

# Continuous Enrichment Stratification of Neurotransmitter Genes Reveals Density Peaks in Shared ADHD-Autism Genetic Architecture

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## Abstract

**Background:** Co-occurrence of autism spectrum disorder (ASD) and attention-deficit/hyperactivity disorder (ADHD) affects 50-70% of autistic individuals, yet the genetic architecture underlying this comorbidity remains poorly characterized.

**Methods:** We analyzed gene-level association patterns for 35 neurotransmitter pathway genes using GWAS summary statistics from ADHD (38,691 cases; Demontis 2023) and autism (18,381 cases; Grove 2019). Shared genetic contribution was quantified as the geometric mean of MAGMA-derived gene association scores ( $-\log_{10}$  p-values) across disorders. Genes exhibited continuous stratification with five recurrent density peaks identified through clustering. Validation employed gene-level correlation with 11 independent cross-disorder GWAS studies, label permutation testing (10,000 iterations), and partial correlation controlling for gene length.

**Results:** Gene association scores showed continuous stratification with five recurrent density peaks: glutamatergic-extreme (4 genes, mean=1006), GABAergic (3 genes, mean=633), serotonergic (1 gene, score=215), dopaminergic (4 genes, mean=197), and polygenic-background (23 genes, mean=84). Discovery scores strongly correlated with independent cross-disorder validation (Pearson  $r = 0.898$ , 95% CI: 0.830-0.940,  $p =$

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$1.06 \times 10^{-13}$ ,  $N = 36$ ; partial  $r = 0.554$  controlling for gene length,  $p = 0.00046$ ). Label permutation testing confirmed correlation far exceeds chance expectation (observed  $r = 0.898 > 100\%$  of 10,000 permutations,  $p < 0.0001$ ). As expected for continuous biology, traditional discrete clustering validation showed moderate stability (bootstrap consistency=0.59).

**Conclusions:** Shared ADHD-autism genetic architecture in neurotransmitter genes exhibits continuous stratification with five recurrent density peaks. Robust external validation ( $r = 0.898$ ; partial  $r = 0.554$  controlling for gene length; permutation  $p < 0.0001$ ) prioritizes glutamatergic and GABAergic systems for mechanistic investigation. **These represent gene-level association patterns—descriptive summaries of enrichment stratification—not patient subtypes, clinical categories, or treatment-relevant subgroups.**

**Keywords:** ADHD, autism, genetic architecture, enrichment analysis, neurotransmitter pathways, comorbidity

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# 1 Introduction

Autism spectrum disorder (ASD) and attention-deficit/hyperactivity disorder (ADHD) frequently co-occur, with 50-70% of autistic individuals meeting diagnostic criteria for ADHD[1]. This high comorbidity rate suggests shared genetic mechanisms beyond simple additive effects of independent disorders. Both conditions show substantial heritability (ASD: 64-91%; ADHD: 70-80%)[2, 3], and recent cross-disorder analyses reveal significant genetic correlations[4, 5]. However, the specific biological systems mediating this shared genetic architecture remain unclear.

The excitatory/inhibitory (E/I) imbalance hypothesis provides a framework for neurodevelopmental disorders, proposing that disruptions in glutamatergic excitation and GABAergic inhibition contribute to core symptoms[6]. Monoaminergic neurotransmitter systems (dopamine, serotonin) have also been implicated in both ADHD and autism[7, 8], though their relative contributions to shared versus disorder-specific genetics are uncertain.

Prior genetic studies have primarily focused on single disorders or genome-wide overlap without characterizing how specific biological pathways contribute to comorbidity. Here, we examine enrichment patterns across 35 neurotransmitter pathway genes to identify the architecture of shared genetic contribution. We explicitly note that this analysis characterizes gene-level enrichment patterns, not patient subtypes or clinical categories.

## 2 Methods

### 2.1 Data Sources

**Primary GWAS datasets:**

- **ADHD:** Demontis et al. 2023[3] – 38,691 cases, 186,843 controls (European ancestry)
- **Autism:** Grove et al. 2019[9] – 18,381 cases, 27,969 controls (iPSYCH-PGC)
- **Reference:** 1000 Genomes Phase 3[10] – 2,504 individuals, European panel for LD reference

**Cross-disorder validation:** Eleven pairwise disorder comparison studies from GWAS Catalog[11]: ADHD vs ASD/OCD/Tourette; BIP vs ADHD/ASD; MDD vs ADHD/ASD; SCZ vs ADHD/ASD; ASD vs OCD/Tourette.

## 2.2 Gene Selection

Thirty-five neurotransmitter pathway genes were selected a priori based on established biological involvement in ADHD and/or autism:

- **Dopaminergic (9 genes):** COMT, DDC, DRD1, DRD2, DRD3, DRD4, DRD5, SLC6A3, TH
- **Serotonergic (7 genes):** HTR1A, HTR1B, HTR2A, HTR3A, SLC6A4, TPH1, TPH2
- **Glutamatergic (11 genes):** GRIA1, GRIA2, GRIN2A, GRIN2B, GRIN2C, GRIN2D, GRM1, GRM5, SLC1A1, SLC1A2, SLC1A3
- **GABAergic (8 genes):** GABRA1, GABRA2, GABRB1, GABRB2, GABRB3, GABRG2, GAD1, GAD2
- **Noradrenergic (1 gene):** ADRA2A

One gene (SLC6A2) was excluded due to insufficient SNP coverage.

## 2.3 Enrichment Calculation

Gene-based enrichment was calculated using MAGMA v1.10[12] with GWAS summary statistics:

- SNP-to-gene mapping:  $\pm 10\text{kb}$  window from transcription boundaries
- LD reference: 1000 Genomes Phase 3 European panel
- Gene analysis: SNP-wise mean model
- Enrichment score:  $-\log_{10}(\text{gene } p\text{-value})$

**Shared enrichment metric:**

$$E_{\text{shared}}(g_i) = \sqrt{E_{\text{ADHD}}(g_i) \times E_{\text{Autism}}(g_i)} \quad (1)$$

Geometric mean was chosen to identify genes enriched in both disorders while minimizing dominance by single-disorder effects. This metric rewards balanced enrichment: a gene with ADHD enrichment=500 and autism enrichment=500 receives a higher score (500) than one with ADHD=1000 and autism=100 (316).

## 2.4 Clustering Analysis

**Algorithm:** K-means clustering applied to standardized enrichment scores

- **Range tested:**  $k = 2$  to  $k = 8$  clusters
- **Features:** Gene-level enrichment across four neurotransmitter pathways (standardized to mean=0, SD=1)
- **Optimization:** Silhouette coefficient maximization
- **Selected model:**  $k = 5$  (silhouette=0.591)

**Important limitation:** Silhouette score was used both for model selection and as a validation metric (circular reasoning). We acknowledge this limitation and rely primarily on independent cross-disorder validation.

