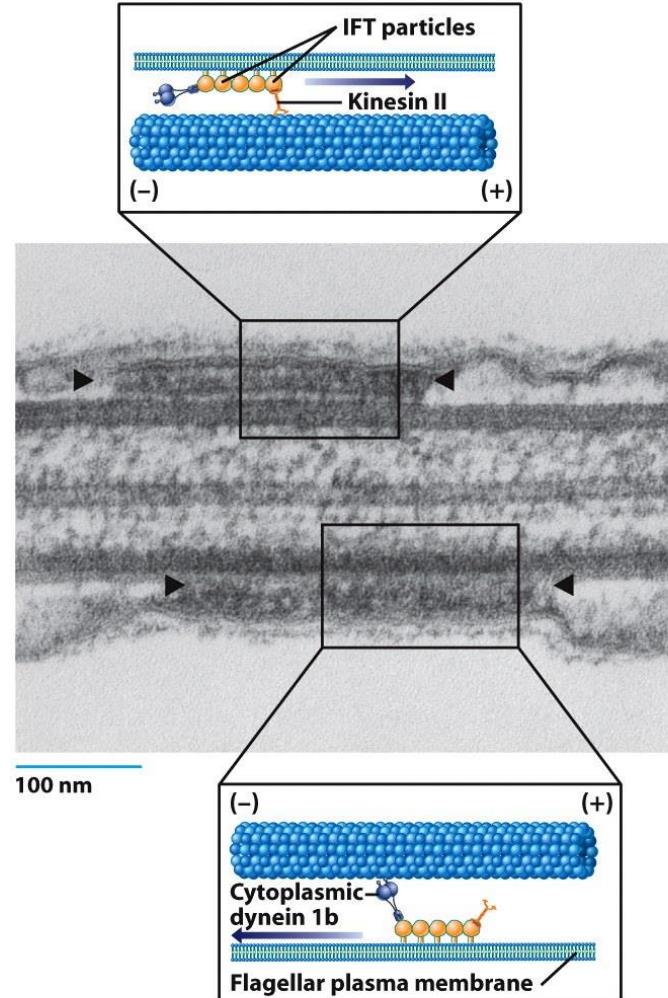
rsula W. Goodenough.

[poll](#)

# Kartagener Syndrome mutations

- cilia have defective or no movement
- you decide to also make  $\text{dynein } 1\beta$  mutation in the cytoplasmic dynein 1b. It is found in the epithelial cilia of the airways. do you see any defect in cilia when this dynein is mutated?

We didn't get to this



# discussion- microtubule dynamics

## lecture thursday: actin filaments and sarcomeres