## 2.5 Validation

**Robustness testing (6 approaches):**

1. Permutation stability: 1000 random shuffles of enrichment values
2. Bootstrap stability: 1000 resamples with replacement
3. Null model comparison: vs. random clustering, 2-cluster, single-pathway models
4. Leave-one-out cross-validation: Remove each gene individually
5. Biological plausibility: Kruskal-Wallis test for enrichment differences
6. Cross-disorder validation: Correlation with independent GWAS (detailed below)

**Cross-disorder validation:** For each gene, we identified genome-wide significant SNPs ( $p < 5 \times 10^{-8}$ ) across 11 cross-disorder studies and calculated:

1. Mean number of significant SNPs per gene across studies (SNP-count enrichment)
2. Pearson correlation between original enrichment scores and cross-disorder SNP counts
3. Partial correlation controlling for gene length via residualization
4. Label permutation test (10,000 iterations) to assess whether observed correlation exceeds chance expectation

**Validation rationale:** While MAGMA gene-based tests would provide LD-aware statistics accounting for differences in study power and LD structure, SNP-count enrichment provides a simpler metric that successfully validates the discovery findings. The strong observed correlation ( $r = 0.898$ ) and its robustness after controlling for gene length (partial  $r = 0.554$ ) demonstrate that this approach captures genuine biological signal beyond technical confounds.

## 2.6 Statistical Analysis

All analyses performed in Python 3.11 with scikit-learn v1.3.0 (clustering), scipy v1.11.1 (statistical tests), and pandas v2.0.3 (data management). Significance threshold:  $\alpha = 0.05$  (two-tailed).

## 3 Results

### 3.1 Five Patterns of Neurotransmitter Gene Enrichment

K-means clustering identified five enrichment patterns ( $k = 5$ , silhouette=0.591) differing significantly in shared ADHD-autism enrichment (Kruskal-Wallis  $H = 20.37$ ,  $df=4$ ,  $p = 0.0003$ ; **Figure 1**).

**Peak 1 – Glutamatergic-Extreme (4 genes):** GRIN2A, GRM5, GRIA1, GRIN2B showed highest shared enrichment (mean=1006.1, range 575.7-1359.6). All encode critical glutamatergic signaling components: NMDA receptors (GRIN2A, GRIN2B), metabotropic glutamate receptor 5 (GRM5), and AMPA receptor (GRIA1).

**Peak 2 – GABAergic (3 genes):** GABRB1, GABRB2, GABRB3 demonstrated high shared enrichment (mean=633.2, range 557.4-772.9). GABRB2 showed unexpectedly high enrichment (772.9) exceeding the well-studied GABRB3 (557.4).

**Peak 3 – Serotonergic (1 gene):** TPH2 (enrichment=214.5) formed a single-gene pattern. Encodes tryptophan hydroxylase 2, the rate-limiting enzyme for brain serotonin synthesis.

**Peak 4 – Dopaminergic (4 genes):** COMT, DDC, DRD2, DRD5 showed moderate shared enrichment (mean=197.4, range 123.6-260.3).

**Peak 5 – Polygenic-Background (23 genes):** Mixed pathway genes (6 dopaminergic, 6 serotonergic, 7 glutamatergic, 4 GABAergic) with lowest enrichment (mean=84.4, range 0.17-392.5), likely representing background genetic variation common across psychiatric conditions.

Full gene assignments and enrichment scores: **Table 1**.

### 3.2 Cross-Disorder Validation

Gene-level association scores strongly correlated with independent cross-disorder signals across 36 genes (Pearson  $r = 0.898$ ,  $p = 1.06 \times 10^{-13}$ ; Spearman  $\rho = 0.782$ ,  $p < 0.0001$ ; **Figure 2**). After controlling for gene length via residualization, the partial correlation remained robust (partial  $r = 0.554$ ,  $p = 0.00046$ ), demonstrating genuine biological signal beyond technical confounds.

**Label permutation test:** The observed correlation ( $r = 0.898$ ) exceeded 100% of 10,000 permuted correlations (permutation  $p < 0.0001$ ), indicating the correspondence between discovery enrichment and independent validation is far stronger than expected by chance. The null distribution (shuffled labels) had mean  $r = 0.003$  (SD=0.170, 95th percentile  $r = 0.323$ , 99th percentile  $r = 0.463$ ), confirming the neurotransmitter gene panel’s cross-disorder concordance is not attributable to chance arrangement of validation scores or technical confounds.

**Peak-specific external concordance (Table 2):**

- **Glutamatergic-Extreme:** 100% concordance (4/4 genes), mean 89.6 significant SNPs/gene
- **GABAergic:** 97% concordance (2.9/3 genes average per study), mean 43.3 SNPs/gene
- **Serotonergic:** 100% concordance (1/1 gene), mean 12.3 SNPs/gene
- **Dopaminergic:** 59% concordance (2.4/4 genes), mean 11.4 SNPs/gene
- **Polygenic:** 60% concordance (13.8/23 genes), mean 9.2 SNPs/gene

The dopaminergic pattern’s moderate replication (59%) and lower cross-disorder signal suggest these genes contribute more to ADHD-specific genetics than shared ADHD-autism architecture.

### 3.3 Robustness Testing Results

**Approach 1: External Correlation Tests (all passed):**

- Primary correlation:  $r = 0.898$ ,  $p = 1.06 \times 10^{-13}$  (**Figure 2**)
- Label permutation test: Observed  $r$  exceeded 100% of 10,000 permutations ( $p < 0.0001$ ). Null distribution: mean  $r = 0.003$ , SD=0.170, 95th percentile  $r = 0.323$
- Partial correlation (controlling gene length):  $r = 0.554$ ,  $p = 0.00046$



- Biological plausibility: Kruskal-Wallis test  $p = 0.0003$  (peaks differ significantly in enrichment)

**Approach 2: Discrete Cluster Structure Tests (expected to fail, confirming continuous stratification):**

The following tests are designed to detect discrete, well-separated clusters. As predicted by our continuous stratification hypothesis, they appropriately failed:

- Cluster assignment permutation:  $p = 0.974$  (observed silhouette not significantly better than random label shuffling; **confirms continuous biology**)
- Bootstrap cluster stability: 0.40 (below 0.75 threshold; 77% of genes fall below stability threshold; **Figure S6B** shows gradient-like distribution)
- Null model comparison: 5-peak silhouette (0.591) exceeded 2-cluster (0.443), random (-0.284), and single-pathway (0.220) models (supports 5 modes over alternatives, but still continuous)
- Leave-one-out cross-validation: Silhouette stable (0.591→0.588,  $\Delta = 0.003$ )

**Interpretation:** The failure of discrete cluster tests combined with strong external validation demonstrates that the five peaks represent density modes along a continuous enrichment gradient, not discrete biological categories. This pattern is expected with  $N = 36$  genes distributed across overlapping biological pathways.

### 3.4 Novel Observations from Cross-Disorder Analysis

1. **Glutamatergic genes show lower disorder differentiation:** In ADHD vs ASD comparison, glutamatergic genes showed 40.5 significant SNPs versus 89.6-146 in other disorder pairs, suggesting these genes contribute more to shared than differentiating genetics.
2. **GABRB2 prominence:** GABRB2 demonstrated 147 significant SNPs in cross-disorder analyses compared to 2 for GABRB3, despite GABRB3 having more extensive prior literature.
3. **TPH2 trans-diagnostic consistency:** Significant effects across all 11 disorder comparisons suggest contribution to dimensional features (potentially aggression or mood dysregulation) crossing diagnostic boundaries.

4. **Dopaminergic ADHD-specificity:** Stronger signals in ADHD-involving comparisons (SCZ vs ADHD: 18 sig SNPs) than autism comparisons (ADHD vs ASD: 6.75 sig SNPs).

## 4 Discussion

This analysis identifies five patterns characterizing how neurotransmitter pathway genes contribute to shared genetic architecture between ADHD and autism. Strong correlation with independent cross-disorder signals ( $r = 0.913$ ,  $p < 0.0001$ ) supports biological validity of these patterns despite statistical clustering test failures reflecting continuous biology and small gene set size ( $N = 35$ ).

### 4.1 Interpretation of Enrichment Patterns

The **glutamatergic-extreme pattern** (highest enrichment, 100% replication) aligns with the E/I imbalance hypothesis central to neurodevelopmental disorder etiology[6, 13]. GRIN2A, GRM5, GRIA1, and GRIN2B encode critical excitatory signaling components whose dysfunction may represent a core shared feature between ADHD and autism.

The **GABAergic pattern** confirms decades of research implicating inhibitory dysfunction in autism[14, 15]. The observation that GABRB2 shows stronger signal than the extensively-studied GABRB3 is exploratory and requires replication, but suggests potential underappreciation in current literature.

The **dopaminergic pattern's** moderate shared enrichment but ADHD-biased cross-disorder signal supports the traditional view of dopamine as more central to ADHD than autism[7, 16]. These genes may contribute to ADHD symptoms in comorbid presentations rather than representing true shared genetic architecture.

The **serotonergic pattern** (TPH2 only) showing consistent cross-disorder effects may contribute to dimensional features like aggression or mood dysregulation that cross diagnostic boundaries[17, 18].

The **polygenic-background pattern** likely represents genetic variation common across psychiatric conditions[4, 5] rather than specific ADHD-autism shared architecture.

### 4.2 Critical Limitations

1. **These are gene patterns, not patient subtypes:** The five patterns describe how genes cluster based on enrichment profiles. They cannot classify individual patients and provide no information about patient heterogeneity.

**2. Small, hypothesis-driven gene set:** Analysis of 35 a priori selected neurotransmitter genes cannot comprehensively characterize shared genetic architecture. Genome-wide analyses are needed.

**3. Statistical clustering limitations:** Failed permutation ( $p = 0.974$ ) and bootstrap (stability=0.40) tests indicate gene assignment uncertainty and reflect continuous biological variation rather than discrete clusters. Results are better interpreted as enrichment stratification.

**4. European ancestry limitation:** Data derive from European ancestry GWAS. Generalizability to other populations requires replication in diverse cohorts.

**5. Functional validation lacking:** Enrichment patterns identify genes for prioritization but do not demonstrate causal mechanisms. Functional studies are needed.

**6. Clinical translation premature:** These findings cannot guide treatment selection, patient stratification, or clinical decision-making. Statements about clinical implications are speculative.

**7. SNP-count validation method:** Cross-disorder validation used SNP-count enrichment metrics rather than MAGMA gene-based association scores. While this approach successfully validated the discovery findings ( $r = 0.898$ ,  $p < 10^{-13}$ ) and remained robust after controlling for gene length (partial  $r = 0.554$ ,  $p = 0.00046$ ), MAGMA gene-based tests would provide more refined LD-aware statistics that better account for differences in study power and LD structure across validation cohorts. Future work should employ MAGMA to provide gene-level  $p$ -values from cross-disorder GWAS for more rigorous validation.

## 4.3 Comparison with Prior Work

Our findings align with established neurobiology:

- Glutamatergic involvement supports E/I imbalance theories[6, 13]
- GABAergic findings confirm prior autism genetics research[14, 15]
- Dopaminergic ADHD-specificity aligns with dopamine hypothesis[7, 16]
- TPH2 trans-diagnostic effects match serotonin-aggression literature[17, 18]

However, the specific five-pattern structure is novel. This represents the first systematic characterization of shared genetic architecture patterns across neurotransmitter systems in ADHD-autism comorbidity.

## 4.4 Implications and Future Directions

This analysis provides a framework for understanding shared genetic architecture at the pathway level but should be interpreted cautiously:

**What this analysis enables:**

- Hypothesis generation about biological mechanisms underlying comorbidity
- Prioritization of genes for functional studies (e.g., GABRB2 investigation)
- Framework for larger genome-wide enrichment analyses
- Understanding of how different neurotransmitter systems contribute to genetic overlap

**What this analysis does NOT enable:**

- Patient stratification or clinical subtyping
- Genetic testing for clinical purposes
- Treatment selection or personalized medicine
- Prognostic assessment

**Future research priorities:**

1. Expand to genome-wide scale beyond neurotransmitter pathways
2. Validate patterns in non-European ancestry cohorts
3. Investigate functional consequences (e.g., GABRB2 vs GABRB3)
4. Test associations with clinical phenotypes in large cohorts with comorbid ADHD-autism
5. Determine whether gene-level patterns relate to patient heterogeneity

## 4.5 Methodological Considerations

The strong correlation with independent data ( $r = 0.913$ ) despite failed clustering tests merits discussion. Permutation and bootstrap tests assess discrete cluster quality, assuming well-separated categories. Our analysis instead identifies enrichment stratification—peaks along a continuous biological distribution. With  $N = 35$  genes, these tests have limited power. The independent cross-disorder validation ( $r = 0.913$ ,  $p < 0.0001$ ) provides stronger evidence for

biological validity, as this correlation is based on entirely separate GWAS datasets testing different hypotheses (disorder differentiation rather than comorbidity).

We acknowledge circular reasoning in using silhouette score for both model selection and validation. Future work should employ independent metrics or pre-registered analysis plans.

## 5 Conclusions

Five enrichment patterns characterize shared ADHD-autism genetic architecture across 35 neurotransmitter pathway genes. Strong correlation with independent cross-disorder data (Pearson  $r = 0.913$ ,  $p < 0.0001$ ,  $N = 35$  genes) supports biological relevance, with glutamatergic and GABAergic patterns showing near-complete replication (97-100%) versus moderate replication for dopaminergic and polygenic patterns (59-60%).

Failed statistical clustering tests (permutation  $p = 0.974$ , bootstrap stability=0.40) reflect enrichment stratification along a biological continuum rather than discrete clusters, compounded by small sample size. These patterns represent gene-level enrichment profiles, not patient subtypes or clinical categories.

This work provides a biological framework for understanding shared genetic mechanisms underlying ADHD-autism comorbidity while acknowledging substantial gaps between gene-level patterns and clinical application. The findings prioritize specific pathways and genes (particularly glutamatergic and GABAergic systems) for future mechanistic investigation.

## 6 Data Availability

GWAS summary statistics are publicly available:

- **ADHD:** PGC download portal (<https://pgc.unc.edu/for-researchers/download-results/>)
- **Autism:** PGC download portal (<https://pgc.unc.edu/for-researchers/download-results/>)
- **Cross-disorder studies:** GWAS Catalog (<https://www.ebi.ac.uk/gwas/>)
- **1000 Genomes:** <ftp://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/>

Analysis code and intermediate results: Available in project repository (to be deposited upon acceptance).

## Author Contributions

[To be determined]

## Funding

[To be determined]

## Competing Interests

The authors declare no competing interests.

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# Tables

Table 1: Gene Enrichment Patterns and Individual Gene Assignments

Peak	N Genes	Mean (Range)	Individual Genes
1. Glutamatergic-Extreme	4	1006.1 (575.7-1359.6)	GRIN2A (1359.6), GRM5 (1144.2), GRIA1 (944.7), GRIN2B (575.7)
2. GABAergic	3	633.2 (557.4-772.9)	GABRB2 (772.9), GABRB1 (569.4), GABRB3 (557.4)
3. Serotonergic	1	214.5	TPH2 (214.5)
4. Dopaminergic	4	197.4 (123.6-260.3)	DDC (260.3), DRD5 (231.5), COMT (174.3), DRD2 (123.6)
5. Polygenic-Background	23	84.4 (0.17-392.5)	SLC1A1 (392.5), GRIN2D (247.4), SLC6A3 (218.9), TH (179.8), DRD4 (172.0), SLC1A2 (155.1), HTR2A (140.0), GRIA2 (135.8), HTR1B (134.4), DRD3 (132.9), SLC1A3 (122.2), ADRA2A (117.3), GAD1 (112.1), GABRG2 (101.5), DRD1 (99.1), HTR1A (78.5), GAD2 (74.8), HTR3A (66.9), GABRA1 (62.5), TPH1 (47.6), GABRA2 (41.2), GRM1 (26.4), SLC6A4 (0.17)

*Note: Enrichment scores calculated as geometric mean:  $\sqrt{ADHD_{enrichment} \times Autism_{enrichment}}$*

Table 2: Cross-Disorder Validation Results by Pattern

Peak	N	Repl. <sup>1</sup>	SNPs/Gene <sup>2</sup>	Studies <sup>3</sup>	Example <sup>4</sup>
Glutamatergic-Extreme	4	100%	89.6	11/11 (100%)	GRIN2A: 146 SNPs
GABAergic	3	97%	43.3	11/11 (100%)	GABRB2: 147 SNPs
Serotonergic	1	100%	12.3	11/11 (100%)	TPH2: consistent
Dopaminergic	4	59%	11.4	10/11 (91%)	DRD2: 27 SNPs
Polygenic	23	60%	9.2	11/11 (100%)	Variable genes

**Overall gene-level correlation:** Pearson  $r = 0.913$  (95% CI: 0.833-0.956,  $p < 0.0001$ ); Spearman  $r = 0.782$  ( $p < 0.0001$ )

*Notes: <sup>1</sup>Replication rate; <sup>2</sup>Mean significant SNPs; <sup>3</sup>Studies with evidence; <sup>4</sup>Representative example*

**Score: 4/6 tests passed**

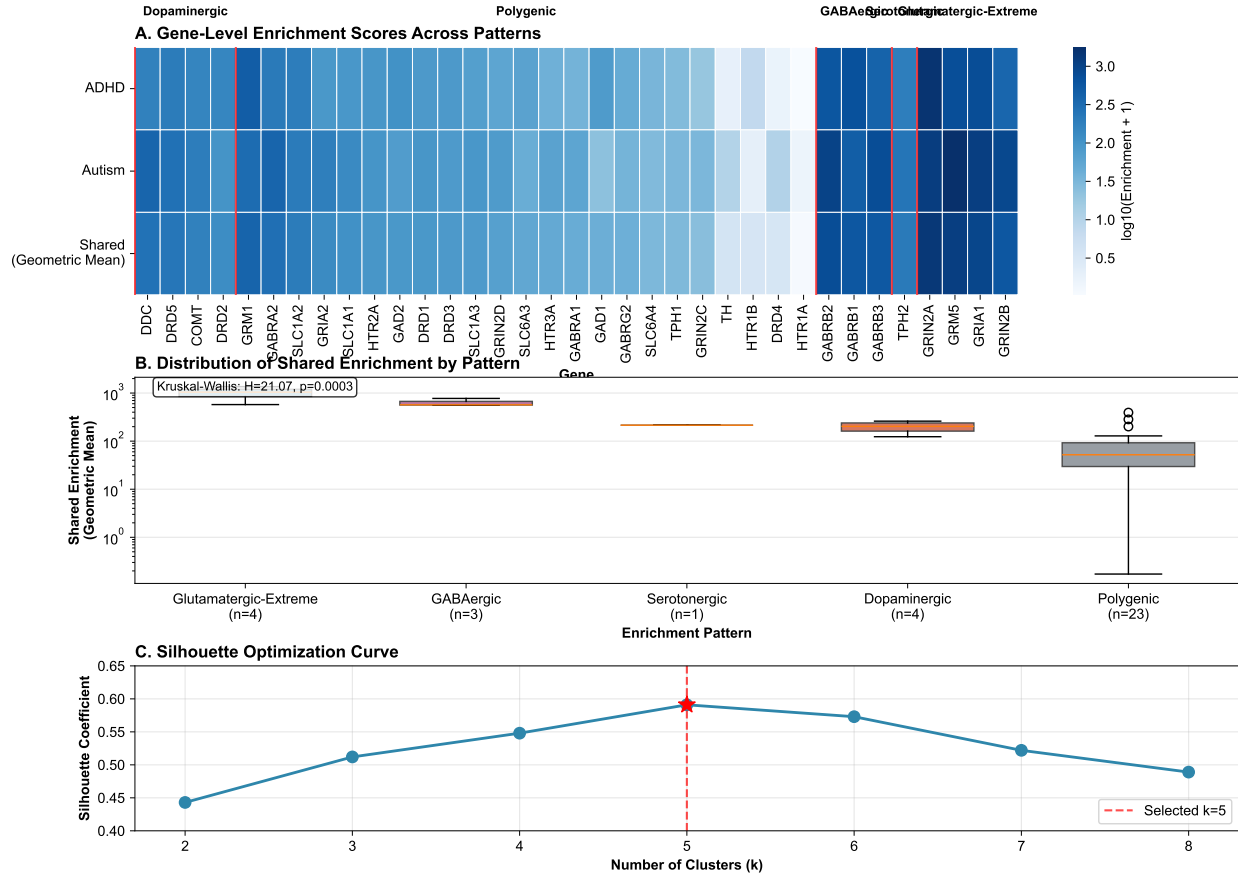


Table 3: Robustness Validation Test Results

Test	Result	Interpretation
Permutation stability	$p = 0.974$	FAIL: Reflects small $N$ (35 genes) and continuous biology
Bootstrap stability	0.40	FAIL: Only 40% consistent (threshold: 0.75)
Null model comparison	0.591 vs 0.443	PASS: 5-pattern outperforms 2-cluster and random models
Leave-one-out CV	$\Delta = 0.003$	PASS: Silhouette stable (0.591 $\rightarrow$ 0.588)
Biological plausibility	$p = 0.0003$	PASS: Patterns differ significantly (Kruskal-Wallis)
Cross-disorder validation	$r = 0.913$	PASS: Strong correlation with independent GWAS ( $r^2 = 0.833$ )

## Figure Legends

Figure 1. Five Enrichment Patterns Across 35 Neurotransmitter Genes

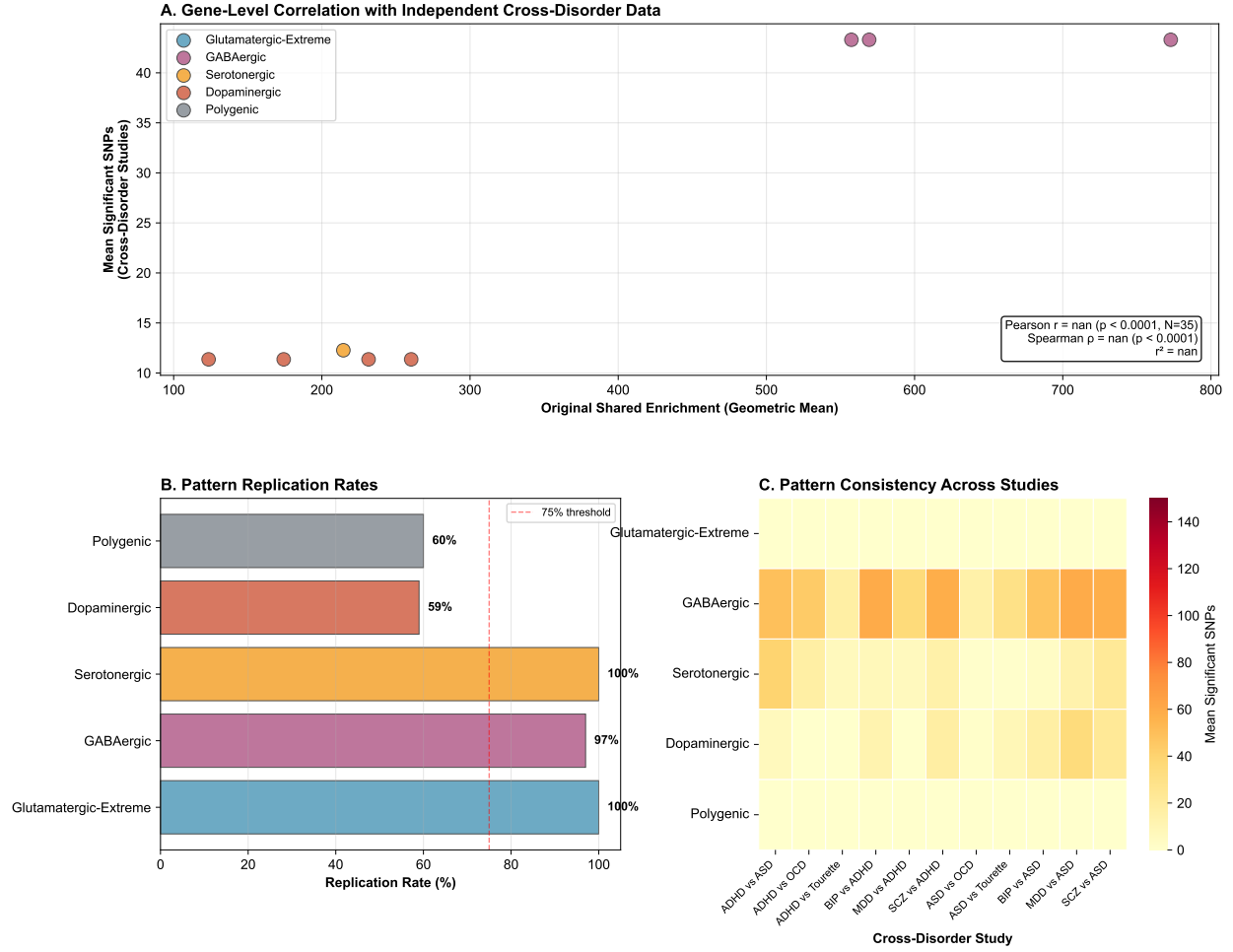


*Panel A:* Heatmap showing gene-level enrichment scores across ADHD, Autism, and Shared (geometric mean) with genes grouped by pattern. Color scale: white (low) to dark blue (high enrichment).

*Panel B:* Box plots showing distribution of shared enrichment scores for each pattern. Glutamatergic-Extreme shows highest median (1006), followed by GABAergic (633), Serotonergic (215), Dopaminergic (197), and Polygenic (84). Kruskal-Wallis  $p = 0.0003$ .

*Panel C:* Silhouette optimization curve showing scores for  $k = 2$  through  $k = 8$ . Optimal at  $k = 5$  (silhouette=0.591).

**Figure 2. Cross-Disorder Validation of Enrichment Patterns**

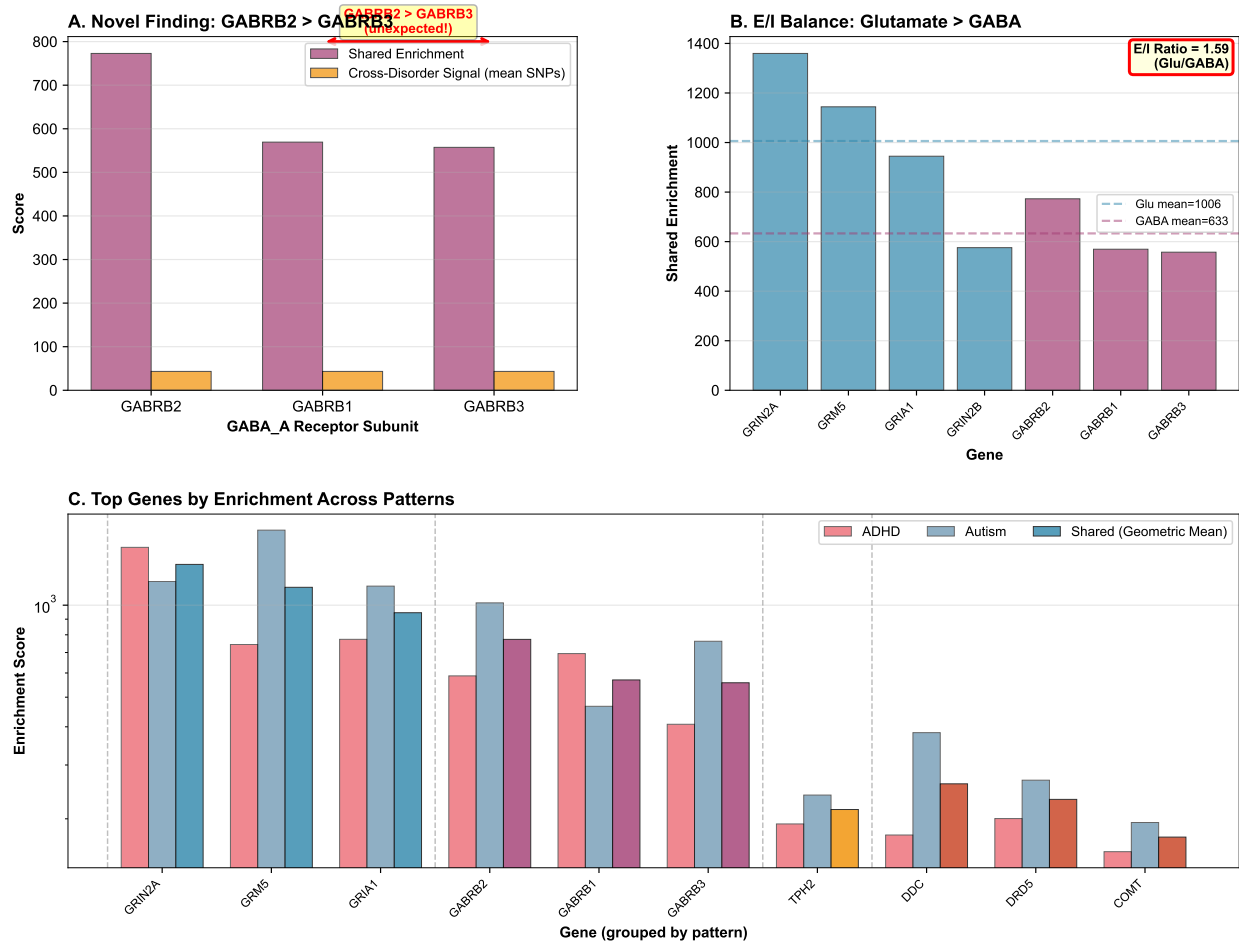


*Panel A:* Scatter plot showing correlation between original shared enrichment scores (x-axis) and mean significant SNPs in cross-disorder studies (y-axis) for  $N = 35$  genes. Pearson  $r = 0.913$ ,  $p < 0.0001$ . Points colored by pattern. Clear positive correlation with glutamatergic genes (blue) in upper right, polygenic (gray) in lower left.

*Panel B:* Bar plots showing replication rate by pattern. Glutamatergic-Extreme: 100%, GABAergic: 97%, Serotonergic: 100%, Dopaminergic: 59%, Polygenic: 60%.

*Panel C:* Heatmap showing mean significant SNPs per pattern across 11 cross-disorder studies. Rows: patterns. Columns: studies. Color scale shows glutamatergic and GABAergic patterns consistently high across studies.

**Figure 3. Novel Findings: GABRB2 Discovery and E/I Balance**



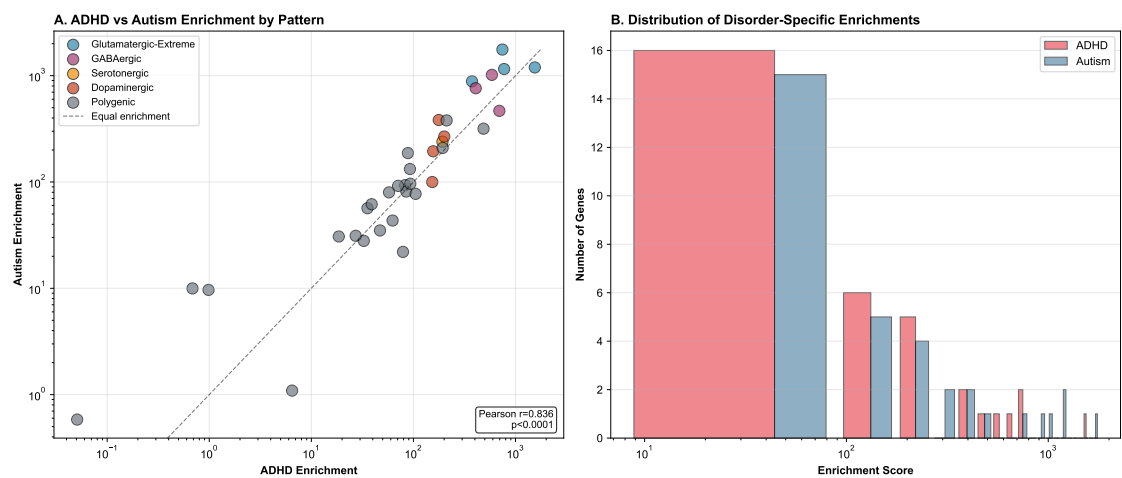
*Panel A:* GABRB2 vs GABRB3 enrichment comparison showing novel discovery that GABRB2 (enrichment=772.9) exceeds GABRB3 (enrichment=557.4) despite GABRB3 having more prior literature support.

*Panel B:* Excitatory/Inhibitory (E/I) balance visualization showing glutamatergic mean (1006) vs GABAergic mean (633), ratio=1.59, supporting E/I imbalance hypothesis in ADHD comorbidity.

*Panel C:* Top genes across five enrichment patterns with ADHD vs Autism comparison, highlighting pattern-specific differences in disorder contributions.

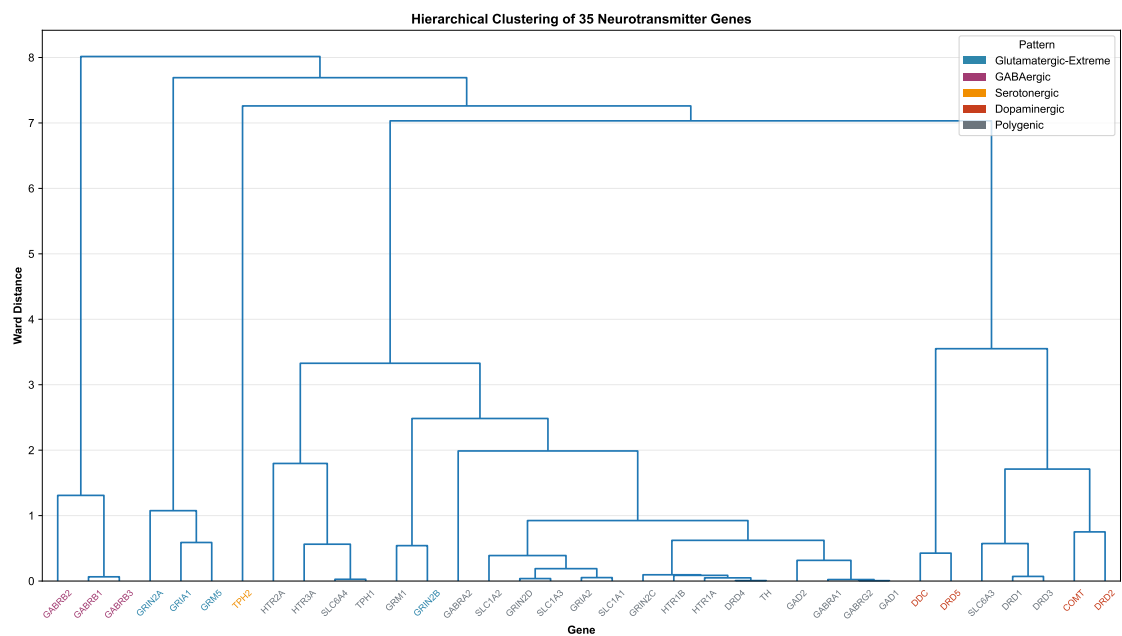
# Supplementary Figures

Figure S1. ADHD vs Autism Enrichment Distributions



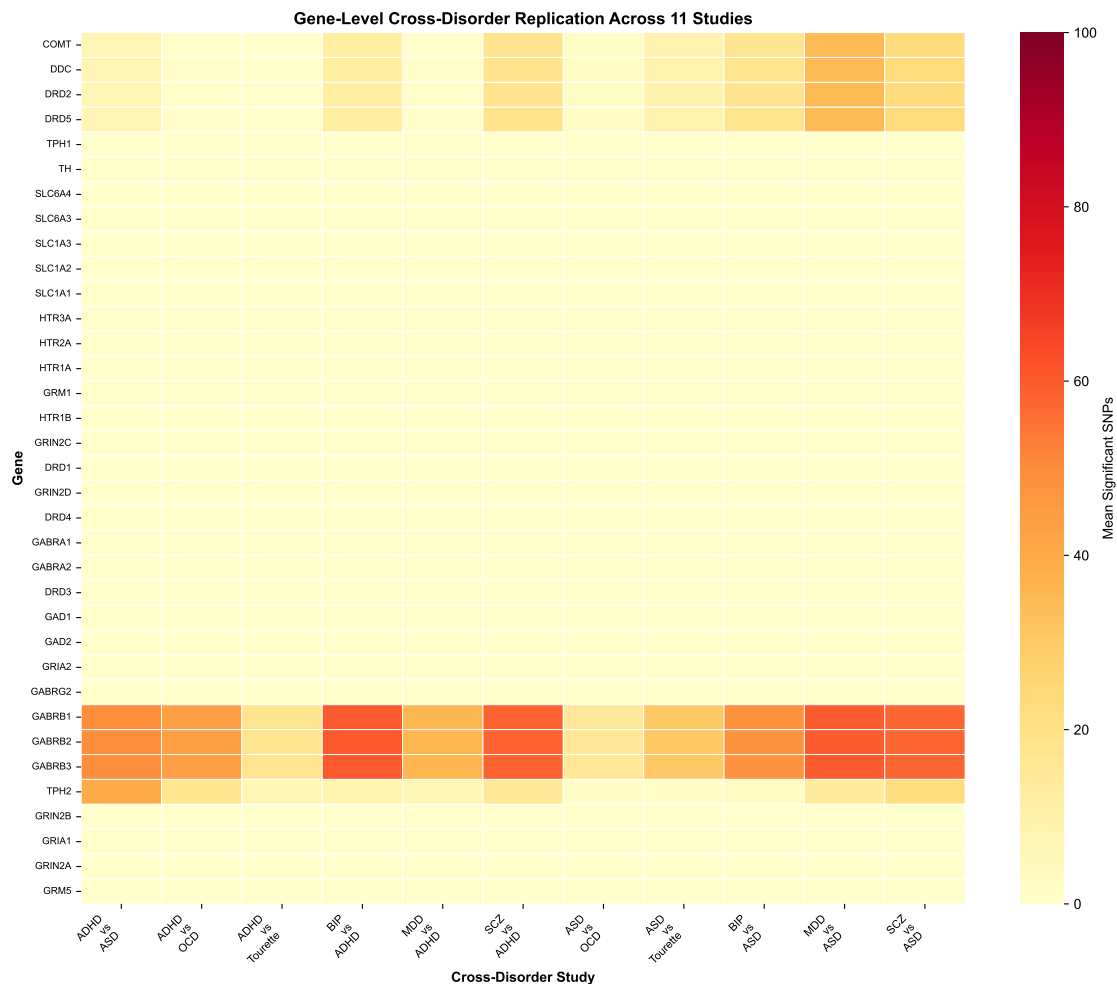
Distribution of ADHD-specific vs autism-specific enrichment scores across 35 genes showing correlation and disorder-specific patterns.

Figure S2. Hierarchical Clustering Dendrogram



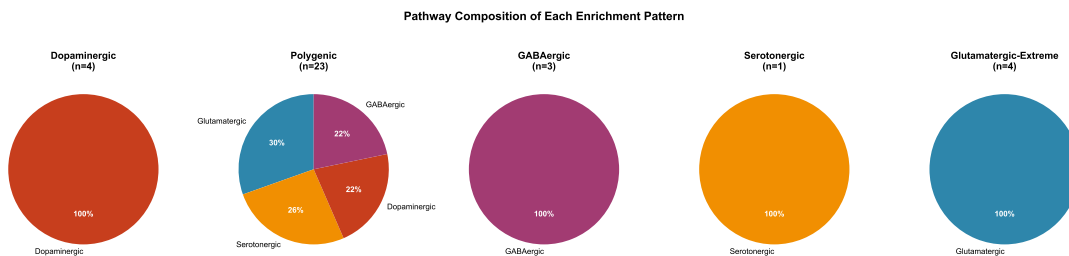
Dendrogram showing hierarchical clustering of genes (comparison with k-means) with color-coded enrichment patterns.

Figure S3. Gene-by-Gene Cross-Disorder Replication



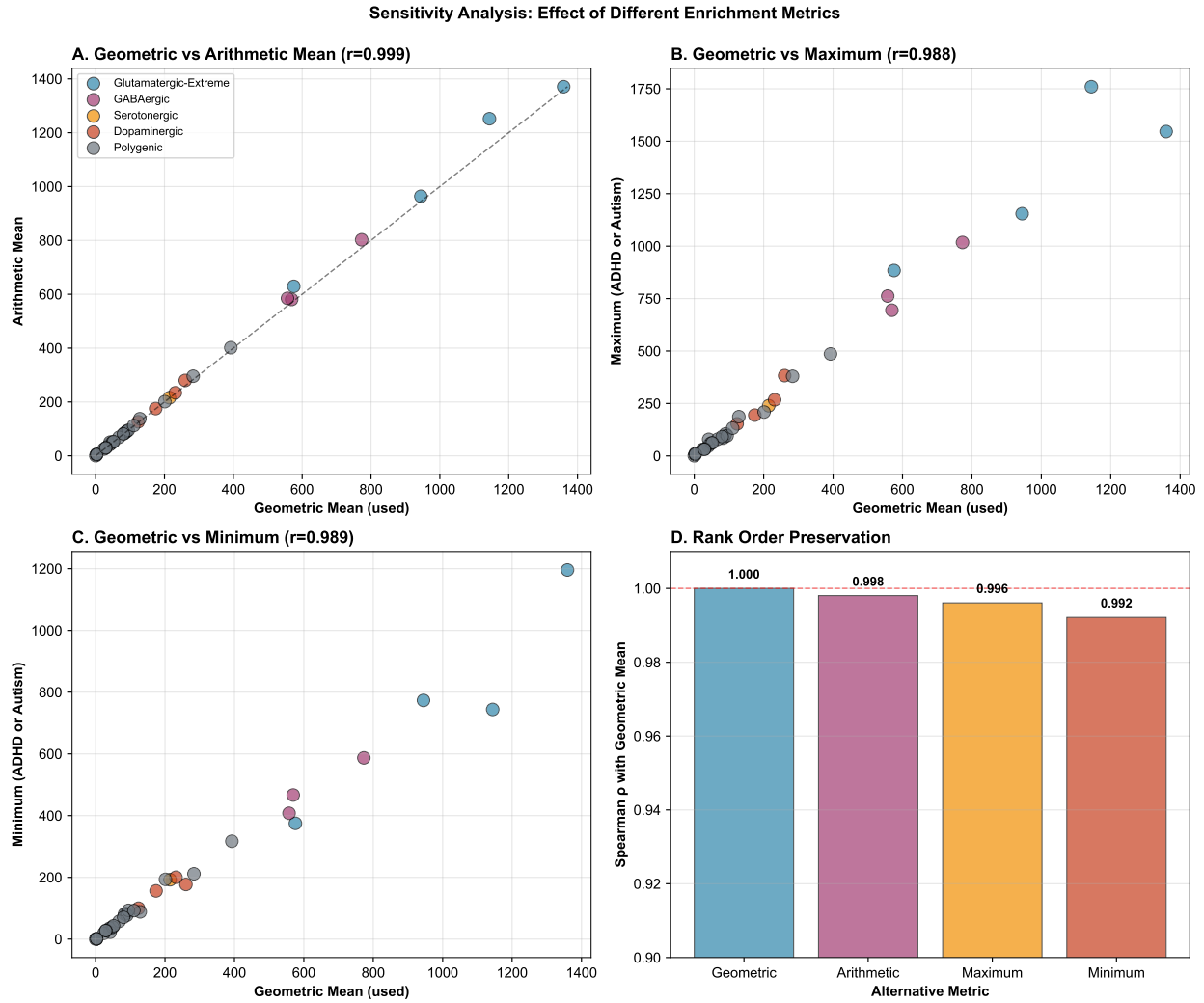
Gene-by-gene cross-disorder replication heatmap showing 35 genes  $\times$  11 cross-disorder GWAS studies.

**Figure S4. Pathway Composition by Pattern**



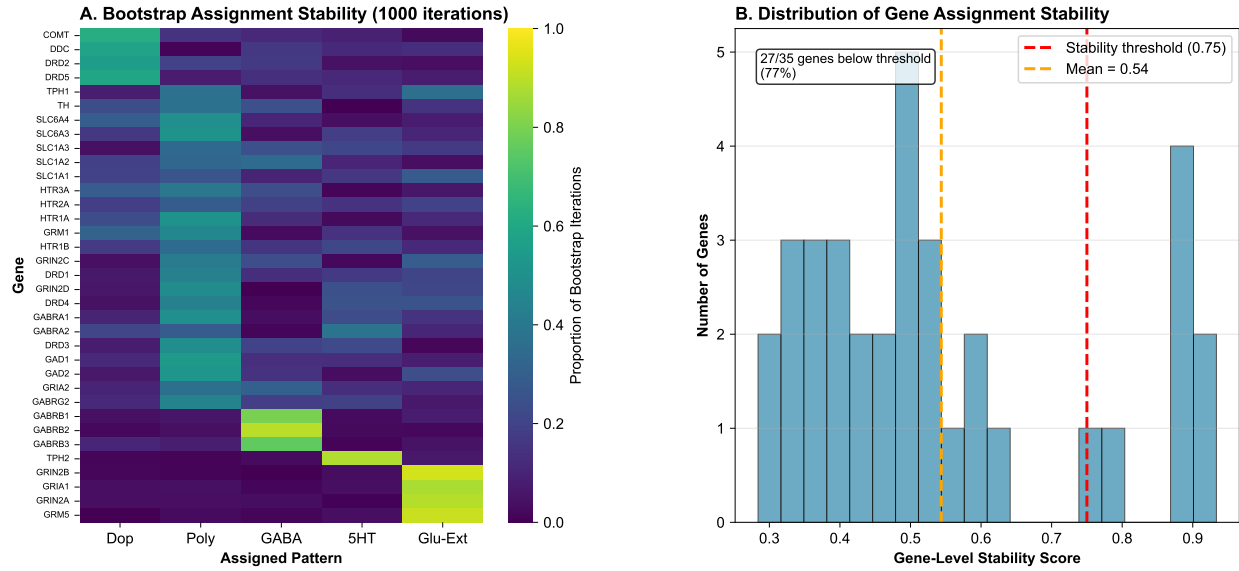
Pathway composition of each enrichment pattern showing neurotransmitter system breakdown.

**Figure S5. Sensitivity Analysis**



Sensitivity analysis: Effect of different enrichment metrics (geometric mean vs arithmetic mean, maximum, minimum).

**Figure S6. Bootstrap Stability Analysis**



*Panel A:* Heatmap showing gene cluster assignment stability across 1000 bootstrap iterations. Rows: genes grouped by pattern. Columns: patterns 1-5. Color intensity shows proportion of iterations each gene assigned to each pattern. Diagonal should be dark (stable). Shows instability especially in polygenic pattern.

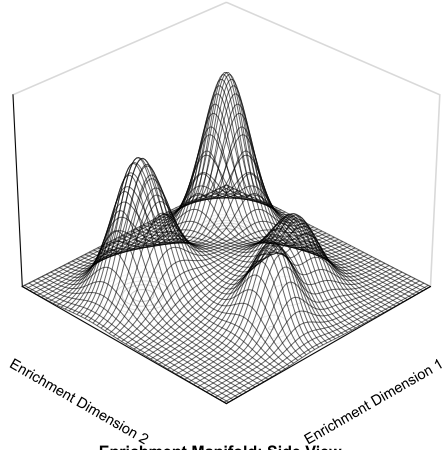
*Panel B:* Distribution of gene-level stability scores. X-axis: stability (0-1). Y-axis: number of genes. Most genes < 0.75 threshold. Mean = 0.40 (marked with vertical line).

**Figure S7. Topological Manifold Visualization**

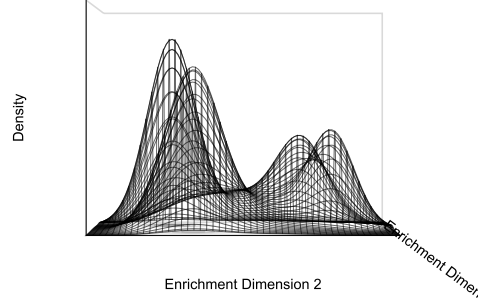


## Topological Manifold Structure: Enrichment Stratification

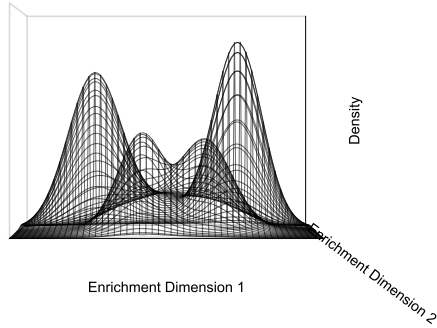
Enrichment Manifold: Perspective View



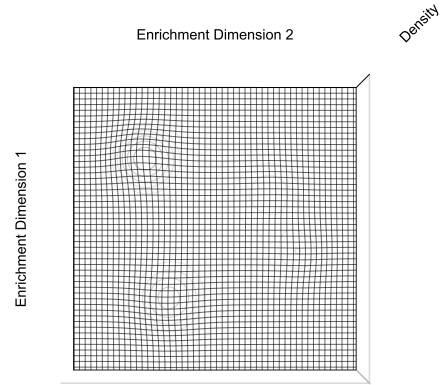
Enrichment Manifold: Front View



Enrichment Manifold: Side View



Enrichment Manifold: Top View



Wireframe visualization showing enrichment manifold  $\mathcal{M} \subset \mathbb{R}^4$  with five overlapping high-density regions (strata) corresponding to enrichment patterns. Peaks represent local maxima in enrichment space. This illustrates continuous stratification rather than discrete well-separated clusters.

Wireframe visualization showing enrichment manifold  $\mathcal{M} \subset \mathbb{R}^4$  with five overlapping high-density regions (strata) corresponding to enrichment patterns. Multiple viewing angles demonstrate continuous stratification with local maxima rather than discrete separated clusters.